

MALAPTERURUS La Cepède, 1803

Malapterurus La Cepède, 1803: 90. Type species: *Silurus electricus* Gmelin, 1789. Type by designation under plenary powers; placed on Official List (ICZN Opinion 93, Direction 56). Gender: Masculine.

Anacanthus Minding 1832: 117. Type species: *Silurus electricus* Gmelin, 1789. Type by monotypy. Preoccupied by *Anacanthus* Gray, 1830, in fishes. Gender: Masculine.

Malopterus Agassiz, 1846: 223. Type species: *Silurus electricus* Gmelin, 1789. Type by being a replacement name. Unjustified emendation of *Malapterurus* La Cepède, 1803. Gender: Masculine.

Remarks: *Malapterus* Jarocki, 1822, *Malapturus* Swainson, 1838, and *Malacopterurus* Gill, 1890, are considered to be incorrect subsequent spellings, and therefore unavailable names, in ICZN Direction 56. Forsskål (1775: 16) applied the generic name of the electric ray, *Torpedo*, to his account of the electric catfish of the Nile.

Malapterurus barbatus Norris, 2002

Malapterurus barbatus Norris, 2002: 88, fig. 49. Type locality: Sierra Leone, Sewa River system, Bagbe River at Yfin, 8°42'N, 11°05'W. Holotype: MNHN 1990-100.

Distribution: Western Guinean rivers from the Kolente River, Sierra Leone, to the Borlor River, Liberia (Norris, 2002).

Malapterurus beninensis Murray, 1855

Malapterurus Beninensis Murray, 1855: 20, pl. Type locality: Old Calabar. Syntypes: BMNH 1856.1.16.1 (2), possibly NMSZ 1855.27 (2).

? *Malapterurus affinis* Günther, 1864: 220. Type locality: Old Calabar. Syntypes: BMNH 1859.11.20.1 (1), BMNH 1864.7.9.1 (1).

Distribution: Coastal plains of west central Africa, from the Volta River, Ghana, to the Shiloango River, Angola (Norris, 2002).

Remarks: Synonymy of *Malapterurus affinis* with *M. beninensis* considered tentative by Norris (2002).

Malapterurus cavalliensis Roberts, 2000

Malapterurus cavalliensis Roberts, 2000: 8. Type locality: Ivory Coast: rivière Cavally directly W of Tiéouléoula about 18 km S of Tai. Holotype: BMNH 2000.3.3.7.

? *Malapterurus thysi* Norris, 2002: 81, fig. 45. Type locality: Ivory Coast, Cess (Nipoué) River at Toyebli, 6°38'N, 8°29'W. Holotype: MRAC 73-10-P-4144.

Distribution: Cess and Cavally Rivers, western Côte d'Ivoire (Norris, 2002).

Malapterurus electricus (Gmelin, 1789)

Silurus electricus Bonnaterre, 1788: 156, pl. 62 (fig. 245). Type locality: Nile and rivers of Africa. No types known.

Silurus electricus Gmelin, 1789: 1354. Type locality: Rosetta, branch of the Nile River, Egypt. Neotype: BMNH 1907.12.2.2547, designated in Poll & Gosse (1969: 5). Placed on Official list as type of *Malapterurus* (ICZN Direction 57).

Distribution: Nile River basin; Western African rivers south to the Niger River basin (Norris, 2002).

Malapterurus leonensis Roberts, 2000

Malapterurus leonensis Roberts, 2000: 9. Type locality: Sierra Leone, Moyamba. Holotype: BMNH 1976.11.12.122.

? *Malapterurus stiassnyae* Norris, 2002: 48, fig. 23. Type locality: Sierra Leone, River Moa drainage, Tiwai Island, 7°33'N, 11°21'W. Holotype: AMNH 59488.

Distribution: Upper Guinean basins of western Africa, between the Bofon River, Guinea, and St. Paul River, Liberia (Norris, 2002).

Malapterurus melanochir Norris, 2002

Malapterurus melanochir Norris, 2002: 109, fig. 65. Type locality: Congo, Congo R. basin, Ruki River at Eala, 00°04'N, 18°20'E. Holotype: MRAC 46166.

Distribution: Central Congo River basin and upper Lualaba River system (Norris, 2002).

Malapterurus microstomus Poll & Gosse, 1969

Malapterurus microstoma Poll & Gosse, 1969: 8, pls. 1, 2 (fig. b), 3 (fig. b). Type locality: Yangambi, Congo R., Zaire. Holotype: MRAC 164701.

Distribution: Congo River basin (Norris, 2002).

Malapterurus minjiriya Saguia, 1987

Malapterurus minjiriya Saguia, 1987: 78, fig. 1. Type locality: Lake Kainji, lower Niger basin, Nigeria. Holotype: BMNH 1979.3.5.405.

Distribution: Niger and Volta River basins; possibly also White Nile and Omo River (Norris, 2002).

Malapterurus monsembeensis Roberts, 2000

Malapterurus monsembeensis Roberts, 2000: 9. Type locality: Upper Congo at Monsembe. Holotype: BMNH 1899.8.22.6.

? *Malapterurus gossei* Norris, 2002: 102, fig. 60. Type locality: Congo, Congo R. basin, Léopoldville (Ouest) (= Kinshasa), 4°20'N, 15°15'E. Holotype: MRAC 88017.

Distribution: Congo River system (Norris, 2002).

Malapterurus occidentalis Norris, 2002

Malapterurus occidentalis Norris, 2002: 73, fig. 40. Type locality: Gambia, Mc Carthy Id. 13°32' [N], 14°46' E [sic, W]. Holotype: BMNH 1901.12.28.77.

Distribution: Middle Gambia River, Gambia, and Geba River, Guinea-Bissau (Norris, 2002).

Malapterurus oguensis Sauvage, 1879

Malapterurus electricus oguensis Sauvage, 1879: 99. Type locality: Lopé, pays des Okanda, fleuve Ogôoué. Holotype: MNHN a-0889. Originally proposed as *Malapterurus electricus* var. *oguensis*. Described in more detail and illustrated in Sauvage (1880c: 45, pl. 1, fig. 3) as *Malapterurus electricus*, var. *ogooensis*.

Distribution: Ivindo River, Ogowe River basin; and Nyanga River and Kouilou-Niari Systems of western Africa.

Malapterurus punctatus Norris, 2002

Malapterurus punctatus Norris, 2002: 91, fig. 51. Type locality: Liberia, Cavally River drainage, unspecified tributary waters of Duobe River at Duo Town, ca. 5°41'N, 8°06'W. Holotype: MRAC 80-36-P-2154.

Distribution: Eastern Guinean rivers, from the St. Paul River, Sierra Leone, to the Cavally River, Liberia (Norris, 2002).

Malapterurus shirensis Roberts, 2000

Malapterurus shirensis Roberts, 2000: 10. Type locality: Chiromo, lower Shire. Holotype: BMNH 1921.9.6.53.

? *Malapterurus zambeziensis* Norris, 2002: 44, fig. 20. Type locality: Zambia/Zimbabwe, Zambezi River drainage, Lake Kariba at Andora Harbor. Holotype: MRAC 187094.

Distribution: Zambezi River basin (Norris, 2002).

Malapterurus tanganyikaensis Roberts, 2000

Malapterurus tanganyikaensis Roberts, 2000: 11. Type locality: Lake Tanganyika. Holotype: BMNH 1936.6.15.1234.

Malapterurus polli Norris, 2002: 111, fig. 67. Type locality: Tanzania: au large de Karema, à 660–700 m de la côte, ± 6°49'N, 30°26'E. Holotype: MRAC 90328.

Distribution: Lake Tanganyika (Norris, 2002).

Malapterurus tanoensis Roberts, 2000

Malapterurus tanoensis Roberts, 2000: 11. Type locality: Unvaiye lagoon, Tano basin, sw. Ghana. Holotype: USNM 316021.

? *Malapterurus murrayi* Norris, 2002: 94, fig. 54. Type locality: Ghana, Tano River near Wiawso, slow pool, 6°12'N 2°29'W. Holotype: BMNH 1969.4.28.29.

Distribution: Ofin and Tano Rivers, Ghana (Norris, 2002).

Malapterurus teugelsi Norris, 2002

Malapterurus teugelsi Norris, 2002: 52, fig. 26. Type locality: Guinea, Ndyarendi, Kogon River, ca. 11°21'N, 14°30'W. Holotype: MRAC 92-59-P-1680.

Distribution: Kogon River basin, Guinea (Norris, 2002).

PARADOXOGLANIS Norris, 2002

Paradoxoglanis Norris, 2002: 117. Type species: *Paradoxoglanis caudivittatus* Norris, 2002. Type by original designation. Gender: Masculine.

Paradoxoglanis caudivittatus Norris, 2002

Paradoxoglanis caudivittatus Norris, 2002: 120, fig. 72. Type locality: Congo, Lomela, N. Sankuru, petits ruisseaux affl. riv. Lomela, 2°00'–2°19'S, 22°58'–23°15'E. Holotype: MRAC 98452.

Distribution: Central portions of the Congo Basin (Norris, 2002).

***Paradoxoglanis cryptus* Norris, 2002**

Paradoxoglanis cryptus Norris, 2002: 128, fig. 78. Type locality: Congo, Aketi, riv. Kagala, affl. riv. Itimbiri, rive droite, 2°44'N, 23°46'E. Holotype: MRAC 88-25-P-2231.

Distribution: Kagal River, Itimbiri basin, Congo System (Norris, 2002).

***Paradoxoglanis parvus* Norris, 2002**

Paradoxoglanis parvus Norris, 2002: 126, fig. 76. Type locality: Congo, Congo R. drainage, Mawuya, riv. Mawuya, terr. de Libenge, 3°14'N 18°41'E. Holotype: MRAC 167535.

Distribution: middle Congo River basin (Norris, 2002).

MOCHOKIDAE Jordan, 1923

Synodontini Bleeker, 1862 (in Bleeker, 1862–63): 6. Type genus: *Synodontis* Cuvier, 1816.

Rhinoglanina Günther, 1864: 4, 10, 216. Type genus: *Rhinoglanis* Günther, 1864.

Mochokidae Jordan, 1923: 150. Type genus: *Mochokus* Joannis, 1835.

Chiloglanidinae Riehl & Baensch, 1990: 396. Type genus: *Chiloglanis* Peters, 1868.

Remarks: The name Simuldentinae, published in Taverne & Aloulou-Triki (1974) is not available as a family group name because it is not based on an available generic name.

Taxonomic summary: Gosse (1986b).

Reviews: Paugy & Roberts (1992, West Africa); Skelton (1993, 2001, Southern Africa); Seegers (1996, Lake Rukwa basin); Skelton, *et al.* (1985, Okavango basin).

Key to genera: Poll & Gosse (1994).

10 genera, 188 species; no named fossil taxa.

***ACANTHOCLEITHRON* Nichols & Griscom, 1917**

Acanthocleithron Nichols & Griscom, 1917: 720. Type species: *Acanthocleithron chapini* Nichols & Griscom, 1917.

Type by monotypy. Gender: Neuter.

***Acanthocleithron chapini* Nichols & Griscom, 1917**

Acanthocleithron chapini Nichols & Griscom, 1917: 721, fig. 26. Type locality: Avakubi [Ituri River], Congo. Holotype: AMNH 6548.

Distribution: Ituri River and Congo River from Kisangani to Stanley Pool (Gosse, 1986b).

***ATOPOCHILUS* Sauvage, 1879**

Atopochilus Sauvage, 1879: 97. Type species: *Atopochilus savorgnani* Sauvage, 1879. Type by monotypy. Gender: Masculine.

***Atopochilus chabanaudi* Pellegrin, 1938**

Atopochilus Chabanaudi Pellegrin, 1938a: 375, fig. 1. Type locality: Brazzaville (Congo). Holotype: MNHN 1937-0111.

Distribution: Congo River at Stanley Pool (Gosse, 1986b).

Remarks: Catalog number for holotype of *Atopochilus Chabanaudi* mistakenly reported as MNHN 37-311 by Bertin & Estève (1950).

***Atopochilus christyi* Boulenger, 1920**

Atopochilus Christyi Boulenger, 1920a: 33, fig. 17. Type locality: Stanley-Falls, ... d'Avakubi sur l'Ituri. Syntypes (2): Possibly BMNH 1919.9.10.282 (1), MRAC 7239 (1), MRAC 7350 (1).

Distribution: Congo River basin (Gosse, 1986b).

***Atopochilus macrocephalus* Boulenger, 1906**

Atopochilus macrocephalus Boulenger, 1906a: 112. Type locality: Fort Don Carlos, in the province of Loanda, at the

junction of the Cambo and Kwango (or Cuango) Rivers, Angola. Holotype: BMNH 1904.5.2.182.

Distribution: Congo River basin (Gosse, 1986b).

***Atopochilus mandevillei* Poll, 1959**

Atopochilus mandevillei Poll, 1959: 104, pl. 18 (figs. 2a–c). Type locality: Stanley Pool. Holotype: MRAC 100503.

Distribution: Congo River at Stanley Pool (Gosse, 1986b).

***Atopochilus pachychilus* Pellegrin, 1924**

Atopochilus pachychilus Pellegrin, 1924c: 487. Type locality: Kanda kanda, dans la Lubilonji, tributaire du Kasai, affluent du Congo. Syntypes (2): MNHN 1924-0213 (1), MRAC 14920 (1).

Distribution: Kasai River, Congo basin; known only from type locality (Gosse, 1986b).

***Atopochilus savorgnani* Sauvage, 1879**

Atopochilus Savorgnani Sauvage, 1879: 98. Type locality: Doumé [Fl. Ogôoué, aux chutes de Doumé (Congo français)]. Holotype: MNHN a-0899. Described in more detail and illustrated in Sauvage (1880c: 43, pl. 3, fig. 3).

Distribution: Upper Ogowe River, Muni River Basin, Equatorial Guinea, and Ntem River, southern Cameroon (Gosse, 1986b).

***Atopochilus vogti* Pellegrin, 1922**

Atopochilus Vogti Pellegrin, 1922c: 350. Type locality: Riv. Wamé, Afrique orientale. Holotype: MNHN 1922-0021.

Distribution: Wami River, Tanzania (Gosse, 1986b; Seegers, 2003).

***BRACHYSYNODONTIS* Bleeker, 1862**

Brachysynodontis Bleeker, 1862 (in Bleeker, 1862–63): 6. Type species: *Synodontis batensoda* Rüppell, 1832. Type by original designation. Gender: Feminine.

Remarks: Treated in Willoughby (1994) as a synonym of *Synodontis*, without comment.

***Brachysynodontis batensoda* (Rüppell, 1832)**

Synodontis batensoda Rüppell, 1832: 6, pl. 3 (fig. 2). Type locality: Nil, bei Kaïro. Holotype: SMF 2636 (dry).

Synodontes Ruppelli Swainson, 1838: 339, fig. 82. Type locality: [No locality stated]. No types known.

Distribution: Nile, Chad, Niger, Senegal and Gambia river basins (Gosse, 1986b).

Remarks: Synonymy follows Grant (2004).

***CHILOGLANIS* Peters, 1868**

Chiloglanis Peters, 1868b: 599. Type species: *Chiloglanis deckenii* Peters, 1868. Type by monotypy. Gender: Masculine.

Reviews: Jubb & Le Roux (1969, southern Africa); Roberts (1989b, Cameroon), Seegers (1996, Lake Rukwa).

Remarks: The name *Chiloglanis athiensis* appears in Copley (1941: 15) but as name only and therefore not an available name.

***Chiloglanis angolensis* Poll, 1967**

Chiloglanis angolensis Poll, 1967: 255, fig. 120. ruisseau à forêt-galerie, traversant la route Chibia-Jau, à la bifurcation Jau-Onguéria, quelques km au sud de la Chibia, environs de Sá de Bandeira, alt. 1.500 m., Angola. Holotype: MD 1875.

Distribution: known only from type locality (Gosse, 1986b).

***Chiloglanis anoterus* Crass, 1960**

Chiloglanis anoterus Crass, 1960: 446, fig. 2 (a–d). Type locality: Upper Pivaan River (Pongola River system), altitude ± 4600 ft. (30°28'E, 27°30'S), Natal. Holotype: AMG P1157.

Distribution: Phongolo and Incomati River system tributaries (Skelton, 1993).

***Chiloglanis asymmetricaudalis* De Vos, 1993**

Chiloglanis asymmetricaudalis De Vos, 1993: 154, figs. 2–4. Type locality: Nyakabuye, rivière Rubyiro, (bassin de la Ruzizi), près de la confluence avec la rivière Ntandwe, Rwanda (2°34'S, 29°02'E). Holotype: MRAC 87-05-P-263.

Distribution: Ruzizi River basin, Lake Tanganyika drainage (De Vos, 1993).

***Chiloglanis batesii* Boulenger, 1904**

Chiloglanis Batesii Boulenger, 1904a: 19. Type locality: South Cameroon; Efulen and stream tributaries of the Lobi River, 15 or 20 miles S. W. of Efulen. Syntypes (5): BMNH 1904.7.1.97–100 (4), BMNH 1904.7.1.127 (1).

Chiloglanis micropogon Poll, 1952: 228, figs. 3–4. Type locality: riv. Nzokwe, affluent del la riv. Ulindi (Territ. Kabare). Holotype: MRAC 91479.

Distribution: Lualaba and Kasai River, Congo Basin; Upper Niger; Meme, Mungo, Lobi Rivers and coastal rivers of Cameroon; Kaduna River, Nigeria (Gosse, 1986b).

Remarks: *Chiloglanis micropogon* has been generally treated as valid, but recently treated by Roberts (1989b: 169) as a synonym of *C. batesii*.

***Chiloglanis benuensis* Daget & Stauch, 1963**

Chiloglanis benuensis Daget & Stauch, 1963: 98, fig. 3. Type locality: Bénoué à Lakdo. Holotype: MNHN 1962-1273.

Distribution: Niger River basin; known only from type locality (Gosse, 1986b).

***Chiloglanis bifurcus* Jubb & Le Roux, 1969**

Chiloglanis bifurcus Jubb & Le Roux, 1969: 17, figs. 9–9a. Type locality: Crocodile River, Incomati River system, Montrose Farm, Nelspruit District, Southern Africa. Holotype: AMG P996.

Distribution: Crocodile-Incomati basin (Skelton, 1993).

Remarks: See Kelynhan & James (1995) for comments on species.

***Chiloglanis brevibarbis* Boulenger, 1902**

Chiloglanis brevibarbis Boulenger, 1902f: 224, pl. 17 (figs. 2, 2a, 2b). Type locality: Mathoia River, in the Kenya district, East Africa. Holotype: BMNH 1902.5.26.19.

Distribution: Tana System and Athi River (Gosse, 1986b).

***Chiloglanis cameronensis* Boulenger, 1904**

Chiloglanis cameronensis Boulenger, 1904a: 18. Type locality: Efulen, South Cameroon. Syntypes (7): BMNH 1904.7.1.90–96 (7).

Distribution: coastal rivers of Cameroon, Equatorial Guinea, and Gabon, Congo River basin (Gosse, 1986b).

***Chiloglanis carnosus* Roberts & Stewart, 1976**

Chiloglanis carnosus Roberts & Stewart, 1976: 282, pl. 8 (figs. b–d). Type locality: Near village of Inga, a few kilometers upstream and on the opposite side of the river from Inga hydroelectric dam. Lat. 5°27.5'S, Long. 13°36.0'E; lower Zaïre or Congo River. Holotype: MCZ 50464.

Distribution: Lower Congo River basin; known only from type locality (Gosse, 1986b).

***Chiloglanis congicus* Boulenger, 1920**

Chiloglanis congicus Boulenger, 1920a: 32, fig. 16. Type locality: Stanley-Falls, Congo Belge. Holotype: MRAC 7159.

Distribution: Stanley Falls, Stanley Pool, Congo River basin (Gosse, 1986b).

***Chiloglanis deckenii* Peters, 1868**

Chiloglanis Deckenii Peters, 1868b: 599, pl. 2. Type locality: Africa orientalis. Syntypes: ZMB 6844 (3), ZMB 16387.

Synodontis eurystomus Pfeffer, 1889: 14. Type locality: Rufu bei Korogwe. Lectotype: ZMH H387, designated by Ladiges et al. (1958: 159).

Distribution: Kenya and Tanzania (Gosse, 1986b).

***Chiloglanis disneyi* Trewavas, 1974**

Chiloglanis disneyi Trewavas, 1974: 365, figs. 6 (left), 8. Type locality: R. Wowe, Mungo system, Cameroon. Holotype: BMNH 1973.5.14.342.

Distribution: Wowe River, Mungo basin; Known only from type locality (Gosse, 1986b).

***Chiloglanis elisabethianus* Boulenger, 1915**

Chiloglanis elisabethianus Boulenger, 1915: 171. Type locality: Rivière Lubumbashi, à Elisabethville. Holotype: MRAC 12126.

Distribution: Luapula-Moero River, Congo River basin (Gosse, 1986b).

***Chiloglanis emarginatus* Jubb & Le Roux, 1969**

Chiloglanis emarginatus Jubb & Le Roux, 1969: 21, figs. 12–12a. Type locality: Lekkerloop River, tributary of the

Komati River of the Incomati River system, on the farm Vergelegen, Carolina District, Southern Africa. Holotype: AMG P953.

Distribution: Phongolo and Incomati Rivers, South Africa and Swaziland, and Pungwe and Zambezi Rivers, Zimbabwe (Skelton, 1993).

***Chiloglanis fasciatus* Pellegrin, 1936**

Chiloglanis fasciatus Pellegrin, 1936a: 58. Type locality: Cubango, Angola. Holotype: MNHN 1936-0158.

Distribution: Zambezi River system, Angola and Botswana (Gosse, 1986b).

***Chiloglanis harbinger* Roberts, 1989**

Chiloglanis harbinger Roberts, 1989b: 175, fig. 13. Type locality: Cameroun, R. Lokunje near Bipindi. Holotype: CAS 60811.

Distribution: Lokunje River basin, southeastern Cameroon (Roberts, 1989b).

***Chiloglanis kalambo* Seegers, 1996**

Chiloglanis kalambo Seegers, 1996: 377, figs. 279–281. Type locality: Kanyele River, a southeastern affluent of the upper Kalambo River, on road from Sumbawanga to Mbala, 34 km N of the Tanzania–Zambia border, Lake Tanganyika drainage, Rukwa region, western Tanzania (8°25'S, 31°30'E). Holotype: MRAC 94-34-P-1079.

Distribution: Upper Kalambo River basin, Tanzania (Seegers, 1996).

***Chiloglanis lamottei* Daget, 1948**

Chiloglanis lamottei Daget, 1948: 38, fig. 12. Type locality: Guinéé Fse: Mt. Nimba, Zié, alt. 475–500 m. Holotype: MNHN 1949-0056.

Distribution: Known only from the type locality (Gosse, 1986b).

***Chiloglanis lufirae* Poll, 1976**

Chiloglanis lufirae Poll, 1976: 97, fig. 52. Type locality: Muye (affluent droit de la Lufira), alt. 800–900 m. [Upemba National Park]. Holotype: MRAC 79-01-P-4728.

Distribution: Lufira River tributary; known only from type locality (Gosse, 1986b).

***Chiloglanis lukugae* Poll, 1944**

Chiloglanis lukugae Poll, 1944a: 7, figs. 3–5. Type locality: Sange, région d'Albertville, Congo belge. Holotype: IRSNB 72; illustrated in Poll (1946: 224, fig. 23).

Distribution: Luapula River basin; tributaries of Lake Tanganyika, Angola, affl. of Cuango River (Gosse, 1986b).

Remarks: Gosse (1986b) stated that the type locality is a small tributary of the Luvua River on road between Albertville and Kiambi but not near Albertville as has been stated elsewhere.

***Chiloglanis macropterus* Poll & Stewart, 1975**

Chiloglanis macropterus Poll & Stewart, 1975: 152, fig. 1. Type locality: Luongo River, at ferry crossing, 53 km. S. of Kawambwa, Zambia. Holotype: ROM 28035.

Distribution: Luongo River, Zambia (Gosse, 1986b).

***Chiloglanis marlieri* Poll, 1952**

Chiloglanis marlieri Poll, 1952: 226, figs. 1–2. Type locality: riv. Ndakirwa à Meshe, affluent de la Luhoho. Holotype: MRAC 91478.

Distribution: Lualaba basin (Gosse, 1986b).

***Chiloglanis mbozi* Seegers, 1996**

Chiloglanis mbozi Seegers, 1996: 224, figs. 161–162. Type locality: Hansekewta River, a brook which is an affluent of the Momba River, draining the Mbozi block via the Msangano trough, 25 km east of Tunduma on the road to Mbeya, Mbeya Region, western Tanzania (9°08'S, 32°52'E). Holotype: MRAC 94-34-P-927.

Distribution: Hansekewta River, Momba River basin, Tanzania (Seegers, 1996).

***Chiloglanis microps* Matthes, 1965**

Chiloglanis microps Matthes, 1965: 188, figs. 3–4. Type locality: riv. Lukima à Kiamakoto (Lufira). Holotype: MRAC 140908.

Distribution: Lufira River basin (Gosse, 1986b).

***Chiloglanis modjensis* Boulenger, 1904**

Chiloglanis modjensis Boulenger, 1904b: 332, pl. 31 (figs. 3, 3a, 3b). Type locality: Modjo River [Webi Shebeli basin]. Syntypes (2): BMNH 1905.7.25.103 (1).

Distribution: Webi Shebeli basin, southern Ethiopia (Gosse, 1986b).

***Chiloglanis neumanni* Boulenger, 1911**

Chiloglanis neumanni Boulenger, 1911a: 481, fig. 359. Type locality: Upper Bubu River, Masailand. Lectotype: BMNH 1905.7.25.49; designated by, and illustrated in, Seegers (1996: 214, fig. 152).

Distribution: Limpopo River system, Cunene, Kafue, Zambezi, Kiavango, and upper Congo Rivers, Lakes Malawi and Kariba (Gosse, 1986b; Skelton, 1993).

***Chiloglanis niger* Roberts, 1989**

Chiloglanis niger Roberts, 1989b: 173, figs. 1f, 2f, 4e–f, 6f, 12. Type locality: Cameroun, Niger basin, R. Menchum below high waterfall near Befang, Bamenda highlands. Holotype: CAS 60809.

Distribution: Menchum River, Niger River basin, Cameroon (Roberts, 1989b).

***Chiloglanis niloticus* Boulenger, 1900**

Chiloglanis niloticus Boulenger, 1900d: 522. Type locality: island of Arko, Soudan. Syntypes (several): BMNH 1907.12.2.2466–73 (8), MNHN 1907-0230 (1), MSNG 14420 (1).

Distribution: Nile and Niger River basins (Gosse, 1986b).

***Chiloglanis normani* Pellegrin, 1933**

Chiloglanis normani Pellegrin, 1933a: 113, fig. on p. 114. Type locality: Danané (Côte d'Ivoire). Syntypes (2): MNHN 1932-0301 (1), NMBA 4253 (1).

Distribution: Cavally River system; known only from type locality (Gosse, 1986b).

***Chiloglanis occidentalis* Pellegrin, 1933**

Chiloglanis occidentalis Pellegrin, 1933a: 112, fig. on p. 112. Type locality: Douékoué, ... Man (Côte d'Ivoire). Syntypes (2): MNHN 1932-0300 (1), NMBA 4252 (1).

Chiloglanis niloticus waterloti Daget, 1954: 304, fig. 115. Type locality: Banamanan, cercle de Kissidougou [Guinea]. Syntypes: MNHN 1935-0224 (1), MNHN 1960-0472 (5), MNHN 1960-0473 (29 or 32), MNHN 1960-0474 (18 or 15).

Distribution: Bafing and Baoule rivers, upper Senegal, upper Niger, Sassandra, upper St. Paul, Kolente and Konkoure Rivers (Gosse, 1986b).

***Chiloglanis paratus* Crass, 1960**

Chiloglanis paratus Crass, 1960: 452, fig. 4 (a–d). Type locality: Concrete wall of Pongola River barrage, altitude ± 1000 ft. (31°30'E, 27°23'S), Natal. Holotype: AMG P1154.

Distribution: Phongolo, Incomati and Limpopo river systems, southern Africa (Skelton, 1993).

***Chiloglanis pojeri* Poll, 1944**

Chiloglanis pojeri Poll, 1944a: 8, figs. 6–8. Type locality: Mambwe, région d'Albertville. Holotype: IRSNB 74; illustrated in Poll (1946: 225, fig. 24).

Distribution: Luvua and Lualaba River, upper Congo Basin; Koki River of Lake Tanganyika (Gosse, 1986b).

***Chiloglanis polyodon* Norman, 1932**

Chiloglanis polyodon Norman, 1932: 184, fig. 3. Type locality: Headwaters of Bagbwe River, Sierra Leone. Holotype: BMNH 1932.5.18.64.

Distribution: Bagbwe River basin, Sierra Leone; known only from type locality (Gosse, 1986b).

***Chiloglanis polypogon* Roberts, 1989**

Chiloglanis polypogon Roberts, 1989b: 168, figs. 1b, 2b, 3c–d, 6a–b, 8. Type locality: Cameroun, Cross basin, high gradient stream along road from Mamfe to Bamenda, 88–94 km SW of Bamenda. Holotype: CAS 60790.

Distribution: Cross River basin, Cameroon (Roberts, 1989b).

***Chiloglanis pretoriae* van der Horst, 1931**

Chiloglanis pretoriae van der Horst, 1931: 248, fig. 2. Type locality: Crocodile River, Pretoria District, Transvaal. Holotype: at TM, current whereabouts unknown.

Chiloglanis pumilus van der Horst, 1931: 250, fig. 4. Type locality: Aapies River and Crocodile River, Pretoria Dist., Transvaal. Syntypes: SAIAB 30011 (1), SAIAB 30012 (1).

Distribution: Incomati, Limpopo, middle and lower Zambezi, Pungwe and Busi basins, southern Africa (Skelton, 1993).

***Chiloglanis reticulatus* Roberts, 1989**

Chiloglanis reticulatus Roberts, 1989b: 165, figs. 1a, 2a, 3a–b, 5, 7. Type locality: Cameroun, Congo basin, R. Mwamedjwel, a very small stream 2–3 km W of Yokadouma. Holotype: CAS 60786.

Distribution: Northwestern portion of the Congo River basin, Cameroon and Congo (Roberts, 1989b).

***Chiloglanis rukwaensis* Seegers, 1996**

Chiloglanis rukwaensis Seegers, 1996: 227, figs. 163–164. Type locality: Chiwanda River (also spelled Chuwanda or Chwanda River), a small river, Momba drainage, 32 km northwest of Tunduma near Chiwanda (or Chuwanda), on the road to Sumbawanga, western Lake Rukwa drainage, Mbeya Region, western Tanzania, (09°10'S, 32°33'E). Holotype: MRAC 94-34-P-932.

Distribution: Chiwanda River, Momba River basin, Tanzania (Seegers, 1996).

***Chiloglanis ruziziensis* De Vos, 1993**

Chiloglanis ruziziensis De Vos, 1993: 162, figs. 6–7. Type locality: Nyakabuye, rivière Rubyiro (bassin de la Ruzizi), pres de la confluence avec la rivière Ntandwe, Rwanda (2°34'S, 29°02'E). Holotype: MRAC 87-05-P-264.

Distribution: Ruzizi River basin, Lake Tanganyika drainage (De Vos, 1993).

***Chiloglanis sanagaensis* Roberts, 1989**

Chiloglanis sanagaensis Roberts, 1989b: 169, figs. 1c, 2c, 3 (e–f), 6 (c–d), 9. Type locality: Cameroun, Sanaga basin, gravel bars and riffles in lower 1–2 km of R. Nchit where it flows into R. Mbam. Holotype: CAS 60794. Distribution: Sanaga River basin (Roberts, 1989b).

***Chiloglanis sardinhai* Ladiges & Voelker, 1961**

Chiloglanis sardinhai Ladiges & Voelker, 1961: 139, pl. 7 (figs. 12 and 13). Type locality: Mujije (der Mujije ist ein Zufluss des Longa, Cuanza-Sul, Angolas). Holotype: ZMH H1317.

Distribution: Longa River basin; known only from type locality (Gosse, 1986b).

***Chiloglanis somereni* Whitehead, 1958**

Chiloglanis somereni Whitehead, 1958: 199, figs. 2–3. Type locality: Waroya River, Nyanza Province, Kenya; at 34°30'E, 0°10'N. Holotype: BMNH 1958.7.18.1.

Distribution: Westward flowing rivers of Kenya into Lake Victoria (Gosse, 1986b).

***Chiloglanis swierstrai* van der Horst, 1931**

Chiloglanis swierstrai van der Horst, 1931: 249, fig. 3. Type locality: Crocodile River, Pretoria District, Transvaal. Holotype: SAIAB 30013.

Chiloglanis engiops Crass, 1960: 451, fig. 3 (a–d). Type locality: Lower Pivaan Rier (Pongola River system), altitude ± 2000 ft. (31°11'E, 27°25'S), Natal. Holotype: AMG P1156.

Distribution: Lowveld and warmer portions of Phongolo, Incomati and Limpopo river basins, southern Africa (Skelton, 1993).

***Chiloglanis trilobatus* Seegers, 1996**

Chiloglanis trilobatus Seegers, 1996: 230, figs. 165–167. Type locality: Piti River, an affluent of the Rungwa River, 63 km S of Rungwa Village on the road from Itigi to Makambako or Mbeya, eastern Lake Rukwa drainage, western Tanzania (07°27'S, 33°25'E). Holotype: MRAC 94-34-P-938.

Distribution: Rungwizi River basin, Lake Rukwa basin, Tanzania (Seegers, 1996).

***Chiloglanis voltae* Daget & Stauch, 1963**

Chiloglanis voltae Daget & Stauch, 1963: 99, fig. 4. Type locality: Bougouri Ba au pont de Nabéré. Holotype: MNHN 1962-1280.

Distribution: Volta and upper Bénoué River systems (Gosse, 1986b).

***EUCHILICHTHYS* Boulenger, 1900**

Euchilichthys Boulenger, 1900d: 522. Type species: *Atopochilus guentheri* Schilthuis, 1891. Type by subsequent designation by Jordan (1920: 488). Gender: Masculine.

***Euchilichthys astatodon* (Pellegrin, 1928)**

Atopochilus astatodon Pellegrin, 1928b: 107, fig. 1. Type locality: Luluaburg Saint-Joseph, Congo belge. Syntypes (6): MNHN 1928-0017 (1), MNHN 1928-0018 (1), MRAC 19942 (1), NMBA 3721 (1), NMBA 3723–24 (2), NMBA 3726 (1).

Distribution: Kasai River (Gosse, 1986b).

Euchilichthys boulengeri Nichols & La Monte, 1934

Euchilichthys boulengeri Nichols & La Monte, 1934: 3, fig. 3. Type locality: Luluabourg, Kasai District, Belgian Congo. Holotype: AMNH 12357.

Distribution: Kasai River (Gosse, 1986b).

Euchilichthys dybowskii (Vaillant, 1892)

Chiloglanis Dybowskii Vaillant, 1892b: 2. Type locality: l'Oubanghi. Syntypes: MNHN 1892-0081 (2).

Euchilichthys habereri Steindachner, 1912: 447. Type locality: Dscha, Süd. Kamerun. Holotype: NMW 10700.

Illustrated and described in more detail in Steindachner (1913: 45, fig. 9 and pl. 1 (figs. 2–2a)).

Distribution: Ja River, Congo basin, Cameroon (Gosse, 1986b).

Euchilichthys guentheri (Schilthuis, 1891)

Atopochilus guntheri Schilthuis, 1891: 86, pl. 6 (fig. 2). Type locality: Stanley Pool. Holotype: BMNH 1899.9.6.6.

Distribution: Congo River basin, up to Bangweulu River system (Gosse, 1986b).

Euchilichthys royauxi Boulenger, 1902

Euchilichthys royauxi Boulenger, 1902d: 46, pl. 13. Type locality: l'Ubangi à Banzylville. Syntypes (3): BMNH 1901.12.26.46–47 (2), MRAC 1186 (1).

Distribution: Congo River basin, up to Moero Lake system (Gosse, 1986b).

HEMISYNODONTIS Bleeker, 1862

Hemisynodontis Bleeker, 1862 (in Bleeker, 1862–63): 6. Type species: *Pimelodus membranaceus* Geoffroy St. Hilaire, 1809. Type by original designation. Gender: Feminine.

Remarks: Treated in Willoughby (1994) as a synonym of *Synodontis*, without comment.

Hemisynodontis membranacea (Geoffroy Saint-Hilaire, 1809)

Pimelodus membranaceus Geoffroy Saint-Hilaire, 1809: pl. 13 (figs. 1–2). Type locality: Fl. Nil (Egypte). Holotype: MNHN 0000-4197. Name available from caption on plate, described as *Synodontis membranaceus* in Geoffroy Saint-Hilaire (1827: 297).

Synodontis Guentheri Vaillant, 1893a: 16. Type locality: Karthoum. Syntypes: BMNH 1862.6.17.160 (1), BMNH 1862.6.17.161–162 (2), BMNH 1865.11.15.13 (1).

Distribution: Nile, Niger, Senegal, Gambia, and Volta River basins, Chad system (Gosse, 1986b).

MICROSYNODONTIS Boulenger, 1903

Microsynodontis Boulenger, 1903e: 26. Type species: *Microsynodontis batesii* Boulenger, 1903. Type by monotypy. Gender: Feminine.

Microsynodontis armatus Ng, 2004

Microsynodontis armatus Ng, 2004h: 10, fig. 6. Type locality: Gabon: Ogooué-Ivindo province, Ivindo River drainage, Balé Creek, 0°31'19"N, 12°47'58"E. Holotype: CU 89392.

Distribution: Ivindo River basin, Gabon (Ng, 2004h: 15).

Microsynodontis batesii Boulenger, 1903

Microsynodontis batesii Boulenger, 1903e: 26, pl. 4. Type locality: Mvile River, southern Cameroon. Syntypes (several; apparently more than 6, from notes found in BMNH register): BMNH 1903.7.28.105–110 (6).

Microsynodontis Christyi Boulenger, 1920a: 32, fig. 15. Type locality: Poko, Congo Belge. Syntypes (4): BMNH 1919.9.10.281 (1), MRAC 7168–70 (3).

Distribution: Ntem River basin, southern Cameroon and northern Gabon, and Campo. Ivindo, Lobé, Nyong and Sanaga River basins, southern and central Cameroon (Ng, 2004h: 9).

Remarks: Ng (2004h: 2) stated that *Microsynodontis christyi* should be treated as valid, but provided no further comments. The BMNH syntype of *M. christyi* has had an incorrect jar label, with the registration number listed as BMNH 1919.9.10.381.

Microsynodontis emarginata Ng, 2004

Microsynodontis emarginatus Ng, 2004h: 15, fig. 8. Type locality: Gabon: Haut-Ogooué Province, Motobo I village, Kiéne creek, 1°32'14.1"S, 13°32'43.5"E. Holotype: CU 89393.

Distribution: Ogooué River basin, Gabon (Ng, 2004h: 19).

Microsynodontis hirsuta Ng, 2004

Microsynodontis hirsutus Ng, 2004h: 20, fig. 10. Type locality: Gabon: Woleu-Ntem province, Ngomo creek, where it crosses Oyem-Minvoul road, 1°41'30.0"N, 11°39'18.9"E. Holotype: CU 87040.
Distribution: Nye River of Ntem River basin, Gabon (Ng, 2004h: 23).

Microsynodontis laevigata Ng, 2004

Microsynodontis laevigatus Ng, 2004h: 24, fig. 11. Type locality: Gabon: Ogooué-Ivindo province, Ivindo River drainage, small creek flowing into Ivindo River, Makokou, 0°35'8"N, 12°51'22"E. Holotype: CU 88265.
Distribution: Ivindo River basin, Gabon (Ng, 2004h: 27).

Microsynodontis lamberti Poll & Gosse, 1963

Microsynodontis lamberti Poll & Gosse, 1963: 61, pl. 4 (fig. 1). Type locality: Rivièrre Lilanda, Yangole, Congo centrale. Holotype: MRAC 137837.

Distribution: Lilanda River, Congo River basin; known only from type locality (Gosse, 1986b).

Microsynodontis nannoculus Ng, 2004

Microsynodontis nannoculus Ng, 2004h: 27, fig. 12. Type locality: Equatorial Guinea: Mami River, a tributary of Kyé River. Holotype: MRAC 173145.

Distribution: Kyé River basin, Ntem River drainage, Equatorial Guinea (Ng, 2004h: 31).

Microsynodontis nasutus Ng, 2004

Microsynodontis nasutus Ng, 2004h: 32, fig. 14. Type locality: Gabon: Woleu-Ntem province, Okano River on rapids 0.5 km S of village of Na, 0°48'35"N, 11°38'47"E. Holotype: CU 89394.

Distribution: Okano River, Ogooué River basin, Gabon (Ng, 2004h: 35).

Microsynodontis notata Ng, 2004

Microsynodontis notatus Ng, 2004h: 35, fig. 15. Type locality: Gabon: Ezanga River, about midway between Lake Ezanga and Ogooué mainstream. Holotype: MRAC 80-51-P-839.

Distribution: Lower Ogooué River, Gabon (Ng, 2004h: 39).

Microsynodontis polli Lambert, 1958

Microsynodontis polli Lambert, 1958: 42, fig. 1. Type locality: Rivièrre Gbin, Guinée Forestière, Guinée Française. Holotype: MRAC 119457.

Distribution: Gbin River, Upper Niger; known only from type locality (Gosse, 1986b).

Microsynodontis vigilis Ng, 2004

Microsynodontis vigilis Ng, 2004h: 39, fig. 16. Type locality: Gabon: Moyen-Ogooué province, 12 km N of Lambaréne, branch of Nzorbang creek near village of same name, 0°34'6.2"N, 10°12'46.4"E. Holotype: CU 87039.

Distribution: Lower Ogooué River basin, Gabon (Ng, 2004h: 43).

MOCHOKIELLA Howes, 1980

Mochokiella Howes, 1980: 165. Type species: *Mochokiella paynei* Howes, 1980. Type by original designation. Gender: Feminine.

Mochokiella paynei Howes, 1980

Mochokiella paynei Howes, 1980: 165, figs. 1, 2b. Type locality: Kassawe Forest Reserve, Sierra Leone. Holotype: BMNH 1979.8.22.1.

Distribution: Kassawe Forest Reserve, Sierra Leone; known only from type locality (Gosse, 1986b).

MOCHOKUS Joannis, 1835

Mochokus Joannis, 1835: [Cl. IV, pl. 8]. Type species: *Mochokus niloticus* Joannis, 1835. Type by monotypy. Gender: Masculine.

Mochocus Günther, 1864: 217. Type species: *Mochokus niloticus* Joannis, 1835. Gender: Masculine. Unjustified emendation of *Mochokus* Joannis, 1835.

Rhinoglanis Günther, 1864: 216. Type species: *Rhinoglanis typus* Günther, 1864. Type by original designation. Gender: Masculine.

Mochokus brevis Boulenger, 1906

Mochocus brevis Boulenger, 1906d: 148. Type locality: Fashoda and Lake No, White Nile. Syntypes (43): BMNH

1907.12.2.2529–2542 (14), BMNH 1907.12.2.2544 (1), MNHN 1907-0233 (2).

Distribution: Nile and Chad basins (Gosse, 1986b).

***Mochokus niloticus* Joannis, 1835**

Mochokus Niloticus Joannis, 1835: [Cl. IV, pl. 8]. Type locality: Nil. Holotype: Whereabouts unknown; not found in MNHN by Boulenger (1900d: 526) or subsequently.

Rhinoglanis typus Günther, 1864: 216, unnumbered figure. Type locality: Gondokoro. Holotype: BMNH 1981.4.13.1.

Rhinoglanis Vannutellii Vinciguerra, 1898: 254. Type locality: lago Rodolfo. Syntypes: MSNG 14419 (4).

Distribution: Nile and Niger basins (Gosse, 1986b).

***SYNODONTIS* Cuvier, 1816**

Synodontis Cuvier, 1816: 203. Type species: *Silurus clarias* Linnaeus, 1758. Type by subsequent designation by Bleeker, 1862 (in Bleeker, 1862–63): 6. Gender: Feminine.

Leiosynodontis Bleeker, 1862 (in Bleeker, 1862–63): 6. Type species: *Synodontis maculosus* Rüppell, 1829. Type by original designation. Gender: Feminine.

Pseudosynodontis Bleeker, 1862 (in Bleeker, 1862–63): 6. Type species: *Synodontis serratus* Rüppell, 1829. Type by original designation. Gender: Feminine.

Revision: Vaillant (1895b, 1896b); Poll (1971, with keys to species).

Reviews: Willoughby (1994, Lake Kainji, Nigeria); Bruwer & van der Bank (2003, southern Africa).

Keys: Skelton & White (1990, southern Africa); Bruwer & van der Bank (2003).

Remarks: The name *Synodentes* published in Minding (1832: 116) is considered an incorrect subsequent spelling and not an available name.

***Synodontis acanthomias* Boulenger, 1899**

Synodontis acanthomias Boulenger, 1899a: 46, pl. 23. Type locality: Boma et Léopoldville [restricted to Boma by lectotype designation]. Lectotype: MRAC 18, designated by Poll (1971: 282).

Synodontis pfefferi Steindachner, 1912: 447. Type locality: Dscha, Nebenfluss des Congo im Bezirke Molundu des südlichen Kamerun. Holotype: NMW 9023. Species illustrated and described in more detail in Steindachner (1913: 43, pl. 8).

Distribution: Congo River basin, except central forest (Gosse, 1986b).

***Synodontis afrofischeri* Hilgendorf, 1888**

Synodontis Afro-Fischeri Hilgendorf, 1888: 77. Type locality: Victoria-Nyanza (Ukerewe-See). Holotype: ZMB 12745. Holotype illustrated in Poll (1971: fig. 28) and Seegers (1996: fig. 171).

Distribution: Lakes Victoria, Nabugabo, Kioga, Ihema; Nile, Kagera, Kingani Malagarasi Rivers (Gosse, 1986b); Piti River, Lake Rukwa basin (Seegers, 1996).

***Synodontis alberti* Schilthuis, 1891**

Synodontis alberti Schilthuis, 1891: 88. Type locality: Kinshasa, Stanley Pool. Lectotype: BMNH 1899.9.6.9, designated by, and illustrated in, Poll (1971: 252, fig. 114).

Distribution: Congo River basin (Gosse, 1986b).

***Synodontis albolineata* Pellegrin, 1924**

Synodontis albolineatus Pellegrin, 1924b: 320, fig. 1. Type locality: Madjingo, rivière Djoua (Gabon). Holotype: MNHN 1924-0147, illustrated in Poll (1971: fig. 68).

Distribution: Ogowe River basin; known only from type locality (Gosse, 1986b).

***Synodontis angelica* Schilthuis, 1891**

Synodontis angelica Schilthuis, 1891: 87. Type locality: Léopoldville, Stanley Pool. Holotype: BMNH 1899.9.6.8; holotype illustrated in Poll (1971: fig. 110, top).

Synodontis tholloni Boulenger, 1901a: 304. Type locality: bassin de l'Ogooué (?). Holotype: MNHN 1890-0030.

Synodontis angelicus zonatus Poll, 1933: 134, fig. 11. Type locality: Lukulu (riv. Lukulu). Holotype: MRAC 37904; holotype illustrated in Poll (1971: fig. 110, bottom). Originally as *Synodontis angelicus* var. *zonatus*.

Synodontis werneri Boeseman, 1957: 146, pl. 5. Type locality: Stanley Pool, Belgian Congo. Holotype: RMNH 20913.

Distribution: Congo River basin, except in central forest (Gosse, 1986b).

***Synodontis annectens* Boulenger, 1911**

Synodontis annectens Boulenger, 1911d: 56. Type locality: Nianimaru, Gambia; Culufi and Geba Rivers, Portuguese Guinea [restricted to Nianimaru by lectotype designation]. Lectotype: BMNH 1901.12.28.60, designated by Poll (1971: 131).

Distribution: Gambia; Geba basin, Guinea Bissau; and Taja basin, Sierra Leone (Gosse, 1986b).

***Synodontis ansorgii* Boulenger, 1911**

Synodontis ansorgii Boulenger, 1911b: 375. Type locality: Geba and Culufi Rivers, Portuguese Guinea [restricted to Geba River, by lectotype designation]. Lectotype: BMNH 1912.4.1.128, designated by, and illustrated in, Poll (1971: 135, fig. 56).

Distribution: Geba River basin, Guinea-Bissau; and Taja River basin, Sierra Leone (Gosse, 1986b).

***Synodontis arnoulti* Roman, 1966**

Synodontis arnoulti Roman, 1966: 141, pl. 7 (fig. 14). Type locality: le Kou, affluent de la Volta Noire, à 30 km au Nord de Bobo-Dioulasso. Holotype: MRAC 141742.

Distribution: Volta River basin (Gosse, 1986b).

***Synodontis aterrima* Poll & Roberts, 1968**

Synodontis aterrimus Poll & Roberts, 1968: 297, figure on p. 298. Type locality: Bokuma, Bassin du Congo. Holotype: MRAC 101466.

Distribution: Central Congo River basin (Gosse, 1986b).

***Synodontis bastiani* Daget, 1948**

Synodontis bastiani Daget, 1948: 35, fig. 10. Type locality: Côte d'Ivoire: Bouaflé. Holotype: MNHN 1949-0055.

Synodontis dageti Poll & Roman, 1967: 185, fig. 3. Type locality: [68 km south of Bolo-Dioulasso], Haut Comoé. Holotype: MRAC 141982.

Synodontis eburneensis Daget, 1965: 473. Type locality: le Bandama au lieu dit Lamto, à mi-chemin entre Toumodi et Tiassalé (Côte d'Ivoire). Holotype: MNHN 1964-0266; holotype illustrated in Poll (1971: fig. 92).

Distribution: Sassandra, Bandama, Bia, and Comoré basins, Côte d'Ivoire (Gosse, 1986b).

Remarks: Synonymy of these three names first proposed by Paugy & Roberts (1992).

***Synodontis batesii* Boulenger, 1907**

Synodontis Batesii Boulenger, 1907b: 50. Type locality: Ja River, South Cameroon. Lectotype: BMNH 1907.5.22.179; designated by, and illustrated in, Poll (1971: 331, fig. 150).

Distribution: Central Congo River basin; Ja River and Nyong River, Cameroon; Muni River, Equatorial Guinea (Gosse, 1986b).

***Synodontis brichardi* Poll, 1959**

Synodontis brichardi Poll, 1959: 100, pl. 17 (figs. 2 a–c). Type locality: Léopoldville, début des rapides à Kinsuka. Holotype: MRAC 100494.

Distribution: Lower Congo River rapids (Gosse, 1986b).

***Synodontis budgetti* Boulenger, 1911**

Synodontis budgetti Boulenger, 1911a: 403, fig. 305. Type locality: Lokoja, Upper Niger [Nigeria]. Holotype: BMNH 1904.1.20.42.

Distribution: Niger and Oueme Rivers, Nokuoue Lake (Gosse, 1986b).

***Synodontis camelopardalis* Poll, 1971**

Synodontis camelopardalis Poll, 1971: 430, fig. 202; pls. 7 (fig. 4), 12 (fig. 14). Type locality: Eala, riv. Ruki. Holotype: MRAC 47654.

Distribution: Central Congo River basin; known only from type locality (Gosse, 1986b).

***Synodontis caudalis* Boulenger, 1899**

Synodontis caudalis Boulenger, 1899a: 44, pl. 22 (fig. 1). Type locality: Boma, Matadi, Léopoldville [restricted to Matadi by lectotype designation]. Lectotype: MRAC 57, designated by, and illustrated in, Poll (1971: 274, fig. 124).

Distribution: (Gosse, 1986b).

***Synodontis caudovittata* Boulenger, 1901**

Synodontis caudovittatus Boulenger, 1901c: 10. Type locality: Mouth of Lake No, White Nile. Lectotype: BMNH 1907.12.2.2360, designated by, and illustrated in, Poll (1971: 104).

Distribution: Nile River basin (Gosse, 1986b).

***Synodontis centralis* Poll, 1971**

Synodontis centralis Poll, 1971: 435, fig. 204; pls. 7(fig. 7), 12 (fig. 12). Type locality: Ndwa village, Kunungu, Zaire. Holotype: MRAC 57198.

Distribution: Central Congo River basin (Gosse, 1986b).

***Synodontis clarias* (Linnaeus, 1758)**

Silurus clarias Linnaeus, 1758: 306. Type locality: habitat in Americae, Africae flaviis. Types: See Remarks, below.

Silurus callarias Bloch & Schneider, 1801: 379. Type locality: in flaviis Americae australis et Nilo. Syntypes: on *Silurus clarias* Linnaeus, and Bloch (1782: pl. 35, figs. 1–2). An unneeded new name for *Silurus clarias* Linnaeus, 1758.

Pimelodus synodontis Geoffroy Saint-Hilaire, 1809: pl. 12 (figs. 5–6). Type locality: Nil. Holotype: MNHN 0000-4194. Name available on caption to plate. Species described as *Synodontis macrodon* by Geoffroy Saint-Hilaire (1827: 295).

Synodontis macrodon Geoffroy Saint-Hilaire, 1827: 295. Type locality: Fl. Nil (Egypte). Holotype: MNHN 0000-4194. New name for *Pimelodus synodontis* Geoffroy Saint-Hilaire, 1809.

Distribution: Nile, Niger, Senegal, Gambia, and Volta River basins, Chad system (Gosse, 1986b).

Remarks: Silfvergrip (1996: 8–9) discussed the specimens that formed the type series for *Silurus clarias*. Two of the four type specimens are extant, neither of which is a species of *Synodontis*. Silfvergrip (1996: 9) selected “Hasselquist’s specimen” (NRM 7044), which was subsequently identified as a specimen of *Bagrus docmak* (Silfvergrip, pers. commun., 2003), as the lectotype of *Silurus clarias*. The nomenclatural impact of this selection would result in *Bagrus docmak* (Forsskål, 1775) becoming a junior synonym of *Silurus clarias*, and the genera *Synodontis* Cuvier, 1816, and *Bagrus* Bosc, 1816, becoming synonyms. Because of the nomenclatural impact of Silfvergrip’s lectotype designation on two relatively well known African catfishes, his designation is not followed here but it left for a more thorough review in the future.

***Synodontis comoensis* Daget & Lévêque, 1981**

Synodontis comoensis Daget & Lévêque, 1981: 45, figs. 1–5. Type locality: rivière Comoé au niveau du bac de Gansé, Parc National de la Comoé (Côte d’Ivoire), 8°39'N, 3°56'W. Holotype: MNHN 1980-1636.

Distribution: Comoé River, Côte d’Ivoire; known only from type locality (Gosse, 1986b).

***Synodontis congica* Poll, 1971**

Synodontis congicus Poll, 1971: 440, fig. 206; pls. 7 (fig. 6), 12 (fig. 16). Type locality: Gangala na Bodio, riv. Dungu. Holotype: MRAC 165695.

Distribution: Congo River basin, except lower Congo, central basin, Luapula-Moero, and high Katanga (Gosse, 1986b).

***Synodontis contracta* Vinciguerra, 1928**

Synodontis contractus Vinciguerra, 1928: 24, pl. 1. Type locality: fiume Rubi a Buta [upper Congo basin]. Lectotype: MSNG 26493, designated by Poll (1971: 383).

Synodontis davidi Axelrod, 1970: 33, figs. Type locality: Stanley Pool near Kinshasha, Congo. Type(s): Whereabouts unknown.

Distribution: Congo River basin (Gosse, 1986b).

***Synodontis courteti* Pellegrin, 1906**

Synodontis Courteti Pellegrin, 1906b: 473. Type locality: Fort-Archambault (Chari) [Soudan]. Holotype: MNHN 1904-0083; illustrated in Poll (1971: 129, fig. 54).

Synodontis thomasi Chabanaud, 1934: 219. Type locality: Afrique équatoriale française. Type(s) (510 mm): Whereabouts unknown.

Distribution: Chad and Niger basins (Gosse, 1986b).

***Synodontis cuangoana* Poll, 1971**

Synodontis cuangoanus Poll, 1971: 446, fig. 208; pl. 7 (fig. 10), pl. 12 (fig. 17). Type locality: Cuango, Cafunfo (8°47'S, 18°01'E). Holotype: MRAC 172253.

Distribution: Congo River basin, south of Kasai, Zaire and Angola (Gosse, 1986b).

***Synodontis decora* Boulenger, 1899**

Synodontis decorus Boulenger, 1899a: 49, pl. 25. Type locality: Léopoldville et Nouvelle-Anvers; [restricted to Nouvelle-Anvers by lectotype designation]. Lectotype: MRAC 154, designated by Poll (1971: 291).

Synodontis vittatus Boulenger, 1920a: 31, pl. 25. Type locality: Stanleyville. Lectotype: MRAC 7119, designated by Poll (1971: 295).

Distribution: Congo River basin, except Luapula River system (Gosse, 1986b).

***Synodontis dekimpei* Paugy, 1987**

Synodontis dekimpei Paugy, 1987: 357, fig. 1. Type locality: rivière Konkouré à Soukya (aval du pont routier Kindia-Telimélé en amont de Tondo) (Guinée). Holotype: MRAC 81-20-P-11.

Distribution: Konkouré River, Guinea (Paugy, 1987).

***Synodontis depauwi* Boulenger, 1899**

Synodontis depauwi Boulenger, 1899a: 45, pl. 22 (fig. 2). Type locality: Léopoldville. Lectotype: MRAC 111, designated by Poll (1971: 278).

Distribution: Congo River at Stanley Pool (Gosse, 1986b).

***Synodontis dhonti* Boulenger, 1917**

Synodontis dhonti Boulenger, 1917b: 367. Type locality: Kilewa Bay [Lake Tanganyika]. Holotype: MRAC 14344; holotype illustrated in Poll (1946: 211, figs. 21–22, and 1971: fig. 168).

Synodontis irsacae Matthes, 1959b: 78. Type locality: Kalundu, nord du Lac Tanganyika. Holotype: MRAC 130315. Distribution: Lake Tanganyika (Gosse, 1986b).

***Synodontis dorsomaculata* Poll, 1971**

Synodontis dorsomaculatus Poll, 1971: 449, fig. 210; pl. 7 (fig. 8), pl. 12 (fig. 18). Type locality: Kadia, Kisale. Holotype: MRAC 71395.

Distribution: Upper Lualaba (Katanga), Congo River basin (Gosse, 1986b).

***Synodontis euptera* Boulenger, 1901**

Synodontis eupterus Boulenger, 1901c: 11. Type locality: Mouth of Lake No, White Nile. Holotype (147 mm SL): BMNH 1907.12.2.2376; holotype redescribed and illustrated in Boulenger (1907a: 370, pl. 68), with locality as Goz-abu-Gumah, White Nile; holotype also illustrated in Poll (1971: fig. 44).

Synodontis macrepipterus Pellegrin, 1922b: 222. Type locality: Fort Crampel (Gribingui). Holotype: MNHN 1922-0031.

Distribution: White Nile, Chad Basin (Gribingui), Niger and Volta Rivers (Gosse, 1986b).

***Synodontis fascipinna* Nichols & La Monte, 1953**

Synodontis fascipinna Nichols & La Monte, 1953: 2, fig. 2. Type locality: Birao, eastern French Equatorial Africa. Holotype: AMNH 19841.

Distribution: Headwaters of the Chari River basin; known only from type locality (Gosse, 1986b).

***Synodontis filamentosa* Boulenger, 1901**

Synodontis filamentosus Boulenger, 1901c: 10. Type locality: Mouth of Lake No, White Nile. Holotype: BMNH 1907.12.2.2377; holotype illustrated in Boulenger (1907: pl. 69) and Poll (1971: fig. 42).

Synodontis augierasi Pellegrin, 1929a: 136, fig. 1. Type locality: [Niger R. at Koulikoro, Mali, w. Africa.] Syntypes: MNHN 1928-0243 (1), MNHN 1928-0244 (1).

Distribution: Nile, Chad, Niger, and Volta basins (Gosse, 1986b).

***Synodontis flavitaeniata* Boulenger, 1919**

Synodontis flavitaeniatus Boulenger, 1919: 161. Type locality: Eala [Congo River basin]. Holotype: MRAC 1369; holotype illustrated in Poll (1971: fig. 170).

Distribution: Central Congo River basin (Gosse, 1986b).

***Synodontis frontosa* Vaillant, 1895**

Synodontis frontosus Vaillant, 1895a: 48. Type locality: Nil [White Nile, Sudan]. Holotype: MNHN 0000-4208. Illustrated in Vaillant (1895b: pl. 10, fig. 8) and described in more detail in Vaillant (1896b: 146).

Synodontis Cternii Vinciguerra, 1898: 247. Type locality: Lago Rodolfo. Holotype: MSNG 14423 (dry).

Distribution: Nile basin; Lakes Rudolf, Albert, Eduard and Kioga; Chad basin; Uebi Giupa (Uebi Shebeli) (Gosse,

1986b).

Synodontis fuelleborni Hilgendorf & Pappenheim, 1903

Synodontis fuelleborni Hilgendorf & Pappenheim, 1903: 265. Type locality: Rukwa-See. Lectotype: ZMB 16309, designated by, and illustrated in, Poll (1971: 320, fig. 146); lectotype also illustrated in Seegers (1996: fig. 177). Distribution: Rukwa Lake and Rufiji basin (Gosse, 1986b).

Synodontis gambiensis Günther, 1864

Synodontis gambiensis Günther, 1864: 214. Type locality: Gambia. Holotype: BMNH 2005.9.9.1; holotype illustrated in Poll (1971: fig. 20).

Synodontis gambiensis latifrons Blache, 1964: 201, fig. 112. Type locality: Lake Tchad basin. Syntypes: Possibly MNHN 1959-0495 (2), MNHN 1959-0500 (5), MNHN 1959-0524 (8).

Distribution: Niger, Chad, Gambia, Geba, Taja, Waange, Volta, and Oueme basins (Gosse, 1986b).

Remarks: Considered to be a synonym of *Synodontis schall* by Lévêque *et al.* (1989: 120) and Paugy (in Lévêque *et al.*, 1992: 561). Register number for holotype of *Synodontis gambiensis* Günther, given in Poll (1971) and Eschmeyer *et al.* (1998) as BMNH 1863.16.4.40, is incorrect; it is not a possible register number in the BMNH system.

Synodontis geledensis Günther, 1896

Synodontis geledensis Günther, 1896b: 220, fig. 2. Type locality: Geledi on the Shebeli. Holotype: BMNH 1895.12.31.69.

Distribution: Uebi Shebeli, Somalia; Lorian Swamps and Uasso Nyiro (Uebi Giuba basin), Kenya (Gosse, 1986b).

Synodontis gobroni Daget, 1954

Synodontis gobroni Daget, 1954: 301, fig. 113. Type locality: Mohti, Markala. Syntypes (4): MNHN 1954-0005 (1), MNHN 1960-0471 (1).

Distribution: Niger River basin (Gosse, 1986b).

Remarks: Treated in Willoughby (1994: 28) as a synonym of *Synodontis guttatus* Günther, 1865.

Synodontis granulosa Boulenger, 1900

Synodontis granulosus Boulenger, 1900c: 480. Type locality: North end of Lake Tanganyika. Lectotype: BMNH 1906.9.6.40, designated by, and illustrated in, Poll (1971: 303, fig. 138).

Distribution: Lake Tanganyika (Gosse, 1986b).

Synodontis greshoffi Schilthuis, 1891

Synodontis greshoffi Schilthuis, 1891: 87. Type locality: Kinshasa, Stanley Pool. Holotype: BMNH 1899.9.6.7; holotype illustrated in Poll (1971: fig. 112).

Synodontis holopercnus Boulenger, 1920a: 28. Type locality: Bosabangi, Avakubi, & Poko, Congo Belge. Syntypes (5): BMNH 1919.9.10.264-265 (2), MRAC 7616 (1), MRAC 7775 (1), MRAC 7829 (1).

Synodontis pantherinus Boulenger, 1920b: 149. Type locality: Ituri à Avakubi. Holotype: MRAC 14513.

Distribution: Congo River basin, except Luapula River system and lower Congo (Gosse, 1986b).

Synodontis guttata Günther, 1865

Synodontis guttatus Günther, 1865a: 452. Type locality: Niger. Holotype: BMNH 1865.4.6.4 (29 inches, dry); holotype illustrated in Boulenger (1911a: fig. 335) and Poll (1971: fig. 24).

Distribution: Lower Niger basin (Gosse, 1986b).

Synodontis haugi Pellegrin, 1906

Synodontis Haugi Pellegrin, 1906a: 470. Type locality: Ngomo (Ogôoué). Holotype: MNHN 1906-0209; holotype illustrated in Poll (1971: fig. 52).

Distribution: Ogowe River basin (Gosse, 1986b).

Synodontis iturii Steindachner, 1911

Synodontis melanostictus iturii Steindachner, 1911c: 534. Type locality: Ituri flüsse. Holotype: NMW 46049; holotype illustrated in Poll (1971: fig. 160). Originally as *Synodontis melanostictus* var. *iturii*.

Synodontis ituriensis Boulenger, 1920a: 28. Type locality: Mawambi sur l'Ituri, Congo Belge. Holotype: MRAC 7576.

Distribution: Ituri River (Gosse, 1986b).

Synodontis katangae Poll, 1971

Synodontis katangae Poll, 1971: 454, fig. 212; pls. 7 (fig. 11), 12 (fig. 19). Type locality: Lukonzolwa, lac Moëro.

Holotype: MRAC 14231.

Distribution: Congo River basin, high Katanga and Luapula-Moero (Gosse, 1986b).

***Synodontis khartoumensis* Abu Gideiri, 1967**

Synodontis khartoumensis Abu Gideiri, 1967: 133, fig. 1. Type locality: Blue Nile, two miles from place where the Blue Nile joins the White Nile to form the main Nile at Khartoum, Sudan. Holotype: at Sudan Natural History Museum.

Distribution: Nile basin and Lake Albert (Gosse, 1986b).

***Synodontis koensis* Pellegrin, 1933**

Synodontis koensis Pellegrin, 1933a: 110, fig. on p. 111. Type locality: Man (rivière Ko). Holotype: NMBA 4500; holotype illustrated in Poll (1971: fig. 72).

Distribution: Nzo River of Sassandra River basin, Côte d'Ivoire (Gosse, 1986b).

***Synodontis laessoei* Norman, 1923**

Synodontis laessoei Norman, 1923c: 696. Type locality: Kokema River, a tributary of the Quanza River, Angola; altitude 4000 ft. Holotype: BMNH 1923.8.15.3; holotype illustrated in Poll (1971: fig. 176).

Distribution: Kolema River of Quanza River basin, Angola; known only from type locality (Gosse, 1986b).

***Synodontis leopardina* Pellegrin, 1914**

Synodontis leopardinus Pellegrin, 1914: 26. Type locality: Pays des Barotsés (Ht.-Zambèze). Holotype: MNHN 1913-0321; holotype illustrated in Poll (1971: fig. 162).

Synodontis jallae Gilchrist & Thompson, 1917: 561. Type locality: Shesheke, S. Rhodesia. Holotype: SAM 14290 [now at AMG].

Distribution: Cunene, Upper Zambezi and Okovango river basins (Skelton, 1993).

***Synodontis leoparda* Pfeffer, 1896**

Synodontis leopardus Pfeffer, 1896: 35. Type locality: Rufu bei Korogwe. Holotype: at ZMH (Gosse, 1986: 132).

Distribution: Coastal rivers of Tanzania and Somalia (Gosse, 1986b).

Remarks: Considered by De Vos (2001a: 49) as a possible synonym of *Synodontis zanzibaricus* Peters, 1868.

***Synodontis levequei* Paugy, 1987**

Synodontis levequei Paugy, 1987: 361, fig. 3. Type locality: rivière Kakrima (basin du Konkouré), près de Kasséri (Guinée). Holotype: MNHN 1987-0931.

Distribution: Kakrima River (Paugy, 1987).

***Synodontis longirostris* Boulenger, 1902**

Synodontis longirostris Boulenger, 1902d: 44, pl. 12 (fig. 2). Type locality: Rivière Yembe à Banzyville. Holotype: MRAC 1243.

Synodontis ovidius Lönnberg & Rendahl, 1920: 173, figs 4–5. Type locality: Mukimbungu, Lower Congo. Holotype: NRM 9913.

Distribution: Congo River basin (Gosse, 1986b).

***Synodontis longispinis* Pellegrin, 1930**

Synodontis Batesi longispinis Pellegrin, 1930: 207. Type locality: la Sangha. Holotype: MNHN 1929-0245; holotype illustrated in Poll (1971: fig. 182). Originally as *Synodontis Batesi* var. *longispinis* nov. var.

Distribution: Sangha River, Congo River basin (Gosse, 1986b).

***Synodontis lufirae* Poll, 1971**

Synodontis lufirae Poll, 1971: 459, fig. 214; pl. 7 (fig. 12). Type locality: Lac de Koni, Lufira. Holotype: MRAC 168400.

Distribution: Lufira River, Congo River basin (Gosse, 1986b).

***Synodontis macrophthalmus* Poll, 1971**

Synodontis macrophthalmus Poll, 1971: 207, fig. 98; pls. 3 (fig. 15), 10 (fig. 5). Type locality: Ampem, Volta Lake (Ghana). Holotype: MRAC 168500.

Distribution: Volta Lake, Ghana (Gosse, 1986b).

***Synodontis macrops* Greenwood, 1963**

Synodontis macrops Greenwood, 1963: 66. Type locality: tributary stream of the Aswa River, Teso district, Uganda.

Holotype: BMNH 1961.12.1.319; holotype illustrated in Poll (1971: fig. 88).

Distribution: Aswa River basin; Nile River system, Uganda (Gosse, 1986b).

***Synodontis macrostigma* Boulenger, 1911**

Synodontis macrostigma Boulenger, 1911a: 432, fig. 325. Type locality: Okovango River. Lectotype: BMNH 1910.5.31.37, designated by Poll (1971: 345). A previous lectotype designation may have been made by Gilchrist & Thompson (1913: 462) in the caption to fig. 109, in which the illustrated specimen is labeled as the type, if the illustrated specimen can be identified.

Distribution: Kafue and Mashi Rivers, Zambezi River basin; Okovango and Cunene Rivers (Gosse, 1986b; Skelton, 1993).

***Synodontis macrostoma* Skelton & White, 1990**

Synodontis macrostoma Skelton & White, 1990: 281, fig. 3. Type locality: Upper Zambezi River at Katima Mulilo, Caprivi, Namibia. Holotype: AMG P11687.

Distribution: Cunene, Okavango, upper Zambezi and Kafur river basins (Skelton, 1993).

***Synodontis manni* De Vos, 2001**

Synodontis manni De Vos, 2001a: 42, figs. 2–3. Type locality: Kenya: Tana River at Baomo area near Baomo village, 1°55'S, 40°08'E. Holotype: NMK FW-600/1.

Distribution: Tana River, Kenya (De Vos, 2001a).

***Synodontis marmorata* Lönnberg, 1895**

Synodontis marmoratus Lönnberg, 1895: 186. Type locality: Muddy rivulet at Bonge [Cameroon]. Lectotype: ZMUU, 38.5 mm SL specimen, designated by, and illustrated in, Poll (1971: 93, fig. 34).

Synodontis marmoratus truncatus Holly, 1927a: 8. Type locality: Njongfluß, Kamerun. Treated by Gosse (1986) as possible synonym of *S. marmoratus*.

Distribution: Sanaga and Nyong Rivers (Gosse, 1986b).

***Synodontis matthesi* Poll, 1971**

Synodontis matthesi Poll, 1971: 462, fig. 216; pls. 7 (fig. 14), 12 (fig. 20). Type locality: Mtera, riv. Ruaha (Tanzania). Holotype: ZMA 109743.

Distribution: Rufiji River basin, Tanzania (Gosse, 1986b).

***Synodontis melanoptera* Boulenger, 1903**

Synodontis melanopterus Boulenger, 1903d: 327, pl. 29 (fig. 1). Type locality: Oguta, Niger Delta. Lectotype: BMNH 1902.10.25.3, designated and illustrated in Poll (1971: 114, fig. 46).

Distribution: Porto Novo, Benin; Niger, Ouémé and Ogun River basins (Gosse, 1986b).

***Synodontis multimaculata* Boulenger, 1902**

Synodontis multimaculatus Boulenger, 1902d: 43, pl. 12 (fig. 1). Type locality: l'Ubangi à Banzyville. Holotype: MRAC 1314; illustrated in Poll (1971: 306, fig. 140).

Distribution: Ubangi River, Congo River basin (Gosse, 1986b).

***Synodontis multipunctata* Boulenger, 1898**

Synodontis multipunctatus Boulenger, 1898d: 497. Type locality: Lake Tanganyika. Holotype: BMNH 1898.9.9.76; illustrated and described in more detail in Boulenger (1898f: 24, pl. 8), with locality as Sumba.

Distribution: Lake Tanganyika (Gosse, 1986b).

***Synodontis nebulosa* Peters, 1852**

Synodontis nebulosus Peters, 1852: 682. Type locality: Tette [Zambezi River, Mozambique]. Holotype: ZMB 3120; illustrated in Peters (1868c: 28, pl. 5, fig. 1).

Distribution: Lower and middle Zambezi River basin (Skelton, 1993).

***Synodontis nigrita* Valenciennes, 1840**

Synodontis nigrita Valenciennes, in Cuvier & Valenciennes, 1840b: 265 (197 in Strasbourg deluxe edition), pl. 441. Type locality: Sénégal. Holotype: MNHN 0000-3051; illustrated in Poll (1971: 59, fig. 14).

Synodontis ornatus Pappenheim, in Pappenheim & Boulenger, 1914: 251, pl. 5 (fig. 2). Type locality: Albert See. Syntypes: ZMB 19097 (8).

Distribution: Nile, Chad, Niger, Senegal, Gambia, Casamance, Geba, Kolente and Volta River basins; coastal rivers from Ghana to Nigeria (Gosse, 1986b).

***Synodontis nigriventris* David, 1936**

Synodontis nigriventris David, 1936b: 417. Type locality: Mongende; Ikengo; Basongo; Koteli; Buta; Budjala; Léopoldville; Flandria [restricted to Buta by lectotype designation]. Lectotype: MRAC 30312, designated by, and illustrated in, Poll (1971: 397, fig. 186).
Distribution: Central Congo River basin (Gosse, 1986b).

***Synodontis nigromaculata* Boulenger, 1905**

Synodontis nigromaculatus Boulenger, 1905b: 645. Type locality: Lake Bangwelo. Holotype: BMNH 1905.11.10.10; holotype illustrated in Poll (1971: fig. 148).

Synodontis melanostictus Boulenger, 1906f: 553, pl. 34. Type locality: Lofu, Lake Tanganyika. Holotype: BMNH 1906.9.8.72.

Synodontis colyeri Boulenger, 1923: 438. Type locality: Mansa River, N. Rhodesia. Holotype: SAM 14529 [now at AMG].

Distribution: Okavango and upper Zambezi River basins; Upper Congo River basin, Zambia; Lake Tanganyika and Kasai River (Skelton, 1993).

***Synodontis njassae* Keilhack, 1908**

Synodontis njassae Keilhack, 1908: 168. Type locality: Njassa-See. Lectotype: ZMB 18191, designated by, and illustrated in, Poll (1971: 337, fig. 154).

Distribution: Lake Malawi (Gosse, 1986b).

***Synodontis notata* Vaillant, 1893**

Synodontis Maculatus Vaillant, 1892b: 2. Type locality: Riv. Oubangui à Bangui. Holotype: apparently MNHN 1892-0080. *Nomen oblitum*, see Remarks.

Synodontis notatus Vaillant, 1893a: 17. Type locality: Riv. Oubangui à Bangui. Holotype: apparently MNHN 1892-0080. Proposed as a replacement for *Synodontis maculatus*, Vaillant, 1892, considered by Vaillant to be effectively preoccupied by *Synodontis maculosus* Rüppell, 1829. Poll (1971: fig. 116) illustrated MNHN 1886-0436 and referred to it as the lectotype. *Nomen protectum*.

Synodontis notatus binotata Pellegrin, 1926: 205. Type locality: Bolobo, Congo belge. Syntypes (2): MRAC 49686, MNHN 1926-0195. Originally as *Synodontis notatus* var. *binotata*.

Synodontis notatus ocellatus Poll, 1938: 404, fig. 9. Type locality: environs de Maka, riv. Londo [Katanga, bassin du Congo]. Holotype: MRAC 49686. Originally as *Synodontis notatus* var. *ocellatus*.

Distribution: Congo River basin (Gosse, 1986b).

Remarks: *Synodontis notata* Vaillant, 1893, has been in wide use since its proposal (see Gosse, 1986b, for list of publications) and considered here to be a *nomen protectum*. The holotype for *Synodontis maculatus* Vaillant, 1892, and *S. notatus* Vaillant, 1893, is listed by Bertin & Estève (1950) and other sources as MNHN 1886-0436, but Gosse (1986: 135) indicated that the holotype was actually MNHN 1892-0080, which was collected by Dybowski from the Oubangui, as stated in Vaillant (1892).

***Synodontis nummifer* Boulenger, 1899**

Synodontis nummifer Boulenger, 1899a: 47, pl. 24. Type locality: Léopoldville. Holotype: MRAC 107.

Distribution: Congo River basin (Gosse, 1986b).

***Synodontis obesus* Boulenger, 1898**

Synodontis obesus Boulenger, 1898a: 415. Type locality: Gaboon and Opobo River, Old Calabar [restricted to Opobo River by lectotype designation]. Lectotype: BMNH 1896.5.5.67, designated by and illustrated in Poll (1971: 96, fig. 36).

Synodontis loppei Pellegrin, 1927: 365. Type locality: Edea [Cameroon]. Syntypes: MHNL R.P.197 (1), MNHN 1927-0278 (1). Illustrated in Pellegrin (1928a: 5, unnumbered figure).

Distribution: Coastal rivers from Ghana to Gabon; basins of the Comoë, Pra, Volta, Momo, Cross, Wouri, Sanaga Nyong, Kribi and Lobé Rivers (Gosse, 1986b).

***Synodontis ocellifer* Boulenger, 1900**

Synodontis ocellifer Boulenger, 1900e: 514. Type locality: Kunchow Creek [River Gambia]. Lectotype: BMNH 1901.7.17.7; designated by, and illustrated in, Poll (1971: 101, fig. 38).

Distribution: Senegal, Gambia, Volta, Chad, and Niger basins (Gosse, 1986b).

Synodontis omias Günther, 1864

Synodontis omias Günther, 1864: 213. Type locality: West Africa [Upper Niger River]. Holotype: BMNH 1863.12.9.3 (skeleton).

Distribution: Niger River basin (Gosse, 1986b).

Synodontis ornatipinnis Boulenger, 1899

Synodontis ornatipinnis Boulenger, 1899b: 111, pl. 43 (fig. 2). Type locality: Coquilhatville et Bikoro (Lac Tumba) [restricted to Coquilhatville by lectotype designation]. Lectotype: MRAC 968, designated by Poll (1971: 299).

Distribution: Congo River system (Gosse, 1986b).

Synodontis ornatissima Gosse, 1982

Synodontis ornatus Boulenger, 1920a: 29, fig. 14. Type locality: Poco, Congo Belge. Holotype: MRAC 7144. Preoccupied by *Synodontis ornatus* Pappenheim, 1914; replaced by *Synodontis ornatissimus* Gosse, 1982.

Synodontis ornatissimus Gosse, 1982: 48. Type locality: Poco, Congo Belge. Holotype: MRAC 7144. Replacement for *Synodontis ornatus* Boulenger, 1920; preoccupied by *Synodontis ornatus* Pappenheim, 1914.

Distribution: Ubangi River and its tributaries, Congo River basin (Gosse, 1986b).

Synodontis pardalis Boulenger, 1908

Synodontis pardalis Boulenger, 1908a: 30. Type locality: a waterfall of the Libi River, near the Ja River (Congo System), South Cameroon. Lectotype: BMNH 1909.4.29.91, designated by, and illustrated in, Poll (1971: 334, fig. 152).

Distribution: Ja River, Congo River basin, Cameroon (Gosse, 1986b).

Synodontis petricola Matthes, 1959

Synodontis petricola Matthes, 1959b: 78. Type locality: lac Tanganyika. Holotype: MRAC 130357; holotype illustrated in Poll (1971: fig. 192).

Distribution: Lake Tanganyika (Gosse, 1986b).

Synodontis pleurops Boulenger, 1897

Synodontis pleurops Boulenger, 1897b: 423. Type locality: Stanley Falls, upper Congo. Holotype: BMNH 1897.9.30.13; holotype illustrated in Poll (1971: fig. 120).

Distribution: Congo River basin, except the Luapula system and the lower Congo River (Gosse, 1986b).

Synodontis polli Gosse, 1982

Synodontis eurystomus Matthes, 1959b: 77. Type locality: lac Tanganyika, confiné au littoral rocheux du lac. Holotype: MRAC 130440. Holotype illustrated in Matthes (1962: pl. 1, fig. c) and Poll (1971: fig. 190). Preoccupied by *Synodontis eurystomus* Pfeffer, 1889; replaced by *Synodontis polli* Gosse, 1982.

Synodontis polli Gosse, 1982: 48. Type locality: Luhanga, Lake Tanganyika. Holotype: MRAC 130440. Replacement for *Synodontis eurystomus* Matthes, 1959.

Distribution: Lake Tanganyika (Gosse, 1986b).

Synodontis polyodon Vaillant, 1895

Synodontis polyodon Vaillant, 1895a: 48. Type locality: Ogôoué. Holotype: MNHN 1886-0433; holotype illustrated in Vaillant (1895b: pl. 9, fig. 1), and described in more detail in Vaillant (1896b: 127).

Distribution: Ogowe River basin (Gosse, 1986b).

Synodontis polystigma Boulenger, 1915

Synodontis polystigma Boulenger, 1915: 170. Type locality: Rivièrre Luapula, à Kasenga; lac Moero, à Lukonzolwa [restricted to lac Moéro by lectotype designation]. Lectotype: MRAC 14174, designated by, and illustrated in, Poll (1971: 356, fig. 164).

Distribution: Luapula-Moëro Rivers, Congo River basin (Gosse, 1986b).

Synodontis pulcher Poll, 1971

Synodontis pulcher Poll, 1971: 465, fig. 218; pls. 7 (fig. 13), 12 (fig. 21). Type locality: Stanley Pool. Holotype: MRAC 168399.

Distribution: Stanley Pool, Congo River basin (Gosse, 1986b).

Synodontis punctifer Daget, 1965

Synodontis punctifer Daget, 1965: 472. Type locality: le Nzo à Guiglo, Côte d'Ivoire. Holotype: MNHN 1964-0265; holotype illustrated in Poll (1971: fig. 90).

Distribution: Nzo River, Sassandra River basin (Gosse, 1986b).

***Synodontis punctulata* Günther, 1889**

Synodontis punctulatus Günther, 1889: 71, pl. 8 (fig. a). Type locality: River Ruva, in the Arusha country. Lectotype: BMNH 1887.11.3.47; designated by, and illustrated in, Poll (1971: 233, fig. 108).

Distribution: Ruwa River, Tanzania; Uebi Shebeli, Somalia (Gosse, 1986b).

Remarks: Considered to be a possible synonym of *Synodontis zanzibaricus* Peters, 1868 (De Vos 2001a: 49).

***Synodontis rebeli* Holly, 1926**

Synodontis rebeli Holly, 1926: 157. Illustrated in Holly (1927b: 215, figs. 5–6). Type locality: Bamfluß, Flußgebiet des Sanaga in Kamerun. Lectotype: NMW 7796: 1; designated by, and illustrated in, Poll (1971: 155, fig. 70).

Distribution: Sanaga River basin, Cameroon (Gosse, 1986b).

***Synodontis resupinata* Boulenger, 1904**

Synodontis resupinatus Boulenger, 1904c: 199, pl. 8. Type locality: Lokoja, Northern Nigeria. Holotype: BMNH 1904.1.20.52.

Distribution: Niger River basin (Gosse, 1986b).

***Synodontis ricardoae* Seegers, 1996**

Synodontis ricardoae Seegers, 1996: 245, figs. 178–180. Type locality: Kazizi, some kilometers northeast of Muze, at the northwestern shore of Lake Rukwa, Rukwa region, western Tanzania (07°37'N, 31°36'E). Holotype: MRAC 94-34-P-1093.

Distribution: Lake Rukwa basin (Seegers, 1996).

***Synodontis robbianus* Smith, 1875**

Synodontis robbianus Smith, 1875: 92, fig. 1. Type locality: Ikorofiong, Old Calabar River. Syntype: BMNH 1874.5.23.3 (1); illustrated in Poll (1971: fig. 26) as holotype.

Distribution: Lower Niger River and Cross River (Gosse, 1986b).

***Synodontis robertsi* Poll, 1974**

Synodontis robertsi Poll, 1974: 442, fig. 1. Type locality: Riv. Lukenie à Elombe, ferry landing, lat. 2°49'S, long. 18°14'E. Holotype: MRAC 189535.

Distribution: Lukenie River, Congo River basin; known only from type locality (Gosse, 1986b).

***Synodontis ruandae* Matthes, 1959**

Synodontis ruandae Matthes, 1959a: 62. Type locality: Riv. Kagera à Rusumu, Ruanda. Holotype: MRAC 130345; holotype illustrated in Poll (1971: 178, fig. 82).

Distribution: Kagera River basin, Nile-Victoria system (Gosse, 1986b).

***Synodontis rufigiensis* Bailey, 1968**

Synodontis rufigiensis Bailey, 1968: 346, fig. 1. Type locality: Lake Lugongwe, at Utete on the lower Rufiji, Tanzania. Holotype: BMNH 1968.6.12.1; holotype illustrated in Poll (1971: 428, fig. 200).

Distribution: Rufiji River basin, Tanzania (Gosse, 1986b).

***Synodontis rukwaensis* Hilgendorf & Pappenheim, 1903**

Synodontis zambezensis rukwaensis Hilgendorf & Pappenheim, 1903: 267. Type locality: Rukwa Sees. Lectotype: ZMB 16311; designated by, and illustrated in, Seegers (1996: 249, fig. 182).

Synodontis maculipinna Norman, 1922: 687. Type locality: Mpanganye, Rufiji River, Tanganyika Territory. Lectotype: BMNH 1922.4.18.27, designated by, and illustrated in, Poll (1971: 376, fig. 174); lectotype also illustrated in Seegers (1996: fig. 184).

Synodontis wamiensis Lohberger, 1930: 91, fig. 1. Type locality: aus dem Wamiflusse und Nebenflüssen [30 km. north of Bahamojo, Tanzania]. Holotype: NMW 18866; illustrated in Seegers (1996: fig. 185).

Distribution: Lake Rukwa basin, Ugalla basin Malagarasi system, and coastal drainages of Tangazni south of the Pangani River (Seegers, 1996).

Remarks: Synonymy based on Seegers (1996).

***Synodontis schall* (Bloch & Schneider, 1801)**

Silurus schall Bloch & Schneider, 1801: 385. Type locality: Habitat Nilum. Holotype: ZMB 3110 (lost; Paepke, 1999: 99).

Pimelodus clarias Geoffroy Saint-Hilaire, 1809: pl. 13 (figs. 3–4). Type locality: Nil, Egypt. Holotype: Possibly

MNHN 0000-4203 (1) or MNHN 0000-4205 (1 of 2). Name available from plate, with illustrated specimen the holotype, if identifiable. Described as *Synodontis clarias* in Geoffroy Saint-Hilaire (1827: 299). Independence of this name and *Silurus clarias* Linnaeus not clearly established, but treated as independent in literature (e. g., Poll, 1971).

Synodontis maculosus Rüppell, 1829: 10, pl. 3 (fig. 1). Type locality: die Märkte von Cairo. Lectotype: SMF 653, designated by Poll (1971: 50).

Synodontis arabi Valenciennes, in Cuvier & Valenciennes, 1840b: 261 (194 in Strasbourg deluxe edition). Type locality: Nil, Egypte. Syntypes: MNHN 0000-4203 (1), MNHN 0000-4205 (2). Unneeded new name for *Silurus schall* Bloch & Schneider, 1801.

Synodontis smithii Günther, 1896b: 222, pl. 9. Type locality: [Not stated; introduction indicates specimen obtained "en route to Lake Rudolph"; BMNH register lists Lake Stephanie [sic, Lac Stefanie, Kenya] and Poll (1971) listed Lake Rudolph, but these are in contradiction to statements in introduction]. Holotype: BMNH 1895.12.31.68.

Distribution: Nile basin, Abaia, Stephanie and Rudolf Lakes, Chad, Niger, Senegal, Uebi Guiba (Uebi Shebeli) basins and possibly Tana and Volta rivers (Gosse, 1986b).

***Synodontis schoutedeni* David, 1936**

Synodontis schoutedeni David, 1936b: 416. Type locality: Kungunu; Koteli; Basongo; Mongende; Budjala; Bumba; Flandria [restricted to Kungunu by lectotype designation]. Lectotype: MRAC 38093; designated by, and illustrated in, Poll (1971: 393, fig. 184).

Distribution: Central Congo River basin (Gosse, 1986b).

***Synodontis serpentis* Whitehead, 1962**

Synodontis serpentis Whitehead, 1962b: 100, figs. 1a, 1b. Type locality: Athi River at Jilore, thirty miles from river mouth, Kenya. Holotype: BMNH 1962.1.19.1.

Distribution: Athi and Tana River basins, Kenya (Gosse, 1986b).

***Synodontis serrata* Rüppell, 1829**

Synodontis serratus Rüppell, 1829: 8, pl. 2 (fig. 1). Type locality: Cairo. Holotype: SMF 2649 (dry).

Synodontis humeratus Valenciennes, in Cuvier & Valenciennes, 1840b: 264 (196 in Strasbourg deluxe edition). Type locality: Nil. No types known; based on a drawing.

Distribution: Nile basin (Gosse, 1986b).

***Synodontis smiti* Boulenger, 1902**

Synodontis smiti Boulenger, 1902d: 45, pl. 12 (fig. 3). Type locality: l'Ubangi à Banzyville. Lectotype: MRAC 1266, designated in Poll (1971: 316).

Synodontis tenuis Nichols & Griscom, 1917: 717, fig. 25. Type locality: Stanleyville, Congo. Holotype: AMNH 6535.

Distribution: Ubangi and Lualaba Rivers, Congo River basin (Gosse, 1986b).

***Synodontis soloni* Boulenger, 1899**

Synodontis soloni Boulenger, 1899b: 110, pl. 43 (fig. 1). Type locality: Congo. Lectotype: MRAC 155385, designated by Poll (1971: 296).

Distribution: Stanley Pool rapids and Libenge, Congo River basin (Gosse, 1986b).

***Synodontis sorex* Günther, 1864**

Synodontis sorex Günther, 1864: 211. Type locality: Chartoum, upper Nile. Lectotype: BMNH 1862.6.17.69, designated by Poll (1971: 65).

Distribution: Nile, Chad, Niger and Volta basins (Gosse, 1986b).

Remarks: Lectotype register number incorrectly listed as BMNH 1862.1.17.69 in Poll (1971) and Eschmeyer *et al.* (1998).

***Synodontis steindachneri* Boulenger, 1913**

Synodontis steindachneri Boulenger, 1913: 69. Type locality: Nyong River, S. Cameroon. Holotype: BMNH 1913.10.29.16; holotype illustrated in Boulenger (1916: fig. 188) and Poll (1971: fig. 162).

Distribution: Nyong River, Cameroon (Gosse, 1986b).

***Synodontis tanganaicae* Borodin, 1936**

Synodontis serratus tanganaicae Borodin, 1936: 9. Type locality: Kasanga, Lake Tanganyika. Lectotype: MCZ 32538; designated by De Vos & Thys van den Audenaerde (1998: 148).

Synodontis lacustricolus Poll, 1953: 157, fig. 18a; pl. 6 (fig. 3). Type locality: Stat. 123, au large de Karéma [Lac Tanganyika]. Holotype: IRSNB 197.

Distribution: Lake Tanganyika (De Vos & Thys van den Audenaerde, 1998: 153).

Remarks: Synonymy first proposed in De Vos & Thys van den Audenaerde (1998: 153), with valid name as *Synodontis tanganyicae*.

***Synodontis tessmanni* Pappenheim, 1911**

Synodontis tessmanni Pappenheim, 1911: 523, fig. 5. Type locality: Akonangi, im Kje [Ntem basin]. Holotype (12.8 cm SL): ZMB 18413; illustrated in Poll (1971: fig. 62), as lectotype, but a holotype was clearly stated (as “Type der Art”) in description.

Synodontis hollyi ntemensis Pellegrin, 1929b: 367. Type locality: Nyabessan (Ntem), Cameroun. Holotype: MNHN 1929-0051. Originally as *Synodontis Hollyi* var. *ntemensis* nov. var.

Distribution: Ntem River, Cameroon (Gosse, 1986b).

***Synodontis thamalakanensis* Fowler, 1935**

Synodontis thamalakanensis Fowler, 1935a: 274, fig. 12. Type locality: Thamalakane River, Maun, Bechuanaland Protectorate. Holotype: ANSP 53245.

Distribution: Okavango and Upper Zambezi River basins (Skelton, 1993).

***Synodontis thysi* Poll, 1971**

Synodontis thysi Poll, 1971: 210, fig. 100; pls. 3 (fig. 16), 10 (fig. 6). Type locality: Mange-Ferry, riv. Little Scarcies ou Kaba (Sierra Leone). Holotype: MRAC 168314.

Distribution: Little Scarcies River, Sierra Leone; known only from type locality (Gosse, 1986b).

***Synodontis tourei* Daget, 1962**

Synodontis tourei Daget, 1962: 113, fig. 38; pl. 11 (figs. 24–25). Type locality: Bafing à Ballay, Guinée. Lectotype: MNHN 1959-0109; designated by Poll (1971: 185).

Distribution: Bafing River, Senegal River basin, Guinea (Gosse, 1986b).

***Synodontis unicolor* Boulenger, 1915**

Synodontis unicolor Boulenger, 1915: 170. Type locality: Lac Moero et rivière Luapula, à Kasenga [restricted to Kasenga by lectotype designation]. Lectotype: MRAC 14223; designated by, and illustrated in, Poll (1971: 359, fig. 166).

Distribution: Luapula-Moero system, Congo River basin (Gosse, 1986b).

***Synodontis vanderwaali* Skelton & White, 1990**

Synodontis vanderwaali Skelton & White, 1990: 284, fig. 5. Type locality: Okavango River, Namibia. Holotype: AMG P5796.6.

Distribution: Cunene, Okavango and upper Zambezi basins (Skelton, 1993).

***Synodontis velifer* Norman, 1935**

Synodontis velifer Norman, 1935a: 219, fig. 3. Type locality: Ejura, Ashanti Forest, Gold Coast. Holotype: BMNH 1934.8.31.146; holotype illustrated in Poll (1971: 164, fig. 74).

Distribution: Sassandra, Bandama and Volta River basins (Gosse, 1986b).

***Synodontis vermiculata* Daget, 1954**

Synodontis vermiculatus Daget, 1954: 292, fig. 109. Type locality: Mopti, riv. Niger (Mali). Neotype: MNHN 1961-1129; designated by Poll (1971: 170).

Distribution: Niger River basin (Gosse, 1986b).

***Synodontis victoriae* Boulenger, 1906**

Synodontis victoriae Boulenger, 1906b: 438. Type locality: Entebbe and Buganga, Lake Victoria [restricted to Buganga by lectotype designation]. Lectotype: BMNH 1906.5.30.190; designated by, and illustrated in, Poll (1971: 120, fig. 50).

Distribution: Victoria Lake basin, Kioga Lake and Victoria Nile, Malagarasi River (Gosse, 1986b).

***Synodontis violacea* Pellegrin, 1919**

Synodontis violaceus Pellegrin, 1919b: 212. Type locality: Gribingui. Lectotype: MNHN 1919-0246, designated by,

and illustrated in, Poll (1971: 149, fig. 66).

Distribution: Chad, Niger and Volta basins (Gosse, 1986b).

***Synodontis voltae* Roman, 1975**

Synodontis voltae Roman, 1975: 45, unnumbered figure. Type locality: dans la Bougouriba, affluent de la Volta Noire. Holotype: MHV V5641.

Distribution: Upper Volta River basin; known only from type locality (Gosse, 1986b).

***Synodontis waterloti* Daget, 1962**

Synodontis waterloti Daget, 1962: 111, fig. 37; pl. 10 (fig. 23). Type locality: Friguiagbé, près Kindia, Guinée. Holotype: MNHN 1935-0223.

Distribution: Waanje and Taja Rivers, Sierra Leone; St. John River basin, Liberia; Cess River basin, Côte d'Ivoire; Pra River basin, Ghana (Gosse, 1986b).

***Synodontis woosnami* Boulenger, 1911**

Synodontis woosnami Boulenger, 1911a: 424, fig. 319. Type locality: Okovango [Botswana]. Holotype: BMNH 1910.5.31.36.

Distribution: Cunene, Okavango and upper Zambezi River basins (Skelton, 1993).

***Synodontis xiphias* Günther, 1864**

Synodontis xiphias Günther, 1864: 215. Type locality: West Africa. Holotype: BMNH 1863.11.9.1 (dry); holotype illustrated in Boulenger (1911a: fig. 339) and Poll (1971: fig. 22).

Synodontis labeo Günther, 1865a: 453. Type locality: West Africa, probably Niger R. Holotype: BMNH 1865.5.9.8 (stuffed); illustrated in Boulenger (1911a: fig. 337).

Distribution: Niger River basin (Gosse, 1986b).

Remarks: *Synodontis labeo* Günther, 1865, is treated by Willoughby (1994: 28) as probably valid.

***Synodontis zambezensis* Peters, 1852**

Synodontis zambezensis Peters, 1852: 682. Type locality: riv. Zambèze, Mozambique. Lectotype: ZMB 3119; designated by, and illustrated in, Poll (1971: 225, fig. 104). Illustrated in Peters (1868c: 31, pl. 5, figs. 2–3).

Synodontis zambesensis Günther, 1864: 214. Type locality: Zambezi. Lectotype: ZMB 3119, designated by, and illustrated in, Poll (1971: 225, fig. 104). Unjustified emendation of *Synodontis zambezensis* Peters, 1852.

Distribution: Pongolo, Lompopo, Pungwe and Zambezi River basins, Rukwa Lake, Luangwe River of Moero basin. (Gosse, 1986b).

***Synodontis zanzibarica* Peters, 1868**

Synodontis zanzibaricus Peters, 1868b: 600. Type locality: Wahrscheinlich von Mombas [Kenya]. Lectotype: ZMB 6846; designated by, and illustrated in, Poll (1971: 229, fig. 106).

Distribution: Mombassa, Kenya (Gosse, 1986b).

Species inquirendae, Synodontis

Synodontis vaillanti Boulenger, 1897b: 424. Type locality: Congo français (Bangui). Holotype: MNHN 1892-0275. Treated by Gosse (1986:148) as doubtfully valid.

Synodontis Hollyi Pellegrin, 1929b: 365. Type locality: Ndjibula ..., Nanga Eboko (Sanaga), Cameroun. Syntypes: MNHN 1929-0047 (1), MNHN 1929-0048 (1), MNHN 1929-0049 (1), MNHN 1929-0050 (1). Type series apparently mixed; with specimens representing both *Synodontis obesus* Boulenger, 1898, and *Synodontis rebeli* Holly, 1926.

Synodontis galinae Kochetov, 1998: 4, 2 figs. Type locality: White Nile. Type(s): Whereabouts unknown.

NEMATOGENYIIDAE Bleeker, 1862

Nematogenyini Bleeker, 1862 (in Bleeker, 1862–63): 16. Type genus: *Nematogenys* Girard, 1855.

Taxonomic summary: de Pinna (2003).

1 genus, 2 species; including 1 named fossil species.

NEMATOGENYS Girard, 1855

Nematogenys Girard, 1855: 198. Type species: *Trichomycterus inermis* Guichenot, 1848. Type by monotypy. Gender: Feminine.

Phylogeny: Arratia & Chang (1975).

† *Nematogenys cuivi* Azpelicueta & Rubilar, 1998

† *Nematogenys cuivi* Azpelicueta & Rubilar, 1998: 476, figs. 2, 3. Type locality: Cura-Mallín Formation, Miocene, Northern slope of the Cerro Rucañanco ($38^{\circ}41'S$, $71^{\circ}11'W$), about 1,250 m a.s.l., Lonquimay, Chile. Holotype: CPUC Lonq (R)/7 (neurocranium with Weberian complex, branchial and hyoid arches, pectoral girdle and spine, some vertebrae, and ribs).

Distribution: Cura-Mallin Formation, south-central Chile (Azpelicueta & Rubilar, 1998).

Nematogenys inermis (Guichenot, 1848)

Trichomycterus inermis Guichenot, 1848: 312, pl. 9 (fig. 2). Type locality: Chile. No types known.

Nematogenys nigricans Philippi, 1866: 716. Type locality: Chile. No types known.

Nematogenys pallidus Philippi, 1866: 716. Type locality: Chile. No types known.

Distribution: Isolated localities in Concepción, Rancagua and Angol, Chile. Formerly widespread throughout much of Central Chile (de Pinna, 2003).

PANGASIIDAE Bleeker, 1858

Pangasini Bleeker, 1858b: 49, 63. Type genus: *Pangasius* Valenciennes, 1840.

Pangasianodonidi Fowler, 1958: 14. Type genus: *Pangasianodon* Chevey, 1931.

Revision: Roberts & Vidthayanon (1991).

Review: Vidthayanon & Roongthongbaisuree (1993, Thailand).

Identification guide: Jayaram (1977d, South Asia).

Phylogeny: Pouyaud *et al.* (2000); Pouyaud *et al.* (2004).

Remarks: Several phylogenetic studies indicate that the species included in this family may constitute a natural group, but the group is likely nested within the Schilbidae. As such, the Pangasiidae may not deserve continued recognition as a family but is so recognized herein due to current usage. See Fumihito (1989) for morphological comparisons among some species.

5 genera, 30 species; including 1 named fossil genus and 2 fossil species.

† *CETOPANGASIUS* Roberts & Jumnongthai, 1999

† *Cetopangasius* Roberts & Jumnongthai, 1999: 177. Type species: †*Cetopangasius chaetobranchus* Roberts & Jumnongthai, 1999. Type by original designation. Gender: Masculine. Also spelled *Cetopagansius* in text. *Cetopangasius* treated here as the valid spelling.

† *Cetopangasius chaetobranchus* Roberts & Jumnongthai, 1999

† *Cetopangasius chaetobranchus* Roberts & Jumnongthai, 1999: 177, figs. 16–21. Type locality: Thailand, North-central Phetchabun Province, Ban Nong Pla; Miocene. Holotype: Department of Natural Resources, Thailand (DNR) TF 5013 (entire skeleton).

Distribution: Thailand; Miocene (Roberts & Jumnongthai, 1999).

HELICOPHAGUS Bleeker, 1858

Helicophagus Bleeker, 1858b: 28, 45. Type species: *Helicophagus typus* Bleeker, 1858. Type by original designation. Gender: Masculine.

Helicophagus leptorhynchus Ng & Kottelat, 2000

Helicophagus leptorhynchus Ng & Kottelat, 2000c: 55, figs. 1, 2a. Type locality: Thailand Ubon Ratchathani Prov-inve, Mun River at Bung Wai, about 7 km W of Ubon Ratchathani ($15^{\circ}12'30"N$, $104^{\circ}47'30"E$). Holotype: USNM 288676.

Distribution: Mekong and Chao Phraya River basins (Kottelat, 2001b).

Helicophagus typus Bleeker, 1858

Helicophagus typus Bleeker, 1858a: 46. Type locality: Palembang, in fluvimine Mussi. Holotype: BMNH 1863.12.4.118; illustrated in Bleeker (1862–63: pl. 79 [= Silur. pl. 31], fig. 2).

Distribution: Sumatra and southeastern Borneo (Roberts & Vidthayanon, 1991; Musikasinthorn *et al.*, 1998).

Helicophagus waandersii Bleeker, 1858

Helicophagus Waandersii Bleeker, 1858b: 175. Type locality: Palembang, in fluviis [Sumatra]. Holotype (340 mm TL): BMNH 1863.12.4.89; illustrated in Bleeker (1862–63: pl. 80 [= Silur. pl. 32]).

Distribution: Mekong and Chao Phraya Rivers, and Sumatra (Roberts & Vidthayanon, 1991).

PANGASIANODON Chevey, 1931

Pangasianodon Chevey, 1931: 538. Type species: *Pangasianodon gigas* Chevey, 1931. Type by monotypy. Gender: Masculine.

Remarks: Treatment of this genus as valid follows Rainboth (1996).

Pangasianodon gigas Chevey, 1931

Pangasianodon gigas Chevey, 1931: 538, figs. 1–2, pl. 1. Type locality: Cambodge. No types known (cast of type in Musée Economique de Phnom-Pehn).

Pangasius paucidens Fang & Chaux, in Chaux & Fang, 1949b: 344, fig. 6. Type locality: Cambodge. Holotype: MNHN 1966-0730.

Distribution: Mekong River basin; stocked in various reservoirs in Thailand (Kottelat, 2001b).

Pangasianodon hypophthalmus (Sauvage, 1878)

Helicophagus hypophthalmus Sauvage, 1878: 235. Type locality: Laos. Lectotype: MNHN a-0745, designated by Kottelat (1984b: 812).

Pangasius sutchi Fowler, 1937: 141, figs. 27–29. Type locality: Bangkok, Siam. Holotype: ANSP 67902.

Distribution: Mekong and Chao Phraya River basin (Kottelat, 2001b); in cultivation elsewhere in Asia, including Viet Nam and Myanmar.

PANGASIUS Valenciennes, 1840

Pangasius Valenciennes, in Cuvier & Valenciennes, 1840b: 45 (34 of Strasbourg deluxe edition). Type species: *Pangasius buchanani* Valenciennes, 1840 (= *Pimelodus pangasius* Hamilton, 1822). Type by monotypy. Gender: Masculine.

Pseudopangasius Bleeker, 1862c: 399. Type species: *Pangasius polyuranodon* Bleeker, 1852. Type by original designation. Gender: Masculine. Also in Bleeker (1862–63: 14, 75).

Neopangasius Popta, 1904: 180. Type species: *Neopangasius nieuwenhuisii* Popta, 1904. Type by monotypy. Gender: Masculine.

Sinopangasius Chang & Wu, 1965: 11, 13. Type species: *Sinopangasius semicultratus* Chang & Wu, 1965. Type by original designation. Gender: Masculine.

Pangasius bocourti Sauvage, 1880

Pangasius (Pseudopangasius) Bocourti Sauvage, 1880b: 229. Type locality: Phnom-Penh [Cambodia]. Holotype: MNHN 0000-9528.

Pangasius altifrons Durand, 1940: 23, pl. 5. Type locality: Tonlé-Sap [Cambodia]. Holotype: at Institut Océanographique Nhatrang.

Distribution: Mekong and Chao Phraya River basins (Roberts & Vidthayanon, 1991; Kottelat, 2001b).

Pangasius conchophilus Roberts & Vidthayanon, 1991

Pangasius conchophilus Roberts & Vidthayanon, 1991: 114, figs. 1b, 2j, 5. Type locality: Thabo, Nongkhai prov. [Thailand]. Holotype: NIFI 2227.

Distribution: Mekong, Bangpakong, and Chao Phraya River basins (Roberts & Vidthayanon, 1991; Kottelat, 2001b).

Pangasius djambal Bleeker, 1846

Pangasius djambal Bleeker, 1846b: 290. Type locality: Batavia, in fluviis. Possible syntypes: BMNH 1863.12.4.81 (1, 400+ mm TL), RMNH 6854 (1), RMNH 31192 (6), RMNH 8069 (1).

Pangasius bedado Roberts, 1999: 110, figs. 1–3. Type locality: Palembang, Sumatra. Holotype: MZB 2598.

Distribution: Java and Borneo (Roberts & Vidthayanon, 1991), Sumatra (Roberts, 1999).

Remarks: Roberts & Vidthayanon (1991:116) designated a neotype (RMNH 6854) for *Pangasius djambal* although several specimens from Bleeker's collections, some of which may be syntypes, were reported on therein. The "neotype" was incorrectly listed as a lectotype by Eschmeyer *et al.* (1998). Synonymy follows Gustiano *et al.* (2004). Register number of syntype of *Pangasius djambal* incorrectly reported as BMNH 1863.12.11.81 in Eschmeyer *et al.* (1998).

***Pangasius elongatus* Pouyaud, Gustiano & Teugels, 2002**

Pangasius elongatus Pouyaud, Gustiano & Teugels, 2002: 248, fig. 5. Type locality: Vietnam: Lower Mekong River Delta. Holotype: MZB 10890.

Distribution: Lower reaches of the Mekong, Chao Phraya, and Bangpakong Rivers (Pouyaud *et al.*, 2002).

***Pangasius humeralis* Roberts, 1989**

Pangasius humeralis Roberts, 1989a: 131, figs. 100, 101c. Type locality: Kapuas basin, fish market at Sintang, Western Borneo (Kalimantan Barat, Indonesia). Holotype: MZB 3680.

Distribution: Kapuas River basin, western Borneo (Roberts, 1989a; Roberts & Vidthayanon, 1991).

† *Pangasius indicus* (Marck, 1876)

† *Brachyspondylus indicus* Marck, 1876a: 412, pl. 24 (fig. 2). Type locality: Padang, Sumatra; Tertiary. Holotype: at Royal Geol. Mus. Dresden (imperfect fish). Also in Marck (1876b), priority of publication not established.

† *Pseudeutropius verbeekii* Günther, 1876: 435, pl. 15 (fig. 2). Type locality: Highlands of Padang, Sumatra; Tertiary. Syntypes (2): BMNH 47527 (postcranial fish).

Distribution: Padang, Sumatra; Tertiary.

Remarks: Gunther (1876: 436) noted that his specimens were likely conspecific with that of Marck, but considered Marck's name inaccurately reflected the geographic distribution and was, therefore, inappropriate. Woodward (1901: 326) lists † *Brachyspondylus indicus*, as a junior synonym of † *Pseudeutropius verbeekii*, which was followed in the literature. Also see Sanders (1934) for summary of the taxonomic history of these names. Roberts & Jumnongthai (1999) treated this as a species of *Pangasius*, apparently following Sanders (1934: 22). Patterson (1993) expressed skepticism of the reported Eocene age of these fossils.

***Pangasius kinabatanganensis* Roberts & Vidthayanon, 1991**

Pangasius kinabatanganensis Roberts & Vidthayanon, 1991: 123, figs. 2P, 10. Type locality: Kinabatangan River at Deramakot [Borneo, Malaysia]. Holotype: FMNH 68042.

Distribution: Kinabatangan River basin, northeastern Borneo (Roberts & Vidthayanon, 1991).

***Pangasius krempfi* Fang & Chaux, 1949**

Pangasius krempfi Fang & Chaux, in Chaux & Fang, 1949b: 343, fig. 5. Type locality: en mer à Bong-Lao [Vietnam]. Holotype: MNHN 1966-0729.

Sinopangasius semicultratus Chang & Wu, 1965: 11, 13, figs. 1–4. Type locality: Off Paihai, Kwangtung, China. Holotype: ASIZB 56-1174.

Distribution: Mekong and Hue River basins, and along coast of South China Sea of Vietnam and Guandong, China (Kottelat, 2001b).

***Pangasius kunyit* Pouyaud, Teugels & Legendre, 1999**

Pangasius kunyit Pouyaud, Teugels & Legendre, 1999: 251, fig. 2. Type locality: Sangasanga Village, 30 km south-east of the town of Samarinda, delta of Mahakam River, East Kalimantan, Indonesia. Holotype: MZB 10009.

Distribution: Mahakam River delta, Borneo, Indonesia (Pouyaud *et al.*, 1999).

***Pangasius larnaudii* Bocourt, 1866**

Pangasius Larnaudii Bocourt, 1866: 15, pl. 1 (figs. 2, 2a). Type locality: Ajuthia. Syntypes: MNHN 0000-1549 (1), MNHN a-9423 (1).

Pangasius taeniura Fowler, 1935b: 98, fig. 19. Type locality: Bangkok, Siam. Holotype: ANSP 61753.

Pangasius burgini Fowler, 1937: 141, figs. 24–26. Type locality: Bangkok, Siam. Holotype: ANSP 67901.

Distribution: Mekong, Meklong and Chao Phraya River basins (Roberts & Vidthayanon, 1991; Kottelat, 2001b).

***Pangasius lithostoma* Roberts, 1989**

Pangasius lithostoma Roberts, 1989a: 132, fig. 102. Type locality: Market at Sintang, Western Borneo (Kalimantan Barat, Indonesia). Holotype: MZB 3678.

Distribution: Kapuas River basin, western Borneo (Roberts, 1989a; Roberts & Vidthayanon, 1991).

***Pangasius macronema* Bleeker, 1851**

Pangasius macronema Bleeker, 1851a: 11. Type locality: Banjermassing, in fluviis. Holotype (116 mm): possibly NMV 45892 (1), RMNH 6855 (10), BMNH 1863.12.4.66 (1).

Pangasius siamensis Steindachner, 1878c: 393. Type locality: Menam-Fluss bei Bangkok. Syntypes: NMW 45469 (1), NMW 76998 (1).

? *Pangasius aequilabialis* Fowler, 1937: 140, figs. 20–23. Type locality: Bangkok, Siam. Holotype: ANSP 67897.

Distribution: Mekong and Chao Phraya River basins, Java, and Borneo (Kottelat, 2001b).

Remarks: BMNH lists BMNH 1863.12.4.66 (1, 144 mm SL) as a type of *Pangasius macronema* but is too large to be the holotype.

***Pangasius mahakamensis* Pouyaud, Gustiano & Teugels, 2002**

Pangasius mahakamensis Pouyaud, Gustiano & Teugels, 2002: 246, fig. 4. Type locality: Indonesia, East Kalimantan Province, Mahakam River at Samarinda. Holotype: MZB 10886.

Distribution: Asia: Mahakam River of eastern Kalimantan, Indonesia (Pouyaud *et al.*, 2002).

***Pangasius mekongensis* Gustiano, Teugels & Pouyaud, 2003**

Pangasius mekongensis Gustiano, Teugels & Pouyaud, 2003: 370, fig. 7. Type locality: [Lower Mekong River, Vietnam]. Holotype: MZB 10847.

Distribution: Mekong River, Vietnam (Gustiano, Teugels & Pouyaud, 2003).

***Pangasius myanmar* Roberts & Vidthayanon, 1991**

Pangasius myanmar Roberts & Vidthayanon, 1991: 131, fig. 17. Type locality: Rangoon, [Myanmar]. Holotype: SU 33787.

Distribution: Myanmar, known only from types (Roberts & Vidthayanon, 1991).

***Pangasius nasutus* (Bleeker, 1863)**

Pseudopangasius nasutus Bleeker, 1863f: 72. Type locality: Bandjermassin, in fluviis. Holotype (270 mm TL): possibly BMNH 1863.12.4.113 (207 mm SL, ~255 mm TL). Also described as new in Bleeker (1863e).

Pangasius ponderosus Myers, in Herre & Myers, 1937: 67, pl. 6. Type locality: Chandra Dam, Perak, Malay Peninsula. Holotype: SU 14162.

Distribution: Sumatra, Borneo and Malay Peninsula (Roberts & Vidthayanon, 1991).

***Pangasius nieuwenhuisii* (Popa, 1904)**

Neopangasius Nieuwenhuisii Popa, 1904: 180. Type locality: le Bo, Bornéo central. Holotype: RMNH 7546; described in more detail, with illustration of holotype, in Popa (1906: 30, pl. 1, figs. 3a, 3b).

Distribution: Mahakam River basin, eastern Borneo (Roberts & Vidthayanon, 1991).

***Pangasius pangasius* (Hamilton, 1822)**

Pimelodus pangasius Hamilton, 1822: 163, 376, pl. 33 (fig. 52). Type locality: Estuaries of Bengal. No types known.

Pachypterus luridus Swainson 1839: 306. No types known. Made available by reference to “Ham. p. 163, f. 62 [sic, 52]” [= Hamilton (1822, 163, pl. 33, fig. 52)]. Unneeded new name for *Pimelodus pangasius* Hamilton, 1822.

Pangasius Buchanani Valenciennes, in Cuvier & Valenciennes, 1840b: 45 (34 of Strasbourg deluxe edition), pl. 425.

Type locality: Estuaries of Bengal. No types known. Unneeded new name for *Pimelodus pangasius* Hamilton, 1822, apparently to avoid tautonymy.

Pangasius pangasius godavarii David, 1962: 151, fig. 3g. Type locality: Godavary River, at Rajahmundry [India]. Syntypes: at Central Inland Fisheries Research Institute’s Tank Fisheries Unit, Bangalore, India.

Pangasius pangasius upiensis Srivastava, 1968: 97, fig. 60. Type locality: Bale-ka-Maidan, R. Rohini, Gorakhpur, Uttar Pradesh, India. Holotype: at Zool. Mus. Gorakhpur Univ., India.

Distribution: Rivers and estuaries of Indian subcontinent (Roberts & Vidthayanon, 1991).

Remarks: Redescribed in Hora (1938b).

***Pangasius polyuranodon* Bleeker, 1852**

Pangasius polyuranodon Bleeker, 1852b: 425. Type locality: Bandjermassing, in fluviis. Holotype (160 mm TL): possibly RMNH 6855.

Pangasius juaro Bleeker, 1852d: 589. Type locality: Palembang, in fluviis. Holotype (336 mm TL): possibly BMNH 1863.12.4.79 (1) or RMNH 6855 (1 of larger specimens).

Distribution: Rivers of Sumatra and northern, western and southern Borneo (Pouyaud *et al.*, 2002).

***Pangasius rheophilus* Pouyaud & Teugels, 2000**

Pangasius rheophilus Pouyaud & Teugels, 2000: 194, figs. 1–2. Type locality: Bahau River, tributary of upper Kayan at Longpujungan, Bulungan Regency, Kalimantan Timur, Indonesia. Holotype: MZB 10010.

Distribution: Bahau River, tributary of upper Kayan at Longpujungan, Kalimantan Timur, Indonesia (Pouyaud & Teugels, 2000).

***Pangasius sabahensis* Gustiano, Teugels & Pouyaud, 2003**

Pangasius sabahensis Gustiano, Teugels & Pouyaud, 2003: 372, fig. 8. Type locality: [Kinabatangan River basin, North Borneo, Sabah State, Malaysia]. Holotype: MZB 10847.

Distribution: Kinabatangan River basin, North Borneo, Sabah State, Malaysia (Gustiano *et al.*, 2003).

***Pangasius sanitwongsei* Smith, 1931**

Pangasius sanitwongsei Smith, 1931: 29, figs. 13–14. Type locality: Menam Chao Phya at Koh Yai, Central Siam. Holotype: Siam Department of Fisheries (apparently lost).

Pangasius beani Smith, 1931: 26. Type locality: Klong Ban Poh, off Lopburi, near Ayuthia, Central Siam. Holotype: USNM 90308.

Distribution: Mekong and Chao Phraya River basins (Roberts & Vidthayanon, 1991; Kottelat, 2001b).

Remarks: Roberts & Vidthayanon (1991: 138) acting as first reviser, selected *Pangasius sanitwongsei* as valid.

Species inquirendae, Pangasius

Pangasius delicatissimus Bleeker, 1863 (in Bleeker, 1862–63): 73, footnote. Type locality: Krawang, Java. No types known, based on a drawing.

Pangasius hoeksi Hardenberg, 1948: 412. Type locality: Kapuas River (W. Borneo). Holotype (275 mm): whereabouts unknown.

***PSEUDOLAIS* Vaillant, 1902**

Pseudolais Vaillant, 1902: 51. Type species: *Pseudolais tetraneuma* Vaillant, 1902. Type by monotypy. Gender: Masculine.

Pteropangasius Fowler, 1937: 142. Type species: *Pangasius cultratus* Smith, 1931. Type by original designation. Gender: Masculine.

***Pseudolais micronemus* (Bleeker, 1847)**

Pangasius micronemus Bleeker, 1847b: 8. Type locality: Java. Syntypes: Possibly BMNH 1863.12.4.82 (1, 250 mm SL), RMNH 6856 (3). Also in Bleeker (1847: 166).

Pangasius rios Bleeker, 1851d: 205. Type locality: Bandjermassing, in fluviis. Holotype (115 mm TL): possibly BMNH 1863.12.4.92 (~ 104 mm SL).

Pseudolais tetraneuma Vaillant, 1902: 52, fig. 3. Type locality: Tepoe, bords du Mahakam, Bornéo central. Holotype: Possibly RMNH 7821.

Pangasius de Zwaani Weber & de Beaufort, 1912: 535, pl. 12 (fig. 3). Type locality: Taluk, Sumatra. Holotype: ZMA 113011; illustration of holotype reproduced in Weber & de Beaufort (1913: fig. 103).

Pangasius tubbi Inger & Chin, 1959: 287, fig. 47. Type locality: Confluence of the Deramakot River with the Kinabatangan River, Kinabatangan District, North Borneo. Holotype: FMNH 68047.

Distribution: Mekong and Hue Rivers, Malay Peninsula, Sumatra, Java, and Borneo (Roberts & Vidthayanon, 1991).

***Pseudolais pleurotaenia* (Sauvage, 1878)**

Pangasius pleurotaenia Sauvage, 1878: 235. Type locality: Laos. Lectotype: MNHN 0000-9529 designated by, and illustrated in, Kottelat (1984b: 813, fig. 8a). One of two syntypes illustrated in Sauvage (1881: 169, pl. 8, fig. 6).

Pangasius cultratus Smith, 1931: 25. Type locality: Tapi River near Bandon, peninsular Siam. Holotype: USNM 90306.

Pangasius fowleri Smith, 1931: 28. Type locality: Lopburi River at Lopburi, Central Siam. Holotype: USNM 90309. Distribution: Mekong, Meklong, Tapi and Chao Phraya River basins (Roberts & Vidthayanon, 1991; Kottelat,

2001b).

PIMELODIDAE

Pimelodini Bonaparte, 1838: 131. Type genus: *Pimelodus* La Cepède, 1803.

Pimelodinae Swainson, 1838: 331, 338. Type genus: *Pimelodus* La Cepède, 1803.

Sorubinae Swainson, 1838: 356. Type genus: *Sorubium* Swainson, 1838.

Hypophthalmini Bleeker, 1862 (in Bleeker, 1862–63): 15. Type genus: *Hypophthalmus* Spix & Agassiz, 1829.

Calophysinae Eigenmann, 1890: 12. Type genus: *Calophysus* Müller & Troschel, 1842.

Luciopimelodinae Driver, 1919: 451. Type genus: *Luciopimelodus* Eigenmann & Eigenmann, 1888.

Pinirampidae Fernández-Yépez, 1965: 12. Type genus: *Pinirampus* Bleeker, 1858.

Brachyplatystomatini Lundberg & Akama, 2005: 496. Type genus: *Brachyplatystoma* Bleeker, 1862.

Remarks: Priority of the publications by Bonaparte and Swainson has not yet been established. Thus, the authorship of the valid family name is not fixed. Calophysidae, based on the unjustified emendation *Calophysus*, has been used on occasion.

Taxonomic summary: Lundberg & Littmann (2003).

Phylogeny: Nass (1991).

Review: Mago-Leccia *et al.* (1986, Venezuela); Castro (1986a, Colombian Amazon and Orinoco basins).

29 genera, 93 species; 4 named fossil species.

***AGUARUNICHTHYS* Stewart, 1986**

Aguarunichthys Stewart, 1986a: 662. Type species: *Aguarunichthys torosus* Stewart, 1986. Type by original designation. Gender: Masculine.

***Aguarunichthys inpai* Zuanon, Rapp Py-Daniel & Jégu, 1993**

Aguarunichthys inpai Zuanon, Rapp Py-Daniel & Jégu, 1993: 258, figs. 2b, 6. Type locality: Brazil: Amazonas State: Solimões River drainage, north of the Marchantaria Island, at about 15 km above confluence with Rio Negro drainage. Holotype: INPA 5398.

Distribution: Middle Amazon River basin, Brazil (Lundberg & Littmann, 2003).

***Aguarunichthys tocantinsensis* Zuanon, Rapp Py-Daniel & Jégu, 1993**

Aguarunichthys tocantinsensis Zuanon, Rapp Py-Daniel & Jégu, 1993: 252, figs. 2a, 4. Type locality: Brazil: Pará State: Tocantins River drainage, rapids above Marabá. Holotype: INPA 5400.

Distribution: Tocantins River basin, Brazil (Lundberg & Littmann, 2003).

***Aguarunichthys torosus* Stewart, 1986**

Aguarunichthys torosus Stewart, 1986a: 663, figs. 7–8. Type locality: Peru, Departamento Amazonas, Río Cenepa, 1.6 km west of Huampami and near Peruvian military camp Chavez Valdivia, about 210 m elev., Approx. 4°28'S, 78°10'W. Holotype: LACM 39651-1.

Distribution: Cenepa River basin, Amazon River drainage, Peru (Lundberg & Littmann, 2003).

***BAGROPSIS* Lütken, 1874**

Bagropsis Lütken, 1874c: 32. Type species: *Bagropsis reinhardti* Lütken, 1874. Type by monotypy. Gender: Feminine.

***Bagropsis reinhardti* Lütken, 1874**

Bagropsis Reinhardti Lütken, 1874a: 32. Type locality: in flumine Rio das Velhas. Syntypes: BMNH 1876.1.10.9 (1), NMW 45905 (1), ZMUC P 29630 (1), ZMUC P 29631 (1), ZMUC P 29632 (1). Species illustrated and described in Lütken (1875: 160 (and p. V of summary), pl. 1, fig. 2).

Distribution: Das Velhas River basin, São Francisco River drainage, Brazil (Lundberg & Littmann, 2003).

***BERGIARIA* Eigenmann & Norris, 1901**

Bergiella Eigenmann & Norris, 1900: 355. Type species: *Pimelodus westermanni* Reinhardt, 1874. Type by original

designation. Gender: Feminine. Preoccupied by *Bergiella* Baker, 1897 (Hymenoptera), replaced by *Bergiaria* Eigenmann & Norris, 1901.

Bergiaria Eigenmann & Norris, 1901: 272. Type species: *Pimelodus westermannii* Reinhardt, 1874. Type by being a replacement name. Gender: Feminine. Replacement for *Bergiella* Eigenmann & Norris, 1900.

***Bergiaria platana* (Steindachner, 1908)**

Bergiella platana Steindachner, 1908d: 111. Type locality: La Plata. Holotype: at NMW.

Distribution: Paraná River basin, Argentina (Lundberg & Littmann, 2003).

***Bergiaria westermannii* (Reinhardt, 1874)**

Pimelodus Westermannii Reinhardt, in Lütken, 1874a: 33. Type locality: in flumine Rio das Velhas. Syntypes: ZMUC P 29638 (1), ZMUC P 29639 (1). Species illustrated and described in more detail in Lütken (1875: 167 (and p. VI of summary), pls. 2 (fig. 4), pl. 3 (fig. 4a–b)).

Distribution: Das Velhas River basin in São Francisco River drainage, Brazil (Lundberg & Littmann, 2003).

***BRACHYPLATYSTOMA* Bleeker, 1862**

Piramutana Bleeker, 1858b: 356. Type species: *Bagrus piramuta* Kner, 1857. Type by monotypy. Gender: Feminine.

Piratinga Bleeker, 1858b: 355. Type species: *Bagrus reticulatus* Kner, 1857. Type by subsequent designation by Bleeker (1862–63): 11. Gender: Feminine.

Brachyplatystoma Bleeker, 1862 (in Bleeker, 1862–63): 10. Type species: *Platystoma vaillanti* Valenciennes, 1840. Type by original designation. Gender: Neuter.

Malacobagrus Bleeker, 1862 (in Bleeker, 1862–63): 11. Type species: *Pimelodes filamentosus* Lichtenstein, 1819. Type by original designation. Gender: Masculine.

Taenionema Eigenmann & Bean, 1907: 662. Type species: *Taenionema steerei* Eigenmann & Bean, 1907. Type by original designation. Gender: Neuter. Preoccupied by *Taenionema* Banks, 1905 (Plecoptera), and by *Taenionema* Bolivar, 1906 (Orthoptera); replaced by *Goslinia* Myers, 1941.

Goslinia Myers, 1941: 88. Type species: *Taenionema steerei* Eigenmann & Bean, 1907. Type by being a replacement name. Gender: Feminine. Replacement for *Taenionema* Eigenmann & Bean, 1907, preoccupied by *Taenionema* Banks, 1905.

Ginesia Fernández-Yépez, 1951: [1]. Type species: *Ginesia cunaguaro* Fernández-Yépez, 1951. Type by original designation. Gender: Feminine.

Merodontotus Britski, 1981: 109. Type species: *Merodontotus tigrinus* Britski, 1981. Type by original designation. Gender: Masculine.

Review: Miranda Ribeiro (1918f, Brazil).

Remarks: Treatment of *Brachyplatystoma* as valid over older names follows Lundberg & Littmann (2003) and Lundberg & Akama (2005). Synonymy of *Merodontodus* and *Goslinia* follows Lundberg & Akama (2005).

***Brachyplatystoma capapretum* Lundberg & Akama, 2005**

Brachyplatystoma capapretum Lundberg & Akama, 2005: 501, fig. 8. Type locality: Brazil, Amazonas State, Rio Tefé sand beach, Lago Mucura, Supiã-Pucu, Tefé, 3°22'S, 64°43'W. Holotype: MZUSP 78481.

Distribution: Amazon River basin of Brazil and Peru (Lundberg & Akama, 2005).

***Brachyplatystoma filamentosum* (Lichtenstein, 1819)**

Pimelodes filamentosus Lichtenstein, 1819: 60. Type locality: Brasilien. Holotype: ZMB 2973.

Platystoma affine Valenciennes, in Cuvier & Valenciennes, 1840b: 24 (18 in the Strasbourg deluxe edition). Type locality: [not stated; apparently from Rio de Janeiro, Brazil]. Holotype: MNHN a-9360 (mounted).

Platystoma gigas Günther, 1872: 450. Type locality: River Huallaga, Upper Amazon [Peru]. Holotype: BMNH 1872.12.3.1 (stuffed).

Piratinga pirá-aíba Goeldi, 1898: 464, 477, pl. (fig. 4). Type locality: [Pará, Brazil]. Holotype: Whereabouts unknown. Name available from caption on plate.

Brachyplatystoma goeldii Eigenmann & Bean, 1907: 661, fig. 1. Type locality: Brazil: Amazon River, between Para and Manaos. Holotype: USNM 52561.

Distribution: Amazon and Orinoco River basins and major rivers of the Guianas and NE Brazil (Lundberg & Litt-

mann, 2003).

***Brachyplatystoma juruense* (Boulenger, 1898)**

Platystoma juruense Boulenger, 1898e: 421, pl. 39. Type locality: Rio Jurua, an affluent of the Amazons, Brazil.

Holotype: BMNH 1897.11.26.11.

Ginesia cunaguaro Fernández-Yépez, 1951: [2], unnumbered figure. Type locality: Río Apure, 3 km east of San Fernando de Apure, Venezuela. Holotype: AFY 51189; current whereabouts unknown.

Distribution: Amazon and Orinoco River basins (Lundberg & Littmann, 2003).

***Brachyplatystoma platynemum* Boulenger, 1898**

Brachyplatystoma platynema Boulenger, 1898b: 477. Type locality: Brazil, Pará. Holotype: BMNH 1898.10.11.20.

Taenionema steerei Eigenmann & Bean, 1907: 662, fig. 2. Type locality: Brazil, Amazon River, between Para and Manaos. Holotype: USNM 52571.

Distribution: Amazon and Orinoco River basins (Lundberg & Littmann, 2003).

† *Brachyplatystoma promagdalena* Lundberg, 2005

† *Brachyplatystoma promagdalena* Lundberg, 2005: 599, figs. 1a, 2a, 3, 4a, 5a, 6a. Type locality: Miocene Villa Vieja Formation of the Honda Group in the southern end of the río Magdalena basin near the town of Villavieja, Huila Department, Colombia at approx. 3°5'N, 75°13'W. Holotype: IGM 183062 (partial vertebral column, Weberian complex).

Distribution: La Venta formation, central Colombia; middle Miocene (Lundberg, 2005).

***Brachyplatystoma rousseauxii* (Castelnau, 1855)**

Bagrus rousseauxii Castelnau, 1855: 32, pl. 14 (fig. 1). Type locality: rivière des Amazones. Holotype: MNHN a-9457 (dry).

Bagrus Goliath Kner, 1857: 379. Type locality: Salto Theotonio [Brazil]. Holotype: at NMW.

Brachyplatystoma paraense Steindachner, 1909a: 195. Type locality: Fischmarkt von Pará [Brazil]. Holotype: NMW 47590. Illustrated and described in more detail in Steindachner (1915e: 65, fig. 4, pl. 13, fig. 5).

Distribution: Amazon and Orinoco River basins and other major rivers of South America east of the Andes mountains (Lundberg & Littmann, 2003).

***Brachyplatystoma tigrinum* (Britski, 1981)**

Merodontotus tigrinus Britski, 1981: 110, figs. 1–2. Type locality: Brazil, Cachoeira do Teotônio, rio Madeira, Território de Rondônia. Holotype: MZUSP 14004.

Distribution: Amazon River basin (Lundberg & Littmann, 2003).

***Brachyplatystoma vaillantii* (Valenciennes, 1840)**

? *Silurus Vaillantii* Cuvier, 1816: 204. Type locality: Brazil. Based on “bagre primus” Marcgravius (1648: 173).

Originally as *Sil. Nob. Vaillantii*.

Platystoma Vaillantii Valenciennes, in Cuvier & Valenciennes, 1840b: 21 (16 of Strasbourg deluxe edition), pl. 423.

Type locality: de Cayenne et de Suriname. Syntypes: MNHN b-0158 (1), MNHN b-0159 (1). Based in part on “bagre primus” Marcgravius (1648: 173).

Bagrus reticulatus Kner, 1857: 376, pl. 1 (fig. 1). Type locality: Salto Theotonio, am Flusse Araguay, Forte do Rio branco und Rio Madeira [Brazil]. Holotype: at NMW.

Bagrus piramuta Kner, 1857: 382. Type locality: Barra do Rio negro und Borba Rio Madeira [Brazil]. Syntypes: at NMW.

Brachyplatystoma parnabyae Steindachner, 1908e: 126. Type locality: Rio Parnabyba. Holotype: at NMW.

Distribution: Amazon and Orinoco River basins and major rivers of the Guianas and NE Brazil (Lundberg & Littmann, 2003).

Remarks: Inclusion of *Silurus Vaillanti* in the synonymy is based on the citation of the same species from Marcgravius (1648) in the text of that species as well as *Platystoma Vaillantii* Valenciennes.

Species inquirenda, Brachyplatystoma

Bagrus punctulatus Castelnau, 1855: 33, pl. 14 (fig. 2). Type locality: de la rivière des Amazones [Brazil]. Holotype: at MNHN, but current whereabouts unknown.

CALOPHYSUS Müller & Troschel, 1842

Calophysus Müller & Troschel, in Müller, 1842b: 310. Type species: *Pimelodes macropterus* Lichtenstein, 1819.

Type by subsequent designation by Bleeker (1862–63: 12). Gender: Masculine. Also as new in Müller (1843: 318).

Callophysus Müller & Troschel, 1848: 629. Type species: *Pimelodes macropterus* Lichtenstein, 1819. Type by subsequent designation by Bleeker (1862–63: 12). Gender: Masculine. Unjustified emendation of *Calophysus*, and a spelling that was in wide use.

Pimeletropis Gill, 1859b: 196. Type species: *Pimeletropis lateralis* Gill, 1859. Type by monotypy. Gender: Feminine.

Pseudocallophysus Bleeker, 1862 (in Bleeker, 1862–63): 12. Type species: *Pimelodus ctenodus* Spix & Agassiz, 1829. Type by original designation. Gender: Masculine.

***Calophysus macropterus* (Lichtenstein, 1819)**

Pimelodes macropterus Lichtenstein, 1819: 59. Type locality: Brasilien. Holotype: ZMB 3055.

Pimelodus ctenodus Spix & Agassiz, 1829: 21, pl. 8a. Type locality: aequitorialis flaviis [Brazil]. Holotype: Whereabouts unknown (Kottelat, 1988).

Pimeletropis lateralis Gill, 1859b: 196. Type locality: Amazon River. Holotype: at Lyceum Natural History, NY (whereabouts unknown).

Distribution: Amazon and Orinoco River basins (Lundberg & Littmann, 2003).

CHEIROCERUS Eigenmann, 1917

Cheirocerus Eigenmann, 1917a: 398. Type species: *Cheirocerus eques* Eigenmann, 1917. Type by original designation. Gender: Masculine.

Sovichthys Schultz, 1944c: 190. Type species: *Sovichthys abuelo* Schultz, 1944. Type by original designation. Gender: Masculine.

Revision: Stewart & Pavlik (1985).

***Cheirocerus abuelo* (Schultz, 1944)**

Sovichthys abuelo Schultz, 1944c: 191, pl. 1 (fig. a). Type locality: Venezuela, Río de Los Pájaros, 3 km above Lago Maracaibo, at depth of 15 ft. Holotype: USNM 121183.

Distribution: Lake Maracaibo basin (Lundberg & Littmann, 2003).

***Cheirocerus eques* Eigenmann, 1917**

Cheirocerus eques Eigenmann, 1917a: 398, pl. 39. Type locality: Villa Bella [Bolivia]. Holotype: FMNH 58255.

Distribution: Amazon River basin (Lundberg & Littmann, 2003).

***Cheirocerus goeldii* (Steindachner, 1908)**

Pimelodina goeldii Steindachner, 1908c: 83. Type locality: Rio Purus [Brazil]. Holotype: NMW 45503.

Pimelodus leptus Eigenmann & Pearson, in Eigenmann & Allen, 1942: 104, pl. 4 (fig. 3). Type locality: Río Pachitea [Peru]. Holotype: CAS 47288.

Distribution: Purus River basin (Lundberg & Littmann, 2003).

DUOPALATINUS Eigenmann & Eigenmann, 1888

Duopalatinus Eigenmann & Eigenmann, 1888b: 136. Type species: *Platystoma emarginatum* Valenciennes, 1840.

Type by monotypy. Gender: Masculine.

***Duopalatinus emarginatus* (Valenciennes, 1840)**

Platystoma emarginatum Valenciennes, in Cuvier & Valenciennes, 1840b: 25 (19 of Strasbourg deluxe edition).

Type locality: rivière de Saint-François. Holotype: MNHN a-9353 (mounted).

Distribution: São Francisco River basin, Brazil (Lundberg & Littmann, 2003).

***Duopalatinus peruanus* Eigenmann & Allen, 1942**

Duopalatinus peruanus Eigenmann & Allen, 1942: 107, pl. 4 (fig. 4). Type locality: Rio Puinagua, mouth of Rio Pacaya [Río Ucayali system, Peru]. Holotype: CAS 63630.

Distribution: Amazon and Orinoco River (Lundberg & Littmann, 2003).

EXALLODONTUS Lundberg, Mago-Leccia & Nass, 1991

Exallodontus Lundberg, Mago-Leccia & Nass, 1991: 843. Type species: *Exallodontus aguanai* Lundberg, Mago-Leccia & Nass, 1991. Type by original designation. Gender: Masculine.

Exallodontus aguanai Lundberg, Mago-Leccia & Nass, 1991

Exallodontus aguanai Lundberg, Mago-Leccia & Nass, 1991: 847, figs. 2–10, 13. Type locality: Venezuela, Territorio Federal Delta Amacuro, Río Orinoco near Los Castillos, 159 n mi from sea buoy at the terminus of the ship navigation channel in Boca Grande, 8°32'N, 62°23'W, 20–30 m. Holotype: MBUCV V-18930.

Distribution: Amazon and Orinoco River basins (Lundberg & Littmann, 2003).

HEMISORUBIM Bleeker, 1862

Hemisorubim Bleeker, 1862 (in Bleeker, 1862–63): 10. Type species: *Platystoma platyrhynchos* Valenciennes, 1840. Type by original designation. Gender: Masculine.

Hemisorubim platyrhynchos (Valenciennes, 1840)

Platystoma platyrhynchos Valenciennes in Cuvier & Valenciennes, 1840b: 27 (20 of Strasbourg deluxe edition). Type locality: [Not stated]. Holotype: MNHN 0000-1203.

Distribution: Amazon, Maroni, Orinoco, and Paraná River basins (Lundberg & Littmann, 2003).

HYPOPTHALMUS Cuvier, 1829

Hypophthalmus Cuvier, 1829: 293. Type species: *Hypophthalmus edentatus* Spix & Agassiz, 1829. Type by subsequent designation by Bleeker (1862–63: 15). Gender: Masculine.

Hypophthalmus Spix & Agassiz, 1829: 16. Type species: *Hypophthalmus edentatus* Spix & Agassiz, 1829. Type by subsequent designation Bleeker (1862–63: 15). Gender: Masculine.

Notophthalmus Hyrtl, 1859: 17. Type species: *Hypophthalmus marginatus* Valenciennes, 1840. Type by monotypy. Gender: Masculine. Preoccupied by *Notophthalmus* Rafinesque, 1820 (Amphibia); apparently not replaced.

Pseudohypophthalmus Bleeker, 1862 (in Bleeker, 1862–63): 15. Type species: *Hypophthalmus fimbriatus* Kner, 1857. Type by original designation. Gender: Masculine.

Review: Lopez-Fernandez & Winemiller (2000, Venezuela).

Phylogeny: Howes (1983a), Lundberg *et al.* (1991).

Hypophthalmus edentatus Spix & Agassiz, 1829

Hypophthalmus edentatus Spix & Agassiz, 1829: 16, pl. 9. Type locality: influviis Brasiliae equitorialis. Syntypes (2): Possibly MHNN 706 (2); see Kottelat (1988: 24) for discussion.

Hypophthalmus Spixii Valenciennes in Cuvier & Valenciennes, 1840b: 231 (172 of Strasbourg deluxe edition). Type locality: la partie la plus septentrionale du Brésil. Possible syntypes: MHNN 706 (2). Unneeded new name for *Hypophthalmus edentatus* Spix & Agassiz, 1829.

Distribution: Amazon and Orinoco River basins and Atlantic coastal rivers of Guyana and Suriname (Lundberg & Littmann, 2003).

Hypophthalmus fimbriatus Kner, 1857

Hypophthalmus fimbriatus Kner, 1857: 444, pl. 9 (fig. 30). Type locality: Rio negro [Brazil]. Syntype: NMW 50519 (1).

Distribution: Amazon River at Santarém and Negro River basin in Brazil and Venezuela (Lundberg & Littmann, 2003).

Hypophthalmus marginatus Valenciennes, 1840

Hypophthalmus marginatus Valenciennes, in Cuvier & Valenciennes, 1840b: 225 (168 of Strasbourg deluxe edition), pl. 439. Type locality: Cayenne, ... Surinam. Syntypes: MNHN a-8961 (1), MNHN a-8963 (1), RMNH 1932 (1), RMNH 1933 (1).

Distribution: Amazon and Orinoco River basins and major rivers of French Guiana and Suriname (Lundberg & Littmann, 2003).

Hypophthalmus oremaculatus Nani & Fuster, 1947

Hypophthalmus oremaculatus Nani & Fuster, 1947: 3, figs. 2–3. Type locality: Puerto Gaboto, Rio Paraná, curso

inferior [Argentina]. Holotype: MACN 3496.
Distribution: Paraná River basin, Brazil and Argentina (Lundberg & Littmann, 2003).

Species inquirendae, Hypophthalmus

Hypophthalmus longifilis Valenciennes in Cuvier & Valenciennes, 1840b: 230 (171 of Strasbourg deluxe edition). Type locality: Surinam. Syntypes: RMNH 1931 (1), RMNH 2974 (1), RMNH 2988 (1).

Hypophthalmus perporosus Cope, 1878: 673. Type locality: Peru, probably Nauta. Holotype: USNM 132589.

Hypophthalmus devall Röhl, 1942: 383. Type locality: Venezuela, ríos Aupre y Orinoco. Type(s): Whereabouts unknown.

IHERINGICHTHYS Eigenmann & Norris, 1900

Iheringichthys Eigenmann & Norris, 1900: 354. Type species: *Pimelodus labrosus* Lütken, 1874. Type by original designation. Gender: Masculine.

Iheringichthys labrosus (Lütken, 1874)

Pimelodus labrosus Lütken, 1874b: 200. Type locality: La Plata. Syntypes: ZMUC P 29633 (1), ZMUC P 29634 (1), ZMUC P 29635 (1), ZMUC P 29636 (1)..

Distribution: Paraná River basin (Lundberg & Littmann, 2003).

Iheringichthys megalops Eigenmann & Ward, 1907

Iheringichthys megalops Eigenmann & Ward, in Eigenmann, McAtee & Ward, 1907: 115, pl. 32 (figs. 3–4). Type locality: Paraguay, Bahia Negra, Río Paraguay. Holotype: CAS 63631.

Distribution: Paraná River basin, Paraguay (Lundberg & Littmann, 2003).

LEIARIUS Bleeker, 1862

Leiarius Bleeker, 1862 (in Bleeker, 1862–63): 10. Type species: *Arius longibarbis* Castelnau, 1855. Type by original designation. Gender: Masculine.

Sciadeoides Eigenmann & Eigenmann, 1888b: 136. Type species: *Sciades marmoratus* Gill, 1870. Type by monotypy. Gender: Masculine. Originally proposed as a subgenus of *Sciades*.

Leiarius marmoratus (Gill, 1870)

Sciades marmoratus Gill, 1870: 95. Type locality: Peru or Ecuador, Amazon system, Río Marañon or Río Napo. Holotype: USNM 8447.

Distribution: Amazon, Essequibo, and Orinoco River basins (Lundberg & Littmann, 2003).

Leiarius pictus (Müller & Troschel, 1849)

Bagrus (Sciades) pictus Müller & Troschel, 1849: 8, pl. 1 (fig. 1). Type locality: unbekannt. Holotype: ZMB 2991.

Distribution: Amazon and Orinoco River basins (Lundberg & Littmann, 2003).

Species inquirendae, Leiarius

Pimelodus arekaima Jardine, in Schomburgk, 1841: 178, pl. 5. Type locality: Upper Essequibo, Rio Branco. No types known.

Arius longibarbis Castelnau, 1855: 36, pl. 15 (fig. 2). Type locality: rivière des Amazones. Holotype: MNHN 0000-1189. Originally as *Arius ? longibarbis*.

Pimelodus multiradiatus Kner, 1857: 414. Type locality: Borba am Rio Madeira und Forte do Rio branco am Rio Facutu [Brazil]. Syntypes: NMW 16515 (1), NMW 16518 (1).

LUCIOPIMELODUS Eigenmann & Eigenmann, 1888

Luciopimelodus Eigenmann & Eigenmann, 1888b: 122. Type species: *Pimelodus pati* Valenciennes, 1835. Type by original designation. Gender: Masculine.

Luciopimelodus pati (Valenciennes, 1835)

Pimelodus Pati Valenciennes, 1835, in Valenciennes, 1835–47: pl. 1 (figs. 7–9). Type locality: Not stated [Corri-

entes, Argentina]. Probable holotype: MNHN a-9419 (1, dry). Name available from plate; mentioned in Valenciennes (1847: 7) from Corrientes. Described in Cuvier & Valenciennes (1840b: 176 (131 of Strasbourg deluxe edition)).

Silurus 11-radiatus Larrañaga, 1923: 386. Type locality: Uruguay. No types known. Corresponds to *Silurus pati* on p. 376.

Distribution: La Plata and Blanco River basins (Lundberg & Littmann, 2003).

MEGALONEMA Eigenmann, 1912

Megalonema Eigenmann, 1912b: 150. Type species: *Megalonema platycephalum* Eigenmann, 1912. Type by original designation. Gender: Neuter.

Megalonema argentina (MacDonagh, 1938)

Perugia argentina MacDonagh, 1938: 157. Type locality: Argentina, rio Paraná, Posadas, Territorio de Misiones. Holotype: MLP 5.V.I.35.

Distribution: Paraná River basin, Argentina (Lundberg & Littmann, 2003).

Megalonema pauciradiatum Eigenmann, 1919

Megalonema pauciradiatum Eigenmann, in Driver, 1919: 455. Type locality: Paraguay, Villa Rica. Holotype: CAS 63672.

Distribution: Paraná River basin, Paraguay (Lundberg & Littmann, 2003).

Megalonema platanum (Günther, 1880)

Pimelodus platanus Günther, 1880a: 10. Type locality: Parana. Holotype: BMNH 1872.6.8.18.

Distribution: Paraná River basin (Lundberg & Littmann, 2003).

Megalonema platycephalum Eigenmann, 1912

Megalonema platycephalum Eigenmann, 1912b: 150, fig. 31, pl. 10 (fig. 2). Type locality: Tumatumari, British Guiana. Holotype: FMNH 53224.

Distribution: Amazon, Essequibo, and Orinoco River basins (Lundberg & Littmann, 2003).

Megalonema psammium Schultz, 1944

Megalonema platycephalum psammium Schultz, 1944c: 216, pl. 2 (fig. b). Type locality: Venezuela, Rio Palmar at the bridge 70 km southwest of Maracaibo. Holotype: USNM 121175.

Distribution: Lake Maracaibo basin, Colombia, Venezuela (Lundberg & Littmann, 2003).

Megalonema xanthum Eigenmann, 1912

Megalonema xanthum Eigenmann, 1912a: 16. Type locality: Girardot [Colombia]. Holotype: FMNH 56032; holotype illustrated in Eigenmann (1922b: 35, pl. 3, fig. 3) as *Perugia xanthus*.

Distribution: Magdalena River basin, Colombia (Lundberg & Littmann, 2003; Maldonado-Ocampo *et al.*, 2005).

PARAPIMELODUS La Monte, 1933

Parapimelodus La Monte, 1933b: 226. Type species: *Pimelodus valenciennis* Lütken, 1874. Type by original designation. Gender: Masculine.

Revision: Lucena *et al.* (1992).

Parapimelodus nigribarbis (Boulenger, 1889)

Pimelodus (Pseudorhamdia) nigribarbis Boulenger, 1889: 266. Type locality: Brazil, Rio Grande do Sul, Camaquam River. Lectotype: BMNH 1889.8.24.6, designated by Lucena *et al.* (1992: 145).

Distribution: Laguna dos Patos basin, Brazil (Lundberg & Littmann, 2003).

Parapimelodus valenciennis (Lütken, 1874)

Pimelodus valenciennis Lütken, 1874b: 200. Type locality: La Plata. Holotype: ZMUC P 29637.

Pimelodus Spegazzinii Perugia, 1891: 632. Type locality: Rio Durazno. Syntypes: MSNG 8026 (2).

Distribution: Paraná River basin (Lundberg & Littmann, 2003).

PERRUNICHTHYS Schultz, 1944

Perrunichthys Schultz, 1944c: 229. Type species: *Perrunichthys perruno* Schultz, 1944. Type by original designation. Gender: Masculine.

Perrunichthys perruno Schultz, 1944

Perrunichthys perruno Schultz, 1944c: 230, pl. 3 (fig. b); fig. 3. Type locality: Venezuela, Río Negro, below the mouth of the Río Yasa, about 75 km. south of Rosario, west side of Lago Maracaibo. Holotype: USNM 121189. Distribution: Lake Maracaibo basin (Lundberg & Littmann, 2003).

PHRACTOCEPHALUS Spix & Agassiz, 1829

Phractocephalus Spix & Agassiz, 1829: 10. Type species: *Phractocephalus bicolor* Spix & Agassiz, 1829. Type by monotypy. Gender: Masculine.

Pirarara Spix & Agassiz, 1829: 23. Type species: *Silurus hemioliopterus* Bloch & Schneider, 1801. Type by monotypy. Gender: Feminine. Appeared first as name in synonymy under *Phractocephalus* and in legend for pl. 6; made available by Bleeker (1862–63: 11) but with authorship to Spix & Agassiz.

Remarks: *Phractocephalus* selected as valid by first reviser action of Agassiz (in Spix & Agassiz, 1831: Conspectus); see Kottelat (1989b: 321) for details.

Phractocephalus hemioliopterus (Bloch & Schneider, 1801)

Silurus hemioliopterus Bloch & Schneider, 1801: 385. Type locality: [in flumine Maranham Brasiliae].

Pimelodus grunniens Humboldt, in Humboldt & Valenciennes, 1821: 172. Type locality: le Bas-Orénoque. No types known.

Phractocephalus bicolor Spix & Agassiz, 1829: 23, pl. 6. Type locality: in fluvio Amazonum [Brazil]. Whereabouts unknown (Kottelat, 1988). As *Pirarara bicolor* on plate.

Distribution: Amazon and Orinoco River basins (Lundberg & Littmann, 2003).

Remarks: See Boeseman (1983b) for comments on taxonomy and nomenclature.

† **Phractocephalus nassi** Lundberg & Aguilera, 2003

† *Phractocephalus nassi* Lundberg & Aguilera, 2003: 101, figs. 3, 5a, 5b, 5e, 5h, 5k, 5l. Type locality: El Mamón oil field, 350 m north of oil well number 1, locality USB 56-FU, 11°15'N, 70°13'W, Urumaco Formation, middle member, Falcón State, northwestern Venezuela. Holotype: MCN.USB OL-2142.

Distribution: Urumaco Formation, Falcón State, Venezuela, upper Miocene (Lundberg & Aguilera, 2003).

Remarks: Described earlier by Lundberg *et al.* (1988) as *Phractocephalus hemioliopterus*.

PIMELODINA Steindachner, 1877

Pimelodina Steindachner, 1877a: 149. Type species: *Pimelodina flavipinnis* Steindachner, 1877. Type by monotypy. Gender: Feminine.

Revision: Stewart (1986a).

Pimelodina flavipinnis Steindachner, 1877

Pimelodina flavipinnis Steindachner, 1877a: 150, pl. 13 (fig. 2) [as *Pimelodus (Pimelodina) flavipinnis*]. Type locality: Amazonenstrom bei Para [Brazil]. Holotype: NMW 45498.

Pimelodina nasus Eigenmann & Eigenmann, 1888b: 120. Type locality: Para [Brazil]. Holotype: MCZ 7490.

Distribution: Amazon and Orinoco River basins (Lundberg & Littmann, 2003).

PIMELODUS La Cepède, 1803

Pimelodus La Cepède, 1803: 93. Type species: *Pimelodus maculatus* La Cepède, 1803. Type by subsequent designation, by Gill (1861b). Gender: Masculine.

Pseudariodes Bleeker, 1862 (in Bleeker, 1862–63): 11. Type species: “*Pseudariodes clarias* = *Silurus clarias* Bl.” [= *Pimelodus Blochii* Valenciennes, 1840, nec. *Silurus clarias* Linnaeus, 1758]. Type by original designation. Gender: Masculine.

Pseudorhamdia Bleeker, 1862 (in Bleeker, 1862–63): 11. Type species: *Pimelodus maculatus* La Cepède, 1803. Type by original designation. Gender: Feminine.

Pimelodus absconditus Azpelicueta, 1995

Pimelodus absconditus Azpelicueta, 1995: 72, figs. 1–2. Type locality: Argentina, Misiones, San Javier, Uruguay River. Holotype: MLP 8781.

Distribution: Paraná River basin (Lundberg & Littmann, 2003).

Pimelodus albicans (Valenciennes, 1840)

Arius albidus Valenciennes, 1835, in Valenciennes, 1835–47: pl. 3, fig. 2. Type locality: Not stated. Holotype: MHN a-9400. Name made available by caption on plate.

Arius albicans Valenciennes, in Cuvier & Valenciennes, 1840b: 80 (60 of Strasbourg deluxe edition). Type locality: Buénos-Ayres. Syntype: MNHN a-9400 (dry). Unneeded new name or *lapsus calami* for *Arius albidus* Valenciennes.

Silurus muticus Larrañaga, 1923: 386. Type locality: Uruguay. No types known.

Distribution: Paraná River basin, Argentina (Lundberg & Littmann, 2003).

Remarks: Treatment of *Pimelodus albicans* as valid follows Lundberg & Littmann (2003), in which *Arius albidus* Valenciennes, 1835, was treated as a *nomen oblitum*.

Pimelodus albofasciatus Mees, 1974

Pimelodus albofasciatus Mees, 1974: 137, pl. 4. Type locality: Suriname, Sipaliwini. Holotype: RMNH 26156.

Distribution: Amazon, Orinoco, upper Corantijn and Sipaliwini River basins (Lundberg & Littmann, 2003).

Pimelodus altissimus Eigenmann & Pearson, 1942

Pimelodus altissimus Eigenmann & Pearson, in Eigenmann & Allen, 1942: 106, pl. 5 (fig. 5). Type locality: Rio Ucayali, near Orellana [Peru]. Holotype: CAS 55369.

Distribution: Amazon River basin (Lundberg & Littmann, 2003).

Pimelodus atrobrunneus Vidal & Lucena, 1999

Pimelodus atrobrunneus Vidal & Lucena, 1999: 123, figs. 1–3. Type locality: Brasil: rio Ligeiro na estrada entre Marcelino Ramos e Maximiliano de Almeida, Marcelino Ramos, 27°38'S–51°52'O, Rio Grande do Sul. Holotype: MCP 19678.

Distribution: Upper Uruguay River basin, Brazil (Lundberg & Littmann, 2003).

Pimelodus blochii Valenciennes, 1840

Pimelodus Blochii Valenciennes, in Cuvier & Valenciennes, 1840b: 188 (139 of Strasbourg deluxe edition). Type locality: Surinam. Syntypes: MNHN b-0072 (1), ZMB 2995 (1). Based in part on *Silurus clarias* of Bloch (1782: pl. 35, figs. 1–2), not *Silurus clarias* Linnaeus, 1758.

Pseudorhamdia macronema Bleeker, 1864a: 79, pl. 13 (fig. 7), pl. 14. Type locality: Surinama. Lectotype: RMNH 3069, designated by Boeseman (1972: 317).

Pseudorhamdia piscatrix Cope, 1870b: 569. Type locality: Pebas [Peru]. Syntypes: ANSP 8386–87 (2).

Distribution: Gulf of Paria, Amazon, Corantijn, Essequibo and Orinoco River basins (Lundberg & Littmann, 2003); Magdalena River basin (Maldonado-Ocampo *et al.*, 2005).

Remarks: Eschmeyer *et al.* (1998) and Lundberg & Littmann (2003) list MNHN b-0072 as lectotype of *Pimelodus blochii*, without citing a published source of the lectotype designation and, instead, apparently relying on a notation in the MNHN catalog to that effect. No published designation has been found, so the two specimens are tentatively retained as syntypes here.

Pimelodus brevis Marini, Nichols & La Monte, 1933

Pimelodus brevis Marini, Nichols & La Monte, 1933: 1, fig. 1. Type locality: Argentina, Rio de la Plata, San Fernando. Holotype: AMNH 12240 (missing).

Distribution: Paraná River basin; Durazno River, Argentina (Lundberg & Littmann, 2003).

Pimelodus coprophagus Schultz, 1944

Pimelodus clarias coprophagus Schultz, 1944c: 203, fig. 2. Type locality: Venezuela, Río Agua Caliente, 2 to 3 km above the southwestern corner of Lago Maracaibo. Holotype: USNM 121150.

Distribution: Lake Maracaibo basin (Lundberg & Littmann, 2003).

Pimelodus fur (Reinhardt, 1874)

Pseudorhamdia fur Reinhardt, in Lütken, 1874c: 33. Type locality: in flumine Rio das Velhas. Syntypes: NMW 44443 (3), NMW 44763 (2), SMNS 2026 (1), ZMB 9181 (2), ZMUC P 29643–ZMUC P 29651 (1 each). Illustrated and described in more detail in Lütken (1875: 169 (and p. VI of summary), pl. 2, fig. 3; pl. 3, fig. 3a).

Distribution: Das Velhas River basin, São Francisco River drainage, Brazil (Lundberg & Littmann, 2003).

Pimelodus garciabarrigai Dahl, 1961

Pimelodus garcia-barrigai Dahl, 1961: 494. Type locality: Colombia, Deep pool in Caño Lozada, about 11 km

above its junction with the Guayabero River. Holotype: ICNMHN 744 (Cala, 1981).

Distribution: Guayabero River basin, Orinoco River drainage, Colombia (Lundberg & Littmann, 2003).

***Pimelodus grosskopfii* Steindachner, 1879**

Pimelodus (Pimelodus) Grosskopfii Steindachner, 1879c: 194. Type locality: Río Cauca [Colombia]. Syntypes: NMW 45781 (4), NMW 45782 (1). Also described as new in Steindachner (1879g: 186); described in more detail and illustrated in Steindachner (1880b: 57, pl. 1 figs. 1, 1a).

Pimelodus longifilis Posada, 1909: 294. Type locality: aus dem Cauca [Colombia]. No types known.

Distribution: Magdalena River basin and Lake Maracaibo basins (Lundberg & Littmann, 2003).

***Pimelodus heraldoi* Azpelicueta, 2001**

Pimelodus heraldoi Azpelicueta 2001: 194, figs. 1–2 Type locality: Brazil, Estado de São Paulo: Município de Pirassununga, rio Mogi Guaçu in Emas. Holotype: MZUSP 22713.

Distribution: Upper Paraná River basin, Brazil (Lundberg & Littmann, 2003).

***Pimelodus jivaro* Eigenmann & Pearson, 1942**

Pimelodus jivaro Eigenmann & Pearson, in Eigenmann & Allen, 1942: 105, pl. 4 (fig. 2). Type locality: Rio Morona [Peru]. Syntypes (5; 85–105 mm): CAS 55891 (3).

Distribution: Upper Amazon River basin, Ecuador, Peru (Lundberg & Littmann, 2003).

***Pimelodus maculatus* La Cepède, 1803**

Pimelodus maculatus La Cepède, 1803: 94. Type locality: Le grand fleuve de la Plata, ... Buénos-Ayres, ainsi qu'à la Encenada. No types known.

Distribution: Paraná and São Francisco River basins (Lundberg & Littmann, 2003).

***Pimelodus microstoma* Steindachner, 1877**

Pimelodus microstoma Steindachner, 1877b: 604, footnote. Type locality: Brazil, von Irisanga, Rio branco und Barra do Rio negro. Syntypes: NMW 45823 (1), NMW 45824 (2).

Distribution: Amazon River basin, Brazil (Lundberg & Littmann, 2003).

***Pimelodus misteriosus* Azpelicueta, 1998**

Pimelodus misteriosus Azpelicueta, 1988: 88, figs. 1–3, 5–6, 8, 10, 12, 14. Type locality: Argentina, Misiones, Canadaria, arroyo Anselmo. Holotype: MLP 9191.

Distribution: Paraná River basin (Lundberg & Littmann, 2003).

***Pimelodus navarroi* Schultz, 1944**

Pimelodus grosskopfii navarroi Schultz, 1944c: 207, pl. 1 (fig. c). Type locality: Venezuela, Río Palmar at the bridge 70 km southwest of Maracaibo. Holotype: USNM 121174.

Distribution: Lake Maracaibo basin (Lundberg & Littmann, 2003).

***Pimelodus ornatus* Kner, 1857**

Pimelodus ornatus Kner, 1857: 411, pl. 6 (fig. 18). Type locality: Surinam, dem Rio negro und Cujaba [Brazil]. Syntypes: NMW 45832 (2), NMW 45843 (1).

Megalonema rhabdostigma Fowler, 1914: 256, fig. 10. Type locality: Rupununi River, British Guiana ... in the highlands of British Guiana, approximately secured in North Latitude 2° to 3°, and West Longitude 50°20'. Holotype: ANSP 39338.

Distribution: Amazon, Corantijn, Essequibo, Orinoco and Paraná River basins and major rivers of the Guianas (Lundberg & Littmann, 2003).

***Pimelodus ortmanni* Haseman, 1911**

Pimelodus ortmanni Haseman, 1911b: 379, pl. 50 (fig. 2). Type locality: Brazil, Porto União da Victoria, Rio Iguassú. Holotype: FMNH 54240.

Distribution: Paraná River basin, Brazil (Lundberg & Littmann, 2003).

***Pimelodus paranaensis* Britski & Langeani, 1988**

Pimelodus paranaensis Britski & Langeani, 1988: 410, figs. 1–2. Type locality: Ilha Solteira, Rio Paraná, SP [Brazil]. Holotype: MZUSP 23089.

Distribution: Upper Paraná River basin, Brazil (Lundberg & Littmann, 2003).

***Pimelodus pictus* Steindachner, 1877**

Pimelodus pictus Steindachner, 1877a: 144. Type locality: aus dem Amazonenstrom auf peruanischen Gebiete, und

aus dem Hyavary. Syntypes: NMW 45859 (3).

Distribution: Amazon and Orinoco River basins (Lundberg & Littmann, 2003).

Pimelodus platicirris Borodin, 1927

Pimelodus platicirris Borodin, 1927c: 2. Type locality: Salto de Pirassunungo, Rio Mogy Guassu, São Paulo Prov., Brazil. Holotype: AMNH 8628.

Distribution: Paraná River basin, Brazil; known only from the holotype (Lundberg & Littmann, 2003).

Pimelodus punctatus (Meek & Hildebrand, 1913)

Megalonema punctatum Meek & Hildebrand, 1913: 77. Type locality: Río Tuyra, Boca de Cupe, Panama. Holotype: FMNH 7577.

Megalonema robustum Meek & Hildebrand, 1913: 78. Type locality: Río Tuyra, Marriganti, Panama. Holotype: FMNH 7578.

Distribution: Tuira River basin, Panama (Lundberg & Littmann, 2003).

Species inquirendae, Pimelodus

Pimelodus rigidus Spix & Agassiz, 1829: 19, pl. 7 (fig. 2). Type locality: Brazil aequatoriali. Holotype: Whereabouts unknown (Kottelat, 1988).

Pseudariodes pantherinus Lütken, 1874b: 192. Type locality: Caracas. Syntypes: BMNH 1876.1.10.10 (1), NMW 45851 (1), USNM 44970 (1), ZMUC P 29640 (1), ZMUC P 29641 (1), ZMUC P 29642 (1).

Piramutana macrospila Günther, 1880a: 10, pl. 2. Type locality: Rio de la Plata. Holotype: BMNH 1878.9.10.14.

PINIRAMPUS Bleeker, 1858

Pinirampus Bleeker, 1858b: 198. Type species: *Pimelodus pinirampus* Agassiz [= *Pimelodus pirinampu* Spix & Agassiz, 1829]. Type by monotypy. Gender: Masculine.

Pirinampus Günther, 1864: 135. Type species: *Pimelodus pirinampu* Spix & Agassiz, 1829. Type by being a replacement name. Gender: Masculine. Unjustified emendation of *Pinirampus* Bleeker, 1858.

Perugia Eigenmann & Norris, 1900: 355. Type species: *Pirinampus* [sic] *agassizii* Steindachner, 1876. Type by original designation. Gender: Feminine.

Pinirampus pirinampu (Spix & Agassiz, 1829)

Pimelodus barbancho Humboldt, in Humboldt & Valenciennes, 1821: 172. Type locality: Guarico, l'Apure et d'autres rivières des steppes de Venezuela. Holotype: at MNHN.

Pimelodus pirinampu Spix & Agassiz, 1829: 20, pl. 8. Type locality: Brasiliae fluviis. Type(s): Whereabouts unknown (Kottelat, 1988).

Pimelodus insignis Jardine, in Schomburgk, 1841: 180, pl. 6. Type locality: Rio Branco [Brazil]. No types known.

Galeichthys araguayensis Castelnau, 1855: 37, pl. 17 (fig. 3). Type locality: Rio Araguay [Brazil]. Holotype: MNHN a-9366 (mounted).

Pinirampus typus Bleeker, 1862 (in Bleeker, 1862–63): 11. Type locality: Brasiliae fluviis. Type(s): Whereabouts unknown. Unneeded replacement for *Pimelodus pinirampus* Spix & Agassiz, 1829, apparently to avoid Stricklandian tautonomy.

Pirinampus Agassizii Steindachner, 1876: 607, pl. 12. Type locality: Pará [Brazil]. Holotype: NMW 45955.

Distribution: Amazon, Essequibo, Orinoco, and Paraná River basins (Lundberg & Littmann, 2003).

Remarks: *Pimelodus barbancho* treated here as a *nomen oblitum*, following Lundberg & Littmann (2003).

PLATYNEMATICHTHYS Bleeker, 1858

Platynemathchys Bleeker, 1858b: 356. Type species: *Bagrus punctulatus* Kner, 1857. Type by monotypy. Gender: Masculine.

Platypogon Starks, 1913: 28. Type species: *Platypogon caerulorostris* Starks, 1913. Type by monotypy. Gender: Masculine.

Platynemathchys notatus (Jardine, 1841)

Pimelodus notatus Jardine, in Schomburgk, 1841: 181, pl. 7. Type locality: Fort St. Joaquim on the Rio Branco [Bra-

zil]. No types known.

Bagrus punctulatus Kner, 1857: 380. Type locality: Forte do Principe am Rio Guaporé und Rio branco [Brazil].

Holotype: at NMW. Preoccupied by *Bagrus punctulatus* Castelnau, 1855; replaced by *Bagrus nigropunctatus* Kner, 1858.

Bagrus nigropunctatus Kner, 1858: 345. Type locality: Forte do Principe am Rio Guapore und Rio branco [Brazil].

Holotype: at NMW. Replacement for *Bagrus punctulatus* Kner, 1857; preoccupied by *Bagrus punctulatus* Castelnau, 1855.

Platypogon caerulorostris Starks, 1913: 29, pl. 5. Type locality: Brazil, Pará. Holotype: SU 22228.

Distribution: Amazon and Orinoco River basins (Lundberg & Littmann, 2003).

PLATYSILURUS Haseman, 1911

Platysilurus Haseman, 1911a: 320. Type species: *Platysilurus barbatus* Haseman, 1911. Type by monotypy. Gender: Masculine.

Platysilurus malarmo Schultz, 1944

Platysilurus malarmo Schultz, 1944c: 234, fig. 3g, pl. 3 (fig. c). Type locality: Venezuela, Lago Maracaibo near the mouth of the Río Concho. Holotype: USNM 121179.

Distribution: Lake Maracaibo basin (Lundberg & Littmann, 2003).

Platysilurus mucosus (Vaillant, 1880)

Platystoma mucosa Vaillant, 1880b: 151. Type locality: à Caldéron (Haute-Amazone). Holotype: MNHN a-1955.

Duoplatinus goeldii Steindachner, 1908b: 65. Type locality: Rio Purus [Brazil]. Holotype: at NMW.

Platysilurus barbatus Haseman, 1911: 320, pl. 52. Type locality: São Antonio de R. Madeira [Brazil]. Holotype: FMNH 60306.

Distribution: Amazon and Orinoco River basins (Lundberg & Littmann, 2003).

Platysilurus olallae (Orcés, 1977)

Duopalatinus olallae Orcés, 1977: 78, fig. 4. Type locality: el bajo Bobonaza [Ecuador]. Holotype: at MEPN (132 mm, catalog number not stated).

Distribution: Ecuador.

Remarks: Not treated in Lundberg & Littmann (2003).

PLATYSTOMATICHTHYS Bleeker, 1862

Platystomatichthys Bleeker, 1862 (in Bleeker, 1862–63): 10. Type species: *Platystoma sturio* Kner, 1857. Type by original designation. Gender: Masculine.

Platystomatichthys sturio (Kner, 1857)

Platystoma sturio Kner, 1857: 395, pl. 3 (figs. 9–9a). Type locality: Rio branco [Brazil]. Holotype: at NMW.

Distribution: Amazon River basin (Lundberg & Littmann, 2003).

PROPIMELODUS Lundberg & Parisi, 2002

Propimelodus Lundberg & Parisi, 2002: 77. Type species: *Pimelodus eigenmanni* van der Stigchel, 1946. Type by original designation.

Propimelodus eigenmanni (van der Stigchel, 1946)

Pimelodus eigenmanni van der Stigchel, 1946: 64. Type locality: Brazil. Holotype: RMNH 15465. Preoccupied by *Pimelodus eigenmanni* Boulenger, 1891 [now *Pimelodella eigenmanni*, in the Heptapteridae]; apparently not replaced.

Distribution: Amazon, Approuague, Kourou, and Oyapock River basins (Lundberg & Littmann, 2003).

Remarks: *Pimelodus eigenmanni* is tentatively treated as valid, despite being a junior primary homonym, following Art. 23.9.5 of the Code of Zoological Nomenclature. Redescribed in Lundberg & Parisi (2002).

PSEUDOPLATYSTOMA Bleeker, 1862

Pseudoplatystoma Bleeker, 1862 (in Bleeker, 1862–63): 10. Type species: *Silurus fasciatus* Linnaeus, 1766. Type by original designation. Gender: Neuter.

Hemiplatystoma Bleeker, 1862 (in Bleeker, 1862–63): 10. Type species: *Platystoma tigrinum* Valenciennes, 1840.

Type by original designation. Gender: Neuter.

Remarks: Eigenmann & Eigenmann (1888b) serve as first revisers in giving precedence to *Pseudoplatystoma* over *Hemiplatystoma*.

***Pseudoplatystoma corruscans* (Spix & Agassiz, 1829)**

Platystoma corruscans Spix & Agassiz, 1829: 26 [pl. 13]. Type locality: in flumine S. Francisci [Brazil]. Holotype: Whereabouts unknown (Kottelat, 1988). As *Sorubim caparary* on plate.

Sorubim caparary Spix in Spix & Agassiz, 1829: pl. 13. Type locality: in flumine S. Francisci [Brazil]. Holotype: Whereabouts unknown (Kottelat, 1988). Name available from caption on plate.

Platystoma coruscans Valenciennes, in Cuvier & Valenciennes, 1840b: 17 (13 in the Strasbourg deluxe edition). Misspelling or emendation of *Platystoma corruscans* Spix & Agassiz.

Silurus macrocephalus Larrañaga, 1923: 386. Type locality: Uruguay. No types known. Appears to correspond to *Silurus mangrullo* on p. 377, which is a *nomen nudum*.

Distribution: São Francisco and Paraná River basins (Lundberg & Littmann, 2003)

Remarks: Agassiz (in Spix & Agassiz, 1831: conspectus) serves as first reviser in giving precedence to *Platystoma corruscans* over *Sorubim caparary*.

***Pseudoplatystoma fasciatum* (Linnaeus, 1766)**

Silurus fasciatus Linnaeus, 1766: 505. Type locality: Brasilia, Surinami. Holotype: Whereabouts unknown.

Platystoma artedii Günther, 1864: 106. Type locality: South America. Syntypes: On *Mystus* no. 6 in Seba (1734–65: 84, pl. 29, fig. 6) and *Mystus* no. 386 of Gronovius (1784: pl. 125).

Pseudoplatystoma fasciatum intermedium Eigenmann & Eigenmann, 1888b: 138. Type locality: Obidos. Holotype: MCZ 7321.

Pseudoplatystoma fasciatum brevifile Eigenmann & Eigenmann, 1889b: 31. Type locality: Goyaz [Brazil]. Holotype: MCZ 7317.

Pseudoplatystoma fasciatum reticulatum Eigenmann & Eigenmann, 1889b: 30. Type locality: Rio Negro [Brazil]. Holotype: MCZ 23813.

Pseudoplatystoma fasciatum nigricans Eigenmann & Eigenmann, 1889b: 31. Type locality: Xingu. Syntypes (2): MCZ 7301 (1), MCZ 7311 (1).

Distribution: Amazon, Corantijn, Essequibo, Orinoco, and Paraná River basins (Lundberg & Littmann, 2003); Magdalena River basin (Maldonado-Ocampo *et al.*, 2005).

***Pseudoplatystoma tigrinum* (Valenciennes, 1840)**

Platystoma truncatum Spix & Agassiz, 1829: 27, pl. 13a. Type locality: Brazil, Japurá and Solimoen. Type (s): Whereabouts unknown (Kottelat, 1988).

Platystoma tigrinum Valenciennes, in Cuvier & Valenciennes, 1840b: 10 (8 of Strasbourg deluxe edition), pl. 422. Type locality: probablement originaire du Brésil. Holotype: MNHN a-9354 (mounted).

Distribution: Amazon and Orinoco River basins (Lundberg & Littmann, 2003).

Remarks: Synonymy follows Lundberg & Littmann (2003) which differs from most previous accounts, in which

Platystoma truncatum was treated as a synonym of *Pseudoplatystoma fasciatum*. Because of this, *Platystoma truncatum* became the senior synonym for a widely used *Pseudoplatystoma tigrinum*. *P. truncatum* was treated as a *nomen oblitum* in Lundberg & Littmann (2003), but without supporting documentation. Use of *P. truncatum* as a valid name has not been found in surveyed compilations of late 19th or 20th century and, therefore, the use of *P. tigrinum* as valid is continued here.

Species inquirendae, Pseudoplatystoma

Platystoma pardalis Valenciennes, 1836, in Valenciennes, 1835–47: pl. 4 (fig. 2). Type locality: Buenos Aires, Argentina. Holotype: MNHN a-8833. Name available from plate, mentioned in Valenciennes (1847: 6), as *Platystoma panthale*. Described in Cuvier & Valenciennes (1840b: 15 (12 of Strasbourg deluxe edition)), as *Platystoma pardale*.

Platystoma orbignianus Valenciennes, 1836, in Valenciennes, 1835–47: pl. 4 (fig. 3). Type locality: Brazil. Holotype: MNHN b-0160. Name available from plate, description in Valenciennes (1847: 6), as *Platystoma orbignyanum*.

Description in Cuvier & Valenciennes (1840b: 12, [9 of Strasbourg deluxe edition]).

Platystoma panthale Valenciennes, 1847: 6. Type locality: [Not stated]. Holotype: MNHN a-8833. New name proposed for *Platystoma pardalis* Valenciennes, 1836, without explanation.

Platystoma punctifer Castelnau, 1855: 40, pl. 19 (fig. 2). Type locality: de l' Amazone. Holotype: MNHN 0000-1582.

SORUBIM Cuvier, 1829

Sorubim Cuvier, 1829: 293. Type species: *Silurus lima* Bloch & Schneider, 1801. Type by subsequent designation, apparently by Kottelat (1988). Gender: Masculine.

Platystoma Spix & Agassiz, 1829: 23. Type species: *Silurus lima* Bloch & Schneider, 1801. Type by subsequent designation by Jordan (1917: 131). Gender: Neuter. Preoccupied by *Platystoma* Meigen, 1803 (Diptera).

Sorubim Spix & Agassiz, 1829: 24. Type species: *Silurus lima* Bloch & Schneider, 1801. Type by subsequent designation by Bleeker (1862–63: 10). Gender: Masculine. First appeared in synonymy of *Platystoma* Spix & Agassiz and therefore not available, but made available because of use as a valid name by Bleeker (1862–63: 10) [ICZN art. 11.6.1]. Preoccupied by *Sorubim* Cuvier, 1829.

Sorubium Swainson, 1838: 356. Type species: *Silurus lima* Bloch & Schneider, 1801. Type by being a replacement name. Unneeded replacement for *Sorubim* Spix & Agassiz, 1829.

Abron Gistel, 1848: X. Type species: *Silurus lima* Bloch & Schneider, 1801. Type by being a replacement name. Gender: Neuter. Replacement for *Platystoma* Valenciennes [= Spix & Agassiz, 1829].

Remarks: *Sorubim* was apparently adopted by Cuvier (1829) from the Spix & Agassiz (1829) manuscript, but the name was published first in the former book.

Revision: Littmann (1998).

Sorubim cuspicaudus Littmann, Burr & Nass, 2000

Sorubim cuspicaudus Littmann, Burr & Nass, 2000: 903, fig. 2b, 3, 4c. Type locality: Colombia, Departamento de Tolima, Río Magdalena drainage, at Puerto Soplaviento. Holotype: FMNH 56223.

Distribution: Magdalena and Sinu River basins, Colombia, and Lake Maracaibo (Lundberg & Littmann, 2003).

Sorubim elongatus Littmann, Burr, Schmidt & Isern, 2001

Sorubim elongatus Littmann, Burr, Schmidt & Isern, 2001: 4, fig. 2. Type locality: Peru: Departamento Loreto, Maynas Province, Ullpa Caño, 50 m upstream of confluence with Moena Caño, Río Itaya drainage, Río Amazonas basin (approx. 3°46'20"S 73°14'17"W). Holotype: SIUC 30303.

Distribution: Amazon, Essequibo, and Orinoco River basins (Lundberg & Littmann, 2003).

Sorubim lima (Bloch & Schneider, 1801)

Silurus lima Bloch & Schneider, 1801: 384. Type locality: in flumine Maranham Brasiliae. Holotype: ZMB 3185 (stuffed).

Sorubim infraoculare Spix, in Spix & Agassiz, 1829: pl. 15. Type locality: Brasiliae aequatorialis fluviis. Holotype: Whereabouts unknown (Kottelat, 1988). Name available from caption on plate.

Platystoma Luceri Weyenbergh, 1877: 10, pl. 3 (figs. 1–3). Type locality: Santa-Fe [Argentina]. Holotype: Whereabouts unknown.

Sorubim latirostris Miranda Ribeiro, 1920: 14, pl. 12. Type locality: Amazonas [Brazil]. Holotype: MNRJ 631.

Distribution: Amazon, Orinoco, Paraná and Parnaíba River basin (Lundberg & Littmann, 2003).

Sorubim maniradii Littmann, Burr & Buitrago-Suárez, 2001

Sorubim maniradii Littmann, Burr & Buitrago-Suárez, 2001: 88, fig. 1. Type locality: Ecuador, Napo State, Río Yasuni, Río Napo drainage, 0°59'06"S 75°25'36"W. Holotype: FMNH 108814.

Distribution: Amazon River basin (Lundberg & Littmann, 2003).

Sorubim trigonocephalus Miranda Ribeiro, 1920

Sorubim trigonocephalus Miranda Ribeiro, 1920: 12, pls. 10, 11. Type locality: Porto Velho [Brazil]. Holotype: MNRJ 882.

Distribution: Madeira and Tapajós River basins, Brazil (Lundberg & Littmann, 2003).

SORUBIMICHTHYS Bleeker, 1862

Sorubimichthys Bleeker, 1862, in Bleeker, 1862–63: 10. Type species: *Sorubim jandia* Spix & Agassiz, 1829. Type

by original designation. Gender: Masculine.

Pteroglanis Eigenmann & Pearson, in Pearson, 1924: 9. Type species: *Pteroglanis manni* Eigenmann & Pearson, 1924. Type by monotypy. Gender: Masculine.

***Sorubimichthys planiceps* (Spix & Agassiz, 1829)**

Platystoma planiceps Spix & Agassiz, 1829: 25, pl. 12. Type locality: Brazil: in fluminae Amazonum, Solimoëns, Rio Negro. Possible syntype or lectotype: MHNN 811(Kottelat, 1988).

Sorubim Pirauáca Spix, in Spix & Agassiz, 1829: pl. 12. Type locality: Brazil: in fluminae Amazonum, Solimoëns, Rio Negro. Possible holotype: MHNN 811(Kottelat, 1988). Name available from caption on plate; as *Platystoma planiceps* in text.

Sorubim jandia Spix & Agassiz, 1829: pl. 14. Type locality: equatorialis fluviis [Brazil]. Holotype: Whereabouts unknown (Kottelat, 1988). Name available from caption on plate; as *Platystoma spatula* in text.

Platystoma spatula Spix & Agassiz, 1829: 26, pl. 14. Type locality: equatorialis fluviis [Brazil]. Types: Whereabouts unknown (Kottelat, 1988).

Sorubimichthys ortoni Gill, 1870: 94. Type locality: Maranon, or Upper Amazon, and Napo Rivers. Holotype: USNM 8238.

Pteroglanis manni Eigenmann & Pearson, in Pearson, 1924: 9, pl. 1 (fig. 1). Type locality: Bolivia, Little Rio Negro, tributary to Rio Beni. Holotype: CAS 59623.

Distribution: Amazon and Orinoco River basins (Lundberg & Littmann, 2003).

Remarks: Lundberg *et al.* (1989) reviewed the nomenclature of this species. Agassiz (in Spix & Agassiz, 1831: conspectus) serves as first reviser in treating *Platystoma planiceps* as valid over *Sorubim Pirauáca*, and *Platystoma spatula* over *Sorubim jandia*. Precedence of *Platystoma planiceps* over *Platystoma spatula* may be by action of Lundberg *et al.* (1989).

***STEINDACHNERIDION* Eigenmann & Eigenmann, 1919**

Steindachneria Eigenmann & Eigenmann, 1888b: 137. Type species: *Steindachneria amblyurus* Eigenmann & Eigenmann, 1888. Type by original designation. Gender: Feminine. Preoccupied by *Steindachneria* Goode & Bean, 1888 (in fishes), replaced by *Steindachneridion* Eigenmann & Eigenmann, 1919.

Steindachneridion Eigenmann & Eigenmann, 1919: 525. Type species: *Steindachneria amblyurus* Eigenmann & Eigenmann, 1888. Type by being a replacement name. Gender: Neuter. Replacement for *Steindachneria* Eigenmann & Eigenmann, 1888.

Revision, with key to species: Garavello (2005).

***Steindachneridion amblyurum* (Eigenmann & Eigenmann, 1888)**

Steindachneria amblyurus Eigenmann & Eigenmann, 1888b: 137. Type locality: Jequitinhonha [Brazil]. Syntypes (2, 35–38 cm): MCZ 7324 (2).

Distribution: Jequitinhonha River basin, Brazil (Garavello, 2005).

***Steindachneridion doceana* (Eigenmann & Eigenmann, 1889)**

Steindachneria doceana Eigenmann & Eigenmann, 1889b: 30. Type locality: Rio Doce [Brazil]. Syntypes (9): MCZ 23792 (1), MCZ 23793 (1), MCZ 23794 (7); one syntype illustrated in Garavello (2005: 614, fig. 6).

Distribution: Doce River basin, Brazil (Garavello, 2005).

† *Steindachneridion iheringi* (Woodward, 1899)

† *Arius iheringi* Woodward, 1899: 64, pl. 2 (fig. 1), pl. 3 (fig. 2). Type locality: Taubaté, Estado do São Paulo, Brazil; Tertiary Lignite. Syntypes: BMNH P.9220 (skull), Museu Paulista (nearly complete specimen), Senckenberg Museum (partial specimen).

Distribution: Paraíba River basin, Estado do São Paulo, Brazil; Tertiary (Santos, 1973).

Remarks: Species assigned to the Pimelodidae and the genus *Steindachneridion* by Santos (1973).

***Steindachneridion melanodermatum* Garavello, 2005**

Steindachneridion melanodermatum Garavello, 2005: 618, fig. 9. Type locality: Brazil, Paraná State, municipality of Quedas do Iguaçu, rio Iguaçu, down river of the Salto Osório dam (approx. 25°35'S 53°05'W). Holotype: MZUSP 87983.

Distribution: Iguaçu River, upper Paraná River Basin, Brazil (Garavello, 2005).

Steindachneridion parahybae (Steindachner, 1877)

Platystoma Parahybae Steindachner, 1877b: 640, pl. 9. Type locality: Rio Parahyba bei Juiz de Fora; Rio Jequitinhonha [Brazil]. Syntypes: MCZ 25521 (1, skeleton), MCZ 7295 (1), MCZ 7323 (2), NMW (?).

Distribution: Paraíba do Sul River basin, Brazil (Garavello, 2005).

Remarks: Garavello (2005: 610) incorrectly reported the catalog number of one lot of the types as MCZ 73231.

Redescribed in Oliveira & de Moraes (1997).

Steindachneridion punctatum (Miranda Ribeiro, 1918)

Steindachneria scripta punctata Miranda Ribeiro, 1918d: 642. Type locality: Itaqui, Rio Grande do Sul [Brazil].

Holotype: MNRJ 1167. Originally *Steindachneria scripta* var. *punctata*.

Distribution: Upper Uruguay and Paraná River basin, Brazil (Garavello, 2005).

Steindachneridion scriptum (Miranda Ribeiro, 1918)

Steindachneria scripta Miranda Ribeiro, 1918d: 640. Type locality: Itaqui [Rio Grande do Sul, Brazil]. Lectotype: MZUSP 2286, designated by Britski (1969).

Distribution: Upper Paraná and Uruguay River basins, Brazil (Garavello, 2005).

† *Steindachneridion silvasantosi* Figueiredo & Costa-Carvalho, 1999

† *Steindachneridion silvasantosi* Figueiredo & Costa-Carvalho, 1999: 685, figs. 1–4. Type locality: Tremembé Formation, Lacustrine sedimentary deposit that outcrops in the mine Nossa Senhora da Guia, Tremembé county, city of Taubaté, São Paulo State [Brazil]; Tertiary (? Oligocene). Holotype: DGM 1291-P; partial skull, including neurocranium and suspensorium.

Distribution: Paraíba River basin, São Paulo State, Brazil; Tertiary (Figueiredo & Costa-Carvalho, 1999).

ZUNGARO Bleeker, 1858

Zungaro Bleeker, 1858b: 196. Type species: *Zungaro humboldtii* Bleeker [= *Pimelodus zungaro* Humboldt, 1821].

Type by absolute tautomy. Gender: Masculine.

Paulicea Ihering, 1898: 108. Type species: *Paulicea jahu* Ihering, 1898. Type by subsequent designation by Eigenmann (1910). Gender: Feminine.

Remarks: Generic synonymy based on Silfvergrip (1992: 306).

Zungaro jahu (Ihering, 1898)

Paulicea jahu Ihering, 1898: 108. Type locality: São Paulo [Brazil]. Types: Whereabouts unknown.

Distribution: Paraná-Paraguay River basin (Lundberg & Littmann, 2003).

Zungaro zungaro (Humboldt, 1821)

Pimelodus zungaro Humboldt, in Humboldt & Valenciennes, 1821: 170, pl. 46 (fig. 1). Type locality: Tomependa, Río Marañon [Amazon River basin, Peru]. Holotype: Whereabouts unknown.

Bagrus flavicans Castelnau, 1855: 31, pl. 13 (fig. 2). Type locality: Brazil, l'Amazone. Holotype: MNHN a-8823 (mounted).

Zungaro humboldtii Bleeker, 1858b: 207. Type locality: Tomedenda, Río Marañon [Amazon River basin, Peru]. Holotype: Whereabouts unknown. Unneeded replacement name for *Pimelodus zungaro*, presumably to avoid tautomy.

Platystoma Lütkeni Steindachner, 1876: 609, pl. 13. Type locality: Laufe des Amazonenstromes gefangen. Syntypes: at NMW.

Distribution: Amazon and Orinoco River basins (Lundberg & Littmann, 2003).

ZUNGPARIOPSIS Steindachner, 1908

Zungpariopsis Steindachner, 1908b: 67. Type species: *Zungpariopsis multimaculatus* Steindachner, 1908. Type by monotypy. Gender: Feminine.

Zungaropsis multimaculatus Steindachner, 1908

Zungaropsis multimaculatus Steindachner, 1908b: 67. Type locality: Rio Xingu. Holotype: NMW 43537.

Remarks: Considered to be a valid species, in a genus near to *Zungaro* by A. Silfvergrip (pers. commun., 2003).

***Species inquirenda*, Pimelodidae**

Pimelodus (Bagrus) maculatus Jardine, in Schomburgk, 1841: 175, fig. Type locality: Most rivers of Guyana, also Rio Negro and Amazon River. No types known. Preoccupied by *Pimelodus maculatus* La Cepède, 1803.

PLOTOSIDAE Bleeker, 1858

Plotosichthyoidei Bleeker, 1858b: 49, 310. Type genus: *Plotosus* La Cepède, 1803.

Taxonomic summaries: Taylor & Gomon (1986, Africa); Tilak (1970a, South Asia); Hoese & Hanley, in Paxton *et al.* (1989, Australia).

Reviews: Allen (1989, Australia); Allen (1991, New Guinea); Ferraris (1999a, western central Pacific).

Identification guide: Jayaram (1982, South Asia).

Remarks: Whitley (1940b: 407) noted that the names *Plotosus laticeps*, *Copidoglanis labiosus*, *Copidoglanis labrosus*, *Copidoglanis levis* and *Copidoglanis curtus*, all found in Saville-Kent (1889), are *nomina nuda* and therefore unavailable. The name *Plotosus flavolineatus* found in Whitley (1941a: 311) is also a *nomen nudum* and not available.

10 genera, 35 species; no named fossil taxa.

Incertae sedis

***Plotosus argenteus* Zietz, 1896**

Plotosus argenteus Zietz, 1896: 410, pl. 16 (fig. 7). Type locality: Barcoo River, Finke River at Hermannsburg and Idracwra, and in Plam Creek and Ilpilla Creek, [Central Australia]; [restricted to Cooper Creek, near Innamincka, (which is apparently the locality listed by Zeitz (1896:411) as Barcoo River) by lectotype designation]. Lectotype: SAMA F1090, designated by Unmack (2001: 57).

Distribution: Cooper Creek, Bulloo and Finke Rivers, internal drainage system, Australia (Allen, 1989).

Remarks: Treated in recent literature as valid either as *Neosilurus argenteus* or *Porochilus argenteus*.

***Copidoglanis rendahli* Whitley, 1928**

Copidoglanis obscurus Rendahl, 1922: 173, fig. 5. Type locality: Glencoe, Hermit Hill, nw. Australia. Lectotype: ZMUC J6341, designated by Pethon (1969: 5). Preoccupied by *Copidoglanis obscurus* Günther, 1864, replaced by *Copidoglanis rendahli* Whitley, 1928.

Copidoglanis rendahli Whitley, 1928: 214. Type locality: Glencoe, Hermit Hill, N.W. Australia. Lectotype: ZMUC J6341 designated by Pethon (1969: 5) for *Copidoglanis obscurus* Rendahl. Replacement for *Copidoglanis obscurus* Rendahl 1922; preoccupied by *Copidoglanis obscurus* Günther, 1864.

Distribution: Widely distributed in northern Australia in isolated localities, including Fitzroy, Ord, Jardin Rivers, and streams of Arnhem Land (Allen, 1989).

Remarks: Treated in recent literature as valid either as *Neosilurus rendahli* or *Porochilus rendahli*.

***ANODONTIGLANIS* Rendahl, 1922**

Anodontiglanis Rendahl, 1922: 168. Type species: *Anodontiglanis dahli* Rendahl, 1922. Type by original designation. Gender: Masculine.

***Anodontiglanis dahli* Rendahl, 1922**

Anodontiglanis dahli Rendahl, 1922: 169, figs. 2–3. Type locality: Glencoe, Northern Territory, Australia. Holotype: ZMUC J361.

Distribution: Northern Australia, including Fitzroy, Daly, East Alligator, Roper, Mitchell and Archer Rivers (Allen, 1989), in lentic and lotic freshwater habitats (Paxton *et al.*, 1989).

***CNIDOGLANIS* Günther, 1864**

Cnidoglanis Günther, 1864: 27. Type species: *Plotosus megastomus* Richardson, 1845. Type by subsequent designation by Jordan (1919b: 332). Gender: Masculine.

Choeroplotosus Kner, 1866: 545. *Choeroplotosus decemfilis* Kner, 1867. Type by subsequent monotypy. Name appeared first without an available species name. Also described as new in Kner (1867: 300). Gender: Masculine.

Neoplotosus Castelnau, 1875: 45. Type species: *Neoplotosus waterhousii* Castelnau, 1875. Type by monotypy. Gender: Masculine.

Ostophycephalus Ogilby, 1899: 155. Type species: *Ostophycephalus duriceps* Ogilby, 1899. Type by original designation. Gender: Masculine.

Remarks: Eschmeyer & Bailey (1990: 92) interpreted the availability of *Choeroplotosus* as follows: "In 1866: 545, Kner listed one species as, "*Choeroplotosus limbatus* (*Plotos. limbatus* ? C. V.)", but in 1867 (p. 300) he described the species as *Choeroplotosus decemfilis* n. sp., with "Syn.? an *Plotosus limbatus* C. V. Apparently the genus can date to Kner (1866: 545) with no definitely-included named species, with species added in 1867."

***Cnidoglanis macrocephalus* (Valenciennes, 1840)**

Plotosus macrocephalus Valenciennes, in Cuvier & Valenciennes, 1840b: 428 (317 in Strasbourg deluxe edition), pl. 449. Type locality: prise à Timor [in error, possibly for Tasmania, Australia]. Holotype: MNHN a-8921.

Plotosus megastomus Richardson, 1845, in Richardson, 1844–48: 31, pl. 21 (figs. 1–3). Type locality: Sydney Cove, Australia. Holotype: BMNH 1974.5.22.1.

Choeroplotosus decemfilis Kner, 1867: 300, pl. 12 (fig. 1 [as *Choeroplotosus limbatus*]). Type locality: Von Sidney. Syntypes (2): NMW 47098 (1), NMW 47100 (1).

Cnidoglanis bostockii Castelnau, 1873: 140. Type locality: Fremantle, Western Australia. Syntypes (2): whereabouts unknown.

Neoplotosus waterhousii Castelnau, 1875: 45. Type locality: Adelaide, South Australia. Holotype: MNHN a-9545.

Ostophycephalus duriceps Ogilby, 1899: 156. Type locality: St. Vincent's Gulf, South Australia. Holotype: SAMA F1093.

Distribution: Kirra, Queensland to Jervis Bay, New South Wales, and Kingston, South Australia to Houtman Abrolhos Islands, Western Australia and Duck River, Tasmania, in near-shore and reef habitats (Paxton *et al.*, 1989).

Remarks: Kowarsky (1976) discussed the valid name for, and distribution of, this species.

***EURISTHMUS* Ogilby, 1899**

Euristhmus Ogilby, 1899: 154. Type species: *Plotosus elongatus* Castelnau, 1878. Type by original designation. Gender: Masculine.

Exilichthys Whitley, 1933: 65. Type species: *Cnidoglanis nudiceps* Günther, 1880. Type by original designation. Gender: Masculine.

***Euristhmus lepturus* (Günther, 1864)**

Cnidoglanis lepturus Günther, 1864: 28. Type locality: Sydney, New South Wales [Australia]. Syntypes (2, one stuffed): BMNH 1864.1.17.33 (1).

Plotosus elongatus Castelnau, 1878a: 237. Type locality: Brisbane River [Queensland, Australia]. Holotype: Whereabouts unknown [possibly MNHN a-2783].

Distribution: Exmouth Gulf, Western Australia to Sydney, New South Wales, and New Guinea, in freshwater, estuarine and near-shore marine habitats (Paxton *et al.*, 1989; Allen, 1997: 60).

***Euristhmus microceps* (Richardson, 1845)**

Plotosus microceps Richardson, 1845, in Richardson, 1844–48: 31, pl. 21 (figs. 4–7). Type locality: North-west coast of Australia. Holotype: BMNH 1846.3.3.2.

Cnidoglanis microcephalus Günther, 1864: 28. Type locality: North-west coast of Australia. Holotype: BMNH 1846.3.3.2. Unjustified emendation of *Plotosus microceps* Richardson, 1845.

Distribution: Shark Bay to Broome, Western Australia, in near-shore habitats over soft bottoms (Paxton *et al.*, 1989).

***Euristhmus nudiceps* (Günther, 1880)**

Cnidoglanis nudiceps Günther, 1880b: 49. Type locality: Arafura Sea. Holotype or lectotype: BMNH 1879.5.14.590; illustrated in Weber & de Beaufort (1913: 232).

Distribution: Shark Bay, Western Australia to Brisbane Queensland, Australia, in near-shore habitats over soft bottoms (Paxton *et al.*, 1989); also New Guinea (Allen, 1997: 60).

Remarks: Paxton *et al.* (1989: 223) list a holotype for *Cnidoglanis nudiceps*, but the Natural History Museum in London lists a second specimen (BMNH 1890.2.26.165) from the Challenger Expedition taken in the Arafura Sea that may also have been examined by Günther. There is no evidence in the original description whether more than one specimen was examined. Therefore, even if the second specimen was originally a syntype, it would have been rendered a paralectotype by the de facto lectotype designation in Paxton *et al.* (1989).

NEOSILUROIDES Allen & Feinberg, 1998

Neosiluroides Allen & Feinberg, 1998: 11. *Neosiluroides cooperensis* Allen & Feinberg, 1998. Type by original designation. Gender: Masculine.

Neosiluroides cooperensis Allen & Feinberg, 1998

Neosiluroides cooperensis Allen & Feinberg, 1998: 12, fig. 2. Type locality: Callamurra Waterhole, Coopers Creek, 11.3 km east of Innamincka, South Australia, ca. 27°41'S, 140°51'E. Holotype: AMS I.18699-001.

Distribution: Coopers Creek system, Lake Eyre drainage (Allen & Feinberg, 1998).

NEOSILURUS Steindachner, 1867

Neosilurus Steindachner, 1867a: 7. Type species: *Neosilurus hyrtlii* Steindachner, 1867. Type by monotypy. Gender: Masculine. Also appeared as new in Steindachner (1867d: 11).

Neosilurus Castelnau, 1878a: 238. Type species: *Neosilurus australis* Castelnau, 1878. Type by monotypy. Gender: Masculine. Preoccupied by *Neosilurus* Steindachner, 1867, in fishes; replaced by *Cainosilurus* Macleay, 1881.

Eumeda Castelnau, 1878b: 143. Type species: *Eumeda elongata* Castelnau, 1878. Type by monotypy. Gender: Feminine.

Cainosilurus Macleay, 1881: 211. Type species: *Neosilurus australis* Castelnau, 1878. Type by being a replacement name. Gender: Masculine. Replacement for *Neosilurus* Castelnau, 1878; preoccupied by *Neosilurus* Steindachner, 1867.

Lambertia Perugia, 1894: 550. Type species: *Lambertia atra* Perugia, 1894. Type by monotypy. Gender: Feminine. Preoccupied by *Lambertia* Robineau-Desvoidy, 1863, in Diptera; replaced by *Lambertichthys* Whitley, 1938.

Anyperistius Ogilby, 1908: 3, 11. Type species: *Anyperistius perugiae* Ogilby, 1908. Type by original designation. Gender: Masculine. Spelled two ways originally: *Anyperistius* and *Anperisteus*. First reviser apparently Eschmeyer & Bailey (in Eschmeyer, 1990) who selected *Anyperistius*.

Lambertichthys Whitley, 1938: 223. Type species: *Lambertia atra* Perugia, 1894. Type by being a replacement name. Gender: Masculine. Replacement for *Lambertia* Perugia, 1894; preoccupied by *Lambertia* Robineau-Desvoidy, 1863, in Diptera.

Neosilurus ater (Perugia, 1894)

Lambertia atra Perugia, 1894: 551. Type locality: Inawi, Papuasia orientale. Syntypes: MSNG 1663 (1), MSNG 8140 (1, missing), ZMA 113360 (1).

Neosilurus mediobarbis Ogilby, 1908: 12. Type locality: ? Queensland, Australia. Holotype: QM (lost: Paxton *et al.*, 1989).

Lambertichthys ater sepikensis Whitley, 1956b: 68. Type locality: Upper Sepik River, New Guinea. Holotype: AMS IA.7278; illustrated in Whitley (1956a: 44, fig. 4).

Distribution: Northern Australia and central-southern New Guinea; in freshwater (Allen, 1989).

Neosilurus brevidorsalis (Günther, 1867)

Copidoglanis brevidorsalis Günther, 1867a: 66. Type locality: Nicol Bay, Cape York, Queensland, Australia. Holotype: BMNH 1867.5.13.11.

Anyperistius perugiae Ogilby, 1908: 11. Type locality: Inawe, St. Joseph River, New Guinea. Type(s): at MSNG and/or BMNH 1965.1.26.1 (1). Based on specimen(s) identified by Perugia (1894: 552) as *Eumeda elongata* (not of Castelnau).

Neosilurus bartoni Regan, 1908c: 153. Type locality: Sogeri, New Guinea. Syntypes: BMNH 1905.8.15.9–10 (2). Distribution: Cape York Peninsula, northern Australia, and central-southern New Guinea; in streams, turbid backwaters, and lagoons (Allen, 1989).

Remarks: BMNH 1965.1.26.1 was sent to BMNH on exchange from MSNG and is apparently a type of *Anyperistius*

perugiae Ogilby, 1908.

***Neosilurus coatesi* (Allen, 1985)**

Tandanus coatesi Allen, 1985: 252, fig. 3. Type locality: Ninar River, ca. 6 km west of Maprik, Papua New Guinea, ca. 3°37'S, 143°00'E. Holotype: WAM P.27839-007.

Distribution: Sepik River basin, in streams of Torracelli Mountains (Allen *et al.*, 1992).

***Neosilurus equinus* (Weber, 1913)**

Copidoglanis equinus Weber, 1913: 527, fig. 11. Type locality: Lorentz-Fluss [New Guinea]. Syntypes (13): ZMA 111105 (7), ZMA 111106 (1), ZMA 111107 (4), ZMA 111108 (2).

Distribution: Southern Papua (formerly Irian Jaya), New Guinea (Allen, 1996).

***Neosilurus gjellerupi* (Weber, 1913)**

Copidoglanis gjellerupi Weber, 1913: 528. Type locality: Kaiserin Augusta-fluss [New Guinea]. Holotype: ZMA 111091.

Distribution: Ramu River basin, Papua New Guinea (Allen *et al.*, 1992) and northern Papua (formerly Irian Jaya), New Guinea (Allen, 1996).

Remarks: The name *Copidoglanis papuensis* that is found in the caption of figures 11 and 12 in Hase (1914: 540) appears to have been published accidentally inasmuch as the species account indicates that the specimens were regarded as having been identified as *Copidoglanis gjellerupi* (Weber, 1913). This was treated by Eschmeyer *et al.* (1998: 1283) as unavailable, because it was not treated as the valid name of a species when proposed.

***Neosilurus gloveri* Allen & Feinberg, 1998**

Neosilurus gloveri Allen & Feinberg, 1998: 13, fig. 3. Type locality: Main Spring, Dalhousie Springs, South Australia, ca. 26°25'S, 135°30'E. Holotype: SAMA F.4159.

Distribution: Dalhousie artesian springs, Finke River basin, southern Australia (Allen & Feinberg, 1998).

***Neosilurus hyrtlui* Steindachner, 1867**

Neosilurus Hyrtlui Steindachner, 1867d: 14, pl. 1 (figs. 3–3a). Type locality: Fitzroy-Flusse bei Rockhampton, Ost-Australien. Syntypes: NMW 45340 (5), NMW 45341 (2).

Silurichthys australis Castelnau, 1875: 45. Type locality: Cape York [Queensland, Australia]. Holotype: Whereabouts unknown.

Neosilurus australis Castelnau, 1878a: 239. Type locality: Freshwater lagoons of Rockhampton, Queensland, Australia. Holotype: Whereabouts unknown.

Eumeda elongata Castelnau, 1878b: 144. Type locality: Brisbane River, Rockhampton [Queensland, Australia]. Holotype: MNHN a-2173.

Neosilurus robustus Ogilby, 1908: 13. Type locality: Keppel Bay, Queensland, Australia. Holotype: Whereabouts unknown, not at QM.

Copidoglanis glencoensis Rendahl, 1922: 170, fig. 4. Type locality: Glencoe R., nw. Australia. Lectotype: ZMUO J5254; Lectotype designated by Pethon (1969: 4).

Neosilurus mortoni Whitley, 1941b: 7, fig. 7. Type locality: Yam Creek (nine miles from Brock's Creek, railway line south from Darwin) Northern Territory of Australia. Holotype: AMS IA.4824.

Distribution: Widely distributed in northern Australia; in freshwater (Allen, 1989).

***Neosilurus idenburgi* (Nichols, 1940)**

Copidoglanis idenburgi Nichols, 1940: 1. Type locality: Bernhard Camp (altitude 75 meters), Idenburg River, Netherland New Guinea. Holotype: AMNH 15034.

Distribution: Ramu River basin, Papua New Guinea (Allen, 1992) and northern Papua (formerly Irian Jaya), New Guinea (Allen, 1996).

***Neosilurus mollespiculum* Allen & Feinberg, 1998**

Neosilurus mollespiculum Allen & Feinberg, 1998: 16, fig. 6. Type locality: Running River, Burdekin River system, Queensland, ca. 19°07'S, 145°50'E, Australia. Holotype: QM I.30685.

Distribution: Isdell, Carson, Drysdale, Katherine, Fergusson, and Burdekin Rivers, northern Australia (Allen & Feinberg, 1998).

***Neosilurus novae-guineae* (Weber, 1907)**

Copidoglanis novae-guineae Weber, 1907: 226. Type locality: Sentani-See, Nord-Neu-Guinea. Syntypes (13):

FMNH 52388 (1), NMW 46696 (1), RMNH 7980 (1), ZMA 112670 (8).

Copidoglanis novae-guineae niger Nichols, 1940: 1. Type locality: Bernhard Camp (altitude 75 meters), Idenburg River, Netherland New Guinea. Holotype: AMNH 15035.

Distribution: Ramu River basin, Papua New Guinea (Allen *et al.*, 1992) and northern Papua (formerly Irian Jaya), New Guinea (Allen, 1996).

***Neosilurus pseudospinosus* Allen & Feinberg, 1998**

Neosilurus pseudospinosus Allen & Feinberg, 1998: 15, figs. 4–5. Type locality: Rocky pool of Ord R. on Old Lissadel Station, Kimberley Dist., Western Australia, ca. 16°40'S, 128°83'E. Holotype: WAM P.28505-007.

Distribution: Ord River, Western Australia (Allen & Feinberg, 1998).

***OLOPLOTOSUS* Weber, 1913**

Oloplotosus Weber, 1913: 521 Type species: *Oloplotosus mariae* Weber, 1913. Type by monotypy. Gender: Masculine.

Key: Allen (1985; 1991).

***Oloplotosus luteus* Gomon & Roberts, 1978**

Oloplotosus luteus Gomon & Roberts, in Roberts, 1978: 47, fig. 25. Type locality: Shallow, turbid backwater of Ok Tedi and lowermost half km of small tributary about 16 km NE of Nigerum, 915 km upriver from Toro Pass, 5°34.1'S, 141°15.0'E; Fly River, Papua New Guinea. Holotype: AMS I.27093-001.

Distribution: Upper Fly River, New Guinea (Roberts, 1978).

***Oloplotosus mariae* Weber, 1913**

Oloplotosus Mariae Weber, 1913: 522, fig. 3. Type locality: Lorentz-Fluss, Sabang; Lorentz-Fluss, Alkmaar [New Guinea]. Syntypes (5): AMNH 9500 (1), ZMA 111103 (1), ZMA 111104 (2).

Distribution: Southern Papua (formerly Irian Jaya), New Guinea (Allen, 1996).

***Oloplotosus torobo* Allen, 1985**

Oloplotosus torobo Allen, 1985: 248, fig. 1. Type locality: Soro River at n. end of Lake Kutubu, ca. 4 km east of Moro landing strip, Papua New Guinea, ca. 6°23'S, 143°15'E, 0–3 m. Holotype: WAM P.28158-003.

Distribution: Soro River, Papua New Guinea (Allen, 1985).

***PARAPLOTOSUS* Bleeker, 1863**

Paraplotosus Bleeker, 1863 (in Bleeker, 1862–63): 100. Type species: *Plotosus albilabris* Valenciennes, 1840. Type by monotypy. Gender: Masculine.

Endorrhysis Ogilby, 1898b: 283. Type species: *Copidoglanis longifilis* Macleay, 1881. Type by original designation. Gender: Feminine.

Revision: Allen (1998).

***Paraplotosus albilabris* (Valenciennes, 1840)**

Plotosus albilabris Valenciennes, in Cuvier & Valenciennes, 1840b: 427 (316 in Strasbourg deluxe edition). Type locality: Batavia. Holotype: MNHN a-9544.

Plotosus macropthalmus Bleeker, 1846a: 179. Type locality: Batavia. Syntypes (size and number not stated): Whereabouts unknown.

Copidoglanis longifilis Macleay, 1881: 207. Type locality: Long Island, Torres Strait, Queensland, Australia. Syntypes: AMS I.16269-001 (2).

Distribution: Indo-Australian Archipelago, New Guinea and Australia; often in coral reef habitats (Allen, 1998).

***Paraplotosus butleri* Allen, 1998**

Paraplotosus butleri Allen, 1998: 129, fig. 3. Type locality: Near Broome, ca. 17°58'S, 122°14'E, Western Australia. Holotype: WAM P.27368-021.

Distribution: Northern Australia, in coastal reefs (Allen, 1998).

***Paraplotosus muelleri* (Klunzinger, 1880)**

Cnidoglanis mülleri Klunzinger, 1880: 411. Type locality: Port Darwin [Australia]. Holotype: SMNS 2519.

Distribution: Dampier Archipelago to the eastern Gulf of Carpentaria, northern Australia; in turbid coastal reefs (Allen, 1998).

PLOTOSUS La Cepède, 1803

Plotosus La Cepède, 1803: 129. Type species: *Platystacus anguillaris* Bloch, 1794. Type by monotypy. Gender: Masculine.

Deportator Gistel, 1848: X. Type species: *Platystacus anguillaris* Bloch, 1794. Type by being a replacement name. Gender: Masculine. Unneeded replacement for *Plotosus* La Cepède, 1803, which was considered by Gistel to be preoccupied by *Plotus* Linnaeus, 1766, in Aves.

Remarks: *Plotosis* (in Duméril, 1856: 340), *Plotoseus* (in Lesson, 1826, in Lesson 1826–31: pl. 31, 1831: 435) and *Plotosius* (in Siebold, 1846: 228) are incorrect subsequent spellings of *Plotosus* and not available names.

Key: Gomon & Taylor (1982).

Plotosus canius Hamilton, 1822

Plotosus canius Hamilton, 1822: 142, 374, pl. 15 (fig. 44). Type locality: rivers of the southern parts of Bengal. No types known.

Plotosus unicolor Valenciennes, in Cuvier & Valenciennes, 1840b: 426 (316 in Strasbourg deluxe edition). Type locality: Java. Holotype: MNHN a-8924.

Plotosus viviparus Bleeker, 1846a: 182. Type locality: Batavia. Syntypes (size and number not stated): possibly RMNH 8066 (1), RMNH 15875 (6), SMNS 10624 (1).

Plotosus horridus Bleeker, 1846a: 183. Type locality: Batavia. Type(s) (size and number not stated): Whereabouts unknown.

Plotosus multiradiatus Bleeker, 1846a: 183. Type locality: Batavia. Type (s) (size and number not stated): Whereabouts unknown.

Distribution: Coastal regions of Thailand, Sundaland, Sulawesi, Moluccas, and India, and into lower Mekong River (Kottelat, 2001b).

Remarks: Synonymy based on Bleeker (1862–63: 99). *Plotosus caesius* in Cuvier (1829, 1836, etc.) and Hyrtl (1859: 5, 6, 17) are considered to be incorrect spellings for *P. canius* and not the proposal of an available name. *Silurus unitius* of Hora (1933: 133) is not available, as it was not treated as a valid species.

Plotosus fisadoha Ng & Sparks, 2002

Plotosus fisadoha Ng & Sparks, 2002: 565, fig. 1. Type locality: Madagascar: southeastern coast of Fanarantsoa Province; Farafangana market. Holotype: UMMZ 235269.

Distribution: Southeastern Madagascar (Ng & Sparks, 2002).

Plotosus limbatus Valenciennes, 1840

Plotosus limbatus Valenciennes in Cuvier & Valenciennes, 1840b: 422 (313 in Strasbourg deluxe edition). Type locality: Côte de Malabar ... Pondichéry; [restricted to Malabar by lectotype designation]. Lectotype: MNHN a-9546, designated by Gomon & Taylor (1982: 9).

Copidoglanis obscurus Günther, 1864: 26. Type locality: Probably from Australia. Holotype: BMNH 1852.9.13.164.

Distribution: Western Indian Ocean and Arabian Sea, in marine and brackish waters (Taylor & Gomon, 1986).

Plotosus lineatus (Thunberg, 1787)

Silurus lineatus Thunberg, 1787: 31, footnote 13. Type locality: [East Indian Seas]. Type(s): ZMUC P 28555. Described in more detail and illustrated in Thunberg (1791: 191, pl. 6).

Platystacus anguillaris Bloch, 1794: 61, pl. 373 (figs. 1, 2). Type locality: Not stated [given as Tranquebar in Bloch & Schneider, 1801]. Lectotype: ZMB 3078, designated by Taylor & Gomon (1986: 160).

Plotosus thunbergianus La Cepède, 1803: 693, 694. Type locality: La mer des grandes Indes. Holotype: ZMUC P 28555. Unneeded new name for *Silurus lineatus* Thunberg, 1787.

Plotoseus ikapor Lesson, 1826, in Lesson 1826–31: pl. 31, fig. 3. Type locality: baie d'Offeac'h de l'ile de Waigou. Holotype: Whereabouts unknown. Name made available by figure caption, with illustrated specimen the holotype; described in Lesson (1831: 132).

Plotosus marginatus Bennett, 1830: 691. Type locality: Sumatra. Syntype: BMNH 1855.12.26.452 (1).

Plotosus vittatus Swainson, 1839: 307. Type locality: Not stated [given as Tranquebar in Bloch & Schneider, 1801]. Unneeded new name for *Platystacus anguillaris* Bloch. Based on "Bl. 373, fig. 1" [= Bloch (1794: pl. 373 (fig. 1)].

Plotosus castaneus Valenciennes, in Cuvier & Valenciennes, 1840b: 421 (312 in Strasbourg deluxe edition). Type locality: Mahé sur la côte malabare. Holotype: MNHN a-8929.

Plotosus lineatus Valenciennes in Cuvier & Valenciennes, 1840b: 412 (306 in the Strasbourg deluxe edition). Type locality: la mer des Indes ... la mer Rouge ... la Séchelles ... la côte de Malabar, Trincomalé, Pondichéry, Amboine ... Célèbes, îles des Amis, îles de la Société, Macao ... Philippines. Syntypes: MNHN a-8930 (1), MNHN a-8931 (2), MNHN a-8932 (10 or 11), MNHN a-8936 (1). Preoccupied in *Plotosus* by *Silurus lineatus* Thunberg, 1787.

Plotosus castaneoides Bleeker, 1851f: 490. Type locality: Rio [=Riouw], in mari. Syntypes (2, 160–170 mm TL): at RMNH.

Plotosus arab Bleeker, 1862a: 111. Type locality: Not stated [given as Tranquebar in Bloch & Schneider, 1801]. Syntypes: at MNHN, ZMB 3078 (1), ZMB 3079 (1). Name made available by reference to *Plotosus anguillaris* of La Cepède (1803: 130, pl. 3 fig. 2), which is based on specimens and *Platystacus anguillaris* Bloch, 1794.

Plotosus brevibarbus Bessednov, 1967: 446, fig. 3. Type locality: 21°10' c. щ, 108°30'в. д, 29 м [Gulf of Tonkin, South China Sea]. Holotype: Mus. TINRO 3667.

Distribution: Indian Ocean and Western Pacific, in marine waters, but sometimes entering freshwaters in East Africa and Madagascar (Taylor & Gomon, 1986); eastern Mediterranean (Golani, 2002).

Remarks: The name *Silurus Arab* in Forsskål (1775: xvi) is not available (M. Kottelat, pers. commun.). The name *Plotosus malignus* appears in the account of *Plotosus lineatus* Valenciennes, but it not treated as valid and is therefore not available from that publication. The name does not appear to have been treated as valid subsequently. See Smith (1941) for nomenclatural comments regarding this species.

***Plotosus nkunga* Gomon & Taylor, 1982**

Plotosus nkunga Gomon & Taylor, 1982: 3, pl. 1. Type locality: South Africa, Transkei, Bashee River (32°15'S, 28°55'E). Holotype: SAM 18457.

Distribution: Southern Africa from Boknes to Boteler Point, and possibly to Zanzibar, in marine waters but also entering freshwater (Taylor & Gomon, 1986).

***Plotosus papuensis* Weber, 1910**

Plotosus papuensis Weber, 1910: 228. Type locality: Lorentz-Fluss [New Guinea]. Syntypes: FMNH 52494 (1), ZMA 111093–97 (5), ZMA 111098–100 (5), ZMA 111513 (1).

Distribution: Southern Papua (formerly Irian Jaya), New Guinea (Allen, 1996).

Species inquirenda, Plotosus

Plotosus abbreviatus Boulenger, 1895a: 247. Type locality: Mouth of Baram River, Sarawak. Holotype: BMNH 1894.8.3.35.

***POROCHELIUS* Weber, 1913**

Porochilus Weber, 1913: 523. Type species: *Porochilus obbesi* Weber, 1913. Type by monotypy. Gender: Masculine.

***Porochilus meraukensis* (Weber, 1913)**

Copidoglanis meraukensis Weber, 1913: 529. Type locality: Merauke, Süd-Neu Guinea. Syntypes: RMNH 10865 (1), ZMA 111092 (1).

Distribution: Southern Papua (formerly Irian Jaya), New Guinea (Allen, 1996).

***Porochilus obbesi* Weber, 1913**

Porochilus Obbesi Weber, 1913: 523, figs. 4–5. Type locality: Lorentz-Fluss, ... bei Sabang, New Guinea. Syntypes (6): ZMA 111101 (3), ZMA 111102 (1), RMNH 10864 (1).

Distribution: Southern Papua (formerly Irian Jaya), New Guinea (Allen, 1996).

***TANDANUS* Mitchell, 1838**

Tandanus Mitchell, 1838: 95. Type species: *Plotosus (Tandanus) tandanus* Mitchell, 1838. Type by monotypy. Gender: Masculine. Originally proposed as a subgenus of *Plotosus*.

Copidoglanis Günther, 1864: 25. Type species: *Plotosus (Tandanus) tandanus* Mitchell, 1838. Type by subsequent

designation by Jordan (1919b: 332) with name was misprinted *Copiodoglanis*. Gender: Masculine.

Tandanus bostocki Whitley, 1944

Plotosus unicolor Castelnau, 1873: 141. Type locality: Interior of Western Australia. Syntype: NMV 51851 (1). Preoccupied by *Plotosus unicolor* Valenciennes, 1840, replaced by *Tandanus bostocki* Whitley, 1944.

Tandanus bostocki Whitley, 1944: 260. Type locality: Interior of Western Australia. Syntype: NMV 51851 (1). Replacement for *Plotosus unicolor* Castelnau, 1873, preoccupied by *Plotosus unicolor* Valenciennes, 1840.

Remarks: The statement in Whitley (1944: 260) that designates a holotype for the species is in error, since he clearly states elsewhere that *Tandanus bostocki* is intended to replace the preoccupied name of Castelnau and, thereby, takes the same specimen as holotype.

Distribution: Coastal drainages of southwestern Australia, from the Frankland River to Moore River (Allen, 1989).

Tandanus tandanus (Mitchell, 1838)

Plotosus (Tandanus) tandanus Mitchell, 1838: 44, 95, pl. 5 (fig. 2). Type locality: Lagoon near Tangulda, Namoi River, New South Wales; river between Gwydir River and MacIntyre River, New South Wales. No types known.

Distribution: Murray-Darling River system and coastal drainages of eastern Australia (Allen, 1989).

Remarks: See Musyl & Keenan (1996) for comments on populational variation that may represent unrecognized additional species.

Species inquirenda, Plotosidae

Plotosus nigricans Valenciennes, in Cuvier & Valenciennes, 1840b: 412, 417 (310 in Strasbourg deluxe edition). Type locality: Not stated. Holotype: at MNHN (skeleton). Species name mentioned in generic account as well as account of *Plotosus lineatus*. Reference made to published comments about the skeleton by Cuvier in Regne Animal, which are adequate to act as an indication, even though Valenciennes never provides a more complete treatment of the species. Gomon & Taylor (1982: 10), disagree and treated the name as a *nomen nudum*, but later argue that a specimen previously treated as the holotype of the species, MNHN a-9433, must not be considered as one of the “original type-specimens.”

PSEUDOPIMELODIDAE Lundberg, Bornbusch & Mago-Leccia, 1991

Pseudopimelodinae Lundberg, Bornbusch & Mago-Leccia, 1991: 204. Type genus: *Pseudopimelodus* Bleeker, 1858.

Remarks: The name Pseudopimelodidae was proposed first by Fernández-Yépez & Martín Salazar (1953) in a manner that made the name available when published, but was rendered unavailable in Art. 13 of the Second Edition of the Code (ICZN, 1961). The name apparently remains unavailable, despite Art. 13.2.1 of the current version of the Code (ICZN, 1999), because the name was treated as unavailable in Ferraris & de Pinna (1999: 8).

Taxonomic summary: Shibatta (2003a).

Phylogeny: Shibatta (1998, 2003b).

5 genera, 29 species; no named fossil taxa.

BATROCHOGLANIS Gill, 1858

Batrachoglanis Gill, 1858: 389. Type species: *Pimelodus raninus* Valenciennes, 1840. Type by original designation. Gender: Masculine.

Remarks: *Batrachoglanis*, first used in Gill (1861c), is either a misspelling or unjustified emendation of *Batrachoglanis* Gill, 1858.

Batrachoglanis acanthochiroides (Güntert, 1942)

Pseudopimelodus acanthochiroides Güntert, 1942: 29. Type locality: Santander, Colombia. Syntypes: NMBA 5277–78 (2).

Pseudopimelodus villosus butcheri Schultz, 1944c: 199, pl. 1 (fig. b). Type locality: Río San Juan near bridge south of Mene Grande, tributary to Rio Motatán, Maracaibo basin, Venezuela. Holotype: USNM 121270.

Distribution: Catatumbo River basin in Lake Maracaibo drainage (Shibatta, 2003a).

Batrochoglanis melanurus Shibatta & Pavanelli, 2005

Batrochoglanis melanurus Shibatta & Pavanelli, 2005: 23, figs. 1–2. Type locality: Córrego Cancela, affluent of rio Cuiabá, rio Paraguai basin; 14°42.501S/ 56°15.850W; Nobres; State of Mato Grosso; Brazil. Holotype: MZUSP 87240.

Distribution: Cuiabá River, Paraguay River basin, Mato Grosso. Brazil (Shibatta & Pavanelli, 2005).

Batrochoglanis raninus (Valenciennes, 1840)

Pimelodus raninus Valenciennes, in Cuvier & Valenciennes, 1840b: 157 (117 in the Strasbourg deluxe edition), pl. 434. Type locality: La Mana ... Rio-Janéiro. Syntypes: MNHN a-9942 (3).

Pseudopimelodus acanthochirius Eigenmann & Eigenmann, 1888b: 122. Type locality: Brazil. Syntypes (5): MCZ 8133 (1), MCZ 8157 (1), MCZ 7732 (2), MCZ 7332 (1).

Distribution: Amazon River basin (Shibatta, 2003a).

Batrochoglanis transmontanus (Regan, 1913)

Pseudopimelodus transmontanus Regan, 1913d: 467. Type locality: Condoto ..., San Juan and the Tamana, and the Durango, W. Ecuador. Syntypes (9): BMNH 1910.7.11.104, BMNH 1910.7.11.105, BMNH 1913.10.1.36, BMNH 1902.5.27.37–40.

Distribution: Baudó, San Juan, Patia and Durango River basins, Pacific slope drainages (Shibatta, 2003a).

Batrochoglanis villosus (Eigenmann, 1912)

Pseudopimelodus villosus Eigenmann, 1912b: 152, fig. 32, pl. 10 (fig. 1). Type locality: Potaro Landing, British Guiana. Holotype: FMNH 53219.

Distribution: Demerara, Essequibo, Orinoco and Amazon River basins (Shibatta, 2003a).

CEPHALOSILURUS Haseman, 1911

Cephalosilurus Haseman, 1911a: 317. Type species: *Cephalosilurus fowleri* Haseman, 1911. Type by monotypy. Gender: Masculine.

Cephalosilurus albomarginatus (Eigenmann, 1912)

Pseudopimelodus albomarginatus Eigenmann, 1912b: 153, pl. 11 (fig. 1). Type locality: Tukeit, British Guiana. Holotype: FMNH 53221.

Distribution: Tukeit, Guyana (Shibatta, 2003a).

Cephalosilurus apurensis (Mees, 1978)

Pseudopimelodus apurensis Mees, 1978b: 253, pl. 1. Type locality: Río Arichuna, near San Pedro, Rincón hondo, Apure, Venezuela. Holotype: RMNH 27644.

Distribution: Arichuna River basin, Apure State, Venezuela (Shibatta, 2003a).

Cephalosilurus fowleri Haseman, 1911

Cephalosilurus fowleri Haseman, 1911a: 317, pl. 46. Type locality: Rio São Francisco, Cidade de Barra, Bahia, Brazil. Holotype: FMNH 54254.

Distribution: São Francisco River basin, Brazil (Shibatta, 2003a).

Cephalosilurus nigricaudus (Mees, 1974)

Pseudopimelodus nigricauda Mees, 1974: 218, figs. 37d, 38d; pls. 11–12. Type locality: Sipaliwini, Suriname. Holotype: RMNH 26739.

Distribution: Sipaliwini River basin, Suriname (Shibatta, 2003a).

LOPHIOSILURUS Steindachner, 1877

Lophiosilurus Steindachner, 1877a: 154. Type species: *Lophiosilurus alexandri* Steindachner, 1877. Type by monotypy. Gender: Masculine.

Lophiosilurus alexandri Steindachner, 1877

Lophiosilurus Alexandri Steindachner, 1877a: 154, pl. 15. Type locality: Aus Brasilien, wahrscheinlich aus dem Amazonenstrome. Holotype: NMW 46123.

Pseudopimelodus Agassizi Steindachner, 1880b: 61. Type locality: Aus Brasilien, wahrscheinlich aus dem Amazonenstrome. Holotype: NMW 46123. Unneeded new name, or *lapsus calami* for *Lophiosilurus alexandri* Steindachner, 1877, originally written as *Pseudop. Agassizi*.

Distribution: São Francisco River basin, Brazil (Shibatta, 2003a).

Remarks: See Pinto & Marzulo (1975) for comments on morphology.

***MICROGLANIS* Eigenmann, 1912**

Microglanis Eigenmann, 1912b: 130, 155. Type species: *Microglanis poecilus* Eigenmann, 1912. Type by original designation. Gender: Masculine.

Key: Bertaco & Cardoso (2005), and Shibatta & Benine (2005), southern Brazil.

***Microglanis ater* Ahl, 1936**

Microglanis ater Ahl, 1936: 109. Type locality: Brazil. Holotype: ZMB 20932.

Distribution: Brazil (Shibatta, 2003a).

***Microglanis cibelae* Malabarba & Mahler, 1998**

Microglanis cibelae Malabarba & Mahler, 1998: 249, figs. 1, 4–6. Type locality: arroio do Ouro, Rio Maquiné, between Maquiné and Barra do Ouro, Rio Grande do Sul, Brasil. Holotype: MZUSP 48653.

Distribution: Coastal drainages in northern Rio Grande do Sul and Santa Catarina States, Brazil (Shibatta, 2003a; Bertaco & Cardoso, 2005).

***Microglanis cottooides* (Boulenger, 1891)**

Pimelodus (Pseudopimelodus) cottooides Boulenger, 1891: 233, pl. 25 (fig. 2). Type locality: Rio Camaquã, Rio Grande do Sul, Brazil. Syntypes: BMNH 1891.3.16.36–45 and BMNH 1891.3.16.106 (lots combined, 12 specimens) and BMNH 1889.8.24.5 (1).

Distribution: Laguna dos Patos and Uruguay River basins, Brazil (Shibatta, 2003a; Bertaco & Cardoso, 2005).

***Microglanis eurystoma* Malabarba & Mahler, 1998**

Microglanis eurystoma Malabarba & Mahler, 1998: 248, fig. 3. Type locality: Rio Uruguay, Itá, Santa Catarina, Brazil. Holotype: MCP 13405.

Distribution: Upper Uruguay River basin, Brazil (Shibatta, 2003a; Bertaco & Cardoso, 2005).

***Microglanis garavelloii* Shibatta & Benine, 2005**

Microglanis garavelloii Shibatta & Benine, 2005: 580, figs. 1–2. Type locality: Brazil, Paraná: Taquari stream, 23°12'24"S 50°56'50"W, Município de Jataizinho. Holotype: MZUSP 88006.

Distribution: Paranapanema and Tietê River basins, Brazil (Shibatta & Benine, 2005).

***Microglanis iheringi* Gomes, 1946**

Microglanis iheringi Gomes, 1946: 9, pl. 1. Type locality: Río Turmero, Aragua, Venezuela. Holotype: FMNH 35350.

Distribution: Turmero River basin, Venezuela (Shibatta, 2003a).

***Microglanis malabarbai* Bertaco & Cardoso, 2005**

Microglanis malabarbai Bertaco & Cardoso, 2005: 62, fig. 2. Type locality: arroio Albino, tributary of rio Ijuí (rio Uruguay drainage), 28°08'10"S, 54°55'28"W, São Pedro do Butiá, Rio Grande do Sul, Brazil. Holotype: MCP 35941.

Distribution: Middle Uruguay River basin, Brazil (Bertaco & Cardoso, 2005).

***Microglanis nigripinnis* Bizerril & Perez-Neto, 1992**

Microglanis nigripinnis Bizerril & Perez-Neto, 1992: 97, figs. 1–2. Type locality: Brazil, Estado do Rio de Janeiro, rio Macacu, Município de Cachoeira do Macacu, Cidade de Papucaia. Holotype: MZUSP 42308.

Distribution: Macacu River basin in Rio de Janeiro State, Brazil (Shibatta, 2003a).

***Microglanis parahybae* (Steindachner, 1880)**

Pseudopimelodus Parahybae Steindachner, 1880b: 60 footnote, pl. 1 (figs. 2–2b). Type locality: Rio Parahyba und von Santa Cruz [Brazil]. Syntypes: MCZ 8161 (25), MCZ 8162 (6), NMW 44433 (9), NMW 44436 (1).

Distribution: Paraíba do Sul River basin, Brazil (Shibatta, 2003a).

***Microglanis pellopterygius* Mees, 1978**

Microglanis pellopterygius Mees, 1978b: 256, pl. 2. Type locality: Tributary stream of Río Aguarico at Santa Cecilia, 0°06'N, 76°51'W, Napo, Ecuador. Holotype: ANSP 130437.

Distribution: Aguarico River basin, Ecuador (Shibatta, 2003a).

***Microglanis poecilus* Eigenmann, 1912**

Microglanis poecilus Eigenmann, 1912b: 155, pl. 12 (fig. 2). Type locality: Below Packeoo Falls, British Guiana.
Holotype: FMNH 46365.

Distribution: Essequibo River basin and rivers of French Guiana (Shibatta, 2003a).

***Microglanis secundus* Mees, 1974**

Microglanis secundus Mees, 1974: 235, fig. 40 e; pl. 14. Type locality: Sipaliwini, Suriname. Holotype: RMNH 26525.

Distribution: Catatumbo River basin; Venezuela and Colombia; Sipaliwini River basin, Suriname (Shibatta, 2003a).

***Microglanis variegatus* Eigenmann & Henn, 1914**

Microglanis variegatus Eigenmann & Henn, in Eigenmann, Henn & Wilson, 1914: 14. Type locality: Near Vinces, Ecuador. Holotype: CAS 17971; holotype illustrated in Eigenmann (1922b: 33, pl. 2, figs. 3–4).

Distribution: Forest pool near Vinces, Ecuador (Shibatta, 2003a).

***Microglanis zonatus* Eigenmann & Allen, 1942**

Microglanis zonatus Eigenmann & Allen, 1942: 89, pl. 3 (figs. 1–2). Type locality: Rio Morona (?). Holotype: CAS 17970.

Distribution: Upper Amazon River basin, Peru (Shibatta, 2003a).

PSEUDOPIMELODUS Bleeker, 1858

Pseudopimelodus Bleeker, 1858b: 196, 204, 207. Type species: *Pimelodus bufonius* Valenciennes, 1840. Type by subsequent designation by Gill (1861c). Gender: Masculine.

***Pseudopimelodus bufonius* (Valenciennes, 1840)**

Pimelodus Bufonius Valenciennes, in Cuvier & Valenciennes, 1840b: 154 (115 in the Strasbourg deluxe edition). Type locality: Cayenne. Holotype: RMNH (lost).

Distribution: Rivers of northeastern South America from Lake Maracaibo basin to eastern Brazil (Shibatta, 2003a) and Magdalena and Cauca River basins, Colombia (Maldonado-Ocampo *et al.*, 2005).

***Pseudopimelodus charus* (Valenciennes, 1840)**

Pimelodus charus Valenciennes, in Cuvier & Valenciennes, 1840b: 159 (118 in the Strasbourg deluxe edition). Type locality: Rio Sabara [Brazil]. No types known; description based on an unpublished illustration, reproduced in Mees (1974, frontispiece).

Distribution: São Francisco River basin, Brazil (Shibatta, 2003a).

***Pseudopimelodus mangurus* (Valenciennes, 1835)**

Pimelodus mangurus Valenciennes, 1835, in Valenciennes, 1835–47: pl. 1 (figs. 4–6). Type locality: Río de la Plata.

Possible holotype: MNHN a-8401(1), MNHN a-9417 (1). Name available from caption on plate, so illustrated specimen is the holotype, if identifiable. Described in Cuvier & Valenciennes (1840b: 156 (116 of Strasbourg deluxe edition)), with locality given as Rio-Janéiro.

Pseudopimelodus roosevelti Borodin, 1927b: 1, fig. 1. Type locality: Parassununga, Estado São Paulo, Brazil. Holotype: AMNH 8638.

Distribution: Uruguay, Paraná, Paraguay and La Plata River basins (Shibatta, 2003a).

***Pseudopimelodus pulcher* (Boulenger, 1887)**

Pimelodus (Pseudopimelodus) pulcher Boulenger, 1887c: 276, pl. 21 (fig. 1). Type locality: Canelos, e. Ecuador. Syntypes: BMNH 1880.12.8.105–107 (3).

Pseudopimelodus variolosus Miranda Ribeiro, 1914: 4, pl. 1 (fig. 2), pl. 2 (figs. 1–2). Type locality: Coxim, Rio Taquary, M. Grosso [Brazil]. Lectotype: MNRJ 818A, designated by Miranda Ribeiro (1953: 404).

Distribution: Upper Amazon River basin (Shibatta, 2003a).

***Pseudopimelodus schultzi* (Dahl, 1955)**

Zungaro zungaro schultzi Dahl, 1955: 13. Type locality: Cereté, Colombia. Holotype: Whereabouts unknown (possibly at ICNMHN).

Distribution: Magdalena River basin, Colombia (Shibatta, 2003a).

***Species inquirenda*, Pseudopimelodidae**

Zungaro mathisoni Fernández-Yépez, 1972a: 22, pl. 41. Type locality: Río Yaracuy drainage, Venezuela. Holotype: author's personal collection; whereabouts unknown.

SCHILBIDAE Bleeker, 1858

Ailichthyoidei Bleeker, 1858b: ix, 49, 248. Type genus: *Ailia* Gray, 1830.

Schilbeini Bleeker, 1858b: 49, 250, 256. Type genus: *Schilbe* Oken, 1817.

Siluranodontinae Regan, 1911: 568. Type genus: *Siluranodon* Bleeker, 1858.

Taxonomic summaries: De Vos (1984b, 1986, Africa).

Reviews: Reizer *et al.* (1980, Senegal), De Vos (1995, Africa); De Vos (1992, West Africa).

Identification guide: Jayaram (1977, South Asia).

Key to genera: Trewavas (1943, Africa); Poll & Gosse (1994, Africa).

Remarks: Although the family group name is often spelled Schilbeidae, the spelling used here, Schilbidae, was considered to be correct by Steyskal (1980: 174) and is followed herein. The monophyly of this family, as currently populated, is unlikely. Species of the Pangasiidae and possibly *Horabagrus* may be nested within the family, and the species of *Neotropius* and *Pseudeutropius* may be more closely related to some species of the Bagridae. See Mo (1991), Pouyaud *et al.* (2000, 2004), Diogo *et al.* (2004) for details. Hardmann (2005) further suggested that African species of the Schilbidae may be more closely related to other African catfishes than to any Asian member of the Schilbidae, in which case the name Ailiidae would be the valid name for the Asian group.

14 genera, 62 species; no named fossil taxa.

Ailia Gray, 1830

Acanthonotus Gray, 1830: pl. 85 (fig. 1). Type species: *Silurus (Acanthonotus) cuvieri* Gray, 1830. Type by monotypy. Gender: Masculine. Name made available by figure caption. Originally proposed as a subgenus of *Silurus*. Subsequently published in Gray (1831: 8), but with *Acanthonotus Hardwickii* as the only included species. Preoccupied by *Acanthonotus* Goldfuss, 1809 (and possibly earlier), in Mammalia.

Ailia Gray, 1830: pl. 85 (fig. 2). Type species: *Malapterus (Ailia) bengalensis* Gray, 1830. Type by monotypy. Gender: Feminine. Originally proposed as a subgenus of *Malapterus*. Name made available by caption of plate; text published later in Gray (1831: 8).

Ailiichthys Day, 1872: 712. Type species: *Ailiichthys punctata* Day, 1872. Type by monotypy. Gender: Masculine. Review: Hora (1941); Jayaram (1963).

Ailia coila (Hamilton, 1822)

Malapterurus coila Hamilton, 1822: 158, 375. Type locality: Fresh water rivers of Bengal. No types known. Unpublished Hamilton illustration reproduced in Gray (1830: pl. 85, fig. 2), as *Malapterus (Ailia) bengalensis*.

Malapterus (Ailia) bengalensis Gray, 1830: pl. 85 (fig. 2). Type locality: India. No types known; name based on previously-unpublished illustration by Hamilton. Name made available by figure caption; brief diagnosis published in Gray (1831: 8), with locality stated as "Rivers Bengal."

Silurus (Acanthonotus) Cuvieri Gray, 1830: pl. 85 (fig. 1). Type locality: India. No types known. Name made available by plate legend; text published later in Gray (1831: 8), as *Acanthonotus Hardwickii*.

Acanthonotus Hardwickii Gray, 1831: 8 [pl. 85 (fig. 1), of Gray, 1830]. Type locality: India. No types known. Unneeded new name for *Silurus (Acanthonotus) Cuvieri* Gray, 1830.

Ailia affinis Günther, 1864: 56. Type locality: Kulu and Assam, India. Syntypes (3, in two lots): BMNH 1860.3.19.867–868 (2), BMNH 2005.5.17.3 (1).

Distribution: Rivers of northern India and Deccan, including Indus, Ganges, Yamuna, Brahmaputra, Mahanadi and Krishna Rivers (Jayaram, 1963).

Ailia punctata (Day, 1872)

Ailiichthys punctata Day, 1872: 713. Type locality: Jumna at and below Delhi, also in the lower Punjab rivers. Syntypes: AMS B.7570 (1), BMNH 1889.2.1.2585–87 (3), BMNH 1889.2.1.2588–90 (3), BMNH 1889.2.1.2591 (1, skeleton), BMNH 1889.2.1.2592–93 (2), NMW 47849 (1), NMW 47863 (1), RMNH 2763 (1), ZMB 11213 (1), ZSI 420 (1, lost).

Distribution: Indus, Yamuna and Ganges Rivers, northern India (Jayaram, 1963).

CLUPISOMA Swainson, 1838

Clupisoma Swainson, 1838: 347, 351, 354. Type species: *Silurus (Clupisoma) argentata* Swainson, 1839. Type by subsequent monotypy. Gender: Neuter. Originally proposed as a subgenus of *Silurus*, without any included species; first species added in Swainson (1839: 306).

Schilbeichthys Bleeker, 1858b: 255, 256. Type species: *Silurus garua* Hamilton, 1822. Type by monotypy. Gender: Masculine.

Key to species: Datta & Karmakar (1980).

Clupisoma bastari Datta & Karmakar, 1980

Clupisoma bastari Datta & Karmakar, 1980: 193, figs. 1–2. Type locality: Indravati River (a tributary of Godavari River) at Lohandigura, 33 kms west of Jagdalpur, Bastar District, Madhya Pradesh, India. Holotype: ZSI FF1342.

Distribution: Godavari River basin, Madhya Pradesh, India (Jayaram, 1999).

Clupisoma garua (Hamilton, 1822)

Silurus garua Hamilton, 1822: 156, 375, pl. 21 (fig. 50). Type locality: Fresh water rivers of the Gangetic provinces. No types known.

Silurus (Clupisoma) argentata Swainson, 1839: 306. Type locality: Fresh water rivers of the Gangetic provinces. No types known. Made available by reference to “Ham. pl. 21, f. 50” [= Hamilton, 1822: pl. 21 (fig. 50)]. Unneeded new name for *Silurus garua* Hamilton, 1822.

Distribution: Throughout northern India, but not south of the Mahanadi River, Pakistan, Bangladesh, and Nepal (Jayaram, 1999).

Clupisoma longianalis (Huang, 1981)

Platytrupius longianalis Huang, 1981: 438, fig. 5. Type locality: Puer county, Xiaoganlanba, Yunnan, China. Holotype: KIZ 735118.

Distribution: Lancangjiang [= Mekong River] (Chu *et al.*, 1999).

Clupisoma montana Hora, 1937

Clupisoma montana Hora, 1937b: 673, figs. 7–8. Type locality: Teesta River, below Darjeeling, India. Type(s): ZSI F 12472/1.

? *Pseudeutropius murius batarensis* Shrestha, 1980: 212, fig. 97. Type locality: Batar, Trisuli river, Nuwakot Dist., Bagmati Zone, Nepal. Holotype (220 mm): at Museum of Zoology Department, Institute of Science, Tribuvanan University, Nepal.

Distribution: Teesta, Jamuna and Sone Rivers, India, Nepal (Jayaram, 1999).

Clupisoma naziri Mirza & Awan, 1973

Clupisoma naziri Mirza & Awan, 1973: 152, fig. 2. Type locality: Indus River at Jinnah Barrage, Pakistan. Holotype: GCM 10.

Distribution: NWFP, Punjab, Azad Kashmir, Indus River basin, Pakistan (Mirza, 2003).

Clupisoma nujiangense Chen, Ferraris & Yang, 2005

Clupisoma nujiangense Chen, Ferraris & Yang, 2005: 566, fig. 1. Type locality: China, Yunnan Province, Baoshan City, Longling County, Mungnuo Township, Sanjiangkou, Nu Jiang (=Salween River), 24°25'18.6"N, 98°58'21.2"E. Holotype: KIZ 200310118.

Distribution: Upper Salween River, Yunnan, China (Chen *et al.*, 2005).

Clupisoma prateri Hora, 1937

Clupisoma prateri Hora, 1937b: 671, fig. 6. Type locality: Burma. Holotype: Asiatic Soc. Bengal 213 or 219 [now at ZSI].

Distribution: Lower and middle reaches of Irrawaddy River, Sittang, Bago and Salween river basins, Myanmar (Ferraris, 2004).

Remarks: Redescribed in Ferraris (2004: 6, fig. 2).

Clupisoma roosae Ferraris, 2004

Clupisoma roosae Ferraris, 2004: 2, fig. 1. Type locality: Myanmar, Kachin State, Myitkyina market. Holotype:

NRM 40030.

Distribution: Upper Irrawaddy River basin, Myanmar (Ferraris, 2004).

***Clupisoma sinense* (Huang, 1981)**

Platytropius sinensis Huang, 1981: 437, figs. 1–4. Type locality: Puer county, Xiaoganlanba, Yunnan, China. Holotype: KIZ 735124.

Distribution: Mekong River basin (Rainboth, 1996), including Lancangjiang (= upper Mekong River of Yunnan) (Chu *et al.*, 1999), and Malay Peninsula (Zakaria-Ismail, 1992; Kottelat, 2001b).

Remarks: Placement of this species in *Clupisoma* follows Ng (1999b) and Chen *et al.* (2005).

***EUTROPIICHTHYS* Bleeker, 1862**

Eutropiichthys Bleeker, 1862b: 398. Type species: *Pimelodus vacha* Hamilton, 1822. Type by original designation.

Gender: Masculine. Also proposed as new in Bleeker (1862–63: 14).

Revision: Hora (1937e).

***Eutropiichthys burmannicus* Day, 1877**

Eutropiichthys burmannicus Day, 1877 (in Day, 1875–78): 490. Type locality: Burma. Possible syntypes: ZSI B.38–39 (2), ZSI B.219 (1, lost), ZSI B.306 (1). Originally as: “Variety *Eutropiichthys burmannicus*,” in the account of *Eutropiichthys vacha*.

Distribution: Irrawaddy, lower Salween and smaller intervening river basins.

***Eutropiichthys murius* (Hamilton, 1822)**

Pimelodus murius Hamilton, 1822: 195, 378. Type locality: Mahananda river [Bengal]. No types known. Unpublished Hamilton illustration reproduced in Hora (1929: pl. 22, fig. 2).

Pachypterus melanurus Swainson, 1839: 306. Type locality: Mahananda river [Bengal]. No types known. Unneeded new name for *Pimelodus murius* Hamilton, 1822.

Distribution: Northern India, Bangladesh and Pakistan (Jayaram, 1999).

***Eutropiichthys vacha* (Hamilton, 1822)**

Pimelodus vacha Hamilton, 1822: 196, 378, pl. 19 (fig. 64). Type locality: Larger fresh water rivers of the Gangetic provinces. No types known.

Pachypterus punctatus Swainson, 1839: 306. Type locality: Larger fresh water rivers of the Gangetic provinces. No types known. Made available by reference to “Ham. p. 196, f. 64” [= Hamilton, 1822: 196, p. 19, fig. 64]. Unneeded new name for *Pimelodus vacha* Hamilton, 1822.

Distribution: Northern India, south to the Mahanadi River, Bangladesh, Nepal and Pakistan (Jayaram, 1999).

***IRVINEIA* Trewavas, 1943**

Irvineia Trewavas, 1943: 165. Type species: *Irvineia voltae* Trewavas, 1943. Type by monotypy. Gender: Feminine.

***Irvineia orientalis* Trewavas, 1964**

Irvineia orientalis Trewavas, 1964: 390, figs. 1, 2, 3 b–c. Type locality: Villaggio Duca delgi Abruzzi, Uebi Scibeli (Shebeli) system. Holotype: MSNG 39551; holotype illustrated in De Vos (1995: 358, fig. 226).

Distribution: Juba-Uebi Shebeli system (De Vos, 1995).

***Irvineia voltae* Trewavas, 1943**

Irvineia voltae Trewavas, 1943: 165, fig. 1. Type locality: Above the Senchi Rapids, River Volta, Gold Coast. Syntypes (3): BMNH 1944.2.9.19–21 (3); one syntype illustrated in De Vos (1995: 362, fig. 229).

Distribution: lower Volta River basin (De Vos, 1995).

***LAIDES* Jordan, 1919**

Laïs Bleeker, 1857b: 473. Type species: *Pangasius hexanema* Bleeker, 1852. Type by monotypy. Gender: Masculine. Preoccupied by *Lais* Gistel, 1848, in Tunicata, replaced by *Laides* Jordan, 1919.

Laides Jordan, 1919b: 293. Type species: *Pangasius hexanema* Bleeker, 1852. Type by being a replacement name. Gender: Masculine. Replacement for *Lais* Bleeker, 1857; preoccupied by *Lais* Gistel, 1848, in Tunicata.

***Laides hexanema* (Bleeker, 1852)**

Pangasius hexanema Bleeker, 1852d: 588. Type locality: Palembang, Batavia. Syntypes (2, 102–152 mm TL): pos-

sibly BMNH 1863.12.4.1007 (1, 157 mm TL), NMV A928 (1).

Distribution: Thailand to Indonesia, in large rivers, including the Mekong (Rainboth, 1996).

***Laides longibarbis* (Fowler, 1934)**

Pangasius longibarbis Fowler, 1934a: 87, fig. 27. Type locality: Me Kong at Chieng Sen, North Siam. Holotype: ANSP 59441.

Distribution: Mekong, Mehklong, and Chao Phraya River basins (Ng, 1999b).

Remarks: Redescribed by Ng (1999b).

***NEOTROPIUS* Kulkarni, 1952**

Pachypterus Swainson, 1838: 346 et seq. Type species: *Silurus atherinoides* Bloch, 1794. Type by subsequent designation by Swain (1882: 281). Gender: Masculine. Originally without any included species. Species first added in Swainson (1839: 306).

Neotropius Kulkarni, 1952: 231. Type species: *Neotropius khavalchor* Kulkarni, 1952. Type by monotypy. Gender: Masculine.

Review: Hora (1941).

Remarks: Swain (1882: 281) and (following Swain) Jordan (1919a: 205) treated *Pachypterus* Swainson as preoccupied in Coleoptera, presumably basing their decision on the following notation in Agassiz (1846: 271): “*Pachypterus Sol. Col.*, 1833.” However, it appears that Agassiz was referring to usage of the name in Dejean, 1834, in which a manuscript by Solier was cited as the source of *Pachypterus*. According to Neave (1940: 515) Dejean’s use of the name was as a *nomen nudum* and *Pachypterus* was not used in a way that would have made it available in Coleoptera until Lucas, 1846, which postdates the first proposal of *Pachypterus*. However, the writings of Swain and Jordan may have stifled use of *Pachypterus* Swainson as valid, and the younger name *Neotropius*, which has been in widespread usage for the past one-half century, is used here. The species listed below are similar overall to those of *Pseudeutropius* and the two genera may not both represent natural groups.

***Neotropius acutirostris* (Day, 1870)**

Pseudeutropius acutirostris Day, 1870c: 618. Type locality: Throughout Burma. Syntypes: AMS B.7967 (1), BMNH 1889.2.1.2413 (1), BMNH 1889.2.1.2462 (1), NMW 48327 (1), ZSI F484 (1), ZSI A.505 (1 lost).

Distribution: Irrawaddy, Sittang, and Bago Rivers, Myanmar.

***Neotropius atherinoides* (Bloch, 1794)**

Silurus Atherinoides Bloch, 1794: 48, pl. 371 (fig. 1). Type locality: Tranquebar. Holotype: ZMB 3013.

Pimelodus angius Hamilton, 1822: 180, 377, pl. 29 (fig. 59). Type locality: Rivers of Bengal. No types known.

Pimelodus urua Hamilton, 1822: 177, 377. Type locality: Rivers and ponds of the northern parts of Bengal. No types known. Unpublished Hamilton illustration reproduced in Hora (1929: pl. 21, fig. 4).

Pachypterus trifasciatus Swainson, 1839: 306. Type locality: Rivers of Bengal. No types known. Name made available by reference to “Ham. P. 180, f. 59” [= Hamilton (1822: 180, pl. 29, fig. 59)]. Unneeded new name for *Pimelodus angius* Hamilton, 1822.

Pseudeutropius atherinoides walkeri Chaudhuri, 1912: 444, pl. 41 (fig. 3). Type locality: Siripur, Saran, Bihar [North India]. Syntypes (2): at ZSI. Originally as *Pseudeutropius atherinoides* var. *walkeri*.

Distribution: Widely distributed in India, except for Assam and Kerala; also in Pakistan, Nepal, and Bangladesh (Jayaram, 1999).

***Neotropius khavalchor* Kulkarni, 1952**

Neotropius khavalchor Kulkarni, 1952: 232, fig. 1. Type locality: Panchaganga River, near Kolhapur, Bombay State, India. Holotype: ZSI F647/2.

Distribution: Krishna River basin, Maharashtra and Andhra Pradesh, India (Jayaram, 1999).

Species inquirenda, Neotropius

Bagrus Exodon Cuvier & Valenciennes, in Valenciennes, 1832b: 385, pl. 4 (fig. 1). Type locality: Bengal. Syntypes: MNHN a-8960 (2), MNHN b-0680 (1). Also in Cuvier & Valenciennes (1840a: 395).

PARAILIA Boulenger, 1899

Parailia Boulenger, 1899b: 105. Type species: *Parailia congica* Boulenger, 1899. Type by monotypy. Gender: Feminine.

Physailia Boulenger, 1901e: 445. Type species: *Physailia pellucida* Boulenger, 1901. Type by monotypy. Gender: Feminine.

Parailia congica Boulenger, 1899

Parailia congica Boulenger, 1899b: 106, pl. 41 (fig. 2). Type locality: Ebinga (rivière du lac Léopold II). Lectotype: MRAC 942, designated by De Vos (1995: 297).

Parailia longifilis Boulenger, 1902d: 37, pl. 10 (fig. 3). Type locality: l'Ubangi à Banzyville. Lectotype: MRAC 1309 (1), designated by De Vos (1995: 295).

Distribution: Congo River basin (De Vos, 1995).

Parailia occidentalis (Pellegrin, 1901)

Ailia occidentalis Pellegrin, 1901: 331. Type locality: Cap Lopez, Congo français. Holotype: MNHN 1885-0404.

Physailia ansorgii Boulenger, 1910: 557. Type locality: Quanza River at Cunga, Angola. Syntypes (2): BMNH 1911.6.1.107 (1), NMW 45490 (1); NMW syntype illustrated in De Vos (1995: 305, fig. 208).

Physailia villiersi Boulenger, 1912b: 17, pl. 17 (fig. 6). Type locality: de la Lucala près de Cabinda, du Chiloango à Mayili, de la Luali à Lundo, et de la Luculla. Syntypes: ANSP 38756–58 (3), BMNH 1912.4.1.419–420 (2), BMNH 1912.4.1.421 (1), BMNH 1912.4.1.422–424 (3), BMNH 1912.4.1.425–429 (5), FMNH 56151 (2), MRAC 1656–1659 (4), MRAC 1660–1661 (2), MRAC 1662 (1), NMW 45489 (16), ZMB 18818 (9).

Distribution: Ogowe, Congo, Quanza, Luculla, Chiloango Rivers (De Vos, 1995).

Parailia pellucida (Boulenger, 1901)

Physailia pellucida Boulenger, 1901e: 445. Type locality: Omdurman, Nile River. Syntypes (numerous): BMNH 1907.12.2.1942–1956 (14 or 15), BMNH 1907.12.2.1940–1941 (2), ? MCZ 32074 (1), MNHN 1907-0215 (3), MNHN 1907-0216 (3).

Distribution: Nile River, Chad Basin, several west African river basins (De Vos, 1995).

Parailia somalensis (Vinciguerra, 1897)

Ailia somalensis Vinciguerra, 1897: 346. Type locality: Lugh, Fl. Ganana, Somaliland. Syntypes: BMNH 1961.5.3.12 (1), MSNG 14612 (4); one MSNG syntype illustrated in de Vos (1995: 321, fig. 212).

Physailia somalensis tanensis Whitehead, 1962a: 101, fig. 2. Type locality: Tana River at Hola, Kenya. Holotype: BMNH 1961.5.3.7.

Distribution: Ganana and lower Tana Rivers (De Vos, 1995).

Parailia spiniserrata Svensson, 1933

Parailia spiniserrata Svensson, 1933: 73, pl. 4 (fig. 2); fig. 24. Type locality: MacCarthy Island Area, Gambia River, British West Africa. Syntypes (about 25): NRM 11144 (4), NRM 14453 (1), NRM 14454 (8), NRM 21144 (2), NRM 31144 (1), NRM 31145 (1), NRM 31146 (4), NRM 31147 (1); one syntype illustrated in De Vos (1995: 326, fig. 215).

Distribution: Gambia, Geba, and Jong River basins (De Vos, 1995).

PAREUTROPIUS Regan, 1920

Ansorgia Boulenger, 1912b: 17. Type species: *Ansorgia vittata* Boulenger, 1912. Type by monotypy. Gender: Feminine. Preoccupied by *Ansorgia* Warren, 1899, in Lepidoptera; replaced by *Ansorgiichthys* Whitley, 1935.

Pareutropius Regan, 1920a: 105. Type species: *Pareutropius micristius* Regan, 1920. Type by monotypy. Gender: Masculine.

Eutropiellus Nichols & La Monte, 1933b: 5. Type species: *Eutropiellus kasai* Nichols & La Monte, 1933. Type by original designation. Gender: Masculine.

Ansorgiichthys Whitley, 1935: 249. Type species: *Ansorgia vittata* Boulenger, 1912. Type by being a replacement name. Gender: Masculine. Replacement for *Ansorgia* Boulenger, 1912; preoccupied by *Ansorgia* Warren, 1899, in Lepidoptera.

Revision: Thys van den Audenaerde (1964, as *Eutropiellus*).

Pareutropius buffei (Gras, 1961)

Eutropiellus buffei Gras, 1961: 406, fig. 3. Type locality: Bas-Ouémé, Bas-Dahomey. Syntypes (5): Lab. d'Hydrobiol. Cotonou, Benin (1), MNHN 1958-0001 (4).

Eutropiellus vandeweyeri Thys van den Audenaerde, 1964: 225, fig. 3. Type locality: Environment de Aba, Lower Nigeria Delta, Nigeria. Holotype: MRAC 140281.

Distribution: Lower Ouémé River, Cross and Niger Rivers (De Vos, 1995).

Pareutropius debauwi (Boulenger, 1900)

Eutropius debauwi Boulenger, 1900a: 138, pl. 50 (fig. 2). Type locality: Uerre. Syntypes: BMNH 1900.12.13.8 (1), MRAC 347 (1).

Ansorgia vittata Boulenger, 1912b: 17, pl. 19 (fig. 2). Type locality: Lower Congo. Syntypes: ANSP 38734–35 (2), BMNH 1912.4.1.414 (1), BMNH 1912.4.1.415–417 (3), BMNH 1912.4.1.418 (1, skeleton), MRAC 1653 (1), MRAC 1654 (1), MRAC 1655 (1), ZMB 18817 (1). Name as *Ansorgia vitata* on p. 17, *A. vittata* on plate legend (p. 28) and on plate; De Vos (1986: 37) cited both spellings and used *Ansorgia vittata*, thereby acting as first reviser.

Ansorgia vittata bistriata Giltay, 1930b: 393, fig. 1. Type locality: Buta. Holotype: MRAC 20584.

Eutropiellus kasai Nichols & La Monte, 1933b: 5, fig. 4. Type locality: Luluabourg, Kasai, Belgian Congo. Holotype: AMNH 12338.

Distribution: Congo River system, Chiloango River, the Kouilou and Ogowe Rivers (De Vos, 1995).

Pareutropius longifilis (Steindachner, 1914)

Eutropius longifilis Steindachner, 1914: 537. Type locality: Deutsch-Ostafrika [Kiperege]. Syntypes: NMW 46507 (2). Species illustrated and described in more detail in Steindachner (1915d: 75, pl. 3, fig. 2), with locality as: Wasserlauf bei dem Dorfe Keperege; one syntype illustrated in De Vos (1995: 347, fig. 222).

Pareutropius micristius Regan, 1920a: 105. Type locality: Tanganyika Territory, Morogoro. Syntypes (7): BMNH 1920.3.8.10–12 (3).

Distribution: eastward flowing rivers north of the Ruvuma, Lake Chiuta, Ruvuma system, Lake Chilwa (De Vos, 1995).

Pareutropius mandevillei Poll, 1959

Pareutropius mandevillei Poll, 1959: 92, pl. 16 (fig. 4a). Type locality: Stanley Pool. Holotype: MRAC 98387.

Distribution: Congo River basin (De Vos, 1995).

PLATYTROPIUS Hora, 1937

Platytropus Hora, 1937a: 352. Type species: *Pseudeutropius siamensis* Sauvage, 1883. Type by original designation. Gender: Masculine.

Nemasiluroides Fowler, 1937: 137. Type species: *Nemasiluroides furcatus* Fowler, 1937. Type by original designation. Gender: Masculine.

Remarks: *Platytropus* Hora was published in January, 1937, and therefore has priority over *Nemasiluroides* Fowler, 1937, which was published on 19 May, 1937. A second paper by Hora (1937c: 39) which has been cited as the original proposal of *Platytropus* (e. g., Eschmeyer, 1990) was not published until November, 1937.

Platytropus siamensis (Sauvage, 1883)

Pseudeutropius siamensis Sauvage, 1883b: 154. Type locality: Mé-Nam, Siam. Holotype: MNHN a-5002.

Nemasiluroides furcatus Fowler, 1937: 138, figs. 16–19. Type locality: Bangkok, Siam. Holotype: ANSP 67893.

Distribution: Chao Phraya River basin, Thailand.

PROEUTROPIICHTHYS Hora, 1937

Proeutropiichthys Hora, 1937a: 353. Type species: *Eutropius macrophthalmus* Blyth, 1860. Type by original designation. Gender: Masculine.

Review: Hora (1941).

? **Proeutropiichthys buchanani** (Valenciennes, 1840)

Bagrus Buchanani Valenciennes, 1840, in Jacquemont, 1835–44: pl. 16 (fig. 3). Type locality: Hindustan, India.

Holotype: MNHN a-5721(1 of 3) or MNHN a-5722 (1 of 3). Name made available by figure caption.

Distribution: Hindustan, India.

Remarks: Illustrated specimen should be treated as the holotype, if identifiable from among the specimens listed herein. Inclusion into *Proeutropiichthys* based on personal observation of types.

? ***Proeutropiichthys goongwaree*** (Sykes, 1839)

Hypophthalmus Goongwaree Sykes, 1839: 163. Type locality: the Mota Mola near Poona, India. No types known.

Also described as new in Sykes (1840: 60); illustrated and described in more detail in Sykes (1841: 369, pl. 64, fig. 3).

Distribution: Southern India (Jayaram, 1999).

Remarks: Placed in *Pseudeutropius* by Günther (1864) and Day (1875–78), then into *Eutropiichthys* by Hora (1937e), where it has remained until now. However, the described species and illustrations are clearly not of a species of either of these genera and more likely that of a species of *Proeutropiichthys*.

Proeutropiichthys macrophthalmos (Blyth, 1860)

Eutropius macrophthalmos Blyth, 1860: 156. Type locality: Tenasserim [Myanmar]. Type(s): Whereabouts unknown.

Proeutropiichthys taakree burmanicus Tilak, 1982: 35, figs. 1, 3, 5. Type locality: Burma. Holotype: ZSI FF771.

Distribution: Irrawaddy, Sittang, and Bago River basins, Myanmar.

Proeutropiichthys taakree (Sykes, 1839)

Hypophthalmus taakree Sykes, 1839: 163. Type locality: Deccan, India. No types known. Also described as new in Sykes (1840: 60); illustrated and described more fully in Sykes (1841: 369, pl. 64, fig. 4).

Schillee Sykesii Jerdon, 1849: 335. Type locality: Cavery River, southern India. No types known.

Pseudeutropius longimanus Günther, 1864: 60. Type locality: India. Holotype: BMNH 1857.6.13.88 (skin).

Pseudeutropius megalops Günther, 1864: 60. Type locality: Godaveri at Mahadespur, Orissa, Central India. Holotype: BMNH 1860.3.19.784.

Distribution: Western Ghats, Kerala, Maharashtra, India (Jayaram, 1999).

PSEUDEUTROPIUS Bleeker, 1862

Pseudeutropius Bleeker, 1862b: 398. Type species: *Eutropius brachypterus* Bleeker, 1858. Type by original designation. Gender: Masculine. Also in Bleeker (1862–63: 14, 74).

Pseudeutropius brachypterus (Bleeker, 1858)

Eutropius brachypterus Bleeker, 1858b: 169. Type locality: Palembang [Sumatra]. Holotype (115 mm TL): Whereabouts unknown; holotype illustrated in Bleeker (1862–63: 71, pl. 75 [= Silur. pl. 27], fig. 1).

Distribution: Kapuas River (Roberts, 1989a); Sumatra.

Pseudeutropius mitchelli Günther, 1864

Pseudeutropius mitchelli Günther, 1864: 59. Type locality: Madras Presidency [India]. Syntypes: BMNH 1863.12.18.134–135 (2).

Distribution: Kerala, India (Jayaram, 1999).

Pseudeutropius moolenburghae Weber & de Beaufort, 1913

Pseudeutropius moolenburghae Weber & de Beaufort, 1913: 249, fig. 100. Type locality: Sumatra (Batang Hari river) [Indonesia]. Syntypes: AMNH 9283 (1), ZMA 112681 (6).

Distribution: Batang Hari River, Sumatra (Weber & de Beaufort, 1913); Kapuas River (Roberts, 1989a).

SCHILBE Oken, 1817

Schilbe Oken, 1817: 1183. Type species: *Silurus mystus* Linnaeus, 1758. Type by monotypy. Gender: Masculine.

Pusichthys Swainson, 1838: 348, et seq. Type species: *Schilbe uranoscopus* Rüppell, 1832. Type by subsequent monotypy. Gender: Masculine. Originally proposed as a subgenus of *Silurus*, without any included species. Species first added in Swainson (1839: 307).

Eutropius Müller & Troschel, 1849: 6. Type species: “*Bagrus schilboides* Val. (*Hypophthalmus niloticus* Rüppell)” [= *Bagrus schilboides* Valenciennes, 1840]. Type by monotypy. Gender: Masculine. Originally proposed as a subgenus of *Bagrus*.

Proeutropius Fowler, 1936b: 307. Type species: *Silurus congensis* Leach, 1818. Type by original designation. Gender: Masculine. Originally proposed as a subgenus of *Eutropius*.

Review: De Vos & Lévéque (1983), west Africa (as *Eutropius*).

Key: De Vos (1984a) Quanza and Bengo rivers, Angola (as *Eutropius*).

***Schilbe angolensis* (De Vos, 1984)**

Eutropius angolensis De Vos, 1984a: 13, figs. 4, 5d. Type locality: Quanza river at Quimbango, Angola, 10°51'S, 17°30'E. Holotype: MRAC 78-6-P-899.

Distribution: Upper Quanza River, Angola (De Vos, 1995).

***Schilbe banguelensis* (Boulenger, 1911)**

Eutropius banguelensis Boulenger, 1911a: 282, fig. 231. Lake Bangwelu. Holotype: BMNH 1907.9.30.8.

Eutropius nasalis Boulenger, 1915: 169. Type locality: Lac Moero. Lectotype: MRAC 14163; lectotype designated by, and illustrated in, De Vos (1995: 124, fig. 158, bottom) as lectoholotype.

Distribution: Chambezi River, Lake Bangweulu, Luapula River and Lake Mweru (De Vos, 1995).

***Schilbe bocagii* (Guimarães, 1884)**

Eutropius Bocagii Guimarães, 1884a: 85, unnumbered pl. Type locality: Dondo (Quanza fl.), Angola. Holotype: MB (destroyed, De Vos, 1995: 128). Illustration of holotype reproduced in De Vos (1984a: 5, fig. 1a).

Eutropius ansorgii Boulenger, 1910: 555. Type locality: Quanza River at Cunga, Angola. Syntypes (3): BMNH 1911.6.1.100–101 (2), NMW 46502 (1).

Eutropius seraoi Boulenger, 1910: 556. Type locality: Bengo River and from the Lucalla River at Lucalla, Angola. Syntypes (10): ANSP 37956 (1), BMNH 1911.6.1.102–105 (4), BMNH 1911.6.1.106 (1), NMW 46524 (2), ZMB 18226 (1); one NMW syntype illustrated in De Vos (1984a: fig. 3, and 1995: 129, fig. 160f).

Eutropius eclipsis Fowler, 1919b: 270, fig. 12. Type locality: Quanza River at Cunga, Angola. Holotype: USNM 42342. Illustration of holotype reproduced in De Vos (1984a: 5, fig. 1b).

Distribution: Lower Quanza River and Bengo Rivers, Angola (De Vos, 1995).

***Schilbe brevianalis* (Pellegrin, 1929)**

Eutropius brevianalis Pellegrin, 1929b: 362. Type locality: Dehane (Nyong), Cameroun. Syntypes: MNHN 1928-0354 (1), MNHN 1928-0355 (1).

Distribution: coastal rivers in Nigeria and Cameroon (De Vos, 1995).

***Schilbe congensis* (Leach, 1818)**

Silurus Congensis Leach, in Tuckey, 1818: 409. Type locality: Lower Congo. Holotype: BMNH 2005.5.17.6.

Eutropius congoensis Boulenger, 1899b: 105. Type locality: Lower Congo. Holotype: BMNH 2005.5.17.6. Unjustified emendation of *Silurus congensis* (De Vos, 1995).

Eutropius congolensis Boulenger, 1901a: 268. Type locality: Lower Congo. Holotype: BMNH 2005.5.17.6. Unjustified emendation of *Silurus congensis* Leach, 1818.

Distribution: Congo River system (De Vos, 1995).

***Schilbe djemeri* (Thys van den Audenaerde & De Vos, 1982)**

Eutropius djemeri Thys van den Audenaerde & De Vos, 1982: 179, figs. 1–2. Type locality: River Djerem, below falls at the rapids, 6–7 km south of Mbakaou, Cameroun. Holotype: MRAC 73-18-P-2310.

Distribution: Upper Sanaga River basin, Cameroun (De Vos, 1995).

***Schilbe durini* (Gianferrari, 1932)**

Eutropius Durini Gianferrari, 1932: 141. Type locality: Lago Tanganyika. Holotype: MSNM 36; illustrated in De Vos (1995: 146, fig. 169).

Distribution: Unknown (De Vos, 1995).

Remarks: Validity of species questioned by De Vos (1986, 1995), who indicated that the type locality is almost certainly wrong.

***Schilbe grenfelli* (Boulenger, 1900)**

Eutropius grenfelli Boulenger, 1900a: 137, pl. 50 (fig. 1). Type locality: Bolobo. Holotype: BMNH 1900.6.23.6.

Eutropius gastratus Nichols & Griscom, 1917: 708, fig. 20. Type locality: Rungu, Congo. Holotype: AMNH 6404.

Eutropius bomae Lönnberg & Rendahl, 1922: 126. Type locality: Boma, lower Congo. Holotype: NRM 6024.

Distribution: Congo River basin, Ogowe, Nyanga, and Ntem-Campo River basins, Nyong and Lokondje basins (De Vos, 1995).

***Schilbe intermedius* Rüppell, 1832**

Schilbe intermedius Rüppell, 1832: 6. Type locality: Nil. Syntypes: BMNH 1850.7.29.14 (1), SMF 202 (1), SMF

2625 (1, dry), SMF 6403 (1, skeleton). BMNH specimen illustrated by Seegers (1996: 184, fig. 129) as holotype.

Schilbe auratus Joannis, 1835: [Cl. IV, pl. 5]. Type locality: Nil. Possible holotype: MNHN b-0346.

Schilbe Senegallus Valenciennes, in Cuvier & Valenciennes, 1840a: 378 (281 of Strasbourg deluxe edition). Type locality: Sénégal. Syntypes (huit pouces): at MNHN.

Bagrus depressirostris Peters, 1852: 682. Type locality: Mozambique. Syntypes: BMNH 1863.4.30.15 (1), ZMB 3015 (2), ZMB 3016 (1). Illustrated and described in more detail in Peters (1868c: 25, pl. 4, fig. 5) as *Eutropius depressirostris*.

Schilbe dispila Günther, 1864: 51. Type locality: River Niger, West Africa; Upper Nile, 500 miles south of Char-toum. Syntypes: BMNH 1852.2.22.17 (1), BMNH unregistered (2).

Schilbe senegalensis Günther, 1864: 51. Type locality: Senegal. Syntypes: at MNHN (not found). Unjustified emen-dation of *Schilbe senegallus* Valenciennes, 1840.

Schilbe senegalensis fasciata Steindachner, 1870: 983, pl. 6 (figs. 1–2). Type locality: Senegal bei St. Louis, Dagana, Podor, und Bakel. Syntypes: NMW 44579–81 (61). Originally as *Schilbe senegalensis* var. *fasciata*.

Schilbe steindachneri Guimaraes, 1884b: 1, pl. 1 (figs. 1–2). Type locality: Cunene. Holotype: MB (destroyed; De Vos, 1995: 162).

Schilbe bouvieri Rochebrune, 1885: 95. Type locality: [Casamance R., Senegal, w. Africa]. Type(s): at Museo Bou-vieri.

Schilbe Emini Pfeffer, 1896: 32. Type locality: Muhale-Bach, Uniamwesi. Holotype: ZMB 12851.

Eutropius lemairii Boulenger, 1900a: 138, pl. 50 (fig. 3). Type locality: Lofoi, Katanga. Syntypes: BMNH 1900.12.13.24 (1), MRAC 293 (1).

Schilbe palmeri Svensson, 1933: 71, pl. 3 (fig. 4). Type locality: Gambia River, Mc Carthy Island area. Syntypes: NRM 11145 (3), NRM 14452 (1), NRM 21145 (1).

Distribution: Widely distributed in Sub-Saharan Africa (De Vos, 1995).

Remarks: See De Vos & Skelton (1990) for discussion of validity and correct name of the species.

***Schilbe laticeps* (Boulenger, 1899)**

Eutropius laticeps Boulenger, 1899b: 105, pl. 41 (fig. 3) [not fig. 2 as stated in text]. Type locality: Kutu, Lac Léopold II. Holotype: MRAC 941.

Distribution: Congo River basin (De Vos, 1995).

***Schilbe mandibularis* (Günther, 1867)**

Eutropius mandibularis Günther, 1867b: 112. Type locality: Bossumprah River, Gold Coast. Holotype: deposited at Liverpool Museum. Holotype considered to be lost by Boulenger (1911: 290) and Trewavas (1943: 168).

Eutropius liberiensis Hubrecht, 1881: 69. Type locality: St. Paul's River, Liberia. Holotype: RMNH 5328.

Eutropius mentalis Boulenger, 1901a: 269. Type locality: River Prah, Gold Coast. Syntypes: BMNH 1899.12.22.8–10 (3).

Distribution: West Africa, from the St. Paul to the Prah River (De Vos, 1995).

Remarks: See De Vos (1983) for comments on taxonomy.

***Schilbe marmoratus* Boulenger, 1911**

Schilbe marmoratus Boulenger, 1911f: 222. Type locality: la rivière Sankuru (Kasaï). Holotype (70 mm): at MHNL.

Schilbe congolensis Steindachner, 1912: 445. Type locality: Moloundou, Dscha, Kamerun. Syntypes: NMW 10698–99 (2). Species described in more detail in Steindachner (1913: 30). Preoccupied in *Schilbe* by *Eutropius congo-lensis* Boulenger, 1901.

Distribution: Congo River basin and Shiloango system (De Vos, 1995).

***Schilbe micropogon* (Trewavas, 1943)**

Eutropius micropogon Trewavas, 1943: 168. Type locality: River Volta, Gold Coast. Syntypes (3): BMNH 1944.2.9.16–17 (2), BMNH 1944.2.9.18 (1).

Distribution: lower course of west African rivers, from Senegal to Cameroon (De Vos, 1995).

***Schilbe moebiusii* (Pfeffer, 1896)**

Eutropius möbiusii Pfeffer, 1896: 30. Type locality: Kingani-Fluss. Syntypes: ZMB 13680 (8, ? now 3); one syntype illustrated in De Vos (1995: 235, fig. 185).

Eutropius moebii Boulenger, 1911a: 293. Type locality: Kingani -Fluss. Syntypes: ZMB 13680 (8, ? now 3). Unjustified emendation of *Eutropius möbiusii* Pfeffer, 1896.

Distribution: Coastal rivers of Tanzania, including the Kingani, Rufiji, and Wani basins (De Vos, 1995).

***Schilbe multitaeniatus* (Pellegrin, 1913)**

Eutropius multitaeniatus Pellegrin, 1913b: 273. Type locality: l'Ogôoué (Ngomo). Syntypes: MNHN 1908-0230 (1), MNHN 1908-0231 (1), MNHN 1913-0266 (1).

Eutropius multilineatus Boulenger, 1916: 293. Unjustified emendation of *Eutropius multitaeniatus* Pellegrin, 1913 (de Vos, 1995).

Eutropius cameronensis Pellegrin, 1929b: 361. Type locality: Akonolinga (Nyong), ... Mfida (Nyong), Nya-bessan (Ntem); Cameroun. Syntypes: MNHN 1928-0351 (1), MNHN 1928-0352 (1), MNHN 1928-0353 (1).

Distribution: Dja, Nyong, Igowe, Nyang, and N'dogo Rivers (De Vos, 1995).

Remarks: See De Vos (1983) for comments on taxonomy.

***Schilbe mystus* (Linnaeus, 1758)**

Silurus mystus Linnaeus, 1758: 305. Type locality: In Nilo. Holotype: NRM 70; illustrated in De Vos (1995: 247, fig. 191).

Hypophthalmus niloticus Rüppell, 1829: 6, pl. 1 (fig. 1). Type locality: Nil. Syntypes: BMNH 1850.7.29.16 (1), possibly RMNH 2985 (1), SMF 752 [or 725] (1), SMF 2637 (1, dry), SMF 6050 (1, skeleton), SMF 6051 (1, skeleton), SMF 6053 (1).

Bagrus Adansonii Valenciennes, in Cuvier & Valenciennes, 1840a: 391 (290 of Strasbourg deluxe edition), pl. 414. Type locality: Sénégal. Holotype: MNHN a-8670 (dry).

Schilbe Hasselquistii Valenciennes, in Cuvier & Valenciennes, 1840a: 377 (280 of Strasbourg deluxe edition). Type locality: Nil. Holotype: MNHN b-0599.

Bagrus schilbeides Valenciennes, in Cuvier & Valenciennes, 1840a: 389 (289 of Strasbourg deluxe edition). Type locality: Nil. Syntypes: MNHN b-0600 (1), MNHN b-0601 (1). Unneeded new name for *Hypophthalmus niloticus* Rüppell, 1829.

Eutropius obtusirostris Günther, 1864: 53. Type locality: India [in error]. Syntypes: BMNH 1842.3.26.28–29 (2).

Eutropius altipinnis Steindachner, 1894: 57, pl. 1 (fig. 1). Type locality: St. Paul -Flusse, bei Soforch Place. Syntypes: NMW 46501 (1), RMNH 5397 (2).

Distribution: Nile River basin and West Africa (De Vos, 1995).

Remarks: De Vos & Skelton (1990: 324, fig. 1) illustrated the holotype of *Silurus mystus* Linnaeus, and discussed nomenclatural history of the specimen but, unfortunately, mislabeled the photographed specimen as NRM 63 (rather than the correct number: NRM 70). De Vos (1995) concluded that *Eutropius obtusirostris* types probably originated in West Africa.

***Schilbe nyongensis* (De Vos, 1981)**

Eutropius nyongensis De Vos, 1981: 968, fig. 4. Type locality: Nyong river, above falls at 12 km. S.-E. of Eseka, Cameroon. Holotype: MRAC 75-4-P-43.

Distribution: Nyong River, Cameroon, known only from type locality (De Vos, 1995).

***Schilbe tumbanus* (Pellegrin, 1926)**

Eutropius tumbanus Pellegrin, 1926: 203. Type locality: Tondu (Lac Tumba); Congo belge. Syntypes (7): MNHN 1926-0161(1), MNHN 1926-0162 (1), MNHN 1926-0163 (1), MRAC 19685-86 (1), MRAC 19686 (1), MRAC 19687 (1), MRAC 19688 (1).

Distribution: Middle Congo River basin, abundant in Lake Tumba (De Vos, 1995).

***Schilbe uranoscopus* Rüppell, 1832**

Schilbe uranoscopus Rüppell, 1832: 4, pl. 1 (fig. 1). Type locality: Ägypten, Nil bei Cairo. Lectotype: BMNH 1850.7.29.22, designated by De Vos (1986: 45).

Schilbe Isidori Valenciennes, in Cuvier & Valenciennes, 1840a: 375 (278 of Strasbourg deluxe edition), pl. 412. Type locality: Egypte. Syntypes (no larger than 10 pouces): at MNHN.

Distribution: Nile River basin, Turkana system, Chad basin, Niger system, and Cross River (De Vos, 1995).

***Schilbe yangambianus* (Poll, 1954)**

Eutropius yangambianus Poll, 1954: 61, fig. 4. Type locality: riv. Lubulu Iles du fleuve. Holotype: MRAC 123625.

Distribution: Congo River basin and one location in upper Zambezi River (Skelton, 1993; De Vos, 1995).

***Schilbe zairensis* De Vos, 1995**

Schilbe zairensis De Vos, 1995: 285, fig. 201. Type locality: Lemfu, Inkisi River, Lower Zaire Basin, Zaire, ca. 5°18'S, 15°13'E. Holotype: MRAC 86-21-P-98.

Distribution: Lower Congo River basin (De Vos, 1995).

***SILONIA* Swainson, 1838**

Silonia Swainson, 1838: 345 et seq. Type species: *Ageniosus (Silonia) lurida* Swainson, 1838. Type by monotypy. Gender: Feminine. Originally proposed as a subgenus of *Ageniosus*.

Silundia Valenciennes, in Cuvier & Valenciennes, 1840b: 48 (36 of Strasbourg deluxe edition). Type species: *Silundia gangetica* Valenciennes, 1840. Type by subsequent designation by Bleeker (1862b: 399). Gender: Feminine.

Silondia Günther, 1864: 65. Type species: *Silundia gangetica* Valenciennes, 1840. Unjustified emendation of *Silundia* Valenciennes, 1840. Gender Feminine.

Silonopangasius Hora, 1937a: 352. Type species: *Ageneiosus childreni* Sykes, 1839. Type by original designation. Gender: Masculine.

***Silonia childreni* (Sykes, 1839)**

Ageneiosus childreni Sykes, 1839: 165. Type locality: Deccan, India. No types known. Also proposed as new in Sykes (1840: 62). Illustrated and described in more detail in Sykes (1841: 375, pl. 66, fig. 3).

Silundia Sykesii Day, 1876: 569. Type locality: Deccan and Kurnool, India. Syntypes: AMS B.8084 (1), NMW 44624 (1), ZSI F1230 (1). Originally as “*Silundia Sykesii*, n.s.? = ? *Ageneiosus Childreni*, Sykes, Bleeker, & Jerdon.”

Distribution: Cauvery, Godavari and Krishna River basins, India (Jayaram, 1999).

***Silonia silondia* (Hamilton, 1822)**

Pimelodus silondia Hamilton, 1822: 160, 375, pl. 7 (fig. 50). Type locality: Gangetic estuaries. No types known.

Ageniosus (Silonia) lurida Swainson, 1838: 345, fig. 85. Type locality: Gangetic estuaries. No types known. Name made available to reference to Ham. pl. 7 fig. 50 [= Hamilton, 1822, pl. 7, fig. 50]. Unneeded new name for *Pimelodus silondia* Hamilton.

Silundia Gangetica Valenciennes, in Cuvier & Valenciennes, 1840b: 49 (36 of Strasbourg deluxe edition), pl. 426. Unneeded new name for *Pimelodus silondia* Hamilton, 1822, apparently to avoid tautonymy.

Distribution: Northern India, Bangladesh, Myanmar and Nepal (Jayaram, 1999).

Remarks: Redescribed in Hora (1938a).

***SILURANODON* Bleeker, 1858**

Siluranodon Bleeker, 1858b: 253, 255, 256. Type species: *Silurus auritus* Geoffroy St. Hilaire, 1809. Type by monotypy. Gender: Masculine.

***Siluranodon auritus* (Geoffroy Saint-Hilaire, 1809)**

Silurus auritus Geoffroy Saint-Hilaire, 1809: pl. 11 (figs. 1–2). Type locality: fl. Nil, Egypt. Possible holotype or syntypes: MNHN a-8954 (4), MNHN a-8956 (1). Name available from caption on plate, with illustrated specimen the holotype, if identifiable. Description of species in Geoffroy St. Hilaire (1827: 291–299).

Distribution: Nile, Chad, Niger, Volta, Comoé Rivers (De Vos, 1995).

Genus *inquirendum*, Schilbidae

Schillee Jerdon, 1849: 335. Type species: apparently not designated; name based on two species. Name credited to Cuvier, which may refer to *Schilbe*, but included species belong to two other genera.

SCOLOPLACIDAE Bailey & Baskin, 1976

Scoloplacinae Bailey & Baskin, 1976: 5. Type genus: *Scolopax* Bailey & Baskin, 1976.

Revision: Schaefer *et al.* (1989).

Phylogeny: Schaefer (1990).

Taxonomic summary: Schaefer (2003a).

1 genus, 4 species; no named fossil taxa.

***SCOLOPLAX* Bailey & Baskin, 1976**

Scolopax Bailey & Baskin, 1976: 5. Type species: *Scolopax dicra* Bailey & Baskin, 1976. Type by original designation. Gender: Feminine.

***Scolopax dicra* Bailey & Baskin, 1976**

Scolopax dicra Bailey & Baskin, 1976: 7, figs. 1–3. Type locality: Isolated Ox-Bow lagoon off the Río Iténez, ca. 400 m southwest of the river at a point opposite Costa Marques (Brazil), Dept. of Beni, Bolivia, 12°28.38'S, 64°16.59'W. Holotype: AMNH 32482.

Distribution: Amazon and Paraguay River basins (Schaefer, 2003a).

***Scolopax distolothrix* Schaefer, Weitzman & Britski, 1989**

Scolopax distolothrix Schaefer, Weitzman & Britski, 1989: 191, figs. 5, 6. Type locality: Small tributary of Rio Batovi, upper Rio Xingu, Mato Grosso, Brazil, approx. 12°58'S, 53°37'W. Holotype: MZUSP 39065.

Distribution: Tocantins/Araguaia, Xingu and Paraguay River basins (Schaefer, 2003a).

***Scolopax dolicholophia* Schaefer, Weitzman & Britski, 1989**

Scolopax dolicholophia Schaefer, Weitzman & Britski, 1989: 196, figs. 9, 10. Type locality: In igarapé, trib. of Rio Tarumãzinho, approximately 45 km north of Manaus, Amazonas, Brazil, 2°42'S, 60°03'W. Holotype: MZUSP 6788.

Distribution: Negro River basin, Brazil (Schaefer, 2003a).

***Scolopax empousa* Schaefer, Weitzman & Britski, 1989**

Scolopax empousa Schaefer, Weitzman & Britski, 1989: 194, figs. 7, 8. Type locality: Rio Ivinheima 70 km upstream from its confluence with Rio Paraná and Rio dos Bandeirantes, Mato Grosso, Brazil, approx. 22°35'S, 53°30'W. Holotype: MZUSP 39075.

Distribution: Amazon and Paraguay/Paraná River basins (Schaefer, 2003a).

SILURIDAE Cuvier, 1816

Siluroïdes Cuvier, 1816: 199. Type genus: *Silurus* Linnaeus, 1758.

Kryptopterini Bleeker, 1862 (in Bleeker, 1862–63): 18, 85. Type genus: *Kryptopterus* Bleeker, 1857.

Phalacronotini Bleeker, 1862 (in Bleeker, 1862–63): 18, 90. Type genus: *Phalacronotus* Bleeker, 1857.

Monophyly: Bornbusch (1991b).

Phylogeny: Bornbusch (1991b; 1995).

Revision: Kobayakawa (1989, *Silurus* sensu lato, including *Pterocryptis*).

Review: Chen (1977, China).

Checklist: Haig (1952, with key to genera).

Identification guide: Jayaram (1977d, South Asia).

Remarks: Although the monophyly of the family appears to be well supported (see Bornbusch, 1991b), the limits of many of the included genera are not clear. As such, the composition of the genera as listed herein are likely to change with additional study.

12 genera, 94 species; 3 named fossil species of uncertain status.

Incertae sedis

***Kryptopterus indicus* Datta, Barman & Jayaram, 1987**

Kryptopterus indicus Datta, Barman & Jayaram, 1987: 29, fig. 1. Type locality: Hornbill Point, Namdapha River, Namdapha Wildlife Sanctuary, Arunachal Pradesh, India. Holotype: ZSI FF1699.

Distribution: Known only from type locality (Jayaram, 1999).

Remarks: Generic placement undetermined. Illustration of type clearly not that of a species of *Kryptopterus*.

BELODONTICHTHYS Bleeker, 1857

Belodontichthys Bleeker, 1857b: 472. Type species: *Belodontichthys macrochir* Bleeker, 1857. Type by monotypy. Gender: Masculine.

Phylogeny: Howes & Fumihito (1991).

Belodontichthys dinema (Bleeker, 1851)

Wallago dinema Bleeker, 1851d: 202. Type locality: Banjermasin. Syntypes (2, 190, 195 mm TL): possibly RMNH 6834 (1 or 2 of 5. Syntype illustrated in Bleeker (1862–63: pl.86 [= Silur. 38], fig. 2) as *Belodontichthys macrochir*.

Belodontichthys macrochir Bleeker, 1857b: 472. Type locality: Banjermasin. Syntypes (2, 190, 195 mm TL): possibly RMNH 6834 (1 or 2 of 5). Unneeded new name for *Wallago dinema* Bleeker, 1851.

Distribution: Laos, Vietnam, Thailand, Malay Peninsula, Sumatra and Borneo (Roberts, 1989a).

Remarks: BMNH 1863.12.4.54 (1, 272 mm SL) is incorrectly registered as a type of *Belodontichthys macrochir* [= *Wallago dinema*], but is too long to be either of the syntypes.

Belodontichthys truncatus Kottelat & Ng, 1999

Belodontichthys truncatus Kottelat & Ng, 1999: 388, fig. 1. Type locality: Chao Phraya River, flood-waters 17.5 km north of Ayuthya, Maharaj Prov., Thailand. Holotype: UMMZ 235105.

Distribution: Mekong and Chao Phraya River basins (Kottelat, 2001b).

CERATOGLANIS Myers, 1938

Ceratoglanis Myers, 1938: 98. Type species: *Hemisilurus scleronema* Bleeker, 1863. Type by original designation. Gender: Masculine.

Ceratoglanis pachynemus Ng, 1999

Ceratoglanis pachynema Ng, 1999c: 390, figs. 3, 5, 6. Type locality: Prachinburi Market, Thailand. Holotype: CAS 96577.

Distribution: Chao Phraya and Mekong River basins (Ng, 1999c).

Ceratoglanis scleronemus (Bleeker, 1863)

Hemisilurus scleronema Bleeker, 1863 (in Bleeker, 1862–63): 93, pl. 101 [= Silur. pl. 53]. Type locality: Java (Krawang). Holotype (399 mm TL): possibly RMNH 2918 (1). Also described as new in Bleeker (1863e and 1863f).

Distribution: Pahang River basin, peninsular Malaysia, Baram, Barito, Kapuas and Rejang river basins, Borneo, Citarum River basin, Java, and the Batang Hari and Siak River basins, Sumatra (Ng, 1999c).

HEMISILURUS Bleeker, 1857

Hemisilurus Bleeker, 1857b: 472. Type species: *Wallago heterorhynchus* Bleeker, 1853. Type by monotypy. Gender: Masculine.

Diastatomystes Vaillant, 1891: 182. Type species: *Diastatomystes chaperi* Vaillant, 1891. Type by monotypy. Gender: Masculine. Also appeared in Vaillant (1893c: 61).

Phylogeny and biogeography: Bornbusch & Lundberg (1989).

Hemisilurus heterorhynchus (Bleeker, 1853)

Wallago heterorhynchus Bleeker, 1853b: 514. Type locality: Moara kompeh, Sumatrae orientalis, in fluviiis. Holotype (326 mm TL): RMNH 6849.

Diastatomystes Chaperi Vaillant, 1891: 182. Type locality: Knapei River, Borneo. Holotype: MNHN 1891-0458. Also described as new in Vaillant (1893c: 61); redescribed and illustrated in Vaillant (1894b: 70, pl. 2, fig. 2).

Distribution: Sumatra, and Borneo (Roberts, 1989a).

Remarks: BMNH 1863.12.4.58 (~205 mm SL) is incorrectly listed as a type of *Wallago heterorhynchus* as the species was named on a single individual and this specimen is too small.

Hemisilurus mekongensis Bornbusch & Lundberg, 1989

Hemisilurus mekongensis Bornbusch & Lundberg, 1989: 435, figs. 1–4. Type locality: Thailand, Ubon Ratchathani

Province, Khong Chiam District, Mun River at Ban Dan, upstream of confluence with Mekong River, 15°18'N, 105°31'E. Holotype: UMMZ 214565.

Distribution: Mekong River basin (Kottelat, 2001b).

***Hemisilurus moolenburghi* Weber & de Beaufort, 1913**

Hemisilurus moolenburghi Weber & de Beaufort, 1913: 212, figs. 84–85. Type locality: Batang Hari, Sumatra. Syntypes: ZMA 113564 (2).

Distribution: Batang Hari, Sumatra, and western Borneo (Roberts, 1989a).

***KRYPTOPTERUS* Bleeker, 1857**

Kryptopterus Bleeker, 1857b: 472. Type species: *Kryptopterus micropus* Bleeker, 1857. Type by subsequent designation by Bleeker (1862: 395, or 1862–63: 18). Gender: Masculine.

Kryptopterichthys Bleeker, 1857b: 472. Type species: *Silurus palembangensis* Bleeker, 1852. Type by subsequent designation by Bleeker (1862: 395, or 1862–63: 18). Gender: Masculine.

Cryptopterus Günther, 1864: 6, 38. Type species: *Kryptopterus micropus* Bleeker, 1857. Type by being a replacement name. Gender: Masculine. Unjustified emendation of *Kryptopterus*. Preoccupied by *Cryptopterus* Kaup, 1860, in fishes.

Cryptopterella Fowler, 1944a: 1. Type species: *Cryptopterella beldti* Fowler, 1944. Type by original designation. Gender: Feminine.

Remarks: First reviser action in making *Kryptopterus* the valid name for this taxon appears to have been accomplished in Haig (1952: 106), in which *Kryptopterichthys* is treated as a junior synonym of *Kryptopterus*, although the names were attributed to Bleeker (1858) which was, until recently, thought to be the source of both names.

***Kryptopterus baramensis* Ng, 2002**

Kryptopterus baramensis Ng, 2002c: 68, figs. 1–2. Type locality: Borneo: Sarawak, Sungai Akah, above Long Tebangan, 300 m below confluence with Sungai Pahang, below falls, 3°22'12"N, 114°56'06"E. Holotype: ROM 72477.

Distribution: Baram River basin, northern Borneo (Ng, 2002c).

***Kryptopterus bicirrhos* (Valenciennes, 1840)**

Silurus bicirrhos Valenciennes, in Cuvier & Valenciennes, 1840a: 367 (272 of Strasbourg deluxe edition), pl. 411. Type locality: Java. Holotype: MNHN a-9932; radiograph of holotype in Roberts (1989a: fig. 113, lower).

Cryptopterus amboinensis Günther, 1864: 40, 429. Type locality: Amboyna [apparently in error]. Syntypes (2): BMNH 1855.3.24.14 (1).

Distribution: Mekong and Chao Phraya River basins, and Sundaland (Kottelat, 2001b).

***Kryptopterus cryptopterus* (Bleeker, 1851)**

Silurus cryptopterus Bleeker, 1851b: 270. Type locality: Bandjarmassing. Holotype (110 mm TL): RMNH 6840 (1 of 16).

Kryptopterus micropus Bleeker, 1857b: 472. Type locality: Bandjarmassing. Holotype (110 mm TL): RMNH 6840 (1 of 16). Unneeded replacement name for *Silurus cryptopterus* Bleeker, 1851, apparently to avoid tautomy.

Distribution: Borneo and Sumatra, and possibly Tapi River (Ng, 2003i: 8).

***Kryptopterus dissitus* Ng, 2001**

Kryptopterus dissitus Ng, 2001a: 198, fig. 1. Type locality: Laos: Champasak Province, Mekong River at Ban Hang Khone, just downstream from Khone falls. Holotype: UMMZ 238017.

Distribution: Chao Phraya and Mekong River basins (Ng, 2001a).

***Kryptopterus eugeneiatus* (Vaillant, 1893)**

Callichrous eugeneiatus Vaillant, 1893c: 61. Type locality: riv. Knapei et Sebroeang, près Smitow, Bornéo. Syntypes: MNHN 1891-0459 (1), MNHN 1891-0460 (1).

Distribution: Kapuas River basin (Roberts, 1989a).

***Kryptopterus geminus* Ng, 2003**

Kryptopterus geminus Ng, 2003i: 2, figs 1, 2a. Type locality: Cambodia: Stung Treng, Mekong River 2 km downstream from mouth of Tonle San on sandbars, Mekong River drainage, 13°31'N, 105°56'E. Holotype: UMMZ

234664.

Distribution: Mekong, Mae Khlong, Bang Pakong and Chao Phraya River basins (Ng, 2003i).

***Kryptopterus hesperius* Ng, 2002**

Kryptopterus hesperius Ng, 2002c: 70, fig. 3. Type locality: Thailand: Kanchanaburi Province, Kwae Noi River at Sai Yok. Holotype: ZMUC P28551.

Distribution: Mae Khlong River basin, western Thailand (Ng, 2002c).

***Kryptopterus lais* (Bleeker, 1851)**

Silurus lais Bleeker, 1851e: 428. Type locality: Sambas, in fluviis. Holotype (125 mm TL): RMNH 6839 (larger of 2).

Distribution: Sambas, Kapuas, Kahajan and Barito rivers, Borneo (Roberts, 1989a).

***Kryptopterus limpok* (Bleeker, 1852)**

Silurus limpok Bleeker, 1852d: 583. Type locality: Palembang, Sumatra. Holotype (175 mm TL): possibly BMNH 1863.12.4.100 (150 mm SL).

Distribution: Mekong and Chao Phraya River basins, Malay Peninsula, Sumatra and Borneo (Kottelat, 2001b).

***Kryptopterus lumholtzi* Rendahl, 1922**

Kryptopterus lumholtzi Rendahl, 1922: 200. Type locality: Bulungan, northeastern Borneo. Holotype: ZMUC J5310.

Distribution: Bulungan, northeastern Borneo (Rendahl, 1922).

***Kryptopterus macrocephalus* (Bleeker, 1858)**

Kryptopterichthys macrocephalus Bleeker, 1858b: 293. Type locality: Sumatra ?; Padang ?, in fluviis. Holotype (113 mm TL): possibly BMNH 1863.12.4.99 (96 mm SL) or BMNH 1863.12.4.101. Illustrated in Bleeker (1862–63: pl. 89 [= Silur. pl. 41], fig. 1).

Distribution: Peninsular Thailand, Malay Peninsula, Sumatra and Borneo (Roberts, 1989a).

***Kryptopterus minor* Roberts, 1989**

Kryptopterus minor Roberts, 1989a: 149, fig. 115. Type locality: Mainstream of Sungai Pinoh at Nanga Saian, 45 km S of Nangapinoh, Western Borneo (Kalimantan Barat, Indonesia). Holotype: MZB 3638. Name spelled *Kryptopterus minimus* on p. 146. As there has not been a first reviser action to fix the name, it is done here, with *Kryptopterus minor* as the valid name.

Distribution: Kapuas River, western Borneo (Roberts, 1989a).

***Kryptopterus mononema* (Bleeker, 1847)**

Silurus mononema Bleeker, 1847b: 8. Type locality: Surakarta, Java. Syntypes: possibly BMNH 1863.12.4.93 (1, 145 mm SL), RMNH 6835 (2). One syntype illustrated in Bleeker (1862–63: 87, pl. 91 [= Silur pl. 43], fig. 1). Also described as new in Bleeker (1847: 166).

Distribution: Java, Indonesia.

***Kryptopterus palembangensis* (Bleeker, 1852)**

Silurus palembangensis Bleeker, 1852d: 584. Type locality: Palembang, in fluviis. Holotype (169 mm TL): Whereabouts unknown.

Distribution: Sumatra.

Remarks: BMNH 1863.12.4.101 (120 mm SL) is incorrectly registered as a type of *Silurus palembangensis* but appears to be too small to be the holotype.

***Kryptopterus paraschilbeides* Ng, 2003**

Kryptopterus paraschilbeides Ng, 2003d: 3, fig. 1. Type locality: Cambodia: Kompong Chhnang, Tonle Sap River, 17 km upstream from Kompong Chhnang. Holotype: UMMZ 238788.

Distribution: Mekong River basin (Ng, 2003d).

***Kryptopterus piperatus* Ng, Wirjoatmodjo & Hadiaty, 2004**

Kryptopterus piperatus Ng, Wirjoatmodjo & Hadiaty, 2004: 92, fig. 1. Type locality: Indonesia, Sumatra: Aceh Selatan: Sungai Lembang in front of camp, Suag Balimbang Research Station, Gunung Leuser National Park. Holotype: MZB 8717.

Distribution: Lembang River, Alas River basin, Sumatra, Indonesia (Ng *et al.*, 2004).

***Kryptopterus sabanus* (Inger & Chin, 1959)**

Ompok sabanus Inger & Chin, 1959: 282. Type locality: Segama River at the Segama Estate near Lahad Datu, Lahad Datu District, East Coast Residency, North Borneo. Holotype: FMNH 44828.

Distribution: Kapuas and Kinabatangan Rivers, northern Borneo (Roberts, 1989a).

Kryptopterus schilbeides (Bleeker, 1858)

Hemisilurus schilbeides Bleeker, 1858b: 297. Type locality: Palembang, Sumatra; Bandjermassin, Borneo. Syntypes (2: 96–99 mm TL): possibly BMNH 1863.12.4.157 (1, 79 mm SL, 91 mm TL), BMNH 1864.5.15.6 (1). Illustrated in Bleeker (1862–63: pl. 90 [=Silur. pl. 42], fig. 4).

Distribution: Lower Mekong River to Indonesia, in rivers, canals, ditches and swamps (Rainboth, 1996).

Remarks: Register number of syntype of *Hemisilurus schilbeides* incorrectly reported as BMNH 1863.12.11.157 in Eschmeyer *et al.* (1998).

Species inquirendae, Kryptopterus

Cryptopterella beldti Fowler, 1944a: 2, unnumbered figure. Type locality: Borneo (aquarium fish). Holotype: ANSP 71571.

MICRONEMA Bleeker, 1857

Micronema Bleeker, 1857b: 472. Type species: *Silurus hexapterus* Bleeker, 1851. Type by monotypy. Gender: Neuter.

Micronema cheveyi (Durand, 1940)

Cryptopterus Cheveyi Durand, 1940: 19, pl. 4. Type locality: Rivière de Kaskos [Cambodia]. Holotype: possibly at Institut Océanographique Nhatrang.

Distribution: Mekong River basin (Chu *et al.*, 1999), in rivers and canals (Rainboth, 1996).

Micronema hexapterus (Bleeker, 1851)

Silurus hexapterus Bleeker, 1851d: 203. Type locality: Bandjermassing. Holotype (145 mm TL): Whereabouts unknown.

Distribution: Thailand to Indonesia, in rivers, streams, and canals (Rainboth, 1996).

Remarks: BMNH 1863.12.4.73 is incorrectly registered as the holotype of *Silurus hexapterus* Bleeker, but it is much larger (159 mm SL) than the listed length of the holotype.

Micronema moorei (Smith, 1945)

Kryptopterus moorei Smith, 1945: 342, fig. 78. Type locality: Menam Chao Phya, near Paknampo, central Thailand. Holotype: USNM 109787.

Distribution: Chao Phraya and lower Mekong River basins, in streams and canals of the floodplain (Rainboth, 1996).

Remarks: Treated as a probable synonym of *Kryptopterus cheveyi* in Kottelat (2001b).

Micronema platypogon (Ng, 2004)

Kryptopterus platypogon Ng, 2004f: 2, figs. 1, 2a. Type locality: Borneo: Sarawak, Rajang River drainage, market at Sibu, 2°17'18.6"N 111°49'49.2"E. Holotype: ZRC 45838.

Distribution: Rajang River basin, northern Borneo (Ng, 2004f).

OMPOK La Cepède, 1803

Ompok La Cepède, 1803: 49. Type species: *Ompok siluroides* La Cepède, 1803. Type by monotypy. Gender: Masculine.

Callichrous Hamilton, 1822: 149. Type species: *Silurus pabda* Hamilton, 1822. Type by subsequent designation by Bleeker (1862: 395 or 1862–63: 17). Gender: Masculine. Originally proposed as a subgenus of *Silurus*.

Pseudosilurus Bleeker, 1857b: 472. Type species: *Wallago leiacanthus* Bleeker, 1853. Type by monotypy. Gender: Masculine.

Silurodes Bleeker, 1857b: 472. Type species: *Silurus macronema* Bleeker, 1851. Type by monotypy. Gender: Masculine. Also in Bleeker (1858b: 255, 256, 271).

Revision: Ng (2003b, *Ompok hypophthalmus* group); Tan & Ng (1996, *Ompok leiacanthus* species group).

Review: Parameswaran *et al.* (1967, Indian species).

***Ompok bimaculatus* (Bloch, 1794)**

Silurus bimaculatus Bloch, 1794: 24, pl. 364. Type locality: Tranquebar. Lectotype: ZMB 2916, designated by, and illustrated in, Paepke (1999: 139, pl. 25, fig. 2).

Callichrous ceylonensis Günther, 1864: 46. Type locality: Ceylon. Syntypes: BMNH 1852.2.19.107–108 (2), BMNH 1853.3.30.60 (1).

Distribution: Widely distributed in south and Southeast Asia, including Mekong River (Kottelat, 2001b), but probably more restricted to just south Asia and Sri Lanka.

Remarks: See Jayaram (1977a) for comments on type locality of *Silurus bimaculatus*, and Rema Devi & Emilyamma (1997) for comments on its identity.

***Ompok binotatus* Ng, 2002**

Ompok binotatus Ng, 2002b: 26, fig. 1. Type locality: Indonesia: Borneo, Kalimantan Barat, Sungai Mandai Kechil near its confluence with Kapuas mainstream, 18 km west southwest of Putussibau, 0°48'N, 112°47'E. Holotype: FMNH 94243.

Distribution: Kapuas River basin, Borneo (Ng, 2002b).

***Ompok borneensis* (Steindachner, 1901)**

Callichrous (Silurodes) borneensis Steindachner, 1901: 445, pl. 18 (fig. 3). Type locality: Baram-Flusse, Borneo. Holotype (8.4 cm): at NMW or SMF.

Distribution: Baram River basin, Borneo (Steindachner, 1901).

***Ompok fumidus* Tan & Ng 1996**

Ompok fumidus Tan & Ng 1996: 537, figs. 3, 4c. Type locality: North Selengor peat swamp forest, 43 km on road from Tanjung Malim to Sungai Besar, Selangor, Malaysia. Holotype: ZRC 15049.

Distribution: peninsular Malaysia (Tan & Ng, 1996).

***Ompok hypophthalmus* (Bleeker, 1846)**

Silurus hypophthalmus Bleeker, 1846a: 149. Type locality: Batavia. Type(s) (size and number not stated): whereabouts unknown.

Silurus macronema Bleeker, 1851: 203. Type locality: Bandjermassing. Holotype (140 mm TL): possibly BMNH 1863.12.4.155 (121 mm SL); illustrated in Bleeker (1862–63: 83, pl. 88 [= Silur. pl. 40], fig. 2), as *Silurodes macronema*.

Distribution: Ciliwung and Brantas River basins, Java, and Barito River basin, southern Borneo (Ng, 2003b).

***Ompok jaynei* Fowler, 1905**

Ompok jaynei Fowler, 1905: 466, fig. 3. Type locality: Baram basin, Baram region of Sarawak, Borneo. Holotype: ANSP 114890.

Distribution: Baram River basin, Borneo (Fowler, 1905).

***Ompok leiacanthus* (Bleeker, 1853)**

Wallago leiacanthus Bleeker, 1853a: 189. Type locality: Marwang, in fluviis [now: East end of Danau Arang Arang, Jambi Prov., Sumatra, Indonesia, by neotype designation]. Neotype: ZRC 38538, designated by Tan & Ng (1996: 532).

Distribution: Sumatra.

***Ompok malabaricus* (Valenciennes, 1840)**

Silurus Malabaricus Valenciennes, in Cuvier & Valenciennes, 1840a: 353 (262 of Strasbourg deluxe edition). Type locality: Malabar. Syntypes: MNHN b-0607 (2).

Distribution: Goa, Kerala, India (Jayaram, 1999).

***Ompok miostomus* Vaillant, 1902**

Wallago miostoma Vaillant, 1902: 44. Type locality: Tepoe, bords du Mahakam, Bornéo central. Syntypes: RMNH 7811 (2); syntype illustrated in Roberts (1982b: fig. 3b).

Distribution: Mahamam River basin, central Borneo (Vaillant, 1902).

***Ompok pabda* (Hamilton, 1822)**

Silurus pabda Hamilton, 1822: 150, 374, pl. 25 (fig. 47). Type locality: ponds and rivers of Bengal. No types known.

Silurus (Callichrus) vittatus Swainson 1839: 306. Type locality: ponds and rivers of Bengal. No types known. Name made available by reference to “Ham. pl. 25, f. 47” [= Hamilton (1822: p. 25, fig. 47)]. Unneeded new name for

Silurus pabda Hamilton, 1822.

Distribution: Brahmaputra and Ganges River basins, India and Bangladesh; Pakistan; Myanmar (Jayaram, 1999).

Ompok pabo (Hamilton, 1822)

Silurus pabo Hamilton, 1822: 153, 375, pl. 22 (fig. 48). Type locality: Brahmaputra River, towards Assam. No types known.

Silurus (Callichrus) erythrogaster Swainson, 1839: 306. Type locality: Brahmaputra River, towards Assam. No types known. Name made available by reference to "Ham. pl. 17 f. 48" [sic pl. 22, fig. 48] (= Hamilton, 1822: pl. 22, fig. 48). Unneeded new name for *Silurus pabo* Hamilton, 1822.

Distribution: Brahmaputra, Ganges, and Yamuna River basins, India and Bangladesh; Pakistan; Myanmar (Jayaram, 1999).

Ompok pinnatus Ng, 2003

Ompok pinnatus Ng, 2003f: 48, fig. 1. Type locality: Cambodia, Tonle Sap at Koompong Chhnang, fishing lot 9 in second channel E of town. Holotype: UMMZ 232679.

Distribution: Chao Phraya River, Thailand and Mekong River basin, Cambodia (Ng, 2003f).

Ompok platyrhynchus Ng & Tan, 2004

Ompok platyrhynchus Ng & Tan, 2004: 2, figs. 1, 2a. Type locality: Borneo: Brunei Darussalam, Temburong district: Belalong sub-basin Sungai Esu, about 15 minutes upstream of Kuala Belalong Field Studies Centre (04°32'17.9"N 115°09'35.2"E). Holotype: ZRC 48678.

Distribution: Temburong River basin, northern Borneo (Ng & Tan, 2004).

Ompok pluriradiatus Ng, 2002

Ompok pluriradiatus Ng, 2002b: 28, fig. 3. Type locality: Indonesia: Borneo: Kalimantan Timur: Mahakam River drainage, a swift blackwater stream entering Mahakam River from the left side downriver of Nuarapahu (0°14'S, 116°07'E). Holotype: MZB 5951.

Distribution: Mahakam River basin, eastern Borneo (Ng, 2002b).

Ompok rhadinurus Ng, 2003

Ompok rhadinurus Ng, 2003b: 1299, figs. 3, 5b. Type locality: Peninsular Malaysia, Selangor, North Selangor Peat Swamp Forest, irrigation canal on western boundary. Holotype: ZRC 14897.

Distribution: Bernam, Endau, Pahang, Pattani and Perak River basins, peninsular Malaysia, Kapuas River basin, Borneo, and Batang Hari, Deli, Indragiri and Musi River basins, Sumatra (Ng, 2003b).

Ompok sindensis (Day, 1877)

Callichrous Sindensis Day, 1877 (in Day, 1875–78): 476, pl. 110 (fig. 1). Type locality: Sind, from the Indus [Pakistan]. Holotype: ZSI 505.

Distribution: Sindh, Pakistan (Mirza, 2003).

Ompok urbaini (Fang & Chaux, 1949)

Cryptopterus urbaini Fang & Chaux, in Chaux & Fang, 1949a: 197, fig. 2. Type locality: Cambodge. Holotype: MNHN 1966-0706.

Distribution: Mekong, Chao Phraya and Pasak River basins, Southeast Asia (Ng, 2003b).

Remarks: Ng (2003b) lists three syntypes, but a holotype was clearly indicated in the description.

Ompok weberi (Hardenberg, 1936)

Callichrous weberi Hardenberg, 1936a: 232. Type locality: Padang Tikarbay, Kapuas River basin, western Borneo. Holotype (79 mm): Whereabouts unknown (see Ng & Siebert, 2002, for comments).

Distribution: Kapuas River, Borneo (Ng & Siebert, 2002).

Remarks: Redescribed in Ng & Siebert (2002).

Species inquirendae, Ompok

Ompok siluroïdes La Cepède, 1803: 49, 50, pl. 1 (fig. 2). Type locality: not stated [Asia]. Holotype: MNHN a-8669.

Silurus lamghur Heckel, 1838: 82, pl. 12 (figs. 5–6). Type locality: Kashmir. Holotype: NMW 44689.

Silurus canio Hamilton, 1822: 151, 374. Type locality: Ponds in the north-east parts of Bengal. No types known. Unpublished Hamilton illustration reproduced in Hora (1929: pl. 20, fig. 5).

- Silurus chechra* Hamilton, 1822: 152, 375. Type locality: Kusi river [India]. No types known.
- Silurus duda* Hamilton, 1822: 152, 375. Type locality: river Kusi [India]. No types known.
- Silurus (Callichrus) affinis* Swainson, 1839: 306. Type locality: Kusi River [India]. No types known. Name made available by reference to "Ham. 152 (duda)" [= Hamilton, 1822: 152]. Unneeded new name for *Silurus duda* Hamilton, 1822.
- Silurus (Callichrus) immaculatus* Swainson, 1839: 306. Type locality: Bengal. No types known. Name made available by reference to "Ham. 151 (canio)" [= Hamilton, 1822: 151]. Unneeded new name for *Silurus canio* Hamilton, 1822.
- Silurus (Callichrus) nebulosus* Swainson, 1839: 306. Type locality: Kusi River [India]. No types known. Name made available by reference to "Ham. 152 (nebulosus)" [=Hamilton, 1822: 152)]. Unneeded new name for *Silurus chechra* Hamilton, 1822.
- Silurus anostomus* Valenciennes, in Cuvier & Valenciennes, 1840a: 363 (269 of Strasbourg deluxe edition), pl. 410. Type locality: Bengale. Syntypes: MNHN a-8949 (2), MNHN a-8950 (2).
- Silurus microcephalus* Valenciennes, in Cuvier & Valenciennes, 1840a: 365 (271 of Strasbourg deluxe edition). Type locality: Bengale. Holotype (6 pouces): at MNHN.
- Silurus Mysoricus* Valenciennes, in Cuvier & Valenciennes, 1840a: 364 (270 of Strasbourg deluxe edition). Type locality: Mysore. Syntypes: MNHN 0000-3107 (2).
- Silurus indicus* M'Clelland & Griffith, in M'Clelland, 1842: 583. Type locality: Loodianah, the Punjab, and Cabool River at Jullalabad. Holotype: Whereabouts unknown, possibly at BMNH.
- Cryptopterus latovittatus* Playfair, 1867: 16. Type locality: Cachar, eastern provinces of British India, within the watershed of the Burhampooter. Syntypes: BMNH 1867.2.14.72–73.
- Pseudosilurus macrophthalmos* Blyth, 1860: 156. Type locality: Tenasserim. Holotype: Whereabouts unknown.
- Callichrous nigrescens* Day, 1870c: 616. Type locality: Throughout the branches of the Irrawaddi, in the Pegu and Sittoung rivers, Burma. Syntypes: AMS B.7636 (1), ZSI A.500 (2, lost).
- Callichrous notatus* Day, 1870c: 616. Type locality: Rivers of Burma. Possible syntypes: AMS B.7982 (2), ZSI 1275 (1), ZSI A.499 (2, lost).
- Callichrous egertonii* Day, 1872: 710. Type locality: Subhimalayan range in the Punjaub. Syntype: AMS B.8065 (1).
- Wallago krattensis* Fowler, 1934b: 335, fig. 1. Type locality: Kratt, southeast Siam. Holotype: ANSP 60177.
- Silurus goae* Haig, 1952: 77, fig. on p. 78. Type locality: Goa, India. Holotype: SU 41889. Referred to *Ompok* rather than *Silurus* by Kobayakawa (1989: 176).

PHALACRONOTUS Bleeker, 1857

Phalacronotus Bleeker, 1857b: 472. Type species: *Silurus phalacronotus* Bleeker, 1851. Type by absolute tautonomy. Gender: Masculine. Type designations of *Silurus leptonema* in Bleeker (1862: 395 and 1862–63: 18) are invalid.

Phalacronotus apogon (Bleeker, 1851)

Silurus apogon Bleeker, 1851c: 67. Type locality: Banjermasin. Syntypes (4, 118–175 mm TL): RMNH 6843 (in part).

Silurus leptonema Bleeker, 1852: 584. Type locality: Palembang, Sumatra. Holotype (268 mm TL): possibly at RMNH.

Silurus micropogon Bleeker, 1855: 418, 419. Type locality: Pontianak; Bandjermasin, Borneo. Apparently an unneeded new name for *Silurus apogon* Bleeker, 1851.

Distribution: Mekong and Chao Phraya river basins, Malay Peninsula, Sumatra and Borneo (Kottelat, 2001b).

Phalacronotus bleekeri (Günther, 1864)

Cryptopterus bleekeri Günther, 1864: 44. Type locality: Siam. Syntypes (2): BMNH 1863.12.4.105, and possibly BMNH 1862.11.1.208.

Micronema bleekeri Bocourt, 1866: 17, pl. 1 (figs. 3, 3a–c). Type locality: [Menam R. at Bangkok]. Syntypes: MNHN 0000-1546 (3).

Distribution: Mekong and Chao Phraya River basins, and Sundaland (Kottelat, 2001b).

Remarks: Günther (1864) cited *Micronema bleekeri* Bocourt in his account of *Cryptopterus bleekeri*. However, he did not cite a Bocourt publication in his list of cited literature (pp. vii–x) and available evidence indicates that

Bocourt's publication did not appear until 1866. Therefore, Günther's name has priority and Bocourt's must be treated as independently proposed and available.

***Phalacronotus micronemus* (Bleeker, 1846)**

Silurus micronemus Bleeker, 1846b: 289. Type locality: Batavia. Holotype: RMNH 6841.

Silurus phalacronotus Bleeker, 1851e: 429. Type locality: Sambas. Holotype (220 mm TL): Whereabouts unknown.

Phalacronotus micruropterus Bleeker, 1857b: 473. Type locality: Sambas, in fluviis. Holotype (220 mm TL): Whereabouts unknown. Unneeded replacement name for *Silurus phalacronotus* Bleeker, 1851, to avoid Stricklandian tautonymy.

Micronema typus Bleeker, 1858b: 300. Type locality: Batavia. Holotype: RMNH 6841. Unneeded new name for *Silurus micronemus* Bleeker, 1846.

Kryptopterus deignani Fowler, 1937: 136, figs. 10–12. Type locality: Me Poon, Siam. Holotype: ANSP 67884.

Distribution: Mekong and Chao Phraya River basins, and Sundaland (Kottelat, 2001b).

Remarks: BMNH 1863.12.4.75 (209 mm SL), BMNH 1863.12.4.95 (222 mm SL, 249 mm TL), and BMNH 1863.12.4.96 (122 mm SL) are incorrectly registered as the types of *Silurus micronemus*.

***Phalacronotus parvanalis* (Inger & Chin, 1959)**

Kryptopterus parvanalis Inger & Chin, 1959: 284, fig. 46. Type locality: Kinabatangan River at Deramakot, Kinabatangan District, North Borneo. Holotype: FMNH 68014.

Distribution: Kinabatangan River at Deramakot, North Borneo, Malaysia (Inger & Chin, 1959).

***PINNIWALLAGO* Gupta, Jayaram & Hajela, 1981**

Pinniwallago Gupta, Jayaram & Hajela, 1981: 291. Type species: *Pinniwallago kanpurensis* Gupta, Jayaram & Hajela, 1981. Type by original designation. Gender: Masculine.

***Pinniwallago kanpurensis* Gupta, Jayaram & Hajela, 1981**

Pinniwallago kanpurensis Gupta, Jayaram & Hajela, 1981: 290, fig. 1. Type locality: 'Bara Tal' near village of Bhitaragaon, Tehsil, Ghatampur, Kanpur, Uttar Pradesh, India. Holotype: ZSI FF1443.

Distribution: Known only from the type locality in Uttar Pradesh, India (Jayaram, 1999).

***PTEROCRYPTIS* Peters, 1861**

Pterocryptis Peters, 1861: 712. Type species: *Pterocryptis gangelica* Peters, 1861. Type by monotypy. Gender: Feminine.

Apodoglanis Fowler, 1905: 463. Type species: *Apodoglanis furnessi* Fowler, 1905. Type by original designation. Gender: Masculine.

Hito Herre, 1924a: 702. Type species: *Hito taytayensis* Herre, 1924. Type by original designation. Gender: Masculine.

Hitoichthys Herre 1924b: 1570. Type species: *Hitoichthys taytayensis* Herre, 1924. Type by original designation. Gender: Masculine.

Penesilurus Herre, 1924a: 703. Type species: *Penesilurus palavanensis* Herre, 1924. Type by original designation. Gender: Masculine. Also described as new in Herre (1924b: 1570); earliest publication not established.

Herklotrella Herre, 1933: 179. Type species: *Herklotrella anomala*, Herre, 1933. Type by original designation. Gender: Feminine. Publication date stated to be Dec. 1933, but perhaps did not appear until later (Eschmeyer, 1998); priority over *Herklotrella* Fowler, 1934, which was published in January, 1934, is not clearly established.

Review: Ng & Freyhof (2001a), Vietnam.

Remarks: The revision by Kobayakawa (1989) included many of these species in an expanded *Silurus* sensu lato. Haig (1952: 82) gave precedence to *Hito* over *Hitoichthys*, after concluding that priority of publication could not be determined.

***Pterocryptis anomala* (Herre, 1933)**

Herklotrella anomala Herre, 1933: 179. Type locality: Hong Kong market. Holotype: SU 26769.

Silurus sinensis Hora, 1937d: 343, figs. 8 (c–d). Type locality: Lunchow, China. Holotype: Originally ZMFMIB 13692, apparently now at ASIZB. Name proposed for specimen identified as *Silurus wynaudensis* in Tchang (1936: 35). Preoccupied by *Silurus sinensis* La Cepède, 1803; replaced by *Silurus gilberti* Hora, 1938.

Silurus gilberti Hora, 1938c: 243. Type locality: Lunchow, China. Holotype: Originally ZMFMIB 13692, apparently now at ASIZB. Replacement name for *Silurus sinensis* Hora, 1937.

Distribution: Rivers draining southeastern China, from Minjiang and Pearl Rivers to vicinity of Hong Kong (Ng & Chan, 2005).

Remarks: Synonymy follows Ng & Chan (2005).

***Pterocryptis berdmorei* (Blyth, 1860)**

Silurichthys Berdmorei Blyth, 1860: 156. Type locality: Tenasserim provinces. Holotype: ZSI 481 (whereabouts unknown).

Silurus morehensis Arunkumar & Tombi Singh, 1997: 73, fig. 1. Type locality: Moreh Bazar, Moreh, Manipur, India. Holotype: MUMF 2211/1A.

Distribution: Southeastern Myanmar and possibly western Thailand (Ng & Freyhof, 2001a), Manipur, India (Arunkumar & Tombi Singh, 1997).

***Pterocryptis bokorensis* (Pellegrin & Chevey, 1937)**

Penesilurus bokorensis Pellegrin & Chevey, 1937: 315. Type locality: [Bokor, Cambodia, elev. 800–1000 m]. Holotype: MNHN 1936-0167.

Distribution: Cambodia.

Remarks: Ng & Kottelat (1998b) indicate that the species is known only from holotype, but Rainboth (1996) implies that it is a food fish, known from upland streams of the Mekong basin in Cambodia.

***Pterocryptis buccata* Ng & Kottelat, 1998**

Pterocryptis buccata Ng & Kottelat, 1998b: 394, figs. 1, 2a, 3a–b. Type locality: Thailand: Kanchanaburi Province, Amphoe Sai Yok, Mae Khlong basin. Holotype: ZRC 41496.

Distribution: Meklong River basin, Kanchanaburi Province, Thailand; in caves and epigean habitats (Ng & Kottelat, 1998b).

***Pterocryptis burmanensis* (Khin-Thant, 1966)**

Silurus burmanensis Khin-Thant, 1966: 219, pls. 1–3. Type locality: Inlé Lake, s. Shan State, 23°35'N, 96°57'E, Burma, elev. 2915 ft. Holotype: Zoological Museum, Art and Sciences University of Rangoon 332.

Distribution: Inle Lake, Myanmar.

***Pterocryptis cochinchinensis* (Valenciennes, 1840)**

Silurus Cochinchinensis Valenciennes, in Cuvier & Valenciennes, 1840a: 352 (262 of Strasbourg deluxe edition). Type locality: [Cochinchine]. Syntypes: MNHN 0000-0573 (1), MNHN b-0602 (1).

Distribution: Coastal Rivers of central Vietnam, between Vinh and An Lao (Ng & Freyhof, 2001a); Xijiang [= West River], Jiulongjiang, Hainan Island (Chu *et al.*, 1999); Nam Xam basin, Laos (Kottelat, 2001b).

***Pterocryptis crenula* Ng & Freyhof, 2001**

Pterocryptis crenula Ng & Freyhof, 2001a: 630, fig. 6. Type locality: Vietnam: Quang Ninh province, Hai Ninh district, torrent at km 5 on road from Bac Phong Sinh to Mong Cai, 21°35'31"N, 107°43'52"E. Holotype: ZRC 46317.

Distribution: coastal rivers of northeastern Vietnam (Ng & Freyhof, 2001a).

***Pterocryptis cucphuongensis* (Mai, 1978)**

Silurus cucphuongensis Mai, 1978: 245, fig. 112. Type locality: Cuc Phuong, Vietnam. Holotype: DVZUT 345.

Distribution: Song Iuong drainage, northern Vietnam (Ng & Freyhof, 2001a).

***Pterocryptis furnessi* (Fowler, 1905)**

Apodoglanis furnessi Fowler, 1905: 463, fig. 2. Type locality: Baram River, Borneo. Holotype: ANSP 114894; holotype illustrated in Bornbusch (1991a: 1071, fig. 1).

Distribution: Niah and Baram River basins, Sarawak (Bornbusch, 1991a).

Remarks: Redescribed in Bornbusch (1991a).

***Pterocryptis gangelica* Peters, 1861**

Pterocryptis gangelica Peters, 1861: 712. Type locality: Ganges River. Holotype: ZMB 4796; holotype illustrated in Bornbusch (1991a: 1077, fig. 4).

Silurus afghana Günther, 1864: 34. Type locality: Afghanistan [in error: Assam, India]. Holotype: BMNH 1860.3.19.755.

Silurus dukai Day, 1873: 239. Type locality: Darjeeling, India. Syntypes: AMS B.7571 (1), ZSI 1118 (1, lost), ZSI 1232 (1, lost).

Distribution: Northern India (Ng & Chan, 2005).

Remarks: Coad (1981b: 16) discussed the mistake in the reported type locality of *Silurus afghana*. Synonymy of *P. gangelica* and *S. afghana* follows Ng & Chan (2005).

***Pterocryptis inusitata* Ng, 1999**

Pterocryptis inusitata Ng, 1999a: 372, figs. 1–2. Type locality: Laos: Mekong Basin, Nam Theun watershed, Nam Ong at Ban Don. Holotype: ZRC 41455.

Distribution: Nam Kading basin, Mekong River system, Laos (Ng, 1999a).

***Pterocryptis taytayensis* (Herre, 1924)**

Hito taytayensis Herre, 1924a: 703. Type locality: Small fresh-water creek, near Taytay, Palawan, Philippines. Holotype: BSMP 9357 (presumed destroyed).

Hitoichthys taytayensis Herre 1924b: 1570. Type locality: Small fresh-water creek, near Taytay, Palawan, Philippines. Holotype: BSMP 9357 (presumed destroyed).

Penesilurus palawanensis Herre, 1924a: 704. Type locality: Lake Manguao, Palawan, Philippines. Holotype: BSMP (considered destroyed). Also described as new in Herre (1924b: 1570), earliest not established.

Distribution: Palawan, Philippines.

***Pterocryptis torrentis* (Kobayakawa, 1989)**

Silurus torrentis Kobayakawa, 1989: 171, fig. 33. Type locality: Lampae stream, Khaoluk village, Trang, Thailand. Holotype: NSMT-P 50234.

Distribution: Trang and Chataburi, Thailand (Kobayakawa, 1989); Myanmar localities reported by Kobayakawa may refer to specimens of *Pterocryptis berdmorei* (Ng & Freyhof, 2001a).

***Pterocryptis verecunda* Ng & Freyhof, 2001**

Pterocryptis verecunda Ng & Freyhof, 2001a: 636, fig. 9. Type locality: Vietnam: Hai Phong province, Cat Ba Island, Stream near entrance of Trung Trang cave, (20°47'17"N, 107°00'04"E). Holotype: ZRC 46316.

Distribution: Cat Ba Island, northeastern Vietnam (Ng & Freyhof, 2001a).

***Pterocryptis wynnaadensis* (Day, 1873)**

Silurus punctatus Day, 1868: 155. Type locality: Stream in Wynnaad, India, elev. 3000 ft. Possible syntypes: AMS B.7990 (1), BMNH 1868.5.14.6–7 (2), BMNH 1889.2.1.2521–2522 (2), NMW 77251 (1), ZMB 11221 (1), ZSI 461 (1), ZSI 1233 (1), ZSI A.480 (1). Preoccupied by *Silurus punctatus* Rafinesque, 1818, and *Silurus punctatus* Cantor, 1842.

Silurus wynnaadensis Day, 1873c: 237. Type locality: in the Wynnaad, in a stream about 3000 ft. above the level of the sea [India]. Possible syntypes: AMS B.7990 (1), BMNH 1868.5.14.6–7 (2), BMNH 1889.2.1.2521–2522 (2), NMW 77251 (1), ZMB 11221 (1), ZSI 461 (1), ZSI 1233 (1), ZSI A.480 (1). Replacement for *Silurus punctatus* Day, 1868.

Distribution: Kerala, India (Ng & Kottelat, 1998b; Jayaram, 1999).

***SILURICHTHYS* Bleeker, 1856**

Silurichthys Bleeker, 1856: 417, 418. Type species: *Silurus phaiosoma* Bleeker, 1851. Type by monotypy. Gender: Masculine.

Revision: Ng & Ng (1998).

***Silurichthys citatus* Ng & Kottelat, 1997**

Silurichthys citatus Ng & Kottelat, 1997: 204, fig. 1. Type locality: Kalimantan Barat, Sungai Sekumpai, a small forest stream flowing into Sungai Pinoh, 0°32'S, 111°39.5'E, Borneo. Holotype: MZB 3670.

Distribution: Kapuas River basin, western Borneo (Ng & Ng, 1998).

***Silurichthys gibbiceps* Ng & Ng, 1998**

Silurichthys gibbiceps Ng & Ng, 1998: 301, figs. 1b, 1j, 2b, 3b, 6. Type locality: Kalimantan Tengah, Sungai Barito basin, Sungai Paku-merah (0°35.171'S, 115°11.398'E) [Indonesia]. Holotype: MZB 6101.

Distribution: Barito River basin, southern Borneo (Ng & Ng, 1998).

***Silurichthys hasseltii* Bleeker, 1858**

Silurichthys Hasseltii Bleeker, 1858: 270. Type locality: Tjisikat, in fluviis. Neotype: RMNH 2992. Illustrated in Bleeker (1862–63: pl. 87 [= Silur. pl. 39], fig. 1).

Distribution: Western Java, Bangka, Bintan, Batam, Singapore, and southern peninsular Malaysia (Ng & Ng, 1998).

Remarks: Species named on illustration that may have been based on RMNH 2992 or on another specimen that was not preserved. In the absence of conclusive evidence that the RMNH specimen was the basis of the illustration, Ng & Ng (1998: 305) selected RMNH 2992 as the neotype of the species.

***Silurichthys indragiriensis* Volz, 1904**

Silurichthys indragiriensis Volz, 1904: 464. Type locality: Kwantang River, near Djapura (Indragiri), Sumatra.

Holotype: NMW 44622.

Distribution: Central Sumatra, Bintan, Bangka, Billiton and peninsular Malaysia (Ng & Ng, 1998).

***Silurichthys marmoratus* Ng & Ng, 1998**

Silurichthys marmoratus Ng & Ng, 1998: 310, figs. 1e, 1f, 2e, 3e, 11. Type locality: Sarawak: Sungai Sebiris, 13.8 km after Kampung Puteh turnoff, towards Lundu on Sematan-Lundu road (1°41'32"N, 109°47'0.8"E). Holotype: ZRC 40293.

Distribution: Northern and western Borneo (Sarawak, Brunei, and Kalimantan Barat) (Ng & Ng, 1998).

***Silurichthys phaiosoma* (Bleeker, 1851)**

Silurus phaiosoma Bleeker, 1851: 428. Type locality: Sambas, in fluviis. Holotype (82 mm TL): RMNH 6831.

Distribution: Northwestern Borneo (Ng & Ng, 1998).

Remarks: BMNH 1863.12.4.109 (102 mm SL) and BMNH 1864.5.15.5 (79 mm SL, 100 mm TL) are registered as types, but are too large to be the holotype.

***Silurichthys sanguineus* Roberts, 1989**

Silurichthys sanguineus Roberts, 1989a: 151, fig. 119. Type locality: Sungai Tekam, a small forest stream, where it enters Kapuas mainstream about 5–6 km upstream from Sanggau, Western Borneo (Kalimantan Barat, Indonesia). Holotype: MZB 3673; holotype illustrated in Ng & Ng (1998: fig. 13).

Distribution: Kapuas River basin, western Borneo; known only from holotype (Ng & Ng, 1998).

***Silurichthys schneideri* Volz, 1904**

Silurichthys schneideri Volz, 1904: 463. Type locality: Upper Langkat, Danau near Sukaranda [Sumatra]. Holotype: NMW 44623.

Silurichthys leucopodus Fowler, 1939: 56, figs. 4–6. Type locality: Waterfall at Trang, Siam. Holotype: ANSP 68463.

Distribution: Northern Sumatra, northern peninsular Malaysia, southern and southeastern Thailand, and southern Cambodia (Ng & Ng, 1998).

***SILURUS* Linnaeus, 1758**

Silurus Linnaeus, 1758: 304. Type species: *Silurus glanis* Linnaeus, 1758. Type by Linnaean tautonymy. On Official List (ICZN Opinion 77 and Direction 56). Gender: Masculine.

Glanis Agassiz, 1857: 333. Type species: *Glanis aristotelis* Agassiz, 1857. Type by monotypy. Gender: Masculine. Preoccupied by *Glanis* Spix & Agassiz, 1829, in fishes.

Parasilurus Bleeker, 1862b: 392, 394. Type species: *Silurus japonicus* Temminck & Schlegel, 1846. Type by original designation. Gender: Masculine. Also in Bleeker (1862–63: 17).

Revision: Kobayakawa (1989), *Silurus* sensu lato, including *Pterocryptis*.

Remarks: The name *Duanensis* appears in the title of Hu *et al.* (2004) as a generic name for a species referred to in the text as a species of *Silurus*. *Duanensis* is considered a *lapsus calami* and, therefore, a *nomen nudum*.

***Silurus aristotelis* Agassiz, 1857**

Glanis Aristotelis Agassiz, 1857: 333. Type locality: Greece. Syntypes: MCZ 7938 (6), USNM 55895 (1).

Silurus (Parasilurus) aristotelis Garman, 1890: 56. Type locality: Acheloüs River, Acarnania, Greece. Syntypes: MCZ 7938 (6), USNM 55895 (1).

Distribution: Balkan Peninsula, Greece (Kobayakawa, 1989).

***Silurus asotus* Linnaeus, 1758**

Silurus Asotus Linnaeus, 1758: 304. Type locality: Asia. Type(s): Whereabouts unknown. Neotype designation by

Chen (1977: 206, 216) apparently valid inasmuch as ten specimens were all designated as a collective neotype (M. Kottelat, pers. commun.).

Silurus inermis Houttuyn, 1782: 338. Type locality: Japanse. No types known. Preoccupied by *Silurus inermis* Linnaeus, 1766; effectively replaced by *Silurus imberbis* Gmelin, 1789.

Silurus dauricus Pallas 1787: 359, pl. 11 (fig. 11). Type locality: Ingoda, Onone et Arguno Dauuriae fluuiis. Type(s) (20" 3" TL): possibly at ZIN or ZMB.

Silurus imberbis Gmelin, 1789:1361. Type locality: Japonia. On *Silurus inermis* Houttuyn and effectively replacing that name. Apparently intended as a new name to avoid the homonymy with *Silurus inermis* Linnaeus, 1766.

Centranodon japonicus La Cepède, 1803: 138, 139. Type locality: Japan. On *Silurus inermis* Houttuyn (1782: 338, no. 27) and *Silurus imberbis* Gmelin.

Silurus punctatus Cantor, 1842: 485. Type locality: Chusan [China]. Syntypes: BMNH 1968.3.11.29 (3); possibly BMNH 1843.7.21.6, BMNH 1843.7.21.24, and BMNH 1843.7.21.25 (not found), also BMNH 1860.3.19.736–737 (2), BMNH 1860.3.19.785–786 (2). Preoccupied by *Silurus punctatus* Rafinesque, 1818; apparently not replaced.

Silurus xanthosteus Richardson, 1845: 133, pl. 56 (figs. 12–14). Type locality: Chusan, Canton, China. Syntypes (3): BMNH 1968.3.11.29 (1) and specimen used for drawing by Reeves.

Silurus japonicus Temminck & Schlegel, 1846, in Temminck & Schlegel, 1843–50: 226, pl. 104 (fig. 1). Type locality: Nagasaki, Japan. Lectotype: RMNH D675, designated by Boeseman (1947: 169).

Silurus cinereus Dabry de Thiersant, 1872: 189, pl. 47 (fig. 1). Type locality: Yang-tsee-kiang [China]. Type(s): possibly at MNHN.

Silurus bedfordi Regan, 1908g: 61, pl. 2 (fig. 3). Type locality: Kimhoa [and] Chong-ju, Corea. Syntypes: BMNH 1907.12.10.66 (1), BMNH 1907.12.10.67 (1).

? *Parasilurus asotus longus* Wu, 1930a: 255, fig. 1. Type locality: Creek at Tian-Tai Mountain, Chekiang [Zhejiang], China. Syntypes (2): possibly at MNHN. Originally as *Parasilurus asotus* var. *longus*.

Distribution: Zhujiang [= Pearl River], Changjiang [=Yangtze River], Huanghe [= Yellow River], Heilongjiang [= Amur River], China (Chu *et al.*, 1999); Southern Hokkaido to Kyushu, Japan, as well as Taiwan, and northern Vietnam (Matsuura *et al.*, 2000); Common throughout Korea (Mori, 1952).

Remarks: Treated as valid as *Parasilurus asotus* in some recent publications.

Silurus biwaensis (Tomoda, 1961)

Parasilurus biwaensis Tomoda, 1961: 348, fig. 1. Type locality: Offshore of Onoé, Lake Biwa-ko, Japan. Holotype: MIKU 34407 [now at FAKU].

Distribution: Lake Biwa, Japan (Matsuura *et al.* 2000); reported from the Ueno Formation, Pliocene of ancient Lake Biwa, by Kobayakawa & Okuyama (1994a, b).

Silurus duanensis Hu, Lan & Zhang, 2004

Silurus duanensis Hu, Lan & Zhang, 2004: 586, 589, figs. 1–2. Type locality: underground rivers of Disu Town (23°34'N, 108°01'E), Du'an County, Guangxi [China]. Holotype: ASIZB 73176.

Distribution: Hongshui River basin, Pearl River drainage, China (Hu *et al.*, 2004).

Silurus glanis Linnaeus, 1758

Silurus glanis Linnaeus, 1758: 304. Type locality: Oriente, minus frequens in Europae lacibus. Syntypes: BMNH 1853.11.12.168 (1, skin), NRM 59 (1). Placed on Official List as type of *Silurus* (ICZN Direction 57).

Silurus silurus Wulff, 1765: 33. Type locality: Germany. No types known.

Silurus glanis aralensis Kessler, 1872: 48. Type locality: Amu-Darya, Syr-Darya, and Zeravshan rivers, cent. Asia. Syntype (3): ZISP 2071 (1). Originally as *Silurus glanis* var. *aralensis*.

? *Silurus chantrei* Sauvage, 1882: 163. Type locality: Fleuve Koura à Tiflis. Syntypes: MNHN a-3932 (2); described in more detail and illustrated in Sauvage (1884b: 19, pl. 1, fig. 1).

Distribution: Danube basin, all Baltic basin drainages, Elbe River, Lake Constance, Murtensee, and Rivers draining the Caspian, Azov and Aral Seas (Lelek, 1987); Lake Urmia, Tedzhzen River and possibly Tigris River basin (Coad, 1995), widely introduced elsewhere in Europe, including Great Britain, France, Italy, the Iberian Peninsula, and rivers flowing into the Arctic Ocean.

Silurus grahami Regan, 1907

Silurus grahami Regan, 1907a: 64. Type locality: Chien Kiung Lake, 30 mi. southeast of Yunnan Fu, China. Holotype: BMNH 1907.5.4.46.

Distribution: Fuxian Lake, Yunnan, China (Chu *et al.*, 1999)

***Silurus lanzhouensis* Chen, 1977**

Silurus lanzhouensis Chen, 1977: 210, pl. 2 (fig. 8). Type locality: [Yellow River system, Lanchow and Inner Mongolia, China]. Syntypes: Mus. Inst. Hydrobiol. Acad. Sinica, Hupei [LAN] 001-003 (3), [NEI] 004-005 (2), [NEI] 769739-42 (4).

Distribution: Changjiang [=Yangtze River], Zhujiang [= Pearl River], Minjiang (Chu *et al.*, 1999).

***Silurus lithophilus* (Tomoda, 1961)**

Parasilurus lithophilus Tomoda, 1961: 350, fig. 2. Type locality: Near Onoé, Lake Biwa-ko, Japan. Holotype: MIKU 34411 [now at FAKU].

Distribution: Lake Biwa and Lake Yogo, Japan (Masuda *et al.*, 1984).

***Silurus mento* Regan, 1904**

Silurus mento Regan, 1904a: 192. Type locality: Sea of Tien [Tien Chih], Yunnan Fu, China, elev. 6000 ft. Syntypes: BMNH 1904.1.26.40–41 (2).

Distribution: Lakes of Dianchi and Yilong, Yunnan, China (Chu *et al.*, 1999)

***Silurus meridionalis* Chen, 1977**

Silurus soldatovi meridionalis Chen, 1977: 209, pl. 2 (fig. 7). Type locality: [Yangtze R. system, China]. Syntypes: ? Mus. Inst. Hydrobiol. Acad. Sinica, Hupei 101–102 (2), 0719 (1), 66.5.194–195 (2), 731419 (1), 731494 (1), 746003 (1), 746008–09 (2).

Distribution: Yangtze River basin, China.

***Silurus microdorsalis* (Mori, 1936)**

Parasilurus microdorsalis Mori, 1936: 671, pl. 24 (fig. 1). Type locality: River Rakuto at Ei-yo, South Chosen [Korea]. Holotype: At Preparatory Department of Keijo Imperial University.

Distribution: Yalujiang [= Amnok River], China (Chu *et al.*, 1999); Upper reaches of the Yalu, Han, Kim, Naktong and Anpyeng Rivers, Korea (Mori, 1952).

***Silurus soldatovi* Nikolsky & Soin, 1948**

Silurus soldatovi Nikolsky & Soin, 1948: 1359, fig. 1. Type locality: [Amur River, Lake Kabar at Elabuga, Khabarovskiy Krai, Russia]. Holotype: ZMMU P-6505.

Distribution: Heilongjiang [=Amur River] China and Russia; Liaohe, China (Chu *et al.*, 1999)

***Silurus triostegus* Heckel, 1843**

Silurus triostegus Heckel, 1843: 1090, pl. 13 (fig. 1). Type locality: dem Tigris bei Mossul. Syntypes (4): at NMW, SMF 2623 (1).

Distribution: Tigris River basin (Coad, 1995); Euphrates River, Turkey (Ünlü & Bozkurt, 1996).

***WALLAGO* Bleeker, 1851**

Wallago Bleeker, 1851b: 265. Type species: *Silurus Müllerii* Bleeker, 1846. Type by subsequent designation by Bleeker (1862: 17, 79). Gender: Masculine.

Silurodon Kner, 1866: 546. Type species: *Silurodon hexanema* Kner, 1866. Type by monotypy. Gender: Masculine. Also described as new in Kner (1867: 305).

Wallagonia Myers, 1938: 98. Type species: *Wallago leerii* Bleeker, 1851. Type by original designation. Gender: Feminine.

Remarks: See Myers (1938: 98, 1948: 19) and Kottelat (2000b: 87) concerning the history of problems surrounding designation of type species of *Wallago*.

Revision, with key to species: Roberts (1982b).

***Wallago attu* (Bloch & Schneider, 1801)**

Silurus attu Bloch & Schneider, 1801: 378, pl. 75. Type locality: in lacubus Malabariae. Holotype: ZMB 8783 (dry, lost). As *Silurus athu* in text and index, *Silurus attu* on plate; first reviser apparently Bleeker (1862–63: 79) in which *Wallago attu* is treated as valid over *Wallago attu* [sic.]).

Silurus boalis Hamilton, 1822: 154, 375, pl. 29 (fig. 49). Type locality: Rivers not only of the Gangetic provinces,

but all over India, and is occasionally found in ponds. No types known.

Silurus (Callichrus) macrostomus Swainson, 1839: 306. Type locality: Rivers not only of the Gangetic provinces, but all over India, and is occasionally found in ponds. No types known. Name made available by reference to "Ham. 154, pl. 29, fig. 49" [= Hamilton, 1822: 154, 375, pl. 29 (fig. 49)]. Unneeded new name for *Silurus boalis* Hamilton, 1822.

Silurus Wallagoo Valenciennes, in Cuvier & Valenciennes, 1840a: 354. Type locality: Bengale, la côte de Coromandel, pays des Birmans. Syntypes: based on specimens at MNHN and literature accounts in Bloch & Schneider (1801), Russell (1803), and Hamilton (1822).

Silurus Müllerii Bleeker, 1846b: 289. Type locality: Batavia. Type(s): Whereabouts unknown.

Wallago Russellii Bleeker, 1854c: 108. Type locality: Calcutta, in fluminae Hooghly ... Batavia, in fluviis. Syntypes (4, 285–485 mm TL): Whereabouts unknown, and numerous literature citations.

Wallago attu valeya Deraniyagala, 1953: 45. Type locality: Yakvala, Ceylon. Holotype: NMSL FF187; illustrated in Deraniyagala (1952: pl. 13, as *Wallagonia attu*).

Distribution: India, Malay Peninsula, Sumatra, Java, and Mekong and Chao Phraya River basins (Kottelat, 2001b).

Remarks: Redescribed in Hora (1939b) as *Wallagonia attu*.

***Wallago leerii* Bleeker, 1851**

Wallago Leerii Bleeker, 1851e: 427. Type locality: Sambas et Palembang, in fluviis. Syntypes (2, 225, 230 mm TL): RMNH 6833 (2 of 4).

Wallago nebulosus Vaillant, 1902: 46. Type locality: Tepoe, bords du Mahakam, Bornéo central. Holotype: RMNH 7812; holotype illustrated by Roberts (1982b: fig. 3a).

Wallagonia tweediei Hora & Misra, in Hora & Gupta, 1941: 18, figs. 2–3. Type locality: Kuala Tahan, Pahang, Malaysia. Holotype: ZRC 350 (plaster cast with head and fins built in); ZSI F13365/1 (right anterior gill arch of holotype).

Distribution: Malay Peninsula, Sumatra and Borneo (Kottelat, 2001b; Ng, 2004a).

***Wallago maculatus* Inger & Chin, 1959**

Wallago maculatus Inger & Chin, 1959: 279. Type locality: Kinabatangan River at Deramakot, Kinabatangan District, North Borneo. Holotype: FMNH 68038.

Distribution: Kinabatangan River, Borneo; known only from type series (Roberts (1982b).

***Wallago micropogon* Ng, 2004**

Wallago micropogon Ng, 2004a: 93, fig. 1. Type locality: Cambodia: Stung Treng morning market (13°30.0'N, 105°58.0'E). Holotype: UMMZ 232320.

Distribution: Mekong River basin of Cambodia, Laos and Vietnam, and middle Chao Phraya River basin (Ng, 2004a).

Species inquirenda, Wallago

Silurodon hexanema Kner, 1866: 546. Type locality: Von Schanghai. Holotype (6 1/3"): at NMW. Described in more detail and illustrated in Kner (1867: 305, pl. 12, fig. 2 [as *Wallago attu* ?]).

Species inquirendae, Siluridae

Silurus sinensis La Cepède, 1803: 58, 82, pl. 2 (fig. 1). Type locality: China. No types known: based on an unpublished drawing.

Silurus sinensis M'Clelland, 1844a: 402. Type locality: China. No types known. Preoccupied by *Silurus sinensis* La Cepède, 1803.

Silurichthys basilewskii Bleeker, 1858b: 256. Type locality: China borealis. Based on specimens identified as *Silurus asotus* in Basilewsky (1855). Originally *Silurichthys* ? *basilewskii*.

Belodontichthys javanensis Hardenberg, 1938: 311. Type locality: Fish market of Batavia. Holotype (215 mm): Whereabouts unknown.

† *Silurus altus* Sytchevskaya, 1989. [No other information; in Gayet & Meunier, 2003].

Distribution: middle Miocene to lower Pliocene, Russia; Lower/Middle Miocene, China; Miocene or Pliocene of Tueirusia Formation, Russia (Gayet & Meunier, 2003).

† *Heterobranchus austriacus* Thenius, 1952. [No other information; in Gayet & Meunier, 2003].

Distribution: Bruun-Vösendorf, Austria, Pannonian (Gayet & Meunier, 2003).

Remarks: Included in Siluridae and probably *Silurus* by Gayet & Meunier (2003), following Gaudant (1994).

† *Silurus glanis atavus* Bogačew, 1924. [No additional information; from Weiler, 1956: 187.]

SISORIDAE Bleeker, 1858

Sisorichthyoidei Bleeker, 1858b: 48, 50. Type genus: *Sisor* Hamilton, 1822.

Glyptosterni Gill, 1861c: 53. Type genus: *Glyptosternon* M'Clelland, 1842.

Erethistides Bleeker, 1862 (in Bleeker, 1862–63): 13. Type genus: *Erethistes* Müller & Troschel, 1849.

Bagarina Günther, 1864: 3, 9, 183. Type genus: *Bagarius* Bleeker, 1854.

Exostomatina Günther, 1864: 264. Type genus *Exostoma* Blyth, 1860.

Continae de Pinna, 1996: 64. Type genus: *Conta* Hora, 1950.

Glyptothoracini de Pinna, 1996: 64. Type genus: *Glyptothorax* Blyth, 1860.

Laguviaini de Pinna, 1996: 65. Type genus: *Laguvia* Hora, 1921.

Pseudecheneidina de Pinna, 1996: 64. Type genus: *Pseudecheneis* Blyth, 1860.

Revision: Hora & Silas (1952a, glyptosternoids).

Review: Chu (1986, western China); Chu & Kuang (1990, Yunnan, China); Chu & Mo (1999, China); Menon (1999, India); Mirza & Hameed (1974, Pakistan).

Phylogeny: de Pinna (1996); Diogo *et al.* (2002, 2003b); Hora & Silas (1952b, glyptosternoids), Chu (1979, glyptosternoids), He (1996b, glyptosternoids), Peng *et al.* (2004, glyptosternoids), Guo *et al.* (2005, glyptosternoids).

Identification guide: Jayaram (1979, South Asia).

Historical biogeography: Hora & Silas (1952b), He (1995, glyptosternoids).

Remarks: In 1996, de Pinna removed several taxa from the Sisoridae and placed them into the family Erethistidae. Those taxa, placed herein within the genera *Conta*, *Erethistes*, *Erethistoides*, *Hara* and *Pseudolaguvia*, together with the subsequently named genera *Ayarnangra* and *Caelatoglanis*, appear instead to be part of a natural subgroup within the Sisoridae that includes the genus *Glyptothorax* and possibly others, but not the glyptosternoid genera (Ferraris & Britz, 2005; pers. obs.; and pers. commun., H. H. Ng, 2005). Therefore, the taxa placed in the Erethistidae by de Pinna are returned to the Sisoridae.

22 genera, 167 species; 1 named fossil species.

Incertae sedis

Laguvia manipurensis Arunkumar, 2000

Laguvia manipurensis Arunkumar, 2000: 194, fig. 1. Type locality: Lairao Maru stream near Moreh, 110 km from Imphal City, Manipur, India. Holotype: MUMF 3001/1A.

Distribution: Yu River basin, Chindwin River basin, Manipur, India (Arunkumar, 2000).

Remarks: Although described as a species of *Laguvia*, neither the description nor the illustration are sufficient to assign this species to genus or even family.

AYARNANGRA Roberts, 2001

Ayarnangra Roberts, 2001: 83. Type species: *Ayarnangra estuaricus* Roberts, 2001. Type by original designation. Gender: Masculine.

Ayarnangra estuaricus Roberts, 2001

Ayarnangra estuaricus Roberts, 2001: 84, figs. 1–3. Type locality: Pathein Chaung (=Ngawan Chaung) near Pathein, lower Ayeyarwaddy basin, Myanmar. Holotype: KUMF 3190.

Distribution: Coastal portions of Irrawaddy and Bago rivers, Myanmar.

BAGARIUS Bleeker, 1854

Bagarius Bleeker, 1854c: 121. Type species: *Pimelodus bagarius* Hamilton, 1822. Type by monotypy. Gender: Masculine.

Revision: Roberts (1983).

Bagarius bagarius (Hamilton, 1822)

Pimelodus bagarius Hamilton, 1822: 186, 378, pl. 7 (fig. 62). Type locality: Ganges River. No types known.

Pimelodus platespogon Valenciennes, 1840, in Jacquemont, 1835–44: pl. 18 (fig. 3). Type locality: l'Inde. Holotype: MNHN 0000-2904 (1 of 2). Name made available by plate caption, therefore the illustrated specimen is the holotype if distinguishable.

Bagarius Buchanani Bleeker, 1854c: 121. Type locality: Calcutta, in flumine Hooghly; Surakata, Javae centrals, in flumine Pepeh. Syntype (3): possibly NMV 46015 (1). Considered to be an unneeded new name for *Pimelodus bagarius* Hamilton by Roberts (1983), to avoid Stricklandian tautonomy.

Distribution: Mekong, Chao Phraya and Ganges River basins (Kottelat, 2001b).

Remarks: Redescribed in Hora (1939a).

† ***Bagarius gigas*** Gunther, 1876

† *Bagarius gigas* Gunther, 1876: 436, pl. 16, fig. 1. Type locality: Highlands of Padang, Sumatra; Tertiary. Holotype BMNH 47513 (Imperfect pectoral arch, etc.).

Distribution: Highlands of Padang, Sumatra; reportedly from Eocene, but age questioned by Gayet & Meunier (2003).

Remarks: See Sanders (1934) for additional information.

Bagarius rutilus Ng & Kottelat, 2000

Bagarius rutilus Ng & Kottelat, 2000b: 10, fig. 3. Type locality: Market in Hanoi, Viet Nam. Holotype: ZRC 40440.

Distribution: Nam Xan and Nam Ma basins, Laos, and Red River basin, Viet Nam and China (Kottelat, 2001b).

Bagarius suchus Roberts, 1983

Bagarius suchus Roberts, 1983: 442, figs. 2c, 4. Type locality: Thailand, Kemrat. Holotype: ANSP 89521.

Distribution: Mekong and Chao Phraya River basins (Kottelat, 2001b).

Bagarius yarrelli (Sykes, 1839)

Bagrus Yarrelli Sykes, 1839: 163. Type locality: Mota Mola at Poona, Deccan, India. Type(s): Whereabouts unknown. Also described as new in Sykes (1840: 60); illustrated and described in more detail in Sykes (1841: 370, pl. 65, fig. 1).

Pimelodus Carnaticus Jerdon, 1849: 341. Type locality: Bowany River, southern India. No types known.

Bagarius lica Volz, 1903a: 557. Type locality: Moresi R. at Palembang, Sumatra. Holotype: at NMBE. Described in more detail in Volz (1903b: 391).

Bagarius Nieuwenhuisii Popta, 1904: 190. Type locality: le Bo, Bornéo central. Holotype: RMNH 7561; described in more detail, with illustration of holotype, in Popta (1906: 66, pl. 4, fig. 14).

Distribution: Widely distributed in southern and southeastern Asia (Kottelat, 2001b).

CAELATOGLANIS Ng & Kottelat, 2005

Caelatoglanis Ng & Kottelat, 2005: 14. Type species: *Caelatoglanis zonatus* Ng & Kottelat, 2005. Type by original designation. Gender: Masculine.

Caelatoglanis zonatus Ng & Kottelat, 2005

Caelatoglanis zonatus Ng & Kottelat, 2005: 14, fig. 2. Type locality: Myanmar, Kayin State, stream "Chon Son" between Kyondaw and Phadaw, about 20 km NW of Payathouzu, (at border with Thailand), 15°25'N, 98°15'E. Holotype: ZRC 49885.

Distribution: Ataran River basin, Myanmar (Ng & Kottelat, 2005).

CONTA Hora, 1950

Conta Hora, 1950: 194. Type species: *Pimelodus conta* Hamilton, 1822. Type by original designation. Gender: Fem-

inine.

***Conta conta* (Hamilton, 1822)**

Pimelodus conta Hamilton, 1822: 191, 378. Type locality: River Mahananda [Bengal]. No types known. Previously unpublished Hamilton illustration reproduced in Hora (1929: pl. 21, fig. 8), and subsequently reproduced in Hora (1950: 297, fig. 4a) and Ng (2005a: 28, fig. 6).

Hara elongata Day, 1872: 704. Type locality: Stream near Garrow hills. Holotype: ZSI 436. Illustration of holotype first published in Day (1877: pl. 92, fig. 5) and reproduced in Ng (2005a: 28, fig. 7).

Distribution: Bhareli and Mahananda Rivers, northeast Bengal, Garo Hills, Meghalaya; and Bangladesh (Jayaram, 1979); also Sarda River, Uttar Pradesh (Tilak, 1987).

***Conta pectinata* Ng, 2005**

Conta pectinata Ng, 2005a: 24, fig. 1. Type locality: India: Assam: Dibrugarh. Holotype: ZRC 49672.

Distribution: Middle Brahmaputra River basin, Assam, India (Ng, 2005a).

***ERETHISTES* Müller & Troschel, 1849**

Erethistes Müller & Troschel, 1849: 12. Type species: *Erethistes pusillus* Müller & Troschel, 1849. Type by monotypy. Gender: Masculine.

***Erethistes maesotensis* Kottelat, 1983**

Erethistes maesotensis Kottelat, 1983: 71, figs. 1–2. Type locality: Thailand: Tak province: Mae Nam Moei, 5 km W of Mae Sot (16°41'N, 98°31'E). Holotype: MHNG 2096.63.

Distribution: Mae Nam Moei, Salween River basin, Thailand (Kottelat, 1983).

***Erethistes pusillus* Müller & Troschel, 1849**

Erethistes pusillus Müller & Troschel, 1849: 12, pl. 1 (fig. 3). Type locality: Assam. Holotype: ZMB 3102; holotype illustration reproduced by Hora (1950: 187, pl. 1, fig. 5, as *Erethistes pussilus*).

Distribution: Ganges River basin, Assam, India.

***ERETHISTOIDES* Hora, 1950**

Erethistoides Hora, 1950: 190. Type species: *Erethistoides montana* Hora, 1950. Type by original designation. Gender: Masculine.

***Erethistoides ascita* Ng & Edds, 2005**

Erethistoides ascita Ng & Edds, 2005a: 240, fig. 1. Type locality: Nepal: Jhapa, Bhadrapur, Mechi River at Bhadrapur, 26°32'17.9"N, 88°6'6.1"E. Holotype: KU 35016.

Distribution: Rivers of lowland plains of southeastern Nepal (Ng & Edds, 2005a).

***Erethistoides cavatura* Ng & Edds, 2005**

Erethistoides cavatura Ng & Edds, 2005a: 243, fig. 5. Type locality: Nepal: Chitawan, Dhungre River at Sauraha. Holotype: OSUS 15572.

Distribution: Narayani River basin, Nepal (Ng & Edds, 2005a).

***Erethistoides montana* Hora, 1950**

Erethistoides montana Hora, 1950: 191, pl. 1 (figs. 10–12). Type locality: Streamlets round about Tangla, Darrang district, Assam [India]. Holotype: ZSI F314/2.

Distribution: Brahmaputra River basin, India (Ng, 2005f).

***Erethistoides pipri* Hora, 1950**

Erethistoides montana pipri Hora, 1950: 193, pl. 1 (figs. 7–9). Type locality: Pipri, Rihand River, Mirzapur district, U. P. [India]. Holotype: ZSI F315/2.

Distribution: Rihand River, Uttar Pradesh, India (Hora, 1950); known only from holotype (Jayaram, 1979).

***Erethistoides sicula* Ng, 2005**

Erethistoides sicula Ng, 2005f: 2, fig. 1. Type locality: India: West Bengal, Schutunga River (tributary of the Mansai River) as Ansole, 26°22'24"N 89°11'17"E. Holotype: UMMZ 243718.

Distribution: Mansai River drainage, Brahmaputra River basin, India (Ng, 2005f: 6).

***EUCHIOLGANIS* Regan, 1907**

Chimarrichthys Sauvage, 1874: 332. Type species: *Chimarrichthys davidi* Sauvage, 1874. Type by monotypy. Gender: Masculine.

Euchiloglanis Regan, 1907c: 158. Type species: *Chimarrichthys davidi* Sauvage, 1874. Type by being a replacement name. Gender: Masculine. Proposed as a replacement for *Chimarrichthys* Sauvage, 1874, then considered to be preoccupied by *Cheimarrichthys* Haast, 1874, in fishes.

Coraglanis Hora & Silas, 1952a: 12. Type species: *Euchiloglanis kishinouyei* Kimura, 1934. Type by original designation. Gender: Masculine.

Remarks: *Chimarrichthys* Sauvage, 1874, is not actually preoccupied by *Cheimarrichthys* Haast, 1874, but the name has apparently not been used as valid since Regan (1907c) proposed *Euchiloglanis* as a replacement. The prevailing usage requirements of the Code require that *Euchiloglanis* be treated as valid here. *Coraglanis* Hora & Silas is generally treated as valid as a monotypic genus for *Coraglanis kishinouyei* in Indian literature. He (1996b) indicated that *Euchiloglanis kishinouyei* shared derived characters uniquely with *Euchiloglanis davidi* and placed the two into *Euchiloglanis*.

***Euchiloglanis davidi* (Sauvage, 1874)**

Chimarrichthys Davidi Sauvage, 1874: 333. Type locality: Yao-Tchy, Thibet oriental. Syntypes: BMNH 1923.3.13.1 (1), MNHN 0000-6273 (3), MNHN 0000-6274 (3).

Distribution: Chinjiang, Sichuan (Chu *et al.*, 1999); eastern Tibet.

***Euchiloglanis kishinouyei* Kimura, 1934**

Euchiloglanis Kishinouyei Kimura, 1934: 178, pl. 6. Type locality: Kwan-hsien, Szechwan Province, China. Holotype: at Laboratory of Biological Department of the Shanghai Science Institute, Shanghai.

Distribution: Jinshajiang [upper Yangtze River basin] (Chu *et al.*, 1999).

***EXOSTOMA* Blyth, 1860**

Exostoma Blyth, 1860: 155. Type species: *Exostoma berdmorei* Blyth, 1860. Type by subsequent designation, by Bleeker (1862–1863: 13). Gender: Neuter.

Remarks: See Regan (1923) for comments on this genus.

***Exostoma berdmorei* Blyth, 1860**

Exostoma Berdmorei Blyth, 1860: 155. Type locality: Tenasserim [Burma]. Syntypes: ASB 597 (2), (now at ZSI). Possible syntype: ASB 600.

Distribution: Eastern Myanmar.

***Exostoma labiatum* (M'Clelland, 1842)**

Glyptosternon labiatus M'Clelland, 1842: 588. Type locality: Mishmee Hills Upper Assam [India]. Holotype: BMNH 1860.3.19.97.

Distribution: Yaluzangbujiang [= upper Brahmaputra River], and Irrawaddy River (Chu *et al.*, 1999).

Species inquirendae, Exostoma

Glyptosternum chaudhurii Hora, 1923b: 41, fig. 7. Type locality: Putao Plains, northern frontier of Burma. Holotype: ZSI F9741/1. Treated as a synonym of either *E. labiatum* or *E. vinciguerrae*.

Glyptosternum stuarti Hora, 1923b: 39, pl. 2 (figs. 4–6). Type locality: Nam-Yak River at Tanja, on the northern frontier of Burma. Holotype: ZSI F9742/1. Treated either as valid or as a synonym of *E. labiatum*.

Exostoma Vinciguerrae Regan, 1905a: 184. Type locality: Khakhyen Hills, upper Burma. Holotype: BMNH 1893.2.16.17. Treated either as valid or as a synonym of *E. labiatum*. See Steinitz (1961) for comments on taxonomy.

***GAGATA* Bleeker, 1858**

Gagata Bleeker, 1858b: 204, 206. Type species: *Pimelodus gagata* Hamilton, 1822. Type by absolute tautonymy. Gender: Feminine.

Callomystax Günther, 1864: 218. Type species: *Pimelodus gagata* Hamilton, 1822. Type by being a replacement name. Gender: Masculine. Unneeded replacement for *Gagata* Bleeker, 1858.

Revision: Hora & Law (1941); Roberts & Ferraris (1998).

***Gagata cenia* (Hamilton, 1822)**

Pimelodus cenia Hamilton, 1822: 174, 376, pl. 31 (fig. 57). Type locality: Northern parts of Bengal, where it frequents rivers. No types known.

Distribution: Ganges, Indus and Mahanadi river basins (Roberts & Ferraris, 1998).

***Gagata dolichonema* He, 1996**

Gagata dolichonema He, 1996a: 380, fig. 1. Type locality: Daojieba of Baoshan County, (24°41'N, 99°10'E), Yunnan Province, China. Holotype: IHASW 791.

Gagata gasawayuh Roberts & Ferraris, 1998: 325, figs. 6–7. Type locality: Myanmar, Tenasserim River mainstream upstream from Kita (or Htee-tah). Holotype: CAS 95544.

Distribution: Upper Salween River basin, China and Irrawaddy, Salween and Tenasserim River basins, Myanmar (Roberts & Ferraris, 1998).

Remarks: Synonymy based on recent examination of specimens from the vicinity of the type locality of *Gagata dolichonema*, and photograph of the holotype.

***Gagata gagata* (Hamilton, 1822)**

Pimelodus gagata Hamilton, 1822: 197, 379, pl. 39 (fig. 65). Type locality: Fresh water rivers and estuaries of Bengal. No types known.

Gagata typus Bleeker, 1863c: 90. Type locality: Fresh water rivers and estuaries of Bengal. No types known. Unneeded new name for *Pimelodus gagata* Hamilton, 1822.

Distribution: Ganges River basin, India and Bangladesh (Roberts & Ferraris, 1998).

***Gagata itchkeea* (Sykes, 1839)**

Phractocephalus itchkeea Sykes, 1839: 164. Type locality: Deccan, India. Type(s): at BMNH. Also described as new in Sykes (1840: 61) Illustrated and described in more detail in Sykes (1841: 373, pl. 67, fig. 1).

Arius pumilus Valenciennes, 1840, in Jacquemont, 1835–44: pl. 18 (fig. 1). Type locality: Inde. Syntypes: MNHN 0000-1208 (8). Name made available from caption on plate.

Distribution: Narmada, Krishna and Cauvery River basins, India (Roberts & Ferraris, 1998).

Remarks: Synonymy of *Arius pumilus* based on examination of types; illustrated specimen of *A. pumilus* is arguably a holotype, but it could not be readily distinguished from remaining specimens.

***Gagata melanoptera* Roberts & Ferraris, 1998**

Gagata melanopterus Roberts & Ferraris, 1998: 330, fig. 10. Type locality: Myanmar: Yangon Division, Hlaing River, 16°53'41"N, 96°05'28"E. Holotype: USNM 348852.

Distribution: Irrawaddy, Rangoon, Sittang and lower Salween River basins, Myanmar (Roberts & Ferraris, 1998).

***Gagata pakistanica* Mirza, Parveen & Javed, 1999**

Gagata pakistanica Mirza, Parveen & Javed, 1999: 1, fig. 1. Type locality: River Indus near Ghazi, Pakistan. Holotype: Disposition not stated in publication, but presumably deposited at GCM.

Distribution: Indus River, Pakistan (Mirza, 2003).

***Gagata sexualis* Tilak, 1970**

Gagata sexualis Tilak, 1970b: 207, fig. 1. Type locality: North Koel River at Daltonganj (Chotanagpur) [South Bihar, India]. Holotype: ZSI F5592/2.

Gagata youssoufi Ataur Rahman, 1976: 5, fig. 1. Type locality: river Meghna near Chandpur, Bangladesh. Holotype: at Museum of Freshwater Fisheries Research Station, Chandpur, Bangladesh.

Distribution: Ganges and Brahmaputra River basins (Roberts & Ferraris, 1998).

Remarks: Synonymy based on Heok Hee Ng (pers. comm., 2003).

***GLARIDOGLANIS* Norman, 1925**

Glaridoglanis Norman, 1925b: 574. Type species: *Exostoma andersonii* Day, 1870. Type by monotypy. Gender: Masculine.

***Glaridoglanis andersonii* (Day, 1870)**

Exostoma andersonii Day, 1870b: 524. Type locality: Hotham; and Ponsee, China. Possible syntypes: ZSI F9173-74/1 [= A.595] (2), ZSI A.596 (2, lost), AMS B.8081 (1).

Distribution: Yaluzangbujiang [= upper Brahmaputra River], and Irrawaddy River, China (Chu *et al.*, 1999).

GLYPTOSTERNON M'Clelland, 1842

Glyptosternon M'Clelland, 1842: 584. Type species: *Glyptosternon reticulatus* M'Clelland, 1842. Type by subsequent designation by Bleeker (1862–63: 12). Gender: Neuter

Glyptosternum Agassiz, 1846: 164. Type species: *Glyptosternon reticulatus* M'Clelland, 1842. Unjustified emendation of *Glyptosternon* M'Clelland, 1842. Gender: Neuter.

Parexostoma Regan, 1905a: 182. Type species: *Exostoma stoliczkae* Day, 1877. Type by subsequent designation, apparently by Jordan (1920: 515). Gender: Neuter.

Remarks: Blyth (1860: 154) restricted *Glyptosternon* to a single species, but did not, by that action, designate that species, *G. reticulatus*, as type. See Regan (1923) for comments on this genus.

Glyptosternon akhtari Silas, 1952

Glyptosternum akhtari Silas, in Hora & Silas, 1952a: 11, pl. 1 (figs. 4–6). Type locality: Bamian River, Oxus watershed, Afghanistan. Holotype: ZSI F643/2.

Distribution: Bamian River, Oxus watershed, Afghanistan (Hora & Silas, 1952a).

Glyptosternon maculatum (Regan, 1905)

Parexostoma maculatum Regan, 1905a: 183. Type locality: Lhasa, Tibet. Syntypes: BMNH 1904.12.28.87–88 (2).

Distribution: Yaluzangbujiang [= upper Brahmaputra River] (Chu *et al.*, 1999).

Glyptosternon malaisei Rendahl & Vestergren, 1941

Glyptosternon malaisei Rendahl & Vestergren, 1941: 213. Type locality: Kambaiti, Kachin State, Irrawaddy River drainage, Burma, 7000 ft. elev. Holotype: NRM 10721.

Distribution: Irrawaddy River drainage, northern Myanmar (Rendahl & Vestergren, 1941).

Glyptosternon reticulatum M'Clelland, 1842

Glyptosternon reticulatus M'Clelland, 1842: 584. Type locality: Sir-i-Chusma, at the source of the Cabul River [Afghanistan]. No types preserved. Name spelled *Glyptosternon retuculatus* and *Glyptosternon reticulatus* in original description; Kullander *et al.* (1999: 146) apparently serve as first revisers in selecting *reticulatus*.

Exostoma stoliczkae Day, 1877b: 782. Type locality: Upper waters of the Indus River. Possible syntypes: AMS I.122 (1), ZSI F497 (1), ZSI 1196–98 (3, lost).

Exostoma gracile Gratzianov, 1907: 58. Type locality: Naryn R. drainage, upper Syr-Darya R. basin, Namangan-skaya Obl., Uzbekistan. Holotype: ZMMU P-785.

Exostoma labrax Gratzianov, 1907: 59. Type locality: Garm dist., mountain spring Kartveng at kishlak Shulmak, Garmskaya Obl., Tadzhikistan. Syntypes: ZMMU P-2034 (2).

Distribution: Upper Jhelum basin and Gilgit River, Indus River drainage, Pakistan and Kashmir (Kullander, 1999).

Remarks: See Steinitz (1961) for comments on taxonomy. Redescribed in Hora (1932).

GLYPTOTHORAX Blyth, 1860

Glyptothorax Blyth, 1860: 154. Type species: *Glyptosternon striatus* M'Clelland, 1842. Type by subsequent designation by Bleeker (1862–1863: 13). Gender: Masculine.

Aglyptosternon Bleeker, 1862 (in Bleeker, 1862–63): 12. Type species: *Silurus coues* Linnaeus, 1766. Type by original designation. Gender: Neuter.

Euclyptosternum Günther, 1864: 183. Type species: *Silurus coues* Linnaeus, 1766. Unjustified emendation of *Aclyp-tosternon* [sic, *Aglyptosternon* Bleeker, 1862]. Gender: Neuter.

Pteroglanis Fowler, 1934a: 92. Type species: *Pteroglanis horai* Fowler, 1934. Type by original designation. Preoccupied by *Pteroglanis* Eigenmann & Pearson, 1924, in fishes, replaced by *Pteropsoglanis* Fowler, 1934. Gender: Masculine.

Pteropsoglanis Fowler, 1934b: 351. Type species: *Pteroglanis horai* Fowler, 1934. Type by being a replacement name. Replacement for *Pteroglanis* Fowler, 1934; preoccupied by *Pteroglanis* Eigenmann & Pearson, 1924. Gender: Masculine.

Sundagagata Boeseman, 1966: 243. Type species: *Sundagagata robusta* Boeseman, 1966. Type by original designation. Gender: Feminine.

Paraglyptothorax Li, 1986: 524. Type species: *Glyptosternum pallozonum* Lin, 1934. Type by original designation.

Gender: Masculine. Originally proposed as a subgenus of *Glyptothorax*.

Superglyptothorax Li, 1986: 524. Type species: *Glyptothorax coheni* Ganguly, Datta & Sen, 1972. Type by original designation. Gender: Masculine. Originally proposed as a subgenus of *Glyptothorax*.

Key: Mo & Chu (1986, Chinese species).

Review: Menon, M. A. S. (1955, India and Burma, with key), Sufi (1963, Pakistan).

Remarks: Extension of the eastern limit of the distribution of this genus into the Black Sea of Turkey noted by Coad & Delmastro (1985).

***Glyptothorax alaknandi* Tilak, 1969**

Glyptothorax brevipinnis alaknandi Tilak, 1969: 42, figs. 8–11. Type locality: Alaknanda River, near Srinagar, dist. Pauri Garhwal, Uttar Pradesh, India. Holotype: ZSI F6154/2.

Distribution: Alaknanda River, Uttar Pradesh, India (Jayaram, 1999).

***Glyptothorax anamalaiensis* Silas, 1951**

Glyptothorax anamalaiensis Silas, 1951b: 370. Type locality: Streams at base of Anamalai Hills, South India. Holotype: ZSI F629/2; illustrated in Silas (1951a: pl. 1, figs 1–3), as *Glyptothorax prox. madraspatanus* Day.

Distribution: Anamalai Hills, Kerala, India (Jayaram, 1999).

***Glyptothorax annandalei* Hora, 1923**

Glyptothorax annandalei Hora, 1923b: 14, pl. 1 (fig. 3). Type locality: Nierolay stream, Bhavani River at the base of Nilgiri Hills [India]. Holotype: ZSI F10135/1.

Distribution: Yaluzanbujiang [= Brahmaputra River], China (Chu & Mo, 1999).

***Glyptothorax armeniacus* (Berg, 1918)**

Glyptosternum armeniacum Berg, 1918: 146. Type locality: River Mukhlassi-darasi, headwaters of Euphrates. Syntypes (5): possibly ZSI F11319/1 (1), ZIN 20806 (4); ZIN syntype illustrated in Berg (1931: pl. 1, figs. 3–5; fig. 6).

Distribution: Tigris River basin, Iran (Coad, 1995); Euphrates River basin, Turkey (Berg, 1918).

***Glyptothorax botius* (Hamilton, 1822)**

Pimelodus botius Hamilton, 1822: 192, 378. Type locality: Northern rivers of Bengal [now India: west Bengal: Hooghly River at Kalna, 23°13'30.0"N, 88°22'39.0"E by neotype designation]. Neotype: ZRC 50223; designated by, and illustrated in, Ng (2005e: 3, fig. 1).

Distribution: Hooghly River, India (Ng, 2005e: 8).

Remarks: Redescribed in Ng (2005e).

***Glyptothorax brevipinnis* Hora, 1923**

Glyptothorax brevipinnis Hora, 1923b: 16, pl. 1 (fig. 4). Type locality: Unknown locality [probably India]. Syntypes: ZSI F10134/1 (4).

Distribution: Not known (Jayaram, 1999).

***Glyptothorax buchanani* Smith, 1945**

Glyptothorax buchanani Smith, 1945: 402, fig. 89. Type locality: The Metum, a small swift affluent of the Mechem, tributary of the Meping, in northern Thailand. Holotype: USNM 117754.

Distribution: Me Ping, Chao Phraya Basin, Thailand (Smith, 1945).

***Glyptothorax burmanicus* Prashad & Mukerji, 1929**

Glyptothorax burmanicus Prashad & Mukerji, 1929: 184, pl. 7 (fig. 3); fig. 5. Type locality: Sankha, a large hill-stream, midway between Kamaing and Mogaung, Myitkyina dist. [Burma]. Holotype: ZSI F10877/1.

Distribution: Irrawaddy and Salween Rivers (Chu & Mo, 1999).

***Glyptothorax callopterus* Smith, 1945**

Glyptothorax callopterus Smith, 1945: 400, fig. 87. Type locality: Waterfall stream on Kao Chong, near Trang, in Peninsular Thailand. Holotype: USNM 109820.

Distribution: Peninsular Thailand.

***Glyptothorax cavia* (Hamilton, 1822)**

Pimelodus cavia Hamilton, 1822: 188, 378. Type locality: Northern rivers of Bengal. No types known.

Euglyptosternum lineatum Day, 1877 (in Day, 1875–78): 500, pl. 116 (fig. 7). Type locality: Jumna, near Sudya in

upper Assam. Possible syntypes: AMS B.7509 (1), ZSI F1312 (1).

Distribution: Ganges River basin.

Glyptothorax coheni Ganguly, Datta & Sen, 1972

Glyptothorax coheni Ganguly, Datta & Sen, 1972: 342, figs. 3–4. Type locality: Subarnarekha River, Chotanagpur Plateau, Bihar, India. Holotype: USNM 205612.

Distribution: Subarnarekha River, Bihar, India (Jayaram, 1999).

Glyptothorax conirostris (Seindachner, 1867)

Glyptosternum conirostre Steindachner, 1867e: 532, pl. 6 (fig. 2). Type locality: Simla. Holotype: at NMW.

Glyptosternum modestum Day, 1872: 714. Type locality: upper portion of the Jumna. Possible syntypes: AMS B.7562 (1), AMS B.7564 (1), ZMB 2765 (1).

Distribution: Jamuna River, Mahananda River basin and Indus River basin (Jayaram, 1999).

Glyptothorax couis (Linnaeus, 1766)

Silurus couis Linnaeus, 1766: 504. Type locality: Syria. Possible syntype: BMNH 1955.6.25.2 (1).

Distribution: Tigris River basin, Iran (Coad, 1995).

Glyptothorax dakpathari Tilak & Husain, 1976

Glyptothorax dakpathari Tilak & Husain, 1976: 229, figs. 1–8. Type locality: Yamuna R., below Barrage, Dakpathar, District Dehra Dun, Uttar Pradesh, India. Holotype: ZSI NRS V-988.

Distribution: Yamuna River, Uttar Pradesh, India (Jayaram, 1999).

Remarks: Treated as a synonym of *Glyptothorax gracilis* by Menon (1999).

Glyptothorax davissinghi Manimekalan & Das, 1998

Glyptothorax davissinghi Manimekalan & Das, 1998: 87, unnumbered figure. Type locality: Karim Puzha, Maancheri, Nilambur Reserve Forest, Kerala, India. Holotype: ZSIC 6008.

Distribution: Kerala, India (Manimekalan & Das, 1998).

Glyptothorax deqinensis Mo & Chu, 1986

Glyptothorax deqinensis Mo & Chu, 1986: 345, 350, fig. 6. Type locality: Deqin county (28°30'N, 99°00'E) Yunnan [China]. Holotype: KIZ 748621.

Distribution: upper Lancangjiang [= Mekong River], China (Chu *et al.*, 1999).

Glyptothorax dorsalis Vinciguerra, 1890

Glyptothorax dorsalis Vinciguerra, 1890: 246, pl. 7 (fig. 4). Type locality: Meetan, Birmania [Myanmar]. Holotype: MSNG 14417.

Distribution: Upper Irrawaddy River basin (pers. obs.); Nujiang [= Salween River], China (Chu *et al.*, 1999).

Glyptothorax exodon Ng & Rachmatika, 2005

Glyptothorax exodon Ng & Rachmatika, 2005: 251, fig. 1. Type locality: Borneo: Kalimantan Barat, Sungai Tekelan. Holotype: MZB 9940.

Distribution: Kapuas River basin, western Borneo (Ng & Rachmatika, 2005).

Glyptothorax fokiensis (Rendahl, 1925)

Glyptosternum fokiensis Rendahl, 1925: 307. Type locality: Lan-Hao, Lien-Cheng-Hsien, Fokien, s. China. Syntypes: NRM 10018 (2).

Glyptosternon punctatum Nichols, 1941: 1, figs. 1–2. Type locality: Kiating (Loshan), Szechwan Prov., China, elev. 1100 ft. Holotype: AMNH 15218.

Distribution: Changjiang [= Yangtze River], rivers east of Yuanjiang (Chu *et al.*, 1999; as *Glyptothorax fukiensis fukiensis*).

Remarks: Redescribed as *Glyptothorax fukiensis* in Xie *et al.* (2001).

Glyptothorax fuscus Fowler, 1934

Glyptothorax fuscus Fowler, 1934a: 89, figs. 31–33. Type locality: Chantaboon, Southeast Siam. Holotype (59 mm): ANSP 59358.

Distribution: Mekong and Chao Phraya River basins (Kottelat, 2001b).

Glyptothorax garhwali Tilak, 1969

Glyptothorax garhwali Tilak, 1969: 37, figs. 1–4. Type locality: Alaknanda River, near Srinagar, dist. Pauri Garhwal, Uttar Pradesh, India. Holotype: ZSI F6152/2.

Distribution: Alaknanda River, Uttar Pradesh, India (Jayaram, 1999).

Glyptothorax gracilis (Günther, 1864)

Glyptosternum gracile Günther, 1864: 186. Type locality: Nepal. Holotype: BMNH 1845.1.9.846.

Distribution: Sikkim, India and Nepal (Jayaram, 1999).

Glyptothorax hainanensis (Nichols & Pope, 1927)

Glyptosternon hainanensis Nichols & Pope, 1927: 333, fig. 7. Type locality: Nodoa, Hainan [China]. Holotype: AMNH 8362.

Distribution: Hainan Island, China (Chu *et al.*, 1999, as *Glyptothorax fukiensis hainanensis*).

Glyptothorax honghensis Li, 1984

Glyptothorax fukiensis honghensis Li, 1984a: 66, fig. 1. Type locality: Hekou, Weishan, Nanjian, Hong River basin, Yunnan Province, China. Syntypes: KIZ 6440430 (1), KIZ 6440474 (1), KIZ 6507022 (1), KIZ 6507133 (1), KIZ 6507134 (1), KIZ 6507137 (1). Name spelled *Glyptothorax fukiensis honghensis* on p. 66 and *Glyptothorax fukiensis honghenensis* on p. 69; *G. honghensis* treated as the valid spelling here, based on presumed etymology of name.

Distribution: Northeastern Laos, and Red River basin of Vietnam and China (Kottelat, 2001b).

Glyptothorax horai (Fowler, 1934)

Pteroglanis horai Fowler, 1934a: 92, figs. 37–40. Type locality: Sop Lao, in Maun Luang, Southeast Shan States [Burma]. Holotype: ANSP 59462.

Distribution: Mekong River basin, Myanmar.

Glyptothorax housei Herre, 1942

Glyptothorax housei Herre, 1942: 117, fig. 1. Type locality: Rapids in a mountain stream in the Anamallai Hills, Puthototam Estate, four miles east of Valparai, Pollachi district, South India. Holotype: SU 36531.

Distribution: Kerala, India (Jayaram, 1999).

Glyptothorax indicus Talwar, 1991

Glyptothorax horai Shaw & Shebbeare, 1936: 188, unnumbered pl. Type locality: Streams of Terai, northern Bengal. Holotype: ZSI F11376/1. Preoccupied in *Glyptothorax* by *Pteroglanis horai* Fowler, 1934; replaced by *Glyptothorax indicus* Talwar, 1991.

Glyptothorax indicus Talwar, in Talwar & Jhingran, 1991: 654, fig. 210. Type locality: Streams of Terai, n. Bengal. Holotype: ZSI F11376/1. Replacement name for *Glyptothorax horai* Shaw & Shebbeare, 1936, preoccupied in *Glyptothorax* by *Pteroglanis horai* Fowler, 1934.

Distribution: Kosi River, Rihand River and rivers of the Vindhya Mountain range, India, as well as the Punjab and Nepal (Jayaram, 1999).

Glyptothorax interspinulus (Mai, 1978)

Glyptosternon interspinulum Mai, 1978: 271, fig. Type locality: Creeks in n. Vietnam. Syntypes: probably at DVZUT.

? *Glyptothorax merus* Li, 1984b: 79, fig. 2. Type locality: Jingdong Co., Yunnan, China. Holotype: KIZ 737159.

Distribution: Nam Ma Basin, Laos, and Red River basin of China and Vietnam (Kottelat, 2001b).

Glyptothorax jalalensis Balon & Hensel, 1970

Glyptothorax jalalensis Balon & Hensel, 1970: 160, fig. 1. Type locality: Kabul River tributary, near Jalal-Abad, Afghanistan. Holotype: SNMB RY 2176.

Distribution: Kabul River basin, Afghanistan (Balon & Hensel, 1970).

Glyptothorax kashmirensis Hora, 1923

Glyptothorax kashmirensis Hora, 1923b: 22, fig. 2. Type locality: Kashmir Valley. Syntypes: ZSI F10270/1 (2).

Distribution: Jhelum River, Kashmir (Kullander, 1999); NWFP, Azad Kashmir (Mirza, 2003).

Remarks: See Rashida *et al.* (1996) for comments on the taxonomic position of this species.

Glyptothorax kurdistanicus (Berg, 1931)

Glyptosternum kurdistanicum Berg, 1931: 1267, fig. 1; pl. 1 (fig. 2). Type locality: Kurdistan, at the village of Germau (or Germav), at the height of 1500 m, ... Germau is situated in latitude 36° southeast of Serdesht on the Western slope of the Sur-kei Range, in the basin of the river Bané, tributary to the Little Zab, which is a tributary to the Tigris R. Holotype: ZIN 20780.

Distribution: Tigris River basin, Iran (Coad, 1995).

***Glyptothorax lampris* Fowler, 1934**

Glyptothorax lampris Fowler, 1934a: 91, figs. 34–36. Type locality: Chieng Mai, North Siam. Holotype: ANSP 59357.

Distribution: Mekong and Chao Phraya River basins, and rivers of southeastern Thailand (Kottelat, 2001b); in medium-sized upland rivers (Rainboth, 1996).

***Glyptothorax laosensis* Fowler, 1934**

Glyptothorax laosensis Fowler, 1934a: 88, figs. 28–30. Type locality: Bua Yai, East Siam. Holotype: ANSP 59412.

Distribution: Mekong and Chao Phraya River basins (Kottelat, 2001b); in small to medium-sized streams (Rainboth, 1996).

***Glyptothorax lonah* (Sykes, 1839)**

Bagrus lonah Sykes, 1839: 164. Type locality: Deccan, India. Holotype: BMNH 1860.3.19.756 or BMNH 1860.3.19.757. Also described as new in Sykes (1840: 61); illustrated and described in more detail in Sykes (1841: 371).

Glyptosternum dekkanense Günther, 1864: 187. Type locality: Dekkan. Holotype: BMNH 1860.3.19.757.

Distribution: Deccan, Godavari and Krishna River basins, India (Jayaram, 1999).

Remarks: Holotype of *Bagrus lonah* Sykes has not been clearly established; see Günther (1864: 187), and Day (1873d: 748) for comments.

***Glyptothorax longicauda* Li, 1984**

Glyptothorax longicauda Li, 1984b: 82, fig. 3. Type locality: Tengchong, Yunnan, China. Holotype: KIZ 764126.

Distribution: Irrawaddy River basin, China (Chu *et al.*, 1999).

***Glyptothorax longjiangensis* Mo & Chu, 1986**

Glyptothorax longiangensis Mo & Chu, 1986: 344, 349, fig. 5. Type locality: Longjiang river (upper tributary of Irrawaddy river), Tengchong County (25°00'N, 98°30'E), Yunnan, China. Holotype: KIZ 764246.

Distribution: Irrawaddy River basin, Yunnan, China (Chu *et al.*, 1999).

***Glyptothorax macromaculatus* Li, 1984**

Glyptothorax macromaculatus Li, 1984b: 82, fig. 7. Type locality: Yangbi Xian, Yunnan, China. Holotype: KIZ 748804.

Distribution: Mekong River basin (Chu *et al.*, 1999; Kottelat, 2001b).

***Glyptothorax madraspatanus* (Day, 1873)**

Glyptosternum madraspatanum Day, 1873b: 526. Type locality: Bowany River, Neilgherries, Madras State, India.

Possible syntypes: AMS B.7759 (1), AMS B.8004 (1), NMW 46582–83 (2), RMNH 2739 (1), ZMB 10798 (1), ZMB 10829 (1), ZMB 11208 (1), ZSI F1235 (1), ZSI F1313 (1).

Distribution: Western Ghats, Anaimalai Hills, Nilgiri Hills, and Cauvery River (Jayaram, 1999).

***Glyptothorax major* (Boulenger, 1894)**

Akysis major Boulenger, 1894a: 246. Type locality: Senah, Tagora and Baram rivers, Sarawak. Syntypes: BMNH 1892.9.2.59 (1), BMNH 1892.10.7.26 (1), BMNH 1893.3.6.173–177 (5), BMNH 1893.3.6.178 (1, skeleton).

Glyptosternum kükenthali Steindachner, 1901: 448, pl. 18 (figs. 5, 5a). Type locality: Baram-Flusse, Borneo. Holotype: SMF 752.

Distribution: Malay Peninsula and western Borneo (Roberts, 1989a).

***Glyptothorax manipurensis* Menon, 1955**

Glyptothorax manipurensis Menon (A. G. K.), 1955: 23, fig. 1. Type locality: Barak River at Karong, Naga Hills, Manipur State, India. Holotype: ZSI F738/2.

Distribution: Barak River, Manipur, India (Jayaram, 1999).

Remarks: Redescribed in Kosygin & Vishwanath (2005).

***Glyptothorax minimaculatus* Li, 1984**

Glyptothorax minimaculatus Li, 1984b: 81, fig. 5. Type locality: Tengchong Xian, Yunnan, China. Holotype: KIZ 764336.

Distribution: Irrawaddy River basin, Yunnan, China (Chu *et al.*, 1999).

***Glyptothorax minutus* Hora, 1921**

Glyptothorax minutus Hora, 1921a: 180, fig. 1. Type locality: Imphal stream near Karong [Kameng], Manipur Valley, India. Syntypes (4): lost (Hora, 1921a: 182).

Distribution: Manipur, India (Hora, 1921a).

***Glyptothorax naziri* Mirza & Naik, 1969**

Glyptothorax naziri Mirza & Naik, 1969: 123, figs. 1–2. Type locality: Zhob River, Baluchistan, Pakistan. Holotype: GCM 6.

Distribution: NWFP, Punjab, Balochistan, Azad Kashmir, Pakistan (Mirza, 2003).

***Glyptothorax nelsoni* Ganguly, Datta & Sen, 1972**

Glyptothorax nelsoni Ganguly, Datta & Sen, 1972: 341, fig. 1. Type locality: Subarnarekha River, Chotanagpur Plateau, Bihar, India. Holotype: USNM 205611.

Distribution: Subarnarekha River, Bihar, India (Jayaram, 1999).

***Glyptothorax nieuwenhuisi* (Vaillant, 1902)**

Glyptosternon Nieuwenhuisi Vaillant, 1902: 72, 162, figs. 14–15. Type locality: Le Bloeoë, Bornéo central. Syntypes: MNHN 1891-0484 (1), MNHN 1891-0485 (1), MNHN 1891-0486 (1), MNHN 1891-0487 (1), MNHN 1903-0189 (1).

Distribution: Bloeoë River, Borneo.

***Glyptothorax obscurus* Li, 1984**

Glyptothorax obscura Li, 1984b: 86, fig. 1. Type locality: Jingdon Xian, Yunnan, China. Holotype: KIZ 737197.

Distribution: Upper Red River basin, China (Li, 1984b).

***Glyptothorax pallozonus* (Lin, 1934)**

Glyptosternum pallozonum Lin, 1934: 228, figs. 7–8. Type locality: Loh Fau Shan, Poh-lo District, Kwangtung, China. Holotype: Fisheries Experiment Station, Canton G10.

Distribution: Dongjiang, Guangdong, China (Chu *et al.*, 1999).

***Glyptothorax panda* Ferraris & Britz, 2005**

Glyptothorax panda Ferraris & Britz, 2005: 376, fig. 1. Type locality: Myanmar, Kachin State: Upper Irrawaddy River drainage, hill stream 8 miles from Kamaing on road to Tanai, vicinity of Myitkyina. Holotype: USNM 384824.

Distribution: Upper Irrawaddy River basin, Myanmar (Ferraris & Britz, 2005).

***Glyptothorax pectinopterus* (M'Clelland, 1842)**

Glyptosternon pectinopterus M'Clelland, 1842: 587. Type locality: The mountains of Simla. No types known.

Distribution: Ganges, Sutlej and upper Indus River basins, south Asia (Kullander, 1999).

***Glyptothorax platypogon* (Valenciennes, 1840)**

Pimelodus platypogon Valenciennes, in Cuvier & Valenciennes, 1840b: 152 (113 of Strasbourg deluxe edition). Type locality: Java. Possible syntypes: MNHN b-0196 (2), MNHN 0000-2903 (2), MNHN 0000-2904 (2); RMNH (not found); SMF 649 (2).

Pimelodus cyanochloros Bleeker, 1847b: 11. Type locality: Java. Syntypes: SMNS 10569 (6). Also described as new in Bleeker (1847a: 168).

Sundagagata robusta Boeseman, 1966: 243, fig. 1. Type locality: River near Buitenzorg, Java. Holotype: RMNH 25264.

Distribution: Java; western Borneo (Roberts, 1989a).

Remarks: *Sundagagata robusta* Boeseman, 1966, was considered by Kottelat *et al.* (1993: 76) to be an abnormal specimen of *Glyptothorax platypogon* (Valenciennes, 1840).

***Glyptothorax platypogonides* (Bleeker, 1855)**

Pimelodus platypogonides Bleeker, 1855a: 272. Type locality: Lahat, in fluviis. Syntypes (4, 70–85 mm TL): possibly BMNH 1863.12.4.154 (1), RMNH 6912 (4), RMNH 15289 (3).

Callomystax Schmidti Volz, 1904: 470. Type locality: Simbolon Gebirge, cent. Sumatra, Indonesia, elev. 1400 m. Syntypes (5): MHNG 683.22 (1), NMBA 2827 (1).

Distribution: Sumatra (Ng & Rachmatika, 2005).

Remarks: Register number of syntype of *Pimelodus platypogonides* incorrectly reported as BMNH 1863.12.11.154 in Eschmeyer *et al.* (1998). Name of species frequently cited as *Glyptothorax platypogonoides*.

Glyptothorax prashadi Mukerji, 1932

Glyptothorax prashadi Mukerji, 1932: 281, fig. 1. Type locality: Kyenchaung, Mergui District, Lower Burma. Holotype: ZSI F11334/1.

Distribution: Southeastern Myanmar and Peninsular Thailand (Jayaram, 1999).

Glyptothorax punjabensis Mirza & Kashmiri, 1971

Glyptothorax conirostris punjabensis Mirza & Kashmiri, 1971: 88, fig. 1. Type locality: Rawal Dam, Rawalpindi, Pakistan. Holotype: GCM 7.

Distribution: Soan River, Jhelum canal, Kabul River, and Siran River, Pakistan, and Kotli, Azad Kashmir (Jayaram, 1999).

Remarks: Treated as a possible synonym of *Glyptothorax kashmirensis* Hora, 1923, by Kullander (1999).

Glyptothorax quadriocellatus (Mai, 1978)

Glyptosternon quadriocellatum Mai, 1978: 272, fig. 121. Type locality: Fast-running creeks, n. Vietnam. possibly at DVZUT.

Glyptosternon minutum Mai, 1978: 274, fig. 122. Type locality: Fast-running creeks, n. Vietnam. Type (s): at DVZUT. Preoccupied in *Glyptothorax* by *Glyptothorax minutus* Hora, 1921; replaced by *Glyptothorax spectrum* Kottelat, 2001.

Glyptothorax spectrum Kottelat, 2001a: 55. Type locality: Fast-running creeks, n. Vietnam. Type (s): at DVZUT.

Replacement name for *Glyptosternon minutum* Mai, 1978, preoccupied in *Glyptothorax* by *Glyptothorax minutus* Hora, 1921.

Distribution: Yuanjiang, China (Chu *et al.*, 1999); northern Viet Nam (Kottelat, 2001a).

Remarks: Kottelat (2001a: 54) synonymized *G. quadriocellatum* and *G. minutum* and selected *G. quadriocellatum* as the valid name.

Glyptothorax saisi (Jenkins, 1910)

Glyptosternum saisi Jenkins, 1910: 128, fig.; pl. 6 (fig. 6). Type locality: Sita Nullah stream, Paresnath Hills, Bihar, w. Bengal, elev. 2150 ft. Holotype: ZSI F2583/1.

Distribution: Bihar, Uttar Pradesh, and Maharashtra, India (Jayaram, 1999).

Remarks: Redescribed in Tilak & Husain (1978).

Glyptothorax siamensis Hora, 1923

Glyptothorax siamensis Hora, 1923a: 168, pl. 12 (figs. 1–3). Type locality: Nakon Sritamarat Hills, Siam. Holotype: ZSI F10548/1.

Distribution: Thailand and Peninsular Malaysia (Ng & Rachimatika, 2005).

Glyptothorax silviae Coad, 1981

Glyptothorax silviae Coad, 1981a: 291, figs. 1–3. Type locality: Iran, Khuzestan, stream 3 km south of Bagh-e Malek, tributary to Rud-e Zard or Ab-e Ala in the drainage of the Jarrahi River, 31°29'N, 49°54'30"E, elev. 660 m. Holotype: NMC 79-0390A.

Distribution: Tigris River and possibly Gulf basins (Coad, 1995).

Glyptothorax sinensis (Regan, 1908)

Glyptosternum sinense Regan, 1908a: 110, pl. 4 (fig. 3). Type locality: Tunting [=Tungting], China. Holotype: BMNH 1907.11.26.4.

Distribution: lower and middle Changjiang [= Yangtze River], China (Chu *et al.*, 1999); also reported from Manipur, India (Kosygin & Vishwanath, 2005).

Remarks: Redescribed as *Glyptothorax sinense* in Xie *et al.* (2001).

Glyptothorax steindachneri (Pietschmann, 1913)

Glyptosternum steindachneri Pietschmann, 1913a: 93. Type locality: dem Tigris, Mosul. Syntypes (2): at NMW.

Distribution: Tigris River basin (Coad, 1995).

Glyptothorax stocki Mirza & Nijssen, 1978

Glyptothorax stocki Mirza & Nijssen, 1978: 79, fig. 1. Type locality: Bhed Nullah, small stream on G.T. road to Rawalpindi, 7 mi. from Lahore, Pakistan. Holotype: ZMA 114763.

Distribution: NWFP, Punjab, Azad Kashmir, Pakistan (Mirza, 2003).

Glyptothorax stoliczkae (Steindachner, 1867)

Glyptosternum Stoličkae Steindachner, 1867e: 533, pl. 6 (fig. 1). Type locality: Simla. Syntypes: NMW 76606 (3). Distribution: Western Himalaya, Shimla, Ganga, India (Jayaram, 1999).

***Glyptothorax striatus* (M'Clelland, 1842)**

Glyptosternon Striatus M'Clelland, 1842: 587, pl. 6 (in part). Type locality: Kasyah mountains. Syntypes (3): BMNH 1860.3.19.95 (1, 78 mm SL).

Distribution: Kashi and Garro Hills, India (Jayaram, 1999).

***Glyptothorax sufii* Bashir & Mirza, 1975**

Glyptothorax telchitta sufii Bashir & Mirza, 1975: 96, fig. 1. Type locality: River Sutlej, Lahore District, Pakistan. Holotype: GCM F-13.

Distribution: Indus River basin, Pakistan (Bashir & Mirza, 1975).

***Glyptothorax sykesi* (Day, 1873)**

Glyptosternum sykesi Day, 1873d: 748. Type locality: Deccan, India. Holotype: BMNH 1860.3.19.756. Name proposed conditionally for specimen in Günther (1864: 187) identified as *Glyptosternum lonah*, which Day contended was not the holotype and, in fact, not even conspecific with *G. lonah*.

Distribution: Peninsular India.

***Glyptothorax telchitta* (Hamilton, 1822)**

Pimelodus telchitta Hamilton, 1822: 185, 378. Type locality: Fresh water rivers of Bengal and Behar [now: India: west Bengal: Hooghly River at Kalna, 23°13'30.0"N, 88°22'39.0"E, by neotype designation]. Neotype: UMMZ 244946; designated by, and illustrated in, Ng (2005e:10, fig. 5). Unpublished Hamilton illustration reproduced in Hora (1929: pl. 21, fig. 3). Name spelled *Pimelodus telchita* on p. 378; first reviser may be Eschmeyer *et al.* (1998), but not yet clearly established to be earliest.

Distribution: Widely distributed in India (Jayaram, 1999).

***Glyptothorax trewavasae* Hora, 1938**

Glyptothorax trewavasae Hora, 1938d: 373, pl. 7 (figs. 3–4). Type locality: Yenna Valley, Satara dist., Maharashtra, India. Holotype: ZSI F9723/1.

Distribution: Krishna River basin, Yenna, Kayana, and Tunga Rivers, India (Jayaram, 1999).

***Glyptothorax trilineatus* Blyth, 1860**

Glyptothorax trilineatus Blyth, 1860: 154. Type locality: Tenasserim [Burma]. Syntypes: ZSI F10380/1 (2).

Glyptothorax trilineatoides Li, 1984b: 87, fig. 5. Type locality: Tengchong Xian, Yunnan, China. Holotype: KIZ 764336.

Distribution: Nujiang [=Salween River], Irrawaddy River (Chu *et al.*, 1999).

***Glyptothorax ventrolineatus* Vishwanath & Linthoingambi, 2005**

Glyptothorax ventrolineatus Vishwanath & Linthoingambi, 2005: 201, fig. 1. Type locality: Iril river, Ukhru district, Manipur, India. Holotype: MUMFL022I.

Distribution: Iril River, Chindwin River basin, Manipur, India (Vishwanath & Linthoingambi, 2005).

***Glyptothorax zanaensis* Wu, He & Chu, 1981**

Glyptothorax zanaensis Wu, He & Chu, 1981: 74, fig. 1 (1-b). Type locality: Qinghai-Xizang plateau region, China.

Syntypes: IHASW 606164–66 (3), IHASW 606168 (1), IHASW 606170–74 (5), IHASW 606178 (1), IHASW 6006651–53 (3).

Glyptothorax longinema Li, 1984b: 81, fig. 6. Type locality: Lushui Xian, Yunnan, China. Holotype: KIZ 741097.

Glyptothorax rubermentus Li, 1984b: 83, 88, fig. 8. Type locality: Baoshan Xian, Yunnan, China. Holotype: KIZ 749356.

Distribution: Mekong and Salween River basins, China (Chu *et al.*, 1999, as *Glyptothorax zanaensis*; Kottelat, 2001b).

***Glyptothorax zhuijiangensis* Lin, 2003**

Glyptothorax zhuijiangensis Lin, 2003: 159, 161. Type locality: Baishuidai Stream in Yaxi Town (22°15'N, 112°59'25"E), Xinhui City, middle-southern Guandong, China. Holotype: ZX 970901 (housed in IHASW).

Distribution: Zhujiang [= Pearl River] China (Lin, 2003).

Species inquirendae, Glyptothorax

Glyptosternum laak Popta, 1904: 190. Type locality: le Howong, Bornéo central. Syntypes: RMNH 7562 (3); described in more detail, with an illustration of one of the syntypes, in Popta (1906: 71, pl. 4, figs. 15a, 15b).

Glyptosternum tionsg Popta, 1904: 191. Type locality: le Kajan, Bornéo central. Syntypes: RMNH 7564 (2); described in more detail, with an illustration of one of the syntypes in Popta (1906: 75, pl. 4, fig. 16).

Glyptothonax conirostre poonaensis Hora, 1938d: 368, pl. 7 (figs. 5–6). Type locality: Mula Mutha R. at Poona, Maharashtra, Bombay State, India. Holotype: ZSI F12126/1. Originally as *Glyptothonax conirostre* var. *poonaensis*. Treated as valid or as a synonym of *Glyptothonax conirostris* (Steindachner, 1867).

GOGANGRA Roberts, 2001

Gangra Roberts & Ferraris, 1998: 333. Type species: *Pimelodus viridescens* Hamilton, 1822. Type by original designation. Gender: Feminine. Preoccupied by *Gangra* Walker, 1862; in Lepidoptera, replaced by *Gogangra* Roberts, 2001.

Gogangra Roberts, 2001: 83. Type species: *Pimelodus viridescens* Hamilton, 1822. Type by being a replacement name. Gender: Feminine. Replacement for *Gangra* Roberts & Ferraris, 1998; preoccupied by *Gangra* Walker, 1862, in Lepidoptera.

Gogangra laevis Ng, 2005

Gogangra laevis Ng, 2005c: 280, fig. 1. Type locality: Bangladesh: Gowain River and Khal at Gowainghat. Holotype: UMMZ 244603.

Distribution: Yamuna and Meghna Rivers, lower Brahmaputra River basin, Bangladesh (Ng, 2005c).

Gogangra viridescens (Hamilton, 1822)

Pimelodus viridescens Hamilton, 1822: 173, 377, pl. 11 (fig. 56). Type locality: Rivers in the northern parts of Bengal. No types known.

Nangra punctata Day, 1877 (in Day, 1875–78): 494, pl. 115 (fig. 8). Type locality: Sone River at Bheer Bhoom in Bengal, India. Syntype: AMS B.7566 (1).

Distribution: Ganges and Brahmaputra River basins (Roberts & Ferraris, 1998; Ng, 2005c).

HARA Blyth, 1860

Hara Blyth, 1860: 152. Type species: “*Hara buchanani* nobis; *Pimelodus hara*, B.H.” [= *Pimelodus hara* Hamilton, 1822]. Type by original designation. Gender: Feminine.

Laguvia Hora, 1921b: 739. Type species: *Pimelodus asperus* M’Clelland, 1844. Type by subsequent designation, by Jordan (1923: 148). Gender: Feminine.

Remarks: Jayaram (1973c, 1999) lists *Laguvia shawi* as the type species of *Laguvia*, by subsequent designation, but it is not stated where this designation was published. If published before 1923, it would have priority over the designation by Jordan (1923). Therefore, the type species as stated above must be considered tentative until the details of the designation of *L. shawi* are clarified. Synonymy of *Laguvia* in *Hara* follows Ng & Kottelat (2005: 20).

Hara filamentosa Blyth, 1860

Hara filamentosa Blyth, 1860: 152. Type locality: Tenasserim [Burma]. Syntypes: ZSI 585 (6).

Distribution: Widely distributed in Myanmar.

Hara hara (Hamilton, 1822)

Pimelodus hara Hamilton, 1822: 190, 378. Type locality: River Kosi [now Hooghly River south of Ranaghat, by neotype designation]. Neotype: UMMZ 244697, designated by, and illustrated in, Ng & Kottelat (2005: 20, fig. 5). Previously unpublished Hamilton illustration reproduced in Hora (1929: pl. 21, fig. 5) and reproduced in Ng & Kottelat (2005: 20, fig. 6).

Pimelodus asperus M’Clelland, 1844a: 404, pl. 24 (fig. 2). Type locality: Chusan, China [now Hooghly River south of Ranaghat, by neotype designation]. Neotype: UMMZ 244697, designated by, and illustrated in, Ng & Kottelat (2005: 19, fig. 5).

Hara buchanani Blyth, 1860: 152. Type locality: River Kosi [now Hooghly River south of Ranaghat, by neotype designation]. Neotype: UMMZ 244697, designated by, and illustrated in, Ng & Kottelat (2005: 20, fig. 5) as

neotype of *Pimelodus hara*. Unneeded new name for *Pimelodus hara* Hamilton, 1822.

Distribution: Bihar, Uttar Pradesh, Orissa, northern Bengal and Assam, India; Bangladesh and Myanmar [questionable] (Jayaram, 1979).

Remarks: Redescribed in Tilak (1978).

***Hara horai* Misra, 1976**

Hara horai Misra, 1976: 245, pl. 9 (figs. 1–3). Type locality: Terai and Duars, n. Bengal. Syntype: ZSI FF955 (formerly ZSI F 11390/1), illustrated in Hora (1950: pl. 2, figs. 1–3) as *Hara hara*.

Distribution: Terai Duars, northern Bengal, India (Misra, 1976).

Remarks: Neotype designation by Tilak & Talwar (1976: 246) [ZSI FF955] invalid inasmuch as that specimen is apparently one of the original syntypes (M. Kottelat, pers. commun.). Redescribed in Tilak (1978).

***Hara jerdoni* Day, 1870**

Hara jerdoni Day, 1870a: 39, pl. 4 (figs. 2a–c). Type locality: Sylhet district. Syntypes (2): ZSI 431 (1).

Distribution: Sylhet district, Bangladesh (Jayaram, 1979).

Remarks: Redescribed in Husain & Tilak (1978).

***Hara saharsai* Datta Munshi & Srivastava, 1988**

Hara saharsai Datta Munshi & Srivastava, 1988: 265, fig. 5. Type locality: Kosi Belt, n. Bihar, India. Syntypes: Bhagalpur Univ., Dept. Zool. Mus. 67 (13). Also appeared as new in Srivastava (1989: 121, fig. 1).

Distribution: northern Bihar, India (Datta Munshi & Srivastava, 1988).

***Hara serrata* Vishwanath & Kosygin, 2000**

Hara serratus Vishwanath & Kosygin, 2000: 143, figs. 2–5. Type locality: Jiri river at Jiribam, Manipur, India.

Holotype: MUMF 2508.

Distribution: Barak and Jiri Rivers, Manipur, India (Vishwanath & Kosygin, 2000).

***MYERSGLANIS* Hora & Silas, 1952**

Myersglanis Hora & Silas, 1952a: 19. Type species: *Exostoma blythii* Day, 1870. Type by original designation. Gender: Masculine.

Key: Vishwanath & Kosygin (1999: 296).

***Myersglanis blythii* (Day, 1870)**

Exostoma Blythii Day, 1870b: 525. Type locality: not stated. Syntypes (2): Asiatic Society of Bengal 599 (2) [now at ZSI] (2), possibly ZSI 2361 (1).

Distribution: Possibly Pharping, Nepal (Regan, 1907c; Hora & Silas, 1952a).

***Myersglanis jayarami* Vishwanath & Kosygin, 1999**

Myersglanis jayarami Vishwanath & Kosygin, 1999: 291, pl. 1. Type locality: Laniye river at Jessami, Manipur, India (94°32'E, 25°38'N). Holotype: MUMF 2138.

Distribution: Lainye River, Chindwin River basin, Manipur, India (Vishwanath & Kosygin, 1999).

***NANGRA* Day, 1877**

Nangra Day, 1877 (in Day, 1875–78): 493. Type species: *Pimelodus nangra* Hamilton, 1822. Type by absolute tau-tonomy. Gender: Feminine.

Revision: Hora & Law (1941); Roberts & Ferraris (1998).

Remarks: See Jayaram (1972a) for comments on the validity of the genus.

***Nangra assamensis* Sen & Biswas, 1994**

Nangra assamensis Sen & Biswas, 1994: 441, fig. 1; pl. 1. Type locality: Brahmaputra River at Neematighat, 14 kms. from Jorhat, Assam [India]. Holotype: ZSI V/F/ERS/53.

Nangra carcharhinoides Roberts & Ferraris, 1998: 338, fig. 16. Type locality: Ganges River at Patna, India. Holotype: CAS 95566.

Distribution: Ganges and Brahmaputra Rivers, India (Roberts & Ferraris, 1998).

***Nangra bucculenta* Roberts & Ferraris, 1998**

Nangra bucculenta Roberts & Ferraris, 1998: 336, fig. 14. Type locality: Ganges River delta, Tangail District, North Central Region, Bangladesh. Holotype: CAS 95564.

Distribution: Ganges River delta, Bangladesh (Roberts & Ferraris, 1998).

***Nangra nangra* (Hamilton, 1822)**

Pimelodus nangra Hamilton, 1822: 193, 378, pl. 11 (fig. 63). Type locality: Kosi River [now: Ganges River at Patna, India, by neotype designation]. Neotype: CAS 96626, designated by Roberts & Ferraris (1998: 341).

Nangra Buchananii Day, 1877 (in Day, 1875–78): 494, pl. 113 (fig. 3). Kosi River [now: Ganges River at Patna, India, by neotype designation]. Neotype: CAS 96626. Replacement for *Pimelodus nangra* Hamilton, 1822, to avoid tautomy.

Distribution: Ganges and Indus River basins (Roberts & Ferraris, 1998).

***Nangra ornata* Roberts & Ferraris, 1998**

Nangra ornata Roberts & Ferraris, 1998: 341, fig. 19. Type locality: Bangladesh: Gowain River and Khal at Gowainghat, northern Sylhet Province (Surma or Meghna watershed). Holotype: UMMZ 233236.

Distribution: Surma or Meghna watershed, Bangladesh (Roberts & Ferraris, 1998).

***Nangra robusta* Mirza & Awan, 1973**

Nangra robusta Mirza & Awan, 1973: 145, fig. 1. Type locality: Indus River at Jinnah Barrage near Kalabagh, Pakistan. Holotype: GCM 9.

Distribution: Indus River, Pakistan (Mirza & Awan, 1973).

***OREOGLANIS* Smith, 1933**

Oreoglanis Smith, 1933: 70. Type species: *Oreoglanis siamensis* Smith, 1933. Type by original designation. Gender: Masculine.

Paroreoglanis Pellegrin, 1936: 244. Type species: *Paroreoglanis delacouri* Pellegrin, 1936. Type by monotypy. Gender: Masculine.

***Oreoglanis delacouri* (Pellegrin, 1936)**

Paroreoglanis delacouri Pellegrin, 1936b: 244. Type locality: Xieng Khouang in upper Laos. Lectotype: MNHN 1936-0031 (103.4 mm SL), designated by Ng & Kottelat (1999: 379).

Distribution: Nam Ngiap basin, Mekong drainage, Laos (Ng & Rainboth, 2001).

***Oreoglanis frenatus* Ng & Rainboth, 2001**

Oreoglanis frenatus Ng & Rainboth, 2001: 7, fig. 6. Type locality: Laos: Xieng Khouang Province, Nam Ka basin, Houay Kheua at Hwy 7 bridge, 19°38'N, 103°28'E. Holotype: UMMZ 236811.

Distribution: Nam Ka Basin, Mekong drainage, Laos (Ng & Rainboth, 2001).

***Oreoglanis hypsiurus* Ng & Kottelat, 1999**

Oreoglanis hypsiurus Ng & Kottelat, 1999: 376, fig. 1. Type locality: Upper Nam Theun, ca. 1 km upstream of confluence with Houay Nuok Lan, 18°04'09"N, 105°29'44"E, Kham-mouan Province, Laos. Holotype: ZRC 40440.

Distribution: Nam Kading basin, Mekong drainage, Laos (Kottelat, 2001b).

***Oreoglanis infulatus* Ng & Freyhof, 2001**

Oreoglanis infulatus Ng & Freyhof, 2001b: 1165, figs. 1–3. Type locality: Vietnam: Ha Tinh Province, stream at Son Kim, a tributary of Song Lam (18°24'25"N, 105°11'10"E). Holotype: ZFMK 35719.

Distribution: Lam River headwaters, central Vietnam (Ng & Freyhof, 2001b).

***Oreoglanis insignis* Ng & Rainboth, 2001**

Oreoglanis insignis Ng & Rainboth, 2001: 15, fig. 8. Type locality: China: Yunnan Province, Baoshan Prefecture, Longchuanjiang and Dajiang, near Qushi. Holotype: KIZ 9810191.

Distribution: Upper Irrawaddy River basin, China and Myanmar, possibly upper Salween basin (Ng & Rainboth, 2001).

***Oreoglanis lepturus* Ng & Rainboth, 2001**

Oreoglanis lepturus Ng & Rainboth, 2001: 17, fig. 10. Type locality: Laos: Bolikamsai Province, Nam Phao about 2 km from Vietnam border, 18°23'N, 105°19'E. Holotype: UMMZ 236814.

Distribution: Nam Phao basin, Mekong drainage, northeastern Laos (Ng & Rainboth, 2001).

***Oreoglanis macronemus* Ng, 2004**

Oreoglanis macronemus Ng, 2004e: 209, fig. 1. Type locality: Laos: Xieng Khouang. Holotype: BMNH 1933.8.19.51.

Distribution: Nam Ngiap or Nam Sen River basins, Xieng Khouang region, Laos (Ng, 2004e).

***Oreoglanis macropterus* (Vinciguerra, 1890)**

Exostoma macropterum Vinciguerra, 1890: 253, pl. 8 (fig. 5). Type locality: Paese dei Catcin, Birmania [Myanmar].

Syntypes: BMNH 1893.2.16.18 (1), MSNG 14410 (3), NMW 46488 (1), RMNH 10236 (1), USNM 44805 (1)

Distribution: Irrawaddy River basin; Salween River, China (Ng & Rainboth, 2001).

Remarks: See Steinitz (1961) for comments on taxonomy.

***Oreoglanis setiger* Ng & Rainboth, 2001**

Oreoglanis setiger Ng & Rainboth, 2001: 23, fig. 12. Type locality: Laos: Louang Namtha Province, Nam Ma Oun, 21°05'N, 101°04'E. Holotype: UMMZ 236813.

Distribution: Nam Oun basin, Mekong River drainage, Laos, and upper Mekong River southwestern China (Ng & Rainboth, 2001).

***Oreoglanis siamensis* Smith, 1933**

Oreoglanis siamensis Smith, 1933: 72, pl. 3 (figs. 1–2); fig. 4. Type locality: Kang River near the base of Doi Angka, northern Siam. Holotype: KUMF 0172.

Distribution: Mae Nam Ping basin, Chao Phraya drainage, northern Thailand (Ng & Rainboth, 2001).

Remarks: Statements in Menon & Yazdani (1968) and Eschmeyer *et al.* (1998) which indicated that ZSI F12233/1 might be the holotype of *Oreoglanis siamensis* are in error. Smith (1933: 74) clearly stated that two specimens were examined, which he identified as the “type” and “topotype,” and that the specimen deposited in the Indian Museum (now ZSI) was the topotype.

***PARACHILOGLANIS* Wu, He & Chu, 1981**

Parachiloglanis Wu, He & Chu, 1981: 76, 79. *Glyptosternum hodgarti* Hora, 1923. Type by original designation.

Gender: Masculine.

***Parachiloglanis hodgarti* (Hora, 1923)**

Glyptosternum hodgarti Hora, 1923b: 38, pl. 2 (figs. 1–3). Type locality: Pharping, Nepal. Holotype: ZSI F1553/1.

Distribution: Yaluzangbujiang [= upper Brahmaputra River] (Chu *et al.*, 1999); Abor Hills, Meghalaya, Darjeeling, Kali River, Teesta valley and Nepal (as *Euchiloglanis hodgarti*) (Jayaram, 1999).

***PAREUCHILOGLANIS* Pellegrin, 1936**

Pareuchiloglanis Pellegrin, 1936b: 245. Type species: *Pareuchiloglanis poilanei* Pellegrin, 1936. Type by monotypy. Gender: Masculine.

Remarks: Included species follows He (1996b).

Review: Ding (2003, western China).

***Pareuchiloglanis anteanalis* Fang, Xu & Cui, 1984**

Pareuchiloglanis anteanalis Fang, Xu & Cui, 1984: 209, fig. 1. Type locality: Wudu County, Gansu Province, China.

Holotype: Shaanxi Inst. Zool. 82VI9565.

Distribution: Wudu County, Gansu Province, China (Fang, *et al.*, 1984).

***Pareuchiloglanis feae* (Vinciguerra, 1890)**

Exostoma Feae Vinciguerra, 1890: 256, pl. 8 (fig. 6). Type locality: Taó, e Iadó, Birmania [Myanmar]. Syntypes: BMNH 1893.2.16.19–20 (2), BMNH 1893.2.16.21–22 (2), BMNH 1893.2.16.23 (1, skeleton), MSNG 14411 (4), MSNG 14412 (3), MSNM 38 (1), NMW 9064–65 (2), RMNH 10237 (3), USNM 44743 (3).

Distribution: Irrawaddy and Salween Rivers, China (Chu *et al.*, 1999).

***Pareuchiloglanis gongshanensis* Chu, 1981**

Pareuchiloglanis gongshanensis Chu, 1981: 28, 31, fig. 2. Type locality: Gongshan Xian, Yunnan, China. Holotype: KIZ 731202.

Distribution: Upper Nujiang [= Salween River], China (Chu *et al.*, 1999).

***Pareuchiloglanis gracilicaudatus* (Wu & Chen, 1979)**

Euchiloglanis gracilicaudata Wu & Chen, 1979: 294, fig. 4. Type locality: Za-Qu, upper Lancang Jiang, Nangqen, Qinghai, China. Syntypes: Qinghai Inst. Biol. (12).

Distribution: Lancangjiang [= Mekong River], China (Chu *et al.*, 1999).

***Pareuchiloglanis kamengensis* (Jayaram, 1966)**

Euchiloglanis kamengensis Jayaram, 1966d: 85, fig. 1. Type locality: Norgum River at Kalaktang, 1370 m alt., Kameng Frontier Division, N.E.F.A. Holotype: ZSI F2105/2.

Distribution: Yaluzangbujiang [= upper Brahmaputra River], Irrawaddy, Nujiang [= Salween River], Lancangjiang [= Mekong River], China (Chu *et al.*, 1999).

***Pareuchiloglanis longicauda* (Yue, 1981)**

Euchiloglanis longicauda Yue, in Zheng, 1981: 183, fig. 151. Type locality: Panyang R. (Hong-Shui R. system), Bama Co., Guangxi Prov., China. Holotype: ASIZB and Fish. Res. Inst., Guangxi Zhuangzu Autonomous Region 750443.

Distribution: Nanpanjiang, Beipanjiang, Hongshuihe, China (Chu *et al.*, 1999).

***Pareuchiloglanis macropterus* Ng, 2004**

Pareuchiloglanis macropterus Ng, 2004g: 3, fig. 1. Type locality: China, Yunnan province, Salween River (Nujiang) drainage; Laowo River, a tributary of Salween River (Nujiang), ca. 5 km to Liuku on Yongping–Liuku Road, 25°50'18.6"N 98°53'46.8"E; 900 m asl. Holotype: ZRC 49124.

Distribution: Salween and Irrawaddy River basins, southern China (Ng, 2004g).

***Pareuchiloglanis macrotremus* (Norman, 1925)**

Euchiloglanis macrotrema Norman, 1925b: 570. Type locality: Ngoi-Tio, Col des Nuages, Tonkin, Vietnam, elev. 4500–6500 ft. Syntypes: BMNH 1925.2.19.5–6 (2).

Distribution: Yuanjiang, China (Chu *et al.*, 1999); Tonkin, Vietnam.

***Pareuchiloglanis myzostomus* (Norman, 1923)**

Euchiloglanis myzostoma Norman, 1923a: 562. Type locality: Yunnan, China. Syntypes: BMNH 1923.2.21.40–49 (10).

Distribution: Lancangjiang [= Mekong River], China (Chu *et al.*, 1999).

***Pareuchiloglanis nebulifer* Ng & Kottelat, 2000**

Pareuchiloglanis nebulifer Ng & Kottelat, 2000b: 11, fig. 4. Type locality: Small creek, trib. of Houai Siam, upstream of Ban Kangpabong, Houaphan Prov., Laos, 20°19'36"N, 104°25'01"E. Holotype: ZRC 45706.

Distribution: Nam Xam and Nam Ma basins, Ma River Drainage, Laos (Kottelat, 2001b).

***Pareuchiloglanis poilanei* Pellegrin, 1936**

Pareuchiloglanis poilanei Pellegrin, 1936b: 246. Type locality: Annam [Vietnam]. Syntypes: MNHN 1936-0005 (1), MNHN 1936-0006 (1), MNHN 1936-0007 (1), MNHN 1936-0008 (1), MNHN 1936-0009 (1), MNHN 1936-0010 (1), MNHN 1936-0011 (1), MNHN 1936-0012 (1), MNHN 1936-0013 (1), MNHN 1936-0014 (1), MNHN 1936-0015 (1), MNHN 1935-0016 (1), MNHN 1936-0017 (1), MNHN 1936-0018 (1), MNHN 1936-0019 (1), MNHN 1936-0020 (4).

Distribution: Central Vietnam.

***Pareuchiloglanis rhabdurus* Ng, 2004**

Pareuchiloglanis rhabdurus Ng, 2004g: 7, fig. 3. Type locality: Vietnam: Ha Giang province, Red River (Song Hong) drainage, Vi Xuyen district, Cao Bo stream (Bac Trao river) near camp 1, 22°45'18.0"N, 104°52'11.4"E. Holotype: AMNH 211153.

Distribution: Song Lo River, Red River basin, northern Vietnam (Ng, 2004g).

***Pareuchiloglanis robustus* Ding, Fu & Ye, 1991**

Pareuchiloglanis robusta Ding, Fu & Ye, 1991: 369, fig. 1. Type locality: a stream (Qing Yijing), Sichuan, China. Holotype: SPNRI 8801.

Distribution: Yangtze River basin, China (Ding *et al.*, 1991).

***Pareuchiloglanis sichuanensis* Ding, Fu & Ye, 1991**

Pareuchiloglanis sichuanensis Ding, Fu & Ye, 1991: 371, fig. 2. Type locality: Baoxing, Sichuan Province, China. Holotype: Sichuan Agric. Univ. 6-1481.

Distribution: Yangtze River basin, China (Ding *et al.*, 1991).

***Pareuchiloglanis sinensis* (Hora & Silas, 1952)**

Euchiloglanis sinensis Hora & Silas, 1952a: 17, fig. 2. Type locality: presumably from some part of China, possibly Yunnan. Holotype: ZSI F12208/1.

Distribution: Jinshajiang, China (Chu *et al.*, 1999).

Pareuchiloglanis songdaensis Nguyen & Nguyen, 2001

Pareuchiloglanis songdaensis Nguyen & Nguyen, 2001: 67, fig. 1. Type locality:[Da River, Muong Te, Lai Chau Town and Phong Tho, Vietnam]. Holotype (115 mm SL): LM 2001.01 H.

Distribution: Da River, Vietnam (Nguyen & Nguyen, 2001).

Pareuchiloglanis songmaensis Nguyen & Nguyen, 2001

Pareuchiloglanis songmaensis Nguyen & Nguyen, 2001: 68, fig. 2. Type locality: [Ma River, Song Ma district, Son La Province, Vietnam]. Holotype (82 mm SL): LM 2001.02H.

Distribution: Ma River, Vietnam (Nguyen & Nguyen, 2001).

Pareuchiloglanis tianquanensis Ding & Fang, 1997

Pareuchiloglanis tianquanensis Ding & Fang, 1997: 17, fig. 1. Type locality: a stream (Upper Qing Yi River), altitude 2010 m., Tianquan County, Sichuan Province [China]. Holotype: SPNRI 920185.

Distribution: Upper Qing Yi River basin, Sichuan, China (Ding & Fang, 1997).

PSEUDECHENEIS Blyth, 1860

Pseudecheneis Blyth, 1860: 154. Type species: *Glyptosternon sulcatus* M'Clelland, 1842. Type by monotypy. Gender: Feminine.

Parapseudecheneis Hora, in Hora & Chabanaud, 1930: 216. Type species: *Pseudecheneis paviei* Vaillant, 1892. Type by monotypy. Gender: Feminine.

Propseudecheneis Hora, 1937d: 348. Type species: *Propseudecheneis tchangi* Hora, 1937. Type by original designation. Gender: Feminine.

Review: Hora (1952); Hora & Chabanaud (1930); Chu (1982), China.

Phylogeny: Chu (1982), Zhou & Zhou (2005).

Pseudecheneis crassicauda Ng & Edds, 2005

Pseudecheneis crassicauda Ng & Edds, 2005b: 2, fig. 1. Type locality: Nepal: Mewa Khola (River), Dhakuta District, 27°0'N, 87°20'E. Holotype: BMNH 1958.9.1.8.

Distribution: Tamur River basin, Kosi drainage, Nepal (Ng & Edds, 2005b).

Pseudecheneis immaculata Chu, 1982

Pseudecheneis immaculatus Chu, 1982: 428, 447, fig. 1. Type locality: Baijixun (upper Lancang River), Weixi County, Yunnan, China. Holotype: KIZ 748742.

Distribution: upper Langcanjiang [= Mekong River], China (Chu *et al.*, 1999).

Pseudecheneis paviei Vaillant, 1892

Pseudecheneis paviei Vaillant, 1892a: 126. Type locality: Near Lai Chau, Tonkin, northern Vietnam. Holotype: MNHN 1892-0049; redescribed and illustrated in Vaillant (1904: 464, pl. 22, fig. 3). Holotype illustrated in Hora & Chabanaud (1930: 217, fig. 2).

Pseudecheneis intermedius Chu, 1982: 430, 447, fig. 2. Type locality: Dongbao (a stream at upper reaches of Babian River, belonging to the Red River drainage), Jingdong County [Yunnan, China]. Holotype: KIZ 737173.

Distribution: Northern Vietnam and Yunnan, China (Kottelat, 2001a).

Remarks: Synonymy follows Ng & Edds (2005b: 12).

Pseudecheneis serracula Ng & Edds, 2005

Pseudecheneis serracula Ng & Edds, 2005b: 6, fig. 3. Type locality: Nepal: Mugu/Bajura, Jhugala, Karnali River, purchased at Jhugala, 29°31'18.0"N, 81°46'48.0"E. Holotype: KU 29554.

Distribution: Karnali and Gandaki River basins, Nepal (Ng & Edds, 2005).

Pseudecheneis sulcata (M'Clelland, 1842)

Glyptosternon sulcatus M'Clelland, 1842: 587, pl. 6 (in part). Type locality: Kasyah Hills. Holotype: BMNH 2005.5.17.5 (76 mm SL).

Distribution: Lancangjiang [= Mekong River], Nujiang [= Salween River], Yaluzangbujiang [= upper Brahmaputra River], Irrawaddy River, China (Chu *et al.*, 1999).

Pseudecheneis sulcatoides Zhou & Chu, 1992

Pseudecheneis sulcatoides Zhou & Chu, 1992: 111, 115, figs. 1–3. Type locality: Yangbi (25°40'N, 99°57'E), [Lan-

cangjiang River (=upper Mekong)] Yunnan Province, China. Holotype: KIZ 839059.

Distribution: Upper Mekong River basin, Yunnan, China (Zhou & Chu, 1992).

Pseudecheneis sympelvica Roberts, 1998

Pseudecheneis sympelvicus Roberts, 1998: 290, fig. 1. Type locality: Nam Veo, tributary of Nam Phao 25 km east of Lak Sao, Nam Theun watershed, Mekong Basin, central Laos. Holotype: ZRC 40359.

Distribution: Nam Theun watershed, Mekong River basin, Laos (Roberts, 1998).

Pseudecheneis tchangi Hora, 1937

Propseudecheneis tchangi Hora, 1937d: 348, fig. 11b. Type locality: Yunnan, China. Holotype: Zoological Museum of Fan Memorial Institute, Beijing, 12016.

Distribution: Red River, Yunnan, China (Hora, 1937d).

PSEUDEXOSTOMA Chu, 1979

Pseudexostoma Chu, 1979: 78, 81. Type species: *Glyptosternum yunnanensis* Tchang, 1935. Type by original designation. Gender: Neuter.

Pseudexostoma brachysoma Chu, 1979

Pseudexostoma yunnanensis brachysoma Chu, 1979: 78, 81, fig. 3. Type locality: Hill stream running into the Nu River, near Laowo, Yunlong Xian [Yunnan, China]. Holotype: KIZ 742093.

Distribution: Nujiang [= Salween River], China (Chu et al., 1999).

Pseudexostoma yunnanense (Tchang, 1935)

Glyptosternum yunnanensis Tchang, 1935b: 174, fig. 1. Type locality: Yunnan, China. Holotype: ASIZB [= ZMFMIB] 12027.

Distribution: Irrawaddy River, China (Chu et al., 1999).

PSEUDOLAGUVIA Misra, 1976

Pseudolaguvia Misra, 1976: 253. Type species: *Glyptothorax tuberculatus* Prashad & Mukerji, 1929. Type by original designation. Gender: Feminine.

Remarks: See Jayaram (1973c) for comments on this genus, but as *Laguvia*.

Pseudolaguvia foveolata Ng, 2005

Pseudolaguvia foveolata Ng, 2005b: 174, fig. 1. Type locality: India: West Bengal: Tista River at Tista barrage, 26°45'10"N, 88°34'11"E. Holotype: UMMZ 244867.

Distribution: Tista River, Brahmaputra River basin, India (Ng, 2005b).

Pseudolaguvia inornata Ng, 2005

Pseudolaguvia inornata Ng, 2005g: 36, fig. 1. Type locality: Bangladesh: Chittagong District, Koilla Khal (creek), 10 km E of Feni-Chittagong highway on road to Ramgarh, 22°55'N, 91°36'E. Holotype: UMMZ 245580.

Distribution: Feni River basin, Bangladesh (Ng, 2005g).

Pseudolaguvia kapuri (Tilak & Husain, 1975)

Laguvia ribeiroi kapuri Tilak & Husain, 1975: 1, figs. 1–3. Type locality: Padhoi river, near Kalsia Ghat, Saharanpur, (U. P.) [India]. Holotype: ZSI/NRS 836.

Distribution: Padhoi River and Dhamola River, Saharanpur, Uttar Pradesh (Jayaram, 1999).

Remarks: Redescribed in Gupta (1982).

Pseudolaguvia muricata Ng, 2005

Pseudolaguvia muricata Ng, 2005g: 41, fig. 3. Type locality: Bangladesh: Sylhet District, Rangapani Khal (creek), 6 km WNW of Sylhet-Shillong highway, 25°10'N, 92°6'E. Holotype: UMMZ 245581.

Distribution: Brahmaputra River basin, Bangladesh (Ng, 2005g).

Pseudolaguvia ribeiroi (Hora, 1921)

Laguvia ribeiroi Hora, 1921b: 741, pl. 29 (fig. 3). Type locality: Khoila River, tributary of Teesta at Jalpaiguri [India]. Holotype: ZSI F 10086/1.

Distribution: Teesta River system, north Bengal; Kosi River system Behar; Rihand River system, Uttar Pradesh, and rivers of Nepal (Jayaram, 1979, 1999).

Pseudolaguvia shawi (Hora, 1921)

Laguvia shawi Hora, 1921b: 740, pl. 29 (fig. 2). Type locality: Mahanadi River below Darjeeling, India. Holotype: ZSI F10085/1.

Distribution: North Bengal, India, and Bangladesh (Jayaram, 1999); Brahmaputra River basin, Assam, India (Arun-kumar, 2000).

***Pseudolaguvia tenebricosa* Britz & Ferraris, 2003**

Pseudolaguvia tenebricosa Britz & Ferraris, 2003: 2, fig. 1. Type locality: Myanmar, Kayin Division, Pathe Chaung, hill stream, 13 miles east of Taungoo, 19°01'11"N, 96°35'33"E. Holotype: USNM 373293.

Distribution: Sittang and upper Irrawaddy River basins, Myanmar (Britz & Ferraris, 2003).

***Pseudolaguvia tuberculata* (Prashad & Mukerji, 1929)**

Glyptothorax tuberculatus Prashad & Mukerji, 1929: 182, pl. 7 (fig. 2); fig. 4. Type locality: Sankha, a large hill-stream, midway between Kamaing and Mogaung, Myitkyina dist., Upper Burma. Holotype: ZSI F10876/1.

Distribution: Myitkyina District, Kachin State, Myanmar (Prashad & Mukerji, 1929).

***SISOR* Hamilton, 1822**

Sisor Hamilton, 1822: 208, 379. Type species: *Sisor rhabdophorus* Hamilton, 1822. Type by monotypy. Gender: Masculine.

Revision: Ng (2003c).

***Sisor barakensis* Vishwanath & Darshan, 2005**

Sisor barakensis Vishwanath & Darshan, 2005: 1952, fig. 1a. Type locality: River Barak, Jiri, Manipur, India. Holotype: MUMF 3131. Holotype illustrated as Image 1 on web version of paper.

Distribution: Barak River, Brahmaputra River basin, India (Vishwanath & Darshan, 2005).

***Sisor chennuah* Ng & Lahkar, 2003**

Sisor chennuah Ng & Lahkar, in Ng, 2003c: 2876, fig. 5. Type locality: India, Assam State, Brahmaputra River drainage, Dibrugarh. Holotype: NRM 40420.

Distribution: Brahmaputra River basin, India and Bangladesh (Ng, 2003c).

***Sisor rhabdophorus* Hamilton, 1822**

Sisor rhabdophorus Hamilton, 1822: 208, 379. Type locality: Northern rivers of Bengal and Behar [now India: West Bengal State, Bhagirathi River at crossing point between Kalna (Barddhaman District) and Nisinghapur (Nadia District) (23°13'33" N, 88°32'41.4"E), by neotype designation]. Neotype: ZRC 45829, designated by Ng (2003c: 2873, 2881).

Distribution: Ganges River basin, West Bengal, India (Ng, 2003c).

Remarks: Name often misspelled *Sisor rhabdophorus*, following the incorrect subsequent spelling in Cuvier & Valenciennes (1840).

***Sisor rheophilus* Ng, 2003**

Sisor rheophilus Ng, 2003c: 2877, fig. 6. Type locality: India: Uttar Pradesh State, Kali Nadi River, near Muzaffarnagar. Holotype: UMMZ 189651.

Distribution: Upper and middle Ganges River basin, Bihar State and Uttar Pradesh, India (Ng, 2003c).

***Sisor tortosus* Ng, 2003**

Sisor tortosus Ng, 2003c: 2878, fig. 7. Type locality: India: Bihar State, Ganges River at Patna. Holotype: CAS 96629.

Distribution: Middle Ganges River basin, Bihar and Deli States, India (Ng, 2003c).

***Species inquirenda*, Sisoridae**

Exostoma Oschanini Herzenstein, 1889a: 70. Upper Amu-Darya R., upper Syr-Darya R. basin (Ugam R, Tashkent), Uzbekistan. Syntypes: BMNH 1888.11.24.1 (1), ZIN 8055 (1), ZIN 8056 (1), ZIN 8057 (2). Also in Herzenstein (1889b:120).

TRICHOMYCTERIDAE Bleeker, 1858

Eremophilini Bonaparte, 1846: 5. Type genus: *Eremophilus* Humboldt, 1805.
Trichomycterini Bleeker, 1858b: 49, 250, 257. Type genus: *Trichomycterus* Valenciennes, 1832.
Vandelliini Bleeker, 1862 (in Bleeker, 1862–63): 17. Type genus: *Vandellia* Valenciennes, 1846.
Stegophilina Günther, 1864: 5, 12, 276. Type genus: *Stegophilus* Reinhardt, 1859.
Pareiodontinae Eigenmann, 1918c: 261. Type genus: *Pareiodon* Kner, 1855.
Tridentinae Eigenmann, 1918c: 275. Type genus: *Tridens* Eigenmann & Eigenmann, 1889.
Glanapteryginae Myers, 1944: 592. Type genus: *Glanapteryx* Myers, 1927.
Sarcoglanidinae Myers & Weitzman, 1966: 278. Type genus: *Sarcoglanis* Myers & Weitzman, 1966.
Trichogeninae Isbrücker, 1986: 276. Type genus: *Trichogenes* Britski & Ortega, 1983.
Copianodontinae de Pinna, 1992b: 179. Type genus: *Copianodon* de Pinna, 1992.
Taxonomic summary: de Pinna & Wosiacki (2003).
Biogeography: Arratia (1990).
Revision: Eigenmann (1918a); Tchernavin (1944, Trichomycterinae).
Phylogeny: Baskin (1973), de Pinna (1989b, Glanapteryginae); de Pinna (1992b); Costa (1994); Bockmann *et al.* (2004).
Review: Miranda Ribeiro (1946, 1951b, Brazilian Stegophilinae); Miranda Ribeiro (1947, Brazilian Vandelliinae);
Miranda Ribeiro (1949b, 1956, 1957, Brazilian Trichomycterinae).
41 genera, 207 species; no named fossil taxa (but see *Propygidium* in section below entitled “Names wrongly treated as Siluriformes”).

***ACANTHOPOMA* Lütken, 1892**

Acanthopoma Lütken, 1892: 57. Type species: *Acanthopoma annectens* Lütken, 1892. Type by monotypy. Gender: Neuter.

***Acanthopoma annectens* Lütken, 1892**

Acanthopoma annectens Lütken, 1892: 53, figures on p. 56. Type locality: Huallaga. Holotype: MTD F5245.

Distribution: Upper and middle Amazon River basin, Brazil and Peru (de Pinna & Wosiacki, 2003).

***AMMOGLANIS* Costa, 1994**

Ammoglanis Costa, 1994: 208. Type species: *Ammoglanis diaphanus* Costa, 1994. Type by original designation.
Gender: Masculine.

***Ammoglanis diaphanus* Costa, 1994**

Ammoglanis diaphanus Costa, 1994: 208, fig. 1. Type locality: Stream trib. to Rio Javaés, Rio Araguaia basin, ca. 40 km north of Araguaçú, 12°24'S, 49°58'W, Tocantin state, Brazil. Holotype: MNRJ 12442.

Distribution: Stream tributary to Javaés River, Araguaia River basin, Brazil (de Pinna & Wosiacki, 2003).

***Ammoglanis pulex* de Pinna & Winemiller, 2000**

Ammoglanis pulex de Pinna & Winemiller, 2000: 257, fig. 1. Type locality: Venezuela: Estado Amazonas: Río Paria Grande at bridge on road between Pto. Ayacucho and Samariapo (5°23'N 67°37'W). Holotype: MBUCV V-29040.

Distribution: Paria Grande River, Pamoni River and Caño Garrapata, Venezuela (de Pinna & Wosiacki, 2003).

***APOMATOCEROS* Eigenmann, 1922**

Apomatoceros Eigenmann, 1922a: 113. Type species: *Apomatoceros allenii* Eigenmann, 1922. Type by monotypy.
Gender: Masculine.

***Apomatoceros allenii* Eigenmann, 1922**

Apomatoceros allenii Eigenmann, 1922a: 113, pls. 3 (figs. 1–4), 4 (fig. 9). Type locality: Río Morona, upper Amazon system, Peru. Holotype: CAS 56174.

Distribution: Amazon River basin, Peru (de Pinna & Wosiacki, 2003).

***BULLOCKIA* Arratia, Chang, Menu-Marque & Rojas, 1978**

Bullockia Arratia, Chang, Menu-Marque & Rojas, 1978: 162, 187. Type species: *Hatcheria maldonadoi* Eigenmann, 1920. Type by monotypy. Gender: Feminine.

Bullockia maldonadoi (Eigenmann, 1920)

Hatcheria Maldonadoi Eigenmann, 1920c: 53. Type locality: lower course of the Rio Nonguen where it passes through the ground of the Agricultural School, at the outskirts of Concepción, Chile. Lectotype: CAS 63842. Illustrated and described in more detail in Eigenmann (1927: 39, pl. 8, figs. 2–2b); with the lectotype established in caption to pl. 8.

Hatcheria bullocki Fowler, 1940b: 180, figs. 14–15. Type locality: Angol, Chile. Holotype: ANSP 69145. Böhlke (1984: 163) noted that stated locality in the original description is Angol, but catalog and label say “El Vergal.” Distribution: Chile (de Pinna & Wosiacki, 2003).

Remarks: The description of *Hatcheria maldonadoi* stated that the Type (= Holotype) was illustrated. However, there was no illustration associated with that publication, so all of the examined specimens need to be treated as syntypes. The illustrated specimen in Eigenmann (1927) was labeled as the type, which is sufficient to be treated as a lectotype designation.

COPIONODON de Pinna, 1992

Copionodon de Pinna, 1992b: 181. Type species: *Copionodon pecten* de Pinna, 1992. Type by original designation. Gender: Masculine.

Copionodon lianae Campanario & de Pinna, 2000

Copionodon lianae Campanario & de Pinna, 2000: 370, figs. 1–2. Type locality: Brazil: Bahia: Rio Grisante (Rio Mucujê basin), approx. 13°08'S 41°17'W. Holotype: MZUSP 42469.

Distribution: Grisante River, tributary to Mucujê River, Paraguaçu River basin, Brazil (de Pinna & Wosiacki, 2003).

Copionodon orthiocarinatus de Pinna, 1992

Copionodon orthiocarinatus de Pinna, 1992b: 188, figs. 3–4. Type locality: Rio Mucujê, trib. of Rio Paraguaçu, Bahia, Brazil, ca. 13°00'S, 41°23'W, elev. 1200 m. Holotype: MZUSP 42463.

Distribution: Mucujê River, tributary of Paraguaçu River, Brazil (de Pinna & Wosiacki, 2003).

Copionodon pecten de Pinna, 1992

Copionodon pecten de Pinna, 1992b: 182, figs. 1–2. Type locality: Rio Mucujê, trib. of R. Paraguaçu, Bahia, Brazil, ca. 13°00'S, 41°23'W, elev. 1200 m. Holotype: MZUSP 42461.

Distribution: Mucujê River, tributary of Paraguaçu River, Brazil (de Pinna & Wosiacki, 2003).

EREMOPHILUS Humboldt, 1805

Eremophilus Humboldt, 1805a: 17. Type species: *Eremophilus mutisii* Humboldt, 1805. Type by monotypy. Gender: Masculine.

Thrichomycterus Humboldt, 1805a: 18. Type species: *Eremophilus mutisii* Humboldt, 1805. Type by monotypy. Gender: Masculine. Given as an alternative name, but never used subsequently, for *Eremophilus* Humboldt.

Trachypoma Giebel, 1871: 97. Type species: *Trachypoma marmoratum* Giebel, 1871. Type by original designation. Gender: Neuter. Preoccupied by *Trachypoma* Günther, 1859, in fishes; apparently not replaced.

Eremophilus mutisii Humboldt, 1805

Eremophilus mutisii Humboldt, 1805a: 18, pl. 6. Type locality: petite rivière de Bogota, qui forme la fameuse catarate de Tequendama, Colombia. No types known.

? *Trachypoma marmoratum* Giebel, 1871: 97. Type locality: obere Amazonenstrom. Type(s): Whereabouts unknown.

Distribution: Bogotá River basin; probably introduced to Ubaté, Chinququirá and Tundama valleys, Colombia (de Pinna & Wosiacki, 2003).

GLANAPTERYX Myers, 1927

Glanapteryx Myers, 1927: 128. Type species: *Glanapteryx anguilla* Myers, 1927. Type by original designation. Gender: Feminine.

Glanapteryx anguilla Myers, 1927

Glanapteryx anguilla Myers, 1927: 129. Type locality: Rapids of São Gabriel, Rio Negro system, Brazil. Holotype: CAS 56048.

Distribution: Negro and Orinoco River basins, Brazil and Venezuela (Nico & de Pinna, 1996; de Pinna & Wosiacki, 2003).

Remarks: Redescribed in de Pinna (1989b), with notes on distribution in Nico & de Pinna (1996).

Glanapteryx niobium de Pinna, 1998

Glanapteryx niobium de Pinna, 1998b: 36, fig. 1. Type locality: Brazil, State of Amazonas, Pico da Neblina National Park, Morro dos Seis Lagos (approx. 0°17'N 66°41'W), Lago Esperança. Holotype: INPA 12421.

Distribution: Morro dos Seis Lagos, Negro River basin, Brazil (de Pinna & Wosiacki, 2003).

GLAPHYROPOMA de Pinna, 1992

Glaphyropoma de Pinna, 1992b: 194. Type species: *Glaphyropoma rodriguesi* de Pinna, 1992. Type by original designation. Gender: Feminine.

Glaphyropoma rodriguesi de Pinna, 1992

Glaphyropoma rodriguesi, 1992b: 196, figs. 7–8. Type locality: Rio Mucujê, trib. of Rio Paraguaçu, Bahia, Brazil, ca. 13°00'S, 41°23'W, elev. 1200 m. Holotype: MZUSP 42465.

Distribution: Mucujê River, tributary of Paraguaçu River, Brazil (de Pinna & Wosiacki, 2003).

HAEMOMASTER Myers, 1927

Haemomaster Myers, 1927: 131. Type species: *Haemomaster venezuelae* Myers, 1927. Type by original designation. Gender: Masculine.

Haemomaster venezuelae Myers, 1927

Haemomaster venezuelae Myers, 1927: 131. Type locality: Venezuela, Playa Matepalma, Orinoco. Holotype: CAS 55882.

Distribution: Amazon and Orinoco River basins, Brazil and Venezuela (de Pinna & Wosiacki, 2003).

Remarks: Redescribed in Schmidt (1985).

HATCHERIA Eigenmann, 1909

Hatcheria Eigenmann, 1909b: 250. Type species: *Hatcheria patagoniensis* Eigenmann, 1909. Type by original designation. Gender: Feminine.

Revision: Arratia & Menu-Marque (1981).

Hatcheria macraei (Girard, 1855)

Thrichomycterus macraei Girard, 1855: 245. Type locality: Near Uspullata, east side of the Cordilleras, Argentina, elev. 7000 ft. Syntypes: MCZ 8298 (1), USNM 1546 (orig. 3, now 1).

Pygidium Burmeisteri Berg, 1895: 128, pl. 2 (fig. 1). Type locality: Provincia Mendoza (Río Mendoza), Argentina. Holotype: at MACN.

Hatcheria patagoniensis Eigenmann, 1909b: 250, pl. 33 (fig. 2), pl. 34 (fig. 1). Type locality: Río Blanco at base of Andes, Patagonia, Argentina, 47°30'S, 72°W. Holotype: Whereabouts unknown.

Hatcheria titcombi Eigenmann, 1918c: 692. Type locality: Rio Comajo; tributary of Lake Traful, tributary to Rio Limay, Argentina. Holotype: CAS 28557; holotype illustrated in Eigenmann (1918a: 284, pl. 44, fig. 2).

Hatcheria pique MacDonagh, 1938: 171, figs. 23–24. Type locality: Río Colorado cerca de su desembocadura, vecino a la estación Pedro Luro del F. C. S., Argentina. Holotype: MLP 15.X.36.

Distribution: Cis-Andean rivers of Argentina, between 29° and 45°30'S (de Pinna & Wosiacki, 2003).

HENONEMUS Eigenmann & Ward, 1907

Henonemus Eigenmann & Ward, in Eigenmann, McAtee & Ward, 1907: 118. Type species: *Stegophilus intermedius* Eigenmann & Eigenmann, 1889. Type by original designation. Gender: Masculine.

Cobitiglanis Fowler, 1914: 268. Type species: *Ochmacanthus taxistigma* Fowler, 1914. Type by original designation. Gender: Masculine. Proposed originally as a subgenus of *Ochmacanthus*.

Henonemus intermedius (Eigenmann & Eigenmann, 1889)

Stegophilus intermedius Eigenmann & Eigenmann, 1889b: 54. Type locality: Goyaz [Brazil]. Holotype: MCZ 9842. Distribution: Araguaia River basin, Brazil (de Pinna & Wosiacki, 2003).

Henonemus macrops (Steindachner, 1882)

Stegophilus macrops Steindachner, 1882b: 178. Type locality: Amazonenstrom. Holotype: at NMW. Species illustrated and described in more detail in Steindachner (1882c: 28, pl. 6, figs. 2, 2a).

Distribution: Amazon River basin, Brazil (de Pinna & Wosiacki, 2003).

Henonemus punctatus (Boulenger, 1887)

Stegophilus punctatus Boulenger, 1887c: 279, pl. 21 (fig. 4). Type locality: Canelos. Holotype: BMNH 1880.12.8.89.

Distribution: Amazon River basin of Brazil, Ecuador and Peru (de Pinna & Wosiacki, 2003).

Henonemus taxistigma (Fowler, 1914)

Ochmacanthus taxistigma Fowler, 1914: 268, fig. 16. Type locality: Rupununi River, British Guiana ... in the highlands of British Guiana, approximately secured in North Latitude 2° to 3°, and West Longitude 50°20'. Holotype: ANSP 39344.

Distribution: Rupununi River basin, Guyana (de Pinna & Wosiacki, 2003).

HOMODIAETUS Eigenmann & Ward, 1907

Homodiaetus Eigenmann & Ward, in Eigenmann, McAtee & Ward, 1907: 117. Type species: *Homodiaetus anisitsi* Eigenmann & Ward, 1907. Type by original designation. Gender: Masculine.

Revision: Koch (2002).

Homodiaetus anisitsi Eigenmann & Ward, 1907

Homodiaetus anisitsi Eigenmann & Ward, in Eigenmann, McAtee & Ward, 1907: 119, pl. 34 (figs. 2–3). Type locality: small creek at Villa Rica, Paraguay. Holotype: CAS 37276; holotype illustrated in Eigenmann (1918a: 351; pl. 56, fig. 3, 5).

Homodiaetus vazferreirai Devincenzi, in Devincenzi & Vaz-Ferreira, 1939: 168, figured. Type locality: río Uruguay, en las inmediaciones de la ciudad de Paysandú, Uruguay. Holotype: MHNM CI 345.

Distribution: Paraná-Paraguay River basin (Koch, 2002).

Homodiaetus banguela Koch, 2002

Homodiaetus banguela Koch, 2002: 42, fig. 4. Type locality: Brasil, Rio de Janeiro: Silva Jardim (rio São João, no cruzamento com a BR-101). Holotype: MCP 19619.

Distribution: São João River, State of Rio de Janeiro, southeastern Brazil (Koch, 2002).

Homodiaetus graciosa Koch, 2002

Homodiaetus graciosa Koch, 2002: 44, fig. 5. Type locality: Brasil, Paraná, Morretes (rio Sagrado, posto florestal). Holotype: MCP 19618.

Distribution: Coastal basins of southeastern Brazil in states of Paraná and São Paulo (Koch, 2002).

Homodiaetus passarellii (Miranda Ribeiro, 1944)

Stegophilus passarellii Miranda Ribeiro, 1944c: 1, fig. on p. 2. Type locality: Córrego do Barro Branco (conhecido atualmente como Roncador) com nascentes na Serra do Barro Branco e desaguando no canal Saracuruna—um dos formadores do rio Estréla (hoje também conhecido como Canal), e que se lança no fundo de baia de Guanabara no local denominado “Boqueirão” estrada de Magé, Estado do Rio de Janeiro [Brazil]. Holotype: MNRJ 3783.

Distribution: Coastal basins of Rio de Janeiro State, southeastern Brazil (Koch, 2002).

ITUGLANIS Costa & Bockmann, 1993

Ituglanis Costa & Bockmann, 1993: 44. Type species: *Pygidium proops parahybae* Eigenmann, 1918. Type by original designation. Gender: Masculine.

Ituglanis amazonicus (Steindachner, 1882)

Trichomycterus amazonicus Steindachner, 1882b: 178. Type locality: Cudajas [Brazil]. Holotype: NMW 43306. Species illustrated and described in more detail in Steindachner (1882c: 29, pl. 6, figs. 4, 4a).

Distribution: Amazon River basin, Brazil, and rivers of French Guiana (de Pinna & Wosiacki, 2003).

Ituglanis bambui Bichuette & Trajano, 2004

Ituglanis bambui Bichuette & Trajano, 2004: 245, fig. 1. Type locality: Brazil: Goiás: São Domingos: Terra Ronca

State Park: Angélica Cave: upper tributary of main subterranean stream, formed by infiltration of epikarstic water, 13°31'W, 46°23'W. Holotype: MZUSP 79860.

Distribution: São Domingos karst area, Tocantins River basin, Brazil (Bichuette & Trajano, 2004).

***Ituglanis eichorniarum* (Miranda Ribeiro, 1912)**

Trichomycterus eichorniarum Miranda Ribeiro, 1912: 27. Type locality: Cáceres, nas margens do Paraguay (M. Grosso) [Brazil]. Lectotype: MNRJ 780A, designated by Miranda Ribeiro (1953: 405).

Distribution: Upper Paraguay River basin, Brazil (de Pinna & Wosiacki, 2003).

***Ituglanis epikarsticus* Bichuette & Trajano, 2004**

Ituglanis epikarsticus Bichuette & Trajano, 2004: 248, fig. 4. Type locality: Brazil: Goiás: São Domingos: Terra Ronca State Park: São Mateus Cave: rimstone dams (travertine basins) fed by infiltration of epikarstic water (13°40'S, 46°22'W). Holotype: MZUSP 79869.

Distribution: São Domingos karst area, Tocantins River basin, Brazil (Bichuette & Trajano, 2004).

***Ituglanis gracilior* (Eigenmann, 1912)**

Pygidium gracilior Eigenmann, 1912b: 213. Type locality: Erukin, British Guiana. Holotype: FMNH 53264; holotype illustrated in Eigenmann (1918a: 326; pl. 50, fig. 3).

Distribution: Guyana (de Pinna & Wosiacki, 2003).

***Ituglanis guayaberensis* (Dahl, 1960)**

Pygidium metae guayaberensis Dahl, 1960b: 307, fig. 2. Type locality: Small pond in a brook trib. to Río Guayabero, ca. 1 km from mouth of the brook In a brook approx. 1500 m. from Camp 1, same locality as the holotype of *Leyvaichthys castaneus*, Orinoco System, Colombia. Holotype: Whereabouts unknown (possibly at ICN-MHN).

Distribution: Guayabero River basin, Orinoco drainage, Colombia (de Pinna & Wosiacki, 2003).

***Ituglanis herberti* (Miranda Ribeiro, 1940)**

Trichomycterus Herberti Miranda Ribeiro, 1940: 60, fig. Type locality: Rio Bodoquena, Pantanal, sur de Mato Grosso [Brazil]. Syntypes (7): MNRJ 1428 (2), MNRJ 1429 (5).

Distribution: Bodoquena River, Paraguay River basin, Brazil (de Pinna & Wosiacki, 2003).

***Ituglanis laticeps* (Kner, 1863)**

Trichomycterus laticeps Kner, 1863: 228, fig. 17. Type locality: Fundort, wie die vorige Art [in reference to *Trichomycterus taenia*, from "Westabhang der Andes im Staate Ecuador"]. Also described in Kner & Steindachner (1864: 54, pl. 6, fig. 2).

Distribution: Ecuador (de Pinna & Wosiacki, 2003).

***Ituglanis macunaima* Datovo & Landim, 2005**

Ituglanis macunaima Datovo & Landim, 2005: 456, fig. 1. Type locality: Brazil, Mato Grosso, Cocalinho; rio Araguaia basin, Corixo da Saudade (Corixinho); 14°17'20.6"S 51°9'12.1"W. Holotype: MZUSP 88452.

Distribution: Araguaia River basin, Brazil (Datovo & Landim, 2005).

***Ituglanis metae* (Eigenmann, 1918)**

Pygidium metae Eigenmann, 1918c: 694. Type locality: Barrigona, Colombia. Holotype: CAS 58138; holotype illustrated in Eigenmann (1918a: 312, pl. 47, fig. 5).

Distribution: Colombia (de Pinna & Wosiacki, 2003).

***Ituglanis nebulosus* de Pinna & Keith, 2003**

Ituglanis nebulosus de Pinna & Keith, 2003: 874, figs. 1, 2. Type locality: French Guiana, Arataye river (Tributary of Approuague river), near the natural preserve "Reserve naturelle des Nouragues" (04°00'22"N, 52°36'34"W). Holotype: MNHN 2001-1128.

Distribution: Approuague River basin, French Guiana (de Pinna & Keith, 2003).

***Ituglanis parahybae* (Eigenmann, 1918)**

Pygidium proops parahybae Eigenmann, 1918a: 332. Type locality: Rio Parahyba, Brazil. Holotype: FMNH 58576. Distribution: Paraíba do Sul and São João River basins, Brazil (de Pinna & Wosiacki, 2003).

***Ituglanis parkoi* (Miranda Ribeiro, 1944)**

Pygidium parkoi Miranda Ribeiro, 1944b: 1, fig. Type locality: rio Iticoá (entre os rios das Pedras e Ituí) que deságua no Javari, afluente do Amazonas, Benjamin Constant, Estados do Amazonas [Brazil]. Holotype: MNRJ

Distribution: Amazon River basin, Brazil (de Pinna & Wosiacki, 2003).

Ituglanis passensis Fernández & Bichuette, 2002

Ituglanis passensis Fernández & Bichuette, 2002: 274, fig. 1. Type locality: Brazil: Goiás: São Domingos: Passa Três cave: subterranean stream in the Tocantins drainage, 13°36'S 46°23'W. Holotype: MCP 27382.

Distribution: Passa Três cave system in São Domingos, Goiás, Brazil (de Pinna & Wosiacki, 2003).

Ituglanis proops (Miranda Ribeiro, 1908)

Trichomycterus proops Miranda Ribeiro, 1908b: [3], fig. 4. Type locality: Rio Ribeira, Iguape, [Brazil]. Lectotype: MNRJ 781A, designated by Miranda Ribeiro (1953: 405).

Distribution: Ribeira de Iguape River basin, Brazil (de Pinna & Wosiacki, 2003).

Ituglanis ramiroi Bichuette & Trajano, 2004

Ituglanis ramiroi Bichuette & Trajano, 2004: 250, fig. 7. Type locality: Brazil: Goiás: São Domingos: Terra Ronca State Park: São Bernardo Cave: side pool fed by small water inlet (13°49'S, 46°21'W), for about 700 m from cave entrance. Holotype: MZUSP 79865.

Distribution: São Domingos karst area, Tocantins River basin, Brazil (Bichuette & Trajano, 2004).

LISTRURA de Pinna, 1988

Listrura de Pinna, 1988: 114. Type species: *Listrura nematopteryx* de Pinna, 1988. Type by original designation.

Gender: Feminine.

Listrura boticario de Pinna & Wosiacki, 2002

Listrura boticario de Pinna & Wosiacki, 2002: 721, fig. 1. Type locality: Brazil, State of Paraná, Município de Guaraqueçaba, pool adjacent to Rio da Figueira (tributary of Rio Morato, itself tributary to Rio Guaraqueçaba, an isolated coastal basin that drains directly into the Baía das Laranjeiras), inside the nature preserve “Reserva Particular do Patrimônio Natural Salto Morato” (25°16'S, 48°12'W, UTM: 7.212.500–7.215.400). Holotype: MZUSP 69573.

Distribution: Da Figueira and Guaraqueçaba River basins, Paraná State, Brazil (de Pinna & Wosiacki, 2003).

Listrura camposi (Miranda Ribeiro, 1957)

Eremophilus camposi Miranda Ribeiro, 1957: 72, fig. Type locality: Rio Poço Grande, [trib. of the Rio Juquiá, Fazenda Poço Grande, Juquiá, ca. 24°15'S, 47°37'W, São Paulo State, Brazil]. Holotype: MZUSP 3426.

Distribution: Poço Grande River, tributary of the Juquiá River, São Paulo State, and Ribeirão da Ilha, Florianópolis, Santa Catarina State, Brazil (de Pinna & Wosiacki, 2003).

Listrura nematopteryx de Pinna, 1988

Listrura nematopteryx de Pinna, 1988: 115, figs. 1–2. Type locality: Small marsh which is source of creek later joining Ribeirão Imbariê, trib. of Rio Estrela, near 58 km mark of old road leading to Petrópolis (Antiga Rio-Petrópolis, also called “Estrada Automóvel Club”), Município de Magé, Localidade de Piabetá, Rio de Janeiro, Brazil (22°36'36"S 43°11'26"W). Holotype: MZUSP 36974.

Distribution: Imbariê Creek, basin, Estrela River system, Piabetá, Rio de Janeiro State, and Picinguaba, São Paulo State, Brazil (de Pinna & Wosiacki, 2003).

Listrura tetraradiata Landim & Costa, 2002

Listrura tetraradiata Landim & Costa, 2002: 152, fig. 1. Type locality: Brazil: Estado do Rio de Janeiro: Município de Saquarema, rio da Represa, a tributary of rio Bom Sucesso, rio Ibicuíba basin of the Lagoa de Araruama system, Palmital, approximately 5 km north from Bacaxá, about 22°50'S, 42°28'W. Holotype: MZUSP 52572.

Distribution: Ibicuíba River, Araruama Lagoon system, Brazil (de Pinna & Wosiacki, 2003).

MALACOGLANIS Myers & Weitzman, 1966

Malacoglanis Myers & Weitzman, 1966: 281. Type species: *Malacoglanis gelatinosus* Myers & Weitzman, 1966. Type by original designation. Gender: Masculine.

Malacoglanis gelatinosus Myers & Weitzman, 1966

Malacoglanis gelatinosus Myers & Weitzman, 1966: 282, figs. 3–4. Type locality: Small caño of the Río Orteguaza, a short distance above its junction with the Río Caquetá, Caquetá Province, Colombia. Holotype: SU 50754.

Distribution: Caquetá River basin, Colombia (de Pinna & Wosiacki, 2003).

MEGALOCENTOR de Pinna & Britski, 1991

Megalocentor de Pinna & Britski, 1991: 115. Type species: *Megalocentor echthrus* de Pinna & Britski, 1991. Type by original designation. Gender: Masculine.

Megalocentor echthrus de Pinna & Britski, 1991

Megalocentor echthrus de Pinna & Britski, 1991: 116, figs. 3, 4 (a–b). Type locality: Rio Madeira, Calama, Praia do Caraparu, Estado do Amazonas, Brazil. Holotype: MZUSP 41879.

Distribution: Amazon and Orinoco River basins (de Pinna & Wosiacki, 2003).

MICROCAMBEVA Costa & Bockmann, 1994

Microcambeva Costa & Bockmann, 1994a: 718. Type species: *Microcambeva barbata* Costa & Bockmann, 1994. Type by original designation. Gender: Feminine.

Microcambeva barbata Costa & Bockmann, 1994

Microcambeva barbata Costa & Bockmann, 1994a: 718, figs. 2–3. Type locality: Brazil: Estado do Rio de Janeiro: rio São João, near Gaviões, Município de Silva Jardim. Holotype: MZUSP 43678.

Distribution: Atlantic coastal drainages of Rio de Janeiro and Espírito Santo states, Brazil (de Pinna & Wosiacki, 2003).

Microcambeva ribeirae Costa, Lima & Bizerril, 2004

Microcambeva ribeirae Costa, Lima & Bizerril, 2004: 3, figs. 1, 2. Type locality: Brazil: Estado de São Paulo: Município de Pedro de Toledo, Rio São Lourençinho, no Bairro São Lourenço. Holotype: MZUSP 84301.

Distribution: Ribeira do Iguape River basin, SE Brazil (Costa *et al.*, 2004).

MIUROGLANIS Eigenmann & Eigenmann, 1889

Miuroglanis Eigenmann & Eigenmann, 1889b: 55. Type species: *Miuroglanis platycephalus* Eigenmann & Eigenmann, 1889. Type by original designation. Gender: Masculine.

Miuroglanis platycephalus Eigenmann & Eigenmann, 1889

Miuroglanis platycephalus Eigenmann & Eigenmann, 1889b: 56. Type locality: Jutahy [Brazil]. Holotype: MCZ 8172.

Distribution: upper Amazon River basin, Brazil (de Pinna & Wosiacki, 2003).

OCHMACANTHUS Eigenmann, 1912

Ochmacanthus Eigenmann, 1912: 213. Type species: *Ochmacanthus flabelliferus* Eigenmann, 1912. Type by original designation. Gender: Masculine.

Gyrinurus Miranda Ribeiro, 1912: 27. Type species: *Gyrinurus batrachostoma* Miranda Ribeiro, 1912. Type by monotypy. Gender: Masculine.

Ochmacanthus alternus Myers, 1927

Ochmacanthus alternus Myers, 1927: 129. Type locality: Venezuela: Caño de Quiribana, near Caicara. Lectotype: CAS 13522.

Distribution: Negro and Orinoco River basins, Brazil and Venezuela (de Pinna & Wosiacki, 2003).

Ochmacanthus batrachostoma (Miranda Ribeiro, 1912)

Gyrinurus batrachostoma Miranda Ribeiro, 1912: 28, pl. Type locality: rio Paraguay em S. Luiz de Cáceres [Mato Grosso, Brazil]. Holotype: MNRJ 786.

Distribution: Paraguay River basin, Brazil (de Pinna & Wosiacki, 2003); la Plata basin, Argentina (Casciotta & Almirón, 1996).

Ochmacanthus flabelliferus Eigenmann, 1912

Ochmacanthus flabelliferus Eigenmann, 1912b: 213. Type locality: Konawaruk, British Guiana. Holotype: FMNH 53263; holotype illustrated in Eigenmann (1918a: 357; pl. 55, fig. 5).

Distribution: River drainages in Guyana and Venezuela (de Pinna & Wosiacki, 2003).

Ochmacanthus orinoco Myers, 1927

Ochmacanthus orinoco Myers, 1927: 130. Type locality: Venezuela: Playa Matepalma, Orinoco. Holotype: CAS 76355.

Distribution: Negro and Orinoco River basins, Brazil and Venezuela (de Pinna & Wosiacki, 2003).

***Ochmacanthus reinhardtii* (Steindachner, 1882)**

Stegophilus Reinhardtii Steindachner, 1882b: 178. Type locality: Amazonenstrom und Rio Iça. Syntypes: NMW 44735 (2), NMW 50603 (3). Species illustrated and described in more detail in Steindachner (1882c: 28, pl. 6, fig. 1).

Distribution: Amazon River basin, Brazil, and drainages in French Guyana (de Pinna & Wosiacki, 2003).

***PARACANTHOPOMA* Giltay, 1935**

Paracanthopoma Giltay, 1935: 1. Type species: *Paracanthopoma parva* Giltay, 1935. Type by original designation.

Gender: Neuter.

Phylogeny: Schmidt (1993).

***Paracanthopoma parva* Giltay, 1935**

Paracanthopoma parva Giltay, 1935: 1, figs. 1–3. Type locality: Rio Catrymany supérieur [Brazil]. Holotype: IRSNB 43.

Distribution: Amazon basin, Brazil, and Essequibo River basin, Guyana (de Pinna & Wosiacki, 2003).

Remarks: Redescribed, with notes on biology, in Schmidt (1993).

***PARASTEGOPHILUS* Miranda Ribeiro, 1946**

Parastegophilus Miranda Ribeiro, 1946: 12. Type species: *Stegophilus maculatus* Steindachner, 1879. Type by original designation. Gender: Masculine.

***Parastegophilus maculatus* (Steindachner, 1879)**

Stegophilus maculatus Steindachner, 1879a: 32. Type locality: La Plata, Prov. of Buenos Aires, Argentina. Holotype: NMW 57222. Species illustrated and described in more detail in Steindachner (1879e: 25, pl. 4, fig. 2).

Distribution: Lower Paraná and Uruguay River basins, Argentina (de Pinna & Wosiacki, 2003).

***Parastegophilus paulensis* (Miranda Ribeiro, 1918)**

Pseudostegophilus paulensis Miranda Ribeiro, 1918e: 727. Type locality: Avanhandava, Rio Tieté, Estado de São Paulo [Brazil]. Holotype: MZUSP 2272.

Pseudostegophilus scarificator Ihering, 1930: 100, pl. 13 (fig. 2). Type locality: São Paulo [Brazil]. Syntypes (2): Whereabouts unknown.

Distribution: Upper Paraná River basin, Brazil (de Pinna & Wosiacki, 2003).

***PARAVANDELLIA* Miranda Ribeiro, 1912**

Paravandellia Miranda Ribeiro, 1912: 28. Type species: *Paravandellia oxyptera* Miranda Ribeiro, 1912. Type by monotypy. Gender: Feminine.

Branchioica Eigenmann, 1918c: 702. Type species: *Branchioica bertoni* Eigenmann, 1918. Type by original designation. Gender: Feminine.

Pleurophysus Miranda Ribeiro, 1918d: 636. Type species: *Pleurophysus hydrostaticus* Miranda Ribeiro, 1918. Type by monotypy. Gender: Masculine.

Parabranchioica Devincenzi & Vaz-Ferreira, 1939: 5. Type species: *Parabranchioica teaguei* Devincenzi & Vaz-Ferreira, 1939. Type by monotypy. Gender: Feminine.

***Paravandellia oxyptera* Miranda Ribeiro, 1912**

Paravandellia oxyptera Miranda Ribeiro, 1912: 29. Type locality: Caceres, margens do Rio Paraguay [Brazil]. Holotype: MNRJ 790.

Branchioica bertoni Eigenmann, 1918c: 703. Type locality: Asuncion, Paraguay. Holotype: CAS 63840; holotype illustrated in Eigenmann (1918a: 368, pl. 53, figs. 3–5) as *Branchioica bertonii*.

Pleurophysus hydrostaticus Miranda Ribeiro, 1918d: 636. Type locality: Rio Claro [Brazil]. Lectotype: MZUSP 2214, designated by Britski (1969: 206).

Vandellia hematophaga Guimarães, 1935: 300, figs. 1–4. Type locality: Río Tietê, Salto, São Paulo [Brazil]. Lecto-

type: SU 16766, designated by Böhlke (1953: 45).

Parabranchioica teaguei Devincenzi & Vaz-Ferreira, 1939: 5, unnumbered figure. Type locality: río Uruguay, en las inmediaciones de la ciudad de Paysandú [Uruguay]. Type(s): at MHNM.

Distribution: Paraná, Paraguay and Uruguay River basins, Brazil, Paraguay, and Uruguay (de Pinna & Wosiacki, 2003).

***Paravandellia phaneronema* (Miles, 1943)**

Branchioica phaneronema Miles, 1943a: 32, fig. 8. Type locality: various points in the Upper Cauca valley. Holotype: lost (Román-Valencia, 1998). Described again in Miles (1943b: 367, figs. 1–3).

Branchioica magdalena Miles, 1943a: 33. Type locality: Río Magdalena, Colombia. Holotype: lost (Román-Valencia, 1998). Described again in Miles (1943b: 368).

Distribution: Magdalena and Cauca River basins, Colombia (de Pinna & Wosiacki, 2003).

Remarks: Although both of these nominal species were described twice in 1943, the text of Miles (1943b) indicates that the names were first made available in Miles (1943a). Román-Valencia (1998) treated *Branchioica magdalena* as a junior synonym of *Paravandellia phaneronema*, thereby acting as first reviser.

***PAREIODON* Kner, 1855**

Pareiodon Kner, 1855a: 160. Type species: *Pareiodon microps* Kner, 1855. Type by monotypy. Gender: Masculine.

Centrophorus Kner, 1859: 167 (footnote). Type species: *Pareiodon microps* Kner, 1855. Type by being a replacement name. Gender: Masculine. Unneeded replacement for *Pareiodon* Kner, 1855, then considered preoccupied by *Parodon*. Preoccupied by *Centrophorus* Müller & Henle, 1837, in Elasmobranchs.

Astomomycterus Guichenot, 1860: 525. Type species: *Trichomycterus pusillus* Castelnau, 1855. Type by original designation. Gender: Masculine.

***Pareiodon microps* Kner, 1855**

Pareiodon microps Kner, 1855a: 160, fig. 2. Type locality: Borba (?) [Río Madeira]. Syntypes: NMW 45486 (2).

Trichomycterus pusillus Castelnau, 1855: 50, pl. 24 (fig. 4). Type locality: l’Araguay et [...] l’Amazone. Syntypes: MNHN 0000-1210 (4).

Distribution: Amazon River basin, Brazil (de Pinna & Wosiacki, 2003).

Remarks: Priority of publication of the two included synonyms may not have yet been established, inasmuch as the precise dates of publication of the two works were not listed in Eschmeyer *et al.* (1998) or other relevant publications. If priority cannot be used to resolve this issue, precedence must be given to *Pareiodon microps* by first reviser action of Eigenmann & Eigenmann (1889: 346).

***PLECTROCHILUS* Miranda Ribeiro, 1917**

Plectrochilus Miranda Ribeiro, 1917: 50. Type species: *Plectrochilus machadoi* Miranda Ribeiro, 1917. Type by monotypy. Gender: Masculine.

***Plectrochilus diabolicus* (Myers, 1927)**

Urinophilus diabolicus Myers, 1927: 132. Type locality: Iquitos, Peru. Holotype: CAS 59940.

Distribution: Amazon River basin, Brazil and Peru (de Pinna & Wosiacki, 2003).

***Plectrochilus machadoi* Miranda Ribeiro, 1917**

Plectrochilus machadoi Miranda Ribeiro, 1917: 50, three unnumbered figures. Type locality: Fluvio Solimões, [Brazil]. Holotype: MNRJ 978.

Vandellia hasemani Eigenmann, 1918a: 363, figs. 33–35, pl. 53 (fig. 3). Type locality: Río Mamoré, Alto Amazonas, Bolivia. Holotype: FMNH 58523.

Distribution: Amazon River basin, Brazil and Peru (de Pinna & Wosiacki, 2003).

***Plectrochilus wieneri* (Pellegrin, 1909)**

Vandellia wieneri Pellegrin, 1909e: 199, unnumbered figure. Type locality: [Río Napo, near mouth of Río Misahualli, Ecuador]. Holotype: MNHN a-9934.

Distribution: Napo River basin, Ecuador (de Pinna & Wosiacki, 2003).

***PSEUDOSTEGOPHILUS* Eigenmann & Eigenmann, 1889**

Pseudostegophilus Eigenmann & Eigenmann, 1889b: 54. Type species: *Stegophilus nemurus* Günther, 1869. Type by original designation. Gender: Masculine.

***Pseudostegophilus haemomyzon* (Myers, 1942)**

Homodiaetus haemomyzon Myers, 1942: 98, fig. 4. Type locality: Río Guarico at Calabozo, Venezuela. Holotype: SU 36500.

Distribution: Orinoco River basin, Venezuela (de Pinna & Wosiacki, 2003).

***Pseudostegophilus nemurus* (Günther, 1869)**

Stegophilus nemurus Günther, 1869: 429. Type locality: Upper Amazon River, Peru. Holotype: BMNH 1869.5.21.9.

Distribution: Amazon River basin, Brazil and Peru (de Pinna & Wosiacki, 2003).

***PYGIDIANOPS* Myers, 1944**

Pygidianops Myers, 1944: 592. Type species: *Pygidianops eigenmanni* Myers, 1944. Type by original designation. Gender: Masculine.

Key: Schaefer *et al.* (2005: 5).

***Pygidianops cuao* Schaefer, Provenzano, de Pinna & Baskin, 2005**

Pygidianops cuao Schaefer, Provenzano, de Pinna & Baskin, 2005: 5, figs. 1, 2. Type locality: Venezuela, Estado Amazonas, Río Cuao at Raudal Guacamaya, 8.1 miles upstream from Raudal El Danto, 05°07.71'N, 67°31.53'W. Holotype: MBUCV V-30917.

Distribution: Cuao River, Orinoco basin, Venezuela (Schaefer *et al.*, 2005).

***Pygidianops eigenmanni* Myers, 1944**

Pygidianops eigenmanni Myers, 1944: 592, pl. 52 (fig. 1), pl. 53 (figs. 3–5). Type locality: Rock pools below São Gabriel Rapids, Rio Negro, Brazil. Holotype: CAS 11120.

Distribution: Negro River basin, Brazil (de Pinna & Wosiacki, 2003).

***Pygidianops magoi* Schaefer, Provenzano, de Pinna & Baskin, 2005**

Pygidianops magoi Schaefer, Provenzano, de Pinna & Baskin, 2005: 10, figs. 5, 6. Type locality: Venezuela, Estado Delta Amacuro, Rio Orinoco at Puerto Cabrian, 8°34.8'N, 62°15.9'W. Holotype: MBUCV V-31035.

Distribution: Lower Orinoco River, between Ciudad Bolívar and Barrancas, Venezuela (Schaefer *et al.*, 2005).

***RHIZOSOMICHTHYS* Miles, 1943**

Bathophilus Miles, 1942: 57. Type species: *Pygidium totae* Miles, 1942. Type by monotypy. Gender: Masculine.

Preoccupied by *Bathophilus* Giglioli, 1882, in fishes; replaced by *Rhizosomichthys* Miles, 1943, and *Bathypygidium* Whitley, 1947.

Rhizosomichthys Miles, 1943b: 369. Type species: *Pygidium totae* Miles, 1942. Type by being a replacement name.

Gender: Masculine. Replacement for *Bathophilus* Miles, 1942; preoccupied by *Bathophilus* Giglioli, 1882.

Bathypygidium Whitley, 1947: 150. Type species: *Pygidium totae* Miles, 1942. Type by being a replacement name.

Gender: Neuter. Replacement for *Bathophilus* Miles, 1942; preoccupied by *Bathophilus* Giglioli, 1882.

***Rhizosomichthys totae* (Miles, 1942)**

Pygidium totae Miles, 1942: 55, unnumbered figure. Type locality: Lago de Tota, Boyacá, Cordillera Oriental, Colombia, 3060 metros. Holotype: ICNMNH 20 [? now ICNMHN 353].

Distribution: Lake Tota basin, Colombia (de Pinna & Wosiacki, 2003).

***SARCOGLANIS* Myers & Weitzman, 1966**

Sarcoglanis Myers & Weitzman, 1966: 279. Type species: *Sarcoglanis simplex* Myers & Weitzman, 1966. Type by original designation. Gender: Masculine.

***Sarcoglanis simplex* Myers & Weitzman, 1966**

Sarcoglanis simplex Myers & Weitzman, 1966: 279, figs. 1–2. Type locality: Rock pools below São Gabriel Rapids, of the Rio Negro (below the town of Uaupés, formerly São Gabriel), Estado de Amazonas, Brazil. Holotype: SU 50189.

Distribution: Upper Negro River basin, Brazil (de Pinna & Wosiacki, 2003).

SCHULTZICHTHYS Myers & Weitzman, 1966

Schultzichthys Dahl, 1960b: 312. Type species: *Schultzichthys gracilis* Dahl, 1960. Type by monotypy. Gender: Masculine.

Schultzichthys bondi (Myers, 1942)

Acanthopoma bondi Myers, 1942: 97, fig. 5. Type locality: Río Apuré at San Fernando de Apuré, Venezuela. Holotype: SU 36498.

Distribution: Amazon and Orinoco River basins (de Pinna & Wosiacki, 2003).

Schultzichthys gracilis Dahl, 1960

Schultzichthys gracilis Dahl, 1960b: 312, figs. 4–5. Type locality: foot of a rapid in Caño Lozada, about 15 km above its junction with the Guayabero River [Río Orinoco system, Colombia]. Holotype: Whereabouts unknown (possibly at ICNMHN).

Distribution: Guayabero River, Orinoco River basin, Colombia (de Pinna & Wosiacki, 2003).

SCLERONEMA Eigenmann, 1918

Scleronema Eigenmann, 1918c: 691. Type species: *Scleronema operculatum* Eigenmann, 1918. Type by original designation. Gender: Neuter.

Scleronema angustirostre (Devincenzi, 1942)

Pygidium angustirostris Devincenzi, in Devincenzi & Teague, 1942: 30, pl. 4 (fig. 3). Type locality: La Cañeda de las Piedras, Uruguay. Holotype: at MHNM (not found).

Distribution: Uruguay (de Pinna & Wosiacki, 2003).

Scleronema minutum (Boulenger, 1891)

Trichomycterus minutus Boulenger, 1891: 235, pl. 26 (fig. 3). Type locality: San Lorenzo district [= São Lourenço das Missões], Brazil. Syntypes: BMNH 1891.3.16.84–86 (3).

Distribution: Rio Grande do Sul State, Brazil (de Pinna & Wosiacki, 2003); la Plata basin, Argentina (Casciotta & Almirón, 1996).

Scleronema operculatum Eigenmann, 1918

Scleronema operculatum Eigenmann, 1918c: 691. Type locality: Cacequy, Brazil. Holotype: FMNH 58080; holotype illustrated in Eigenmann (1918a: 281, pl. 44, fig. 1).

Distribution: Rio Grande do Sul State, Brazil (de Pinna & Wosiacki, 2003).

SILVINICHTHYS Arratia, 1998

Silvinichthys Arratia, 1998: 356. Type species: *Trichomycterus mendozensis* Arratia, Chang, Menu-Marque & Rojas, 1978. Type by monotypy. Gender: Masculine.

Silvinichthys bortayro Fernández & de Pinna, 2005

Silvinichthys bortayro Fernández & de Pinna, 2005: 101, figs. 1–2. Type locality: Argentina, Provincia de Salta, Departamento Capital, artificial well near Río Arenales at San Luis, 1273 m elevation, 12 m depth, 24°50'S, 65°30'W. Holotype: FML 2590.

Distribution: Artificial wells in Salta Province, Argentina (Fernández & de Pinna, 2005).

Silvinichthys mendozensis (Arratia, Chang, Menu-Marque & Rojas, 1978)

Trichomycterus mendozensis Arratia, Chang, Menu-Marque & Rojas, 1978: 170, figs. 8–10. Type locality: freshwater of the preandean range of Mendoza, Argentina ..., where it occupies a restricted environment between 1,500 to 1,700 m above sea level. Holotype: FFSUC IC 118-27.

Distribution: Mendoza River basin, Argentina (de Pinna & Wosiacki, 2003).

STAUROGLANIS de Pinna, 1989

Stauroglanis de Pinna, 1989a: 6. Type species: *Stauroglanis gouldingi* de Pinna, 1989. Type by original designation. Gender: Masculine.

Stauroglanis gouldingi de Pinna, 1989

Stauroglanis gouldingi de Pinna, 1989a: 7, figs. 1–3. Type locality: Cachoeira do Aracu, R. Daraá (R. Negro drainage system), Estado do Amazonas, Brazil. Holotype: MZUSP 31088.

Distribution: Daraá River, Negro River basin, Brazil (de Pinna & Wosiacki, 2003).

STEGOPHILUS Reinhardt, 1859

Stegophilus Reinhardt, 1859: 5 Type species: *Stegophilus insidiosus* Reinhardt, 1859. Type by monotypy. Gender: Masculine.

Stegophilus insidiosus Reinhardt, 1859

Stegophilus insidiosus Reinhardt, 1859: 5, pl. 2. Type locality: Rio das Velhas, Rio São Francisco system, Brazil.

Syntypes: BMNH 1875.5.22.1 (1), ZMUC P 30152 (1), ZMUC P 30153 (1).

Distribution: São Francisco River basin, Brazil (de Pinna & Wosiacki, 2003).

Stegophilus panzeri (Ahl, 1931)

Henonemus panzeri Ahl, 1931: 210, fig. 3. Type locality: Rio Capim [Brazil]. Holotype: at ZMB (orig. no. 656).

Distribution: Lower Amazon River basin, Brazil (de Pinna & Wosiacki, 2003).

Stegophilus septentrionalis Myers, 1927

Stegophilus septentrionalis Myers, 1927: 130. Type locality: Venezuela: Santa Barbara, Orinoco. Holotype: CAS 64597.

Distribution: Orinoco River basin, Venezuela (de Pinna & Wosiacki, 2003).

STENOLICMUS de Pinna & Starnes, 1990

Stenolicmus de Pinna & Starnes, 1990: 77. Type species: *Stenolicmus sarmientoi* de Pinna & Starnes, 1990. Type by original designation. Gender: Masculine.

Stenolicmus sarmientoi de Pinna & Starnes, 1990

Stenolicmus sarmientoi de Pinna & Starnes, 1990: 77, fig. 1–2. Type locality: Río Matos, trib. of Río Apere, 48 km east of San Borja, Ballivia Prov., Depto. Beni, Bolivia, 14°55'S, 66°17'W. Holotype: USNM 301664.

Distribution: Upper Apere River basin, Bolivia (de Pinna & Wosiacki, 2003).

TRICHOCENES Britski & Ortega, 1983

Trichogenes Britski & Ortega, 1983: 211. Type species: *Trichogenes longipinnis* Britski & Ortega, 1983. Type by original designation. Gender: Masculine.

Trichogenes longipinnis Britski & Ortega, 1983

Trichogenes longipinnis Britski & Ortega, 1983: 212, figs. 1–2. Type locality: Cachoeira do Amor, km 3 da estrada Parati-Ubatuba, SP [Brazil] Holotype: MZUSP 16099.

Distribution: Atlantic coastal drainages, northern São Paulo State, Brazil (de Pinna & Wosiacki, 2003).

TRICHOMYCTERUS Valenciennes, 1832

Trichomycterus Valenciennes, 1832a: 348. Type species: *Trichomycterus nigricans* Valenciennes, 1832. Type by monotypy. Gender: Masculine.

Review: Fernández (1999, Argentina).

Trichomycterus albinotatus Costa, 1992

Trichomycterus albinotatus Costa, 1992: 102, figs. 1–2. Type locality: Estado do Rio de Janeiro: Visconde de Maua, 1 km O de la Ville de Maromba, rio Preto, bassin du rio Paraíba [22°30'S 44°15'], Brazil. Holotype: MZUSP 42312.

Distribution: Preto River, Paraíba do Sul River basin, Brazil (de Pinna & Wosiacki, 2003).

Trichomycterus alternatus (Eigenmann, 1918)

Pygidium alternatum Eigenmann, 1918c: 700. Type locality: Rio Doce, Brazil. Holotype: FMNH 58082; holotype illustrated in Eigenmann (1918a: 336, pl. 52, fig. 3).

Pygidium florense Miranda Ribeiro, 1943: 1, fig. Type locality: rio das Flores, próximo a Ipiabas (estaçao de Pandiá Calógeras), Estado do Rio de Janeiro [Brazil]. Holotype: MNRJ 3751.

Pygidium travassosi Miranda Ribeiro, 1949a: 145, fig. 2. Type locality: Rio das Pedras, Fazenda Penedo, Agulhas Negras, Estado do Rio, Rio de Janeiro, Brazil. Holotype: MNRJ 5424.

Distribution: Atlantic coastal river basins, Rio de Janeiro and Espírito Santo States, Brazil (de Pinna & Wosiacki,

2003), Jequitinhonha River basin, Minas Gerais (Triques & Vono, 2004).

***Trichomycterus alterus* (Marini, Nichols & La Monte, 1933)**

Pygidium alterum Marini, Nichols & La Monte, 1933: 2, fig. 2. Type locality: Rio de los Sauces, La Rioja, Argentina. Holotype: AMNH 12241.

Distribution: Humahuaca, Los Sauces River and Valle Guanchin (La Rioja), Argentina (de Pinna & Wosiacki, 2003).

***Trichomycterus areolatus* Valenciennes, 1846**

Trichomycterus areolatus Valenciennes, in Cuvier & Valenciennes, 1846: 492. Type locality: la rivière de San Jago, Santiago. Syntypes: MNHN 0000-3167 (3), MNHN a-6310 (2).

Trichomycterus maculatus Valenciennes, in Cuvier & Valenciennes, 1846: 493. Type locality: Santiago du Chili. Syntypes: MNHN 0000-3166 (1), MNHN 0000-4236 (10).

Trichomycterus marmoratus Philippi, 1866: 714. Type locality: Chile. No types known.

Trichomycterus palleus Philippi, 1866: 715. Type locality: Chile. No types known.

Trichomycterus tigrinus Philippi, 1866: 714. Type locality: Chile. No types known.

Distribution: Pacific draining rivers of Central Chile (de Pinna & Wosiacki, 2003).

***Trichomycterus arleoi* (Fernández-Yépez, 1972)**

Pygidium arleoi Fernández-Yépez, 1972a: 21, pl. 34. Type locality: Estación -140 — de la Cuenca del Rio Yaracuy, Venezuela. Holotype: collection of author, whereabouts unknown.

Distribution: Yaracuy River basin, Venezuela (de Pinna & Wosiacki, 2003).

***Trichomycterus auroguttatus* Costa, 1992**

Trichomycterus auroguttatus Costa, 1992: 105, fig. 6. Type locality: Rio de Janeiro Estado do Rio de Janeiro: 1 km à l'ouest de la ville de Visconde de Mauá, rio Marimbondo, affluent du rio Preto, bassin du rio Paraíba, Brazil. Holotype: MZUSP 43341.

Distribution: Marimbondo River, Preto River system, Paraíba do Sul River basin, Brazil (de Pinna & Wosiacki, 2003).

***Trichomycterus bahianus* Costa, 1992**

Trichomycterus bahianus Costa, 1992: 105, fig. 5. Type locality: Estado da Bahia: Una, 6,5 km au sud-est de la ville de São José, dans um ruisseau affluent du ribeirão Caveira, bassin du rio Una, Bahia, Brazil. Holotype: MZUSP 43340.

Distribution: Ribeirão Caveira tributary, Una River basin, State of Bahia, Brazil (de Pinna & Wosiacki, 2003).

***Trichomycterus bananeui* (Eigenmann, 1912)**

Pygidium bananeui Eigenmann, 1912a: 19. Type locality: Bernal Creek near Honda, Colombia. Holotype: FMNH 56025; holotype illustrated in Eigenmann (1918a: 318, pl. 48, fig. 1).

Distribution: River drainages of Tolima and Cundinamarca, Colombia (Maldonado-Ocampo *et al.*, 2005).

Remarks: Treated as a junior synonym of *Trichomycterus bogotensis* (Eigenmann, 1912), in de Pinna & Wosiacki (2003), who also apparently acted as first reviser, but treated as a valid species in Maldonado-Ocampo *et al.* (2005).

***Trichomycterus barbouri* (Eigenmann, 1911)**

Pygidium barbouri Eigenmann, 1911: 214, pl. 32. Type locality: Río Beni, e. Bolivia. Holotype: MCZ 29313.

Distribution: Amazon River basin, Bolivia, and la Plata River basin, Argentina (Fernández, 2000b); Beni River basin, Bolivia (de Pinna & Wosiacki, 2003).

Remarks: Redescribed in Fernández (2000).

***Trichomycterus belensis* Fernández & Vari, 2002**

Trichomycterus belensis Fernández & Vari, 2002: 739, fig. 1. Type locality: Argentina, Provincia de Catamarca, Departamento Belén, stream tributary to Laguna Blanca, 11 km northeast from Belén on provincial road 43 along route from Belén to Antogasta de la Sierra, near Los Nacimientos de San Antonio (approximately 26°30'S, 67°03'W). Holotype: FML 2530.

Distribution: Laguna Blanca basin, Argentina (de Pinna & Wosiacki, 2003).

***Trichomycterus bogotensis* (Eigenmann, 1912)**

Pygidium bogotense Eigenmann, 1912a: 18. Type locality: On the plains of Bogota, at an elevation of nearly nine thousand feet, ..., Chapinero, Colombia. Holotype: FMNH 56030; holotype illustrated in Eigenmann (1918a:

315, pl. 49, fig. 3).

Distribution: Upper Cauca, Calima and San Juan rivers, Colombia (Maldonado-Ocampo *et al.*, 2005).

***Trichomycterus bomboizanus* (Tortonese, 1942)**

Pygidium bomboizanum Tortonese, 1942: 113, pl. 1. Type locality: Rio Bomboiza (Ecuador merid.) Ecuador. Holotype: MZUT 3553.

Distribution: Bomboiza River basin, Ecuador (de Pinna & Wosiacki, 2003).

***Trichomycterus borellii* Boulenger, 1897**

Trichomycterus borellii Boulenger, 1897c: 3. Type locality: Mission d'Aguairenda, Tala, and Lesser, Argentina.

Syntypes: BMNH 1897.1.27.26 (1), ZMUT 1396 (6), ZMUT 1397 (2), ZMUT 13998 (1).

Pygidium Schmidti Berg, 1897: 266. Type locality: Río de Belén (Provincia de Catamarca), Argentina. Syntypes: BMNH 1898.9.23.3 (1), MACN 2361 (1), MACN 4595 (1), MACN 5164 (2), MACN 5176 (1).

Distribution: Salta, Catamarca and Mendoza, Argentina; Aguairenda, Bolivia (de Pinna & Wosiacki, 2003).

***Trichomycterus boylei* Nichols, 1956**

Pygidium boylei Nichols, 1956: 1, fig. 1. Type locality: dry country at Tilcara, Argentina, elevation 8000 feet, Argentina. Holotype: AMNH 20299.

Distribution: Grande River basin, Argentina (de Pinna & Wosiacki, 2003).

***Trichomycterus brasiliensis* Reinhardt, 1874**

Trichomycterus brasiliensis Reinhardt, in Lütken, 1874c: 29. Type locality: in Rio das Velhas. Syntypes: NMW 85270 (3), SMNS 2021 (1), ZMB 9171 (2), ZMUC P 30154–ZMUC P 30166 (1 each).

Trichomycterus brasiliensis tristis Lütken, 1875: 137. Type locality: Rio das Velhas, Minas Gerais, Brazil. Holotype: ZMUC P 30167.

Distribution: Upper São Francisco River, State of Minas Gerais and in smaller adjoining basins in Southeastern Brazil (de Pinna & Wosiacki, 2003).

***Trichomycterus caliensis* (Eigenmann, 1912)**

Pygidium caliense Eigenmann, 1912a: 18. Type locality: Cali, Colombia. Holotype: FMNH 56029.

Distribution: Calima River basin, Colombia (de Pinna & Wosiacki, 2003).

***Trichomycterus candidus* (Miranda Ribeiro, 1949)**

Eremophilus candidus Miranda Ribeiro, 1949b: 2, unnumbered plate. Type locality: pequeno córrego que cai no ribeirão Espírito Santo, afluente do Rio Claro e este do Sapucaí, que deságua no Rio Grande — Município de Conceição Aparecida, Estado de Minas Gerais, Brazil. Holotype: MNRJ 5209.

Distribution: Grande River basin, Minas Gerais State, Brazil (de Pinna & Wosiacki, 2003).

Remarks: Redescribed as *Eremophilus candidus* in Barbosa & Costa (2003a).

***Trichomycterus castroi* de Pinna, 1992**

Trichomycterus castroi de Pinna, 1992a: 90, figs. 1–3. Type locality: Branch of the Rio Iguaçu, near the point crossed by Curitiba-Paranaguá Road, State of Paraná, Brazil, ca. 25°26'S, 49°06'W. Holotype: MZUSP 36964.

Distribution: Iguaçu River basin, Brazil (de Pinna & Wosiacki, 2003).

***Trichomycterus catamarcensis* Fernández & Vari, 2000**

Trichomycterus catamarcensis Fernández & Vari, 2000: 990. Type locality: Provincia de Catamarca, Departamento Belén, stream tributary to Laguna Blanca, 11 km northeast from Belén on provincial road 43 along route from Belén to Antofagasta de la Sierra, near Los Nacimientos de San Antonio (approximately 26° 30'S, 67°03'W), elevation 3500m, Argentina. Holotype: FML 2507.

Distribution: Laguna Blanca basin in Catamarca Province, Argentina (de Pinna & Wosiacki, 2003).

***Trichomycterus caudofasciatus* Alencar & Costa, 2004**

Trichomycterus caudofasciatus Alencar & Costa, 2004: 5, fig. 2. Type locality: Brazil: Estado de Minas Gerais: Município de Alto Caparaó, Rio Caparaó, Rio Itabapoana basin, Alto Caparaó, 20°25'53.9"S, 41°51'56.8"W. Holotype: UFRJ 6002.

Distribution: Upper Itabapoana River basin, southeastern Brazil (Alencar & Costa, 2004).

***Trichomycterus celsae* Lasso & Provenzano, 2003**

Trichomycterus celsae Lasso & Provenzano, 2003: 1140, fig. 1. Type locality: Venezuela, Estado Bolívar, río Kukenán, cabeceras, valle entre los tepuyes Roraima y Kukenán, Gran Sabana (05°06'30"N, 60°49'48"W). Holotype:

type: MHNLS 6453.

Distribution: Kukenán River, Caroní River basin, Venezuela; known only from type locality (Lasso & Provenzano, 2003).

***Trichomycterus chaberti* Durand, 1968**

Trichomycterus chaberti Durand, 1968: 344, fig. 3. Type locality: Umayalanta [Cave], Bolivia. Holotype: MNHN 1968-0217.

Distribution: Umayalanta Cave system, Bolivia (de Pinna & Wosiacki, 2003).

***Trichomycterus chapmani* (Eigenmann, 1912)**

Pygidium chapmani Eigenmann, 1912a: 18. Type locality: Boquia, Colombia. Holotype: FMNH 56027; holotype illustrated in Eigenmann (1918a: 309, fig. 13; pl. 47, fig. 2).

Distribution: Upper Cauca River basin, and Dagua, Calima and San Juan Rivers, Colombia (Maldonado-Ocampo *et al.*, 2005).

***Trichomycterus chiltoni* (Eigenmann, 1920)**

Pygidium chiltoni Eigenmann, 1920c: 54. Type locality: lower course of the Rio Nonguen where it passes through the ground of the Agricultural School, at the outskirts of Concepción, Chile. Lectotype: CAS 57596. Also described as new in Eigenmann (1927: 40, pl. 8, figs. 1, 1a, and pl. 13, figs. 5–6), with lectotype indicated in caption of pl. 8: “Type: 15059 I. U. M., 168 mm.”

Distribution: Western drainages in central Chile (de Pinna & Wosiacki, 2003).

***Trichomycterus chungaraensis* Arratia, 1983**

Trichomycterus chungaraensis Arratia, 1983: 67, fig. 2. Type locality: Streams of Vertiente de Mal Paso, Chungará Lake, 4500 m above sea level, North Chile, South America, Chile. Holotype: FFSUC IC 290878A.

Distribution: Streams of Chungará Lake, Chile (de Pinna & Wosiacki, 2003).

***Trichomycterus concolor* Costa, 1992**

Trichomycterus concolor Costa, 1992: 107, fig. 9. Type locality: ruisseau 20 km au sud de la ville de Garapuava, bassin du rio São Francisco, Minas Gerais, Brazil. Holotype: MZUSP 43347.

Distribution: Upper São Francisco River basin, Brazil (de Pinna & Wosiacki, 2003).

***Trichomycterus conradi* (Eigenmann, 1912)**

Pygidium conradi Eigenmann, 1912b: 212. Type locality: Amatuk, British Guiana. Holotype: FMNH 53721; holotype illustrated in Eigenmann (1918a: 325; pl. 50, fig. 2).

Distribution: River drainages in Guyana and Venezuela (de Pinna & Wosiacki, 2003).

***Trichomycterus corduvensis* Weyenbergh, 1877**

Trichomycterus corduvensis Weyenbergh, 1877: 11, pl. 3 (figs. 1–2). Type locality: Rio Primero, y en las acequias de Córdoba, Argentina. Syntype: MSNG 9020 (1).

Distribution: Primeiro River basin, Argentina (de Pinna & Wosiacki, 2003).

***Trichomycterus davisi* (Haseman, 1911)**

Pygidium davisi Haseman, 1911b: 380, pl. 77; fig. 1. Type locality: Serrinha Paraná, Rio Iguaçu system, Brazil. Holotype: FMNH 60309; holotype illustrated in Eigenmann (1918a: 334; pl. 51, fig. 5).

Distribution: Iguaçu and Ribeira de Iguaape River basins, Brazil (de Pinna & Wosiacki, 2003).

***Trichomycterus diabolus* Bockmann, Casatti & de Pinna, 2004**

Trichomycterus diabolus Bockmann, Casatti & de Pinna, 2004: 227, figs. 1–2. Type locality: Brazil: São Paulo: Rio Paranapanema basin, município de Teodoro Sampaio, Morro do Diabo State Park, Córrego Carlos (22°35'28.0"S, 52°14'38.1"W). Holotype: MZUSP 78860.

Distribution: Paranapanema River basin, southeastern Brazil (Bockmann *et al.*, 2004).

***Trichomycterus dispar* (Tschudi, 1846)**

Pygidium dispar Tschudi, 1846: 22, pl. 3. Type locality: ... meisten Flüssen der Cordillera; ich habe ein Exemplar ahf einer Höhe von mehr als 14000 Fussü. M. gefangen; aber immer nur auf dem Hochlande zwischen den beiden Gebirgsketten und am Ostabhang der Anden wie an dem der Cordillera, Peru. Syntype: MHNN 767 (1).

Distribution: Peruvian Andes (de Pinna & Wosiacki, 2003).

***Trichomycterus dorsotriatus* (Eigenmann, 1918)**

Pygidium dorsotriatum Eigenmann, 1918c: 695. Type locality: Villavicencio, Colombia. Holotype: FMNH 58096;

holotype illustrated in Eigenmann (1918a: 320, pl. 48, fig. 3). Appeared only as *Pygidium dorsotriatum* in Eigenmann (1918c) but as *Pygidium dorsostriatum* in later Eigenmann publications (e. g., Eigenmann, 1918a: 320); *dorsotriatum* regarded as a typographical error in Eschmeyer *et al.* (1998) and de Pinna & Wosiacki (2003), but no corrigendum by Eigenmann has been reported.

Distribution: River drainages in Villavicencio, Colombia (de Pinna & Wosiacki, 2003).

***Trichomycterus duellmani* Arratia & Menu-Marque, 1984**

Trichomycterus duellmani Arratia & Menu-Marque, 1984: 510, figs. 14–15. Type locality: Río Tupiza, 12.5 km SE Tupiza, Bolivia, ... 22°33'S, 65°45'W. Holotype: KU 20191.

Distribution: Tupiza River basin, Bolivia (de Pinna & Wosiacki, 2003).

***Trichomycterus emanueli* (Schultz, 1944)**

Pygidium emanueli emanueli Schultz, 1944c: 259, pl. 5 (fig. c). Type locality: Río Chama at Estanques, Estado de Mérida, Venezuela. Holotype: USNM 121223.

Distribution: Chama River basin, Venezuela (de Pinna & Wosiacki, 2003).

***Trichomycterus fassli* (Steindachner, 1915)**

Pygidium fassli Steindachner, 1915a: 200. Type locality: Rio Songo im Distrikt Nord-Yungas, Bolivia. Syntypes (4): NMW 44470 (1). Illustrated and described in more detail as *Pygidium fasslii* in Steindachner (1915e: 97, pl. 13, figs. 1–2).

Distribution: Songo River basin, Bolivia (de Pinna & Wosiacki, 2003).

***Trichomycterus gabrieli* (Myers, 1926)**

Pygidium gabrieli Myers, 1926: 151. Type locality: São Gabriel rapids, Rio Negro, in rock-pools, Amazon system, Brazil. Syntypes: CAS 64583 (4), SU 36556 (1).

Distribution: Upper Negro River, Brazil (de Pinna & Wosiacki, 2003).

***Trichomycterus giganteus* Lima & Costa, 2004**

Trichomycterus giganteus Lima & Costa, 2004: 1, fig. 1. Type locality: Brazil: Estado do Rio de Janeiro: Município do Rio de Janeiro, Campo Grande, Rio Guandu-Mirim, Rio Guandu basin. Holotype: UFRJ 5999.

Distribution: Upper Guandu River basin, southeastern Brazil (Lima & Costa, 2004).

***Trichomycterus goeldii* Boulenger, 1896**

Trichomycterus goeldii Boulenger, 1896b: 154. Type locality: Colonia Alpina, Organ Mts in the Province Rio Janeiro, at an altitude of nearly 2600 feet, Brazil. Syntypes: BMNH 1896.4.4.7–8 (2).

Distribution: Mountain ranges of coastal basins of Rio de Janeiro State, Brazil (de Pinna & Wosiacki, 2003).

***Trichomycterus gorgona* Fernández & Schaefer, 2005**

Trichomycterus gorgona Fernández & Schaefer, 2005: 69, figs. 1–3. Type locality: Colombia, Departamento Cauca, Isla Gorgona, freshwater stream near northeast end of island, 02°59'N, 78°11'30"W. Holotype: ANSP 149946.

Distribution: Gorgona Island, Pacific coast of Colombia (Fernández & Schaefer, 2005).

***Trichomycterus guaraquessaba* Wosiacki, 2005**

Trichomycterus guaraquessaba Wosiacki, 2005: 51, fig. 2. Type locality: rio Bracinho, Fazenda Salto Dourado, Município de Guaraqueçaba, Paraná, Brazil. Holotype: MPEG 7916.

Distribution: Bracinho River, Atlantic coastal basin, southern Brazil (Wosiacki, 2005).

***Trichomycterus guianensis* (Eigenmann, 1909)**

Pygidium guianensis Eigenmann, 1909a: 11. Type locality: Aruataima Falls, Upper Potaro, British Guiana. Holotype: FMNH 52676; holotype illustrated in Eigenmann (1918a: 325, pl. 50, fig. 1) as *Pygidium guianense*.

Distribution: Rivers of the Guianas and Venezuela (de Pinna & Wosiacki, 2003).

***Trichomycterus hasemani* (Eigenmann, 1914)**

Pygidium hasemani Eigenmann, 1914a: 48. Type locality: Santarem, Pará, Brazil. Holotype: FMNH 56424; holotype illustrated in Eigenmann (1918a: 326, pl. 50, fig. 4).

Distribution: Amazon River basin (de Pinna & Wosiacki, 2003).

***Trichomycterus heterodontus* (Eigenmann, 1918)**

Pygidium heterodontum Eigenmann, 1918c: 692. Type locality: Rio Mendoza, Palmira, Argentina, 900 m, Argentina. Holotype: CAS 58139; holotype illustrated in Eigenmann (1918a: 296, pl. 44, fig. 4).

Distribution: Mendoza River basin, Argentina (de Pinna & Wosiacki, 2003).

Trichomycterus iheringi (Eigenmann, 1918)

Pygidium iheringi Eigenmann, 1918c: 697. Type locality: São Paulo in coastal streams and Parana basin, Brazil.

Holotype: CAS 64585; holotype illustrated in Eigenmann (1918a: 330, pl. 50, fig. 5) and Wosiacki (2005: 58, fig. 3).

Distribution: Ribeira do Iguape River basin, Brazil (de Pinna & Wosiacki, 2003).

Remarks: Redescribed in Wosiacki (2005: 56).

Trichomycterus immaculatus (Eigenmann & Eigenmann, 1889)

Pygidium immaculatum Eigenmann & Eigenmann, 1889b: 52. Type locality: Juiz de Fora, Rio Parahybuna, São Matheos, Goyaz [Brazil]. Syntypes (14): MCZ 8266 (1), MCZ 8300 (10), MCZ 8302 (1), MCZ 8305 (1), MCZ 8307 (1). Species illustrated in Eigenmann (1918a: 334, pl. 52, fig. 1).

Distribution: Paraíbuna River, Paraíba do Sul River basin, Brazil (de Pinna & Wosiacki, 2003).

Trichomycterus itacambirussu Triques & Vono, 2004

Trichomycterus itacambirussu Triques & Vono, 2004: 165, fig. 2. Type locality: Brazil: Minas Gerais: County of Grão Mogol, Rio Jequitinhonha basin: drainage of Rio Itacambiruçu: Córrego do Cambral, 42°45'W 16°33'S. Holotype: MZUSP 58493.

Distribution: Jequitinhonha River basin, Minas Gerais, Brazil (Triques & Vono, 2004).

Trichomycterus itacarambiensis Trajano & de Pinna, 1996

Trichomycterus itacarambiensis Trajano & de Pinna, 1996: 86, figs. 1–2. Type locality: Creek inside Olhos d'Água cave, Município de Itacarambi, Minas Gerais, Brazil, 15°06'06"S, 44°09'30"W. Holotype: MZUSP 42469.

Distribution: Olhos d'Água Cave in State of Minas Gerais, Brazil (de Pinna & Wosiacki, 2003).

Trichomycterus itatiayae Miranda Ribeiro, 1906

Trichomycterus brasiliensis itatiayae Miranda Ribeiro, 1906: 178, pl. 1 (figs. a–c). Type locality: ... do Itatiaya, ... em afluente do Parahyba, Rio de Janeiro State, Brazil. Lectotype: MNRJ 792, designated by Caramaschi & Caramaschi (1991: 223).

Distribution: Rivers in Itatiaia Mountains, upper reaches of Paraíba do Sul River basin, Brazil (de Pinna & Wosiacki, 2003).

Remarks: Type locality was interpreted by Caramaschi & Caramaschi (1991: 223) as: Ribeirão da Tapera (tributary of the Ribeirão Bonito, Rio Paraíba do Sul drainage), Mont-Serrat, Itatiaia (ca. 22°27'S, 44°50'W, ca. 800 m a.s.l.), Rio de Janeiro State, Brazil.

Trichomycterus jacupiranga Wosiacki & Oyakawa, 2005

Trichomycterus jacupiranga Wosiacki & Oyakawa, 2005: 469, figs. 5–6. Type locality: rio do Queimado, Parque Estadual de Jacupiranga, Cajati [São Paulo, Brazil]. Holotype: MZUSP 67818.

Distribution: Queimado River, São Paulo, Brazil (Wosiacki & Oyakawa, 2005).

Trichomycterus jequitinhonhae Triques & Vono, 2004

Trichomycterus jequitinhonhae Triques & Vono, 2004: 167, fig. 3. Type locality: Brazil: Minas Gerais: County of Coronel Murta: Rio Jequitinhonha basin: Córrego Laranjeiras, 42°18'W 16°45'S. Holotype: MZUSP 58497.

Distribution: Jequitinhonha River basin, Minas Gerais, Brazil (Triques & Vono, 2004).

Trichomycterus johnsoni (Fowler, 1932)

Pygidium johnsoni Fowler, 1932a: 367, figure on p. 364. Type locality: Descalvados, Matto Grosso, Brazil. Holotype: ANSP 53873.

Distribution: Paraná River basin, Mato Grosso, Brazil, and Corrientes, Argentina (de Pinna & Wosiacki, 2003).

Trichomycterus knerii Steindachner, 1882

Trichomycterus Knerii Steindachner, 1882a: 142. Type locality: Canelos [Ecuador]. Syntype: NMW 43328 (1). Species illustrated and described in more detail in Steindachner (1882d: 81, pl. 5, figs. 1, 1a).

Distribution: Canelos, Ecuador (de Pinna & Wosiacki, 2003).

Trichomycterus landinga Triques & Vono, 2004

Trichomycterus landinga Triques & Vono, 2004: 162, fig. 1. Type locality: Brazil: Minas Gerais: County of Coronel Murta: Rio Jequitinhonha basin: Córrego Moquém, 42°35'W 16°45'S. Holotype: MZUSP 58496.

Distribution: Jequitinhonha River basin, Minas Gerais, Brazil (Triques & Vono, 2004).

Trichomycterus latidens (Eigenmann, 1918)

Pygidium latidens Eigenmann, 1918c: 693. Type locality: Small creek near mouth of Rio Calima, Colombia. Holotype: IU 13801 (whereabouts unknown); holotype illustrated in Eigenmann (1918a: 312, pl. 47, fig. 4).

Distribution: Calima River basin, Colombia (de Pinna & Wosiacki, 2003).

***Trichomycterus latistriatus* (Eigenmann, 1918)**

Pygidium latistriatum Eigenmann, 1918c: 696. Type locality: Quebrada de Pinchote, Santander, Colombia. Holotype: FMNH 58449; holotype illustrated in Eigenmann (1918a: 321, pl. 48, fig. 4).

Distribution: Gaira, Cesar and Ariguani rivers, Magdalena River basin, Colombia (Maldonado-Ocampo *et al.*, 2005).

***Trichomycterus laucaensis* Arratia, 1983**

Trichomycterus laucaensis Arratia, 1983: 74, fig. 8. Type locality: System of Lauca River, Parinacota, 4,390 m above sea level, northern Chile. Holotype: FFSUC IC 160878A.

Distribution: Lauca River basin, Chile (de Pinna & Wosiacki, 2003).

***Trichomycterus lewi* Lasso & Provenzano, 2003**

Trichomycterus lewi Lasso & Provenzano, 2003: 1144, fig. 7. Type locality: Venezuela, Estado Bolívar, río Kukenán, cabeceras, valle entre los tepuyes Roraima y Kukenán, Gran Sabana (05°06'30"N, 60°49'48"O). Holotype: MHNLS 6454.

Distribution: Kukenán River, Caroni River basin, Venezuela; known only from type locality (Lasso & Provenzano, 2003).

***Trichomycterus longibarbatus* Costa, 1992**

Trichomycterus longibarbatus Costa, 1992: 104, fig. 4. Type locality: près de la ville de Santa Tereza, Espírito Santo, Brazil. Holotype: MZUSP 43339.

Distribution: Near Santa Tereza, Espírito Santo State, Brazil (de Pinna & Wosiacki, 2003).

***Trichomycterus maracaiboensis* (Schultz, 1944)**

Pygidium banneui maracaiboensis Schultz, 1944c: 262, pl. 6 (fig. b). Type locality: Río San Juan near bridge, south of Mene Grande, tributary to Río Motatán, Maracaibo basin, Venezuela. Holotype: USNM 121227.

Distribution: San Juan River, tributary to Motatán River, Lake Maracaibo basin, Venezuela (de Pinna & Wosiacki, 2003).

***Trichomycterus mboycey* Wosiacki & Garavello, 2004**

Trichomycterus mboycey Wosiacki & Garavello, 2004: 8, fig. 5. Type locality: Brazil: State of Paraná: município de Foz do Jordão, Jordão reservoir, rio Jordão near at its mouth, tributary of rio Iguaçu (rio Paraná Basin), 25°45'S, 52°10'W. Holotype: MPEG 6695.

Distribution: Jordão River, Iguaçu River basin, Brazil (Wosiacki & Garavello, 2004).

***Trichomycterus meridae* Regan, 1903**

Trichomycterus meridae Regan, 1903b: 624. Type locality: Merida, Venezuela, and from the Rio Albireggas above Merida, altitude 3500 metres, Venezuela. Syntypes: BMNH 1903.4.28.35-38 (4), BMNH 1906.6.30.77-79 (3), USNM 133136 (1).

Distribution: Albirregas River basin, Venezuela (de Pinna & Wosiacki, 2003).

***Trichomycterus migrans* (Dahl, 1960)**

Pygidium migrans Dahl, 1960b: 309, fig. 3. Type locality: in front of Camp 1, in the Guayabero River [Orinoco System, Colombia]. Holotype: ICNMHN 399.

Distribution: Guayabero River, Orinoco River basin, Colombia (de Pinna & Wosiacki, 2003).

***Trichomycterus mimonha* Costa, 1992**

Trichomycterus mimonha Costa, 1992: 106, fig. 7. Type locality: 2 km au nord de la ville de Piquete, rio Benfica, bassin du rio Paraíba, São Paulo, Brazil. Holotype: MZUSP 43343.

Distribution: Benfica River, Paraíba do Sul River basin, Brazil (de Pinna & Wosiacki, 2003).

***Trichomycterus mirissumba* Costa, 1992**

Trichomycterus mirissumba Costa, 1992: 107, fig. 8. Type locality: Visconde de Mauá, 1 km à l'est de la ville de Maromba, rio Preto, près du confluent avec le ruisseau Santa Clara, bassin du rio Paraíba, Rio de Janeiro, Brazil. Holotype: MZUSP 43345.

Distribution: Preto River, Paraíba do Sul River basin, Brazil (de Pinna & Wosiacki, 2003).

***Trichomycterus mondolfi* (Schultz, 1945)**

Pygidium mondolfi Schultz, 1945: 29, fig. 1. Type locality: Quebrado Chacaito near Caracas but in Estado de Miranda, Río Tuy system, Venezuela. Holotype: USNM 120377.

Distribution: Tuy River basin, Caribbean Drainage, Venezuela (de Pinna & Wosiacki, 2003).

***Trichomycterus motatanensis* (Schultz, 1944)**

Pygidium emanueli motatanensis Schultz, 1944c: 260, pl. 6 (fig. a). Type locality: Río San Juan at the bridge south of Mene Grande, Motatán system, Maracaibo basin Venezuela. Holotype: USNM 121232.

Distribution: San Juan River, Motatán drainage, Lake Maracaibo basin, Venezuela (de Pinna & Wosiacki, 2003).

***Trichomycterus naipi* Wosiacki & Garavello, 2004**

Trichomycterus naipi Wosiacki & Garavello, 2004: 2, fig. 2. Type locality: Brazil: State of Paraná: município de Tijucas do Sul, rio São João, tributary of rio da Várzea (Iguacu Basin), near Lagoinha, 25°55'S, 49°11'W. Holotype: MPEG 6699.

Distribution: da Várzea River, Iguacu River basin, Brazil (Wosiacki & Garavello, 2004).

***Trichomycterus nigricans* Valenciennes, 1832**

Trichomycterus nigricans Valenciennes, 1832a: 348. Type locality: ruisseaux de Sainte-Catherine du Brésil, Santa Catarina, Brazil. Holotype: apparently MNHN b-0251.

Distribution: Coastal drainages in Santa Catarina State, Brazil (de Pinna & Wosiacki, 2003).

Remarks: Species redescribed by Arratia (1998) with MNHN b-0251 listed as the holotype.

***Trichomycterus nigromaculatus* Boulenger, 1887**

Trichomycterus nigromaculatus Boulenger, 1887b: 349. Type locality: Andes of Columbia. Syntypes: BMNH 1880.2.26.16–17 (2).

Distribution: Colombian Andes (de Pinna & Wosiacki, 2003).

***Trichomycterus pantherinus* Alencar & Costa, 2004**

Trichomycterus pantherinus Alencar & Costa, 2004: 2, fig. 1. Type locality: Brazil: Estado do Espírito Santo: Município de Sant Leopoldina, below waterfall near Fazenda Sete Quedas, Rio da Prata, Rio Santa Maria da Vitória basin, 20°03'15.9"S, 40°32'20.5"W. Holotype: UFRJ 6001.

Distribution: Santa Maria da Vitória River basin, Southeastern Brazil (Alencar & Costa, 2004).

***Trichomycterus paolencis* (Eigenmann, 1918)**

Pygidium paolence Eigenmann, 1918c: 698. Type locality: São Paulo in the Parana basin and (?) in coastal streams. ... Alto da Serra, Rio Tieté, São Paulo. ... Mogi das Cruzes, Rio Tieté, São Paulo, Brazil. Holotype: FMNH 58085; holotype illustrated in Eigenmann (1918a: 332, pl. 51, fig. 3).

Distribution: Paraná River basin, São Paulo State, Brazil (de Pinna & Wosiacki, 2003).

***Trichomycterus papilliferus* Wosiacki & Garavello, 2004**

Trichomycterus papilliferus Wosiacki & Garavello, 2004: 5, figs. 3, 4. Type locality: Brazil: State of Paraná: município de Foz do Jordão, Jordão reservoir, rio Jordão near at its mouth, tributary of rio Iguaçu (rio Paraná Basin), 25°45'S, 52°10'W. Holotype: MPEG 6692.

Distribution: Jordão River, Iguaçu River basin, Brazil (Wosiacki & Garavello, 2004).

***Trichomycterus paquequerensis* (Miranda Ribeiro, 1943)**

Pygidium paquequerense Miranda Ribeiro, 1943: 2, fig. Type locality: rio Paquequer Grande, Estado do Rio de Janeiro [Brazil]. Holotype: MNRJ 1159.

Distribution: Paquequer River, Paraíba do Sul River basin, Brazil (de Pinna & Wosiacki, 2003).

***Trichomycterus piurae* (Eigenmann, 1922)**

Pygidium punctulatum piurae Eigenmann, 1922b: 63. Type locality: Piura, Peru. Holotype: CAS 58119.

Distribution: Piura River basin, Peru (de Pinna & Wosiacki, 2003).

***Trichomycterus plumbeus* Wosiacki & Garavello, 2004**

Trichomycterus plumbeus Wosiacki & Garavello, 2004: 13, fig. 8. Type locality: Brazil: State of Paraná: município de Foz do Jordão, Jordão reservoir, rio Jordão near at its mouth, tributary of rio Iguaçu (rio Paraná Basin), 25°45'S, 52°10'W. Holotype: MPEG 6689.

Distribution: Jordão River, Iguaçu River basin, Brazil (Wosiacki & Garavello, 2004).

***Trichomycterus potschi* Barbosa & Costa, 2003**

Trichomycterus potschi Barbosa & Costa, 2003b: 282, fig. 1. Type locality: Brazil: Estado do Rio de Janeiro: rio das Cachoeiras, serra do Matutu, Município de Mangaratiba. Holotype: MCP 29061.

Distribution: Coastal river basins between Mangaratiba and Itaguaí, Rio de Janeiro State, Brazil (Barbosa & Costa, 2003b).

***Trichomycterus pradensis* Sarmento-Soares, Martins-Pinheiro, Aranda & Chamon, 2005**

Trichomycterus pradensis Sarmento-Soares, Martins-Pinheiro, Aranda & Chamon, 2005: 291, fig. 1. Type locality: Brazil: Bahia: Jucuruçu, rio Jucuruçu, 2 km before the city of Jucuruçuon road Itamaraju-Jucuruçu, middle of rio Jucuruçu basin, 16°50'10"S 40°08'40"W. Holotype: MNRJ 28483.

Distribution: Jucuruçu River, a coastal river basin of SE Brazil (Sarmento-Soares *et al.*, 2005).

***Trichomycterus pseudosilvinichthys* Fernández & Vari, 2004**

Trichomycterus pseudosilvinichthys Fernández & Vari, 2004: 876, figs. 1–2. Type locality: Argentina, Provincia de La Rioja, Departamento Chilecito, Río Amarillo at Famatina, a small assemblage of houses near Fundición de Oro Santa Florentina (28°55'S, 67°31'W), on east slope of Sierra de Famatina. Holotype: FML 2588.

Distribution: La Rioja province, Argentina (Fernández & Vari, 2004).

***Trichomycterus punctatissimus* Castelnau, 1855**

Trichomycterus punctatissimus Castelnau, 1855: 49, pl. 24 (fig. 3). Type locality: De l'Araguay. Holotype: MNHN b-0610.

Distribution: Araguaia River basin, Brazil (de Pinna & Wosiacki, 2003).

***Trichomycterus punctulatus* Valenciennes, 1846**

Trichomycterus punctulatus Valenciennes, in Cuvier & Valenciennes, 1846: 488, pl. 552. Type locality: la rivière de Lima, Peru. Syntypes: MNHN 0000-3168 (5).

Distribution: Western Peru (de Pinna & Wosiacki, 2003).

***Trichomycterus ramosus* Fernández, 2000**

Trichomycterus ramosus Fernández, 2000a: 350, fig. 1. Type locality: Catamarca: Departamento Belém: Laguna Blanca, 3680 m elevation, approximately 26°30'S, 67°03'W, Argentina. Holotype: FML 2070.

Distribution: Laguna Blanca basin, Catamarca Province, Argentina (de Pinna & Wosiacki, 2003).

***Trichomycterus regani* (Eigenmann, 1918)**

Pygidium regani Eigenmann, 1918c: 696. Type locality: Tado, Rio San Juan, Colombia. Holotype: CAS 64591; holotype illustrated in Eigenmann (1918a: 323, pl. 48, fig. 5).

Distribution: San Juan River basin, Colombia (de Pinna & Wosiacki, 2003).

***Trichomycterus reinhardti* (Eigenmann, 1918)**

Pygidium reinhardti Eigenmann, 1918c: 699. Type locality: Burmier on the Rio Itabira, a tributary of the Rio das Velhas, Brazil. Holotype: FMNH 58081; holotype illustrated in Eigenmann (1918a: 333, pl. 51, fig. 4).

Distribution: Upper São Francisco River basin, Brazil (de Pinna & Wosiacki, 2003).

***Trichomycterus retropinnis* Regan, 1903**

Trichomycterus retropinnis Regan, 1903b: 624. Type locality: St. Augustin, Andes of Colombia, elev. 5000 feet. Syntypes (2): BMNH 1899.8.21.12–13 (2).

Distribution: Upper Magdalena River basin, Colombia (Maldonado-Ocampo *et al.*, 2005).

***Trichomycterus riojanus* (Berg, 1897)**

Pygidium riojanum Berg, 1897: 269. Type locality: un arroyo de la Cordillera de La Rioja, Argentina. Holotype: MACN 5175 (destroyed, Fernández & Schaefer, 2003: 357).

Distribution: La Rioja Range, Argentina (de Pinna & Wosiacki, 2003).

***Trichomycterus rivulatus* Valenciennes, 1846**

Trichomycterus rivulatus Valenciennes, in Cuvier & Valenciennes, 1846: 495. Type locality: ruisseaux qui se jettent dans le lac de Titicaca, vaste mer alpine, peuplée par les Orestias sans ventrales, ou dans les affluens de l'Apurimac, l'une des sources de l'Amazone ... du Guasacoma, Peru. Syntype: MNHN b-0586 (1).

Trichomycterus barbatula Valenciennes, in Cuvier & Valenciennes, 1846: 498. Type locality: Guasacoma et du Rio de Pontezualo près coroico, par une hauteur de terize à quatorze milles pieds et à una latitude de seize à dix-sept degrés nord. Syntypes: MNHN 0000-4077 (1), MNHN 0000-4078 (5) and MNHN b-0587 (3).

Trichomycterus gracilis Valenciennes, in Cuvier & Valenciennes, 1846: 497. Type locality: Rio de Azangaro près de

Guasacona, dans le Rio de Pontezualo près de Coroico, et enfin dans le lac de la Compucila dans les Andes, à l'ouest de Cuzco, par la hautes de quatorze mille pieds, Peru. Syntypes: MNHN 0000-3129 (2), MNHN 0000-4063 (2), MNHN a-9766 (6), MNHN b-0588 (2).

Trichomycterus incae Valenciennes, in Cuvier & Valenciennes, 1846: 496. Type locality: Rio Guatanai à Cuzco, Peru. Syntype: MNHN a-8986 (1).

Trichomycterus pentlandi Castelnau, 1855: 49, pl. 24 (fig. 1). Type locality: lac situé près de la mission de Sarayacu, qui communiqua avec la rivière d'Ucayale, Peru. Holotype: MNHN b-0608.

Trichomycterus pictus Castelnau, 1855: 50, pl. 24 (fig. 2). Type locality: grand lac de Titicaca. Syntypes: MNHN b-0609 (3).

Trichomycterus pardus Cope, 1874b: 132. Type locality: Upper Amazon. Possible holotype: ANSP 22004.

Trichomycterus poeyanus Cope, 1877: 47. Type locality: Arequipa, Peru. Syntypes: ANSP 21382-83 (2).

Pygidium oroyae Eigenmann & Eigenmann, 1889b: 51. Type locality: Pochachara, Oroya River [Brazil]. Syntypes (8): MCZ 3955 (8).

Trichomycterus eigenmanni Boulenger, 1898c: 8. Type locality: Cumbaca [Brazil]. Holotype: MCZ 8301.

Pygidium quechuorum Steindachner, 1900: 207. Type locality: Arequipa, Río Chile, Südperu. Syntypes (5): at NMW.

Pygidium tiraquae Fowler, 1940c: 92, fig. 52. Type locality: Tiraque, Cochabamba Department, Bolivia. Holotype: ANSP 69126.

Pygidium atochaeanum Allen, in Eigenmann & Allen, 1942: 156, pl. 13 (figs. 3-5). Type locality: Rio de Atocha, Atocha, Bolivia. Holotype: CAS 64576.

Distribution: High-altitude lakes and streams in the central Andean range (including Lakes Titicaca and Poopó), from Lake Junin in the north to Chilean region of Tarapacá in the south, spanning Western Bolivia, Peru and Northern Chile (de Pinna & Wosiacki, 2003).

***Trichomycterus roigi* Arratia & Menu-Marque, 1984**

Trichomycterus roigi Arratia & Menu-Marque, 1984: 494, figs. 1-10. Type locality: Río Pastos Chicos, Jujuy, north of Argentina; 23°24'S — 66°35'W. Holotype: MLP 29-8-83-1.

Distribution: Pastos Chicos River basin, Argentina (de Pinna & Wosiacki, 2003).

***Trichomycterus romeroi* (Fowler, 1941)**

Pygidium romeroi Fowler, 1941c: 4, figs. 6-8. Type locality: Honda, Colombia. Holotype: ANSP 69331.

Distribution: Honda River basin, Colombia (de Pinna & Wosiacki, 2003).

***Trichomycterus santaeritae* (Eigenmann, 1918)**

Pygidium santae-ritae Eigenmann, 1918a: 341, pl. 52 (fig. 5). Type locality: Santa Rita, Rio Preto, Brazil. Holotype: FMNH 58577.

Distribution: Preto River, Paraíba do Sul River basin, Brazil (de Pinna & Wosiacki, 2003).

***Trichomycterus spegazzinii* (Berg, 1897)**

Pygidium Spegazzinii Berg, 1897: 267. Type locality: Río de Cachi (Provincia de Salta), ..., a una altura de 2500 a 2800 metros sobre el nivel del mar, Argentina. Syntypes: BMNH 1898.9.23.1-2 (2), MACN 4925 (19), MACN 5173 (2), SMF 831 (1).

Distribution: Provinces of Salta and Catamarca in Argentina (de Pinna & Wosiacki, 2003).

***Trichomycterus spelaeus* Do Nascimento, Villarreal & Provenzano, 2001**

Trichomycterus spelaeus Do Nascimento, Villarreal & Provenzano, 2001: 21, fig. 1. Type locality: Cueva Punto Fijo, en el caserío Punto Fijo a 7,5 km al N. del Cerro Yolanda. 590 m.s.n.m., cuenca del Río Guasare, Edo. Zulia, Venezuela (10°57'10"N 72°28'06"E). Holotype: MBUCV V-29602.

Distribution: Punto Fijo Cave, upper Guasare River basin, Venezuela (de Pinna & Wosiacki, 2003).

***Trichomycterus spilosoma* (Regan, 1913)**

Pygidium spilosoma Regan, 1913d: 468. Type locality: Rio Sipi and Rio Tamana, Colombia. Syntypes (3, 130-250 mm TL): BMNH 1910.7.11.106-107 (2), BMNH 1910.7.11.108 (1, skeleton), BMNH 1910.7.11.15 (1).

Distribution: Sipi and Tamana River basins, Colombia (de Pinna & Wosiacki, 2003).

***Trichomycterus stawiarski* (Miranda Ribeiro, 1968)**

Pygidium stawiarski Miranda Ribeiro, 1968: 1. Type locality: Pequeno córrego pertencente á bacia do Rio Paraná —

localidade de Bituruna —Est. do Paraná, Paraná, Brazil. Holotype: MNRJ 9739.

Distribution: Iguaçu River basin, Brazil (de Pinna & Wosiacki, 2003).

***Trichomycterus stellatus* (Eigenmann, 1918)**

Pygidium stellatum Eigenmann, 1918a: 308, pl. 47 (fig. 1). Type locality: Quebrada Sarjento, Colombia. Holotype: FMNH 58101.

Distribution: Rivers of Andean eastern cordillera, Colombia (de Pinna & Wosiacki, 2003).

***Trichomycterus straminius* (Eigenmann, 1918)**

Pygidium straminium Eigenmann, 1918c: 694. Type locality: Quebrada del Mango, Santander, Colombia. Holotype: FMNH 58105; holotype illustrated in Eigenmann (1918a: 313, pl. 49, fig. 1) as *Pygidium stramineum*.

Distribution: Andean streams of Cundinamarca and Santander, Colombia (Maldonado-Ocampo *et al.*, 2005).

***Trichomycterus striatus* (Meek & Hildebrand, 1913)**

Pygidium striatum Meek & Hildebrand, 1913: 78. Type locality: Río Cana at Cana, Panama. Holotype: FMNH 7579.

Pygidium septentrionale Behre, 1928: 309, pl. 18. Type locality: small streams tributary to Rio Chiriquí del Tire above Caldera, Pacific slope of Panama, altitude about 4,000 feet ... Quebrada Salão, Panama, ca. elev. 4000 ft. Holotype: FMNH 59522.

Distribution: Rivers of Panama and Costa Rica (de Pinna & Wosiacki, 2003); Catatumbo, Magdalena, Cauca and Dagua rivers, Colombia (Maldonado-Ocampo *et al.*, 2005).

***Trichomycterus taczanowskii* Steindachner, 1882**

Trichomycterus Taczanowskii Steindachner, 1882b: 177. Type locality: Rio Huambo, Rio de Tortora [Peru]. Syntype: NMW 43387 (1). Species illustrated and described in more detail in Steindachner (1882c: 22, pl. 4, figs. 1, 1a, 1b).

Distribution: Peru (de Pinna & Wosiacki, 2003).

***Trichomycterus taenia* Kner, 1863**

Trichomycterus taenia Kner, 1863: 228, fig. 16. Type locality: Vom Westabhang der Andes im Staate Ecuador, Ecuador. Holotype: at NMW. Also appeared in Kner & Steindachner (1864: 52, pl. 6, fig. 1).

Distribution: Ecuadorian Andes (de Pinna & Wosiacki, 2003); Magdalena and Patia River basins, Colombia (Maldonado-Ocampo *et al.*, 2005).

***Trichomycterus taeniops* Fowler, 1954**

Pygidium tenue Fowler, 1945c: 6, figs. 7–9. Type locality: Acobamba, near Tarma at 2000 meters elevation, Rio Ucayali basin, Peru. Holotype: ANSP 71638. Preoccupied in *Trichomycterus* by *Trichomycterus tenuis* Weyenbergh, 1877; replaced by *Trichomycterus taeniops* Fowler, 1954.

Trichomycterus taeniops Fowler, 1954: 36, fig. 635. Type locality: Acobamba, near Tarma, at 2000 meters elevation, Rio Ucayali basin, Peru, elev. 2000 m, Peru. Replacement for *Pygidium tenue* Fowler, 1945; preoccupied in *Trichomycterus* by *Trichomycterus tenuis* Weyenbergh, 1877.

Distribution: Ucayali River basin (elevation 2000 m), Peru (de Pinna & Wosiacki, 2003).

***Trichomycterus taroba* Wosiacki & Garavello, 2004**

Trichomycterus taroba Wosiacki & Garavello, 2004: 10, fig. 6. Type locality: Brazil: State of Paraná: município de Foz do Jordão, Jordão reservoir, rio Jordão near at its mouth, tributary of rio Iguaçu (rio Paraná Basin), 25°45'S, 52°10'W. Holotype: MPEG 6689.

Distribution: Jordão River, Iguaçu River basin, Brazil (Wosiacki & Garavello, 2004).

***Trichomycterus tenuis* Weyenbergh, 1877**

Trichomycterus tenuis Weyenbergh, 1877: 12, pl. 3 (figs. a–c). Type locality: una pequeña laguna, en la Sierra de Córdoba, cerca de la Villa Cruz-del-eje, Argentina. Syntypes: MSNG 8852 (2).

Distribution: Córdoba Sierra, Argentina (de Pinna & Wosiacki, 2003).

***Trichomycterus transandianus* (Steindachner, 1915)**

Pygidium taenia transandianum Steindachner, 1915e: 100, pl. 12 (fig. 6). Type locality: Gebirgsbach im Cañon del Gallo, einem rechten Seitentale des Rio Combeima in der Zentral-Cordillere, Columbian, in einer Höhe von 1800 m. Syntypes (2): NMW 44475. Originally as *Pygidium taenia* Var. *nova transandianum*.

Distribution: Combeima River basin, central Andean cordillera, Colombia (de Pinna & Wosiacki, 2003).

Trichomycterus trefauti Wosiacki, 2004

Trichomycterus trefauti Wosiacki, 2004: 3, fig. 2. Type locality: riacho Andrequicé, tributary of rio Paraúna, itself a tributary of rio das Velhas (rio São Francisco basin), approximately 18°30'S, 43°30'W, Município de Trinta Réis, Minas Gerais, Brazil. Holotype: MZUSP 79911.

Distribution: Upper São Francisco River basin, Brazil (Wosiacki, 2004).

Trichomycterus triguttatus (Eigenmann, 1918)

Pygidium triguttatum Eigenmann, 1918a: 339, pl. 52 (fig. 4). Type locality: Jacarehy, São Paulo, Brazil. Holotype: FMNH 58670.

Distribution: São Paulo State, Brazil (de Pinna & Wosiacki, 2003).

Trichomycterus tupinamba Wosiacki & Oyakawa, 2005

Trichomycterus tupinamba Wosiacki & Oyakawa, 2005: 466, figs. 2–4. Type locality: Brazil, São Paulo: rio Betari, tributary of rio Ribeira de Iguape, in front of the camping area of Núcleo Santana, Parque Estadual Turístico do Alto Ribeira. Holotype: MZUSP 62382.

Distribution: Betari and Iporanga rivers, Ribeira de Iguape River basin, Brazil (Wosiacki & Oyakawa, 2005).

Trichomycterus unicolor (Regan, 1913)

Pygidium unicolor Regan, 1913d: 468. Type locality: Condoto, Colombia. Syntypes: BMNH 1913.10.1.42–43 (2). Distribution: San Juan River basin, Colombia (de Pinna & Wosiacki, 2003).

Trichomycterus variegatus Costa, 1992

Trichomycterus variegatus Costa, 1992: 103, fig. 3. Type locality: Estado de Minas Gerais: ville de São Roque de Minas, rio do Peixe, affluent du haut rio São Francisco [Brazil]. Holotype: MZUSP 42316.

Distribution: Upper São Francisco River basin, Brazil (de Pinna & Wosiacki, 2003).

Trichomycterus venulosus (Steindachner, 1915)

Pygidium venulosum Steindachner, 1915a: 199. Type locality: Paramo de Cruz verde, östliche Cordillere, Columbien, in 3000 m. Syntypes: NMW 44476 (2). Species illustrated and described in more detail in Steindachner (1915e: 85, pl. 121, figs. 3–4).

Distribution: Streams of eastern Andean cordillera, Colombia (Maldonado-Ocampo *et al.*, 2005).

Trichomycterus vermiculatus (Eigenmann, 1918)

Pygidium vermiculatum Eigenmann, 1918c: 699. Type locality: Juiz de Fora, Rio Parahyba, Brazil. Holotype: FMNH 58077; holotype illustrated in Eigenmann (1918a: 335, pl. 52, fig. 2).

Distribution: Paraíba do Sul River, State of Minas Gerais, Brazil (de Pinna & Wosiacki, 2003).

Trichomycterus vittatus Regan, 1903

Trichomycterus vittatus Regan, 1903b: 623. Type locality: Marcapata Valley, E. Peru. Holotype: BMNH 1902.5.29.210.

Distribution: Eastern Peru (de Pinna & Wosiacki, 2003).

Trichomycterus weyrauchi (Fowler, 1945)

Pygidium weyrauchi Fowler, 1945c: 7, figs. 10–12. Type locality: Acobamba, near Tarma, at 2200 meters elevation, Rio Ucayali drainage, Peru. Holotype: ANSP 71639.

Distribution: Ucayali River basin (elev. 2900 m), Peru (de Pinna & Wosiacki, 2003).

Trichomycterus yuska Fernández & Schaefer, 2003

Trichomycterus yuska Fernández & Schaefer, 2003: 355, figs. 1–2. Type locality: Argentina: Provincia de Catamarca, Departamento Tinogasta, Arroyo Aguas Calientes, 4050 m elevation. Holotype: FML 2535.

Distribution: Known only from the type locality, a high elevation stream in western Argentina (Fernández & Schaefer, 2003).

Trichomycterus zonatus (Eigenmann, 1918)

Pygidium zonatum Eigenmann, 1918a: 330, pl. 51 (fig. 1). Type locality: Agua Quente, São Paulo, Brazil. Holotype: FMNH 58573.

Trichomycterus cubataonis Bizerril, 1994: 618, figs. 1–3. Type locality: Rio Cubatão, Joinville, Estado de Santa Catarina [Brazil]. Holotype: MNRJ 12490.

Distribution: Coastal rivers from Santa Catarina to São Paulo States, Brazil (de Pinna & Wosiacki, 2003).

TRIDENS Eigenmann & Eigenmann, 1889

Tridens Eigenmann & Eigenmann, 1889b: 53. Type species: *Tridens melanops* Eigenmann & Eigenmann, 1889.

Type by original designation. Gender: Masculine.

Tridens melanops Eigenmann & Eigenmann, 1889

Tridens melanops Eigenmann & Eigenmann, 1889b: 53. Type locality: Iça [Brazil]. Syntypes (27): CAS 64598 (1), BMNH 1889.11.14.73 (1), MCZ 8137 (1), MCZ 1566385 (6), MCZ 156639 (2, c&s), MZUSP uncat [ex. MCZ 156638] (1), USNM 41522 (1), USNM 120296 (3).

Distribution: Amazon River basin, Brazil (de Pinna & Wosiacki, 2003).

TRIDENSIMILIS Schultz, 1944

Tridensimilis Schultz, 1944c: 266. Type species: *Tridensimilis venezuelae* Schultz, 1944. Type by original designation. Gender: Masculine.

Tridensimilis brevis (Eigenmann & Eigenmann, 1889)

Tridens brevis Eigenmann & Eigenmann, 1889b: 54. Type locality: Tabatinga [Brazil]. Holotype: MCZ 8160. Distribution: Amazon River basin, Brazil (de Pinna & Wosiacki, 2003).

Tridensimilis venezuelae Schultz, 1944

Tridensimilis venezuelae Schultz, 1944c: 267, pl. 6 (fig. c). Type locality: Río Negro, below the mouth of the Río Yasa, Maracaibo basin, Venezuela. Holotype: USNM 121290.

Distribution: Orinoco River basin, Venezuela (de Pinna & Wosiacki, 2003).

TRIDENTOPSIS Myers, 1925

Tridentopsis Myers, 1925b: 84. Type species: *Tridentopsis pearsoni* Myers, 1925. Type by original designation. Gender: Feminine.

Tridentopsis cahuali Azpelicueta, 1990

Tridentopsis cahuali Azpelicueta, 1990: 982, figs. 1–4. Type locality: Estancia El Bagual, Formosa Province, Argentina, 26°10'53"S, 58°56'39"W. Holotype: MLP 5-IX-89-1.

Distribution: Paraguay River basin, Argentina (de Pinna & Wosiacki, 2003).

Tridentopsis pearsoni Myers, 1925

Tridentopsis pearsoni Myers, 1925b: 84. Type locality: Lagoons at Lake Rogoagua, Bolivia. Holotype: CAS 28258. Distribution: Upper Amazon River basin, Bolivia (de Pinna & Wosiacki, 2003).

Tridentopsis tocantinsi La Monte, 1939

Tridentopsis tocantinsi La Monte, 1939: 1. Type locality: Rio Tocantins, northeastern Brazil. Holotype: AMNH 13967.

Distribution: Tocantins River basin, Brazil (de Pinna & Wosiacki, 2003).

TYPHLOBELUS Myers, 1944

Typhlobelus Myers, 1944: 593. Type species: *Typhlobelus ternetzi* Myers, 1944. Type by original designation. Gender: Masculine.

Key: Schaefer *et al.* (2005).

Typhlobelus guacamaya Schaefer, Provenzano, de Pinna & Baskin, 2005

Typhlobelus guacamaya Schaefer, Provenzano, de Pinna & Baskin, 2005: 14, figs. 7, 8. Type locality: Venezuela, Estado Amazonas, Río Cuao at Raudal Guacamaya, 8.1 miles upstream from Raudal El Danto, 05°07.71'N, 67°31.53'W. Holotype: MBUCV V-30936.

Distribution: Cuao River, Orinoco River basin, Venezuela (Schaefer *et al.*, 2005).

Typhlobelus lundbergi Schaefer, Provenzano, de Pinna & Baskin, 2005

Typhlobelus lundbergi Schaefer, Provenzano, de Pinna & Baskin, 2005: 10, figs. 5, 6. Type locality: Venezuela, Estado Delta Amacuro, Río Orinoco at Los Castillos, upstream of east end of Caño Limón, 8°31.2'N, 62°35.1'W. Holotype: MBUCV V-31040.

Distribution: Lower Orinoco River, between Ciudad Bolívar and Los Castillos, Venezuela (Schaefer *et al.*, 2005).

Typhlobelus macromycterus Costa & Bockmann, 1994

Typhlobelus macromycterus Costa & Bockmann, 1994b: 68, figs. 1–3. Type locality: Brazil: Estado do Pará, Rio Tocantins near Tucuruí. Holotype: MNRJ 12129.

Distribution: Tocantins River near Tucuruí, Pará State, Brazil (de Pinna & Wosiacki, 2003).

***Typhlobelus ternetzi* Myers, 1944**

Typhlobelus ternetzi Myers, 1944: 593, pl. 52 (fig. 2), pl. 53 (figs. 6–8). Type locality: Rock pools below São Gabriel Rapids, Rio Negro, Brazil. Holotype: CAS 11118.

Distribution: Upper Negro River basin, Brazil (de Pinna & Wosiacki, 2003).

***VANDELLIA* Valenciennes, 1846**

Vandellia Valenciennes, in Cuvier & Valenciennes, 1846: 386. Type species: *Vandellia cirrhosa* Valenciennes, 1846.

Type by monotypy. Gender: Feminine.

Urinophilus Eigenmann, 1918a: 358. Type species: *Vandellia sanguinea* Eigenmann, 1918. Type by subsequent designation by Eigenmann (1920d: 441). Gender: Masculine. Originally proposed without any included species; species first assigned, and a type species designated, in Eigenmann (1920d).

***Vandellia beccarii* Di Capriacco, 1935**

Vandellia beccarii Di Capriacco, 1935: 59. Type locality: Rockstone, in flumine Essequibo dicto, in Guiana Britanica. Holotype: MZUF 5506.

Distribution: Orinoco River basin and rivers of Guyana (de Pinna & Wosiacki, 2003).

Remarks: Redescribed in Schmidt (1987).

***Vandellia cirrhosa* Valenciennes, 1846**

Vandellia cirrhosa Valenciennes, in Cuvier & Valenciennes, 1846: 386, pl. 547. Type locality: [Probably from America]. Syntypes (2): MNHN a-6308.

Vandellia gigantea Cornalia, 1849: 15, figs. 4–5. Type locality: [Fl. Amazonum et Napo]. Type(s): lost (Cagnolaro & Violani, 1988).

Vandellia plazaii Castelnau, 1855: 51, pl. 28 (fig. 1). Type locality: rio Ucayale (Pérou). Holotype: MNHN a-6309.

Vandellia Balzanii Perugia, 1897: 23. Type locality: Rio Beni, Missioni Mosetenes, Bolivia. Holotype: MSNG 8848.

Urinophilus erythrurus Eigenmann, 1922b: 114, pls. 3 (figs. 5–7), 4 (10–16). Type locality: Río Morona, Peru. Holotype: CAS 64599.

Distribution: Amazon River basin (de Pinna & Wosiacki, 2003).

Remarks: *Vandellia gigantea* Cornalia, 1849, was treated as a *nomen oblitum* by Cagnolaro & Violani (1988), but considered to be available, albeit a junior synonym, in de Pinna & Wosiacki (2003).

***Vandellia sanguinea* Eigenmann, 1918**

Vandellia sanguinea Eigenmann, 1918c: 701. Type locality: San Antonio de Rio Madeira. Holotype: FMNH 58086; holotype illustrated in Eigenmann (1918a: 365, pl. 53, fig. 2).

Distribution: Amazon, Orinoco and Essequibo River basins (de Pinna & Wosiacki, 2003).

Genus *inquirendum*, Trichomycteridae

Pygidium Meyen, 1835, in Meyen, 1834–35: 475. Type species: *Pygidium fuscum* Meyen, 1835. Type by monotypy. Gender: Neuter. Familial assignment of this genus is uncertain (cf. Tchernavin, 1944).

Remarks: The family group name Pygidiidae was proposed in Eigenmann & Eigenmann (1888a: 649) on *Pygidium* Meyen, 1835.

Species *inquirenda*, Trichomycteridae

Pygidium fuscum Meyen, 1835, in Meyen, 1834–35: 369. Type locality: Peru. Holotype: Whereabouts unknown. Familial assignment of this species is uncertain (see Tchernavin, 1944).

UNPLACED NAMES

SILURIFORMES *Incertae sedis*

† **BACHMANNIA** Dolgopol de Saez, 1945

† *Bachmannia* Dolgopol de Saez, 1945: 453. Type species: † *Bachmannia chabutensis* Dolgopol de Saez, 1945. Type by Monotypy. Gender: Feminine.

† ***Bachmannia chabutensis*** Dolgopol de Saez, 1945

† *Bachmannia chabutensis* Dolgopol de Saez, 1945: 453, figure on p. 453. Type locality: de la laguna del Hunco y del Mirador de Chubut [Argentina]; Tertiary. Holotype: MLP 40-V-17-1a (impression).

† *Arius argentinus* Dolgopol de Saez, 1945: 455, figure on p. 455. Type locality: La Laguna del Huncho y del Niradore, Chabut, Argentina; Tertiary. Holotype: MLP 40-V-17-3 (impression of cranium and partial vertebral column).

Distribution: Miocene of Argentina (Gayet & Meunier, 2003).

Remarks: Synonymy from Gayet & Meunier (2003), which appears to be based on Arratia & Cione (1996).

CONORHYNCHOS Bleeker, 1858

Conostome Duméril, 1856: 484. Type species: *Pimelodus conirostris* Cuvier, 1836. Type by original designation. Gender: Feminine. Possibly preoccupied by *Conostoma* Hodgson, 1842, in birds and not treated as valid in recent times. Considered a *nomen oblitum*.

Conorhynchos Bleeker, 1858: 191, 205, 209. Type species: *Pimelodus conirostris* Cuvier, 1836. Type by monotypy. Gender: Neuter. New name for *Conostome* Duméril, which is apparently preoccupied by *Conostoma* Hodgson, 1842, in birds. Name often misspelled *Conorhynchus* following the spelling in Bleeker (1862-63).

Conorhynchus Bleeker, 1863 (in Bleeker, 1862-63): 12. Type species: *Pimelodus conirostris* Cuvier, 1836. Type by original designation. New spelling for *Conorhynchos*. Preoccupied by *Conorhynchus* Motschousky, 1860, in Coleoptera.

Conorhynchichthys Regan, 1908, in Regan, 1906-08: 192. Type species: *Pimelodus conirostris* Cuvier, 1836. Type by being a replacement name. Gender: Masculine. Replacement for *Conorhynchus* Bleeker, 1863 [= *Conorhynchos* Bleeker, 1858], preoccupied by *Conorhynchus* Motschousky, 1860.

Remarks: Initially included in the Pimelodidae, but that placement questioned in Reis *et al.* (2003).

Conorhynchos conirostris (Cuvier, 1836)

Pimelodus conirostris Cuvier, 1836: 541, footnote 12. Type locality: rivière de Saint-François. Holotype: MNHN a-9413. Described and illustrated in Cuvier & Valenciennes (1840b: 204 (156 in Strasbourg deluxe edition), pl. 436).

Conorhynchus glaber Steindachner, 1877b: 637, pl. 8. Type locality: einem flusse bei Porto Seguro [apparently in error]. Holotype: at NMW.

Distribution: São Francisco River basin, Brazil.

† **EOPEYERIA** Whitley, 1947

† *Ariopsis* Peyer, 1928: 43. Type species: † *Ariopsis aegyptiacus* Peyer, 1928. Type by monotypy. Gender: Feminine. Preoccupied by *Ariopsis* Gill, 1861, in Recent fishes. Replaced by † *Peyeria* Whitley, 1940.

† *Peyeria* Whitley, 1940a: 242. Type species: † *Ariopsis aegyptiacus* Peyer, 1928. Type by being a replacement name. Gender: Feminine. Replacement for † *Ariopsis* Peyer, 1928, preoccupied by *Ariopsis* Gill, 1861, in Recent fishes. But, † *Peyeria* Whitley preoccupied by † *Peyeria* Weiler, 1935, in fossil Chondrichthyes.

† *Eopeyeria* Whitley, 1947: 150. Type species: † *Ariopsis aegyptiacus* Peyer, 1928. Type by being a replacement name. Gender: Feminine. Replacement for † *Peyeria* Whitley, 1940, preoccupied by † *Peyeria* Weiler, 1935, in fossil Chondrichthyes.

† ***Eopeyeria aegyptiaca*** (Peyer, 1928)

† *Ariopsis aegyptiacus* Peyer, 1928: 43, pl. 5, pl. 6 (fig. 1). Type locality: Qasr-es-Sagha-Stufe, Norden des Fajum. Ägyptens; Eocene. Holotype: at Bayerische Staatssammlung, München.

Distribution: Fajum, Egypt; Eocene.

† **FAJUMIA** Stromer, 1904

† *Fajumia* Stromer, 1904: 3. Type species: † *Fajumia schweinfurthi* Stromer, 1904. Type by monotypy. Gender:

Feminine.

Remarks: Placed by Jordan (1923) into the Bagridae, but placed in the Ariidae by Peyer (1928). Gayet & Meunier (2003) place this genus as Siluriformes *incertae sedis*, possibly belonging within the Arioida.

† ***Fajumia menoni*** Sahni & Mishra, 1975

† *Fajumia menoni* Sahni & Mishra, 1975: 7, pl. 1 (figs. 3 a–c). Type locality: Bluish-grey stage (Middle Eocene) at Gabhatad, Western India; Middle-Eocene. Holotype: LUVF 11140 (cranium).

Distribution: Western India, middle Eocene (Sahni & Mishra, 1975).

Remarks: Placed in the Bagridae by Sahni & Mishra (1975), following placement of genus in that family by Jordan (1923).

† ***Fajumia misrai*** Sahni & Mishra, 1975

† *Fajumia misrai* Sahni & Mishra, 1975: 8, pl. 1 (figs. 4–5). Type locality: Bluish-grey shales of Babia Stage (Middle Eocene) at Nareda, Western India. Holotype: LUVF 11142 (cranium).

Distribution: Western India; middle Eocene (Sahni & Mishra, 1975).

Remarks: Placed in the Bagridae by Sahni & Mishra (1975), following placement of genus in that family by Jordan (1923).

† ***Fajumia schweinfurthi*** Stromer, 1904

† *Fajumia Schweinfurthi* Stromer, 1904: 3, pl. 1 (figs. 1–2). Type locality: Mitteleocän am Nordrande des Fajûm, Aegypten. Holotype: in Munich (?) at Bayerische Staatssammlung, München (nearly complete cranium).

Distribution: Fajum, Egypt; middle Eocene.

† ***Fajumia stromeri*** Peyer, 1928

† *Fajumia Stromeri* Peyer, 1928: 33, fig. 3, pl. 4 (fig. 1). Type locality: Qasr-es-Sagha-Stufe, Norden des Fajum, Ägyptens; Eocene. Holotype: at Bayerische Staatssammlung, München.

Distribution: Fajum, Egypt; middle Eocene.

HORABAGRUS Jayaram, 1955

Horabagrus Jayaram, 1955b: 261. Type species: *Pseudobagrus brachysoma* Günther, 1864. Type by original designation. Gender: Masculine.

Remarks: *Horabagrus* was initially placed in the Bagridae and has subsequently been suggested to be more closely related to species of the Schilbidae (K. C. Jayaram, pers. comm.) or as a separate family that also includes some species currently placed in the Schilbidae (de Pinna, 1998a, Hardmann, 2005).

Horabagrus brachysoma (Günther, 1864)

Pseudobagrus brachysoma Günther, 1864: 86. Type locality: Cochinchina. Holotype: BMNH 1862.9.18.5 (skin).

Pseudobagrus chryseus Day, 1865b: 290. Type locality: Kurriavanoor River, Kurriapudnam or Cochin, India. Holotype: Possibly AMS B.7922 (1), BMNH 1865.7.17.5-6 (2), MCZ 4280 (1), NMW 44177 (1), RMNH 6870 (1), ZMB 9575 (1), ZSI 475 (1, lost). Also described and illustrated in Day (1865a: 185, pl. 13, fig. 2).

Distribution: Vernbanad Lake and estuaries of Kerala and southwestern Karnataka, India (Pethiyagoda & Kottelat, 1994).

Remarks: See Jayaram (1952) for taxonomic comments.

Horabagrus nigricollaris Pethiyagoda & Kottelat, 1994

Horabagrus nigricollaris Pethiyagoda & Kottelat, 1994: 110, fig. 13. Type locality: India: Kerala: Chalakudy River, 26 km upstream of Chalakudy town, near Vettilappara. Holotype: AMS I.34198-001.

Distribution: Chalakudy River, Kerala, India (Pethiyagoda & Kottelat, 1994).

PHREATOBADIUS Goeldi, 1905

Phreatobius Goeldi, 1905: 549. Type species: *Phreatobius cisternarum* Goeldi, 1905. Type by monotypy. Gender: Masculine.

Remarks: Originally treated as a genus of the Trichomycteridae, in recent years this genus has been placed within the Heptapteridae (e. g., Bockmann & Guazzelli, 2003), following Buckup (1988) and the unpublished study by Bockmann (1998). However, Muriel-Cunha & de Pinna (2005) suggests that the placement of this genus and its single named species is unsettled. The family-group name Phreatobinae was proposed for this genus by Reichel

(1927: 383).

***Phreatobius cisternarum* Goeldi, 1905**

Phreatobius cisternarum Goeldi, 1905: 549. Type locality: Wasser einer Binnenlandzisterne tief im Innern der Mündung des Amazonenstromes vorgelagerten Rieseninsel Marajó [Brazil]. Syntypes (2): MHNG 2623.30 (1), plus one of the following: FMNH 58580 (1), MHNG 1213.97 (1), or MHNG 1505.91 (2 entire specimens plus one sectioned and mounted on microscopic preparations).

Distribution: Marajó island, mouth of the Amazon River, Brazil, in cisterns (Bockmann & Guazzelli, 2003).

Remarks: See Reichel (1927), Carvalho (1967) and Muriel-Cunha & de Pinna (2005) for further information on this enigmatic species.

† *PLIOSILURUS* Weiler, 1956.

† *Pliosilurus* Weiler, 1956: 180. Type species: † *Pliosilurus primus* Weiler, 1956. Type by monotypy. Gender: Masculine.

Remarks: Listed as † *Pliosilurus primus* gen. & sp. nov., which, if published before 1931, would have made the type designation by original designation, but not after 1930.

† *Pliosilurus primus* Weiler, 1956

† *Pliosilurus primus* Weiler, 1956: 180, figs. 1–2; pl. 11. Type locality: Willershausen bek Göttingen [Pliocene, Germany]. Holotype: Geologischen Instituts der Universität Göttingen 4836 (head, pectoral girdle, portion of vertebral column and anal fin).

Distribution: Germany; Pliocene.

† *RHINEASTES* Cope, 1872

† *Rhineastes* Cope, 1872b: 486. Type species: † *Rhineastes peltatus* Cope, 1872. Type by original designation. Gender: Masculine.

Remarks: † *Rhineastes* has generally been placed in the Ariidae, but Lundberg (1992: 396) considered that placement in error and, instead, treated the genus as *Incatae sedis* in the Siluriformes, which is followed here.

† *Rhineastes peltatus* Cope, 1872

† *Rhineastes peltatus* Cope, 1872b: 486. Type locality: Bridger Formation, South Bitter Creek, Wyoming. Holotype USNM 3984 (supraoccipital bone and dorsal spine); holotype illustrated in Cope (1884b: 63, pl. 5, figs. 1–2).

Distribution: Bridger Formation, Wyoming; known only from type locality (Lundberg, 1975).

† *Rhineastes smithii* Cope, 1872

† *Rhineastes smithii* Cope, 1872b: 486. Type locality: Bridger Formation, South Bitter Creek, Wyoming. Holotype: USNM 3978 (pectoral spine); holotype illustrated in Cope (1884b: 64, pl. 5, figs. 5–11).

Distribution: Bridger Formation, Wyoming (Lundberg, 1975).

Remarks: Inclusion in † *Rhineastes* considered tentative by Lundberg (1975).

Species inquirenda*, † *Rhineastes

† *Rhineastes radulus* Cope, 1873: 639. Type locality: Bridger Formation at Cottonwood Creek, Wyoming; Eocene. Holotype: USNM 4099 (fragments); holotype illustrated in Cope (1884b: 67, pl. 5, figs. 14–17). Identity uncertain (Lundberg, 1975).

† *SOCNOPAEA* Stromer, 1904

† *Socnopaea* Stromer, 1904: 6. Type species: † *Socnopaea grandis* Stromer, 1904. Type by monotypy. Gender: Feminine.

Remarks: Jordan (1923) placed this genus and † *Fajumia* into the Bagridae, but Peyer (1928) placed both genera in the Ariidae.

† *Socnopaea grandis* Stromer, 1904

† *Socnopaea grandis* Stromer, 1904: 6, pl. 1 (fig. 3). Type locality: Mitteleocän am Nordrande des Fajûm, Aegypten. Holotype: in Munich (partial cranium).

Distribution: Fajum, Egypt; middle Eocene.

† *Socnopaea horai* Sahni & Mishra, 1975

† *Socnopaea horai* Sahni & Mishra, 1975: 10, pl. 2 (fig. 1). Type locality: Ossiferus gypseous shales of Babia stage (Middle Eocene) at Harudi, Western India. Holotype: LUVP 11145 (cranium).

Genera inquirendae, Siluriformes

Tachysurus La Cepède, 1803: 150. Type species: *Tachysurus sinensis* La Cepède, 1803. Type by monotypy. Gender: Masculine. The type species is considered to be an unrecognizable siluriform by Wheeler & Baddokwya (1981) and Taylor (1986a).

Laimumena Sauvage, 1884a: 147. Type species: *Laimumena barbonica* Sauvage, 1884. Type by monotypy. Gender: Feminine. Description of type species is not recognizable to genus or even family.

Trachymochlus Hoedeman, 1961: 137. Type species: *Trachymochlus cupido* Hoedeman, 1961. Type by monotypy. Gender: Masculine.

Species inquirendae, Siluriformes

Silurus undecimalis Linnaeus, 1758: 305. Type locality: Not stated. Type(s): originally part of collection at Museum Adolphi Friderici but whereabouts unknown, apparently not at NRM. Type locality stated as "Surinami" in Linnaeus (1764: 97).

Silurus luvur Molina, 1782: 346. Type locality: Chile. No types known.

Silurus schilby Sonnini, 1799: 256, pl. 23 (fig. 1). Type locality: Nil. On p. 295 there is note about *Silurus schilbe niloticus* of Hasselquist that suggests that this may not be intended to be the proposal of a new name.

Silurus gurgu Bloch & Schneider, 1801: 388. Type locality: in Nilo. No types known, based on literature account. Proposed conditionally.

Silurus minutus Schneider, in Bloch & Schneider, 1801: 389. Type locality: India. No types known, based on unpublished literature source. Proposed conditionally.

Tachysurus sinensis La Cepède, 1803: 150, 151, pl. 5 (fig. 2). Type locality: China. No types known; based on a Chinese painting; illustration in La Cepède reproduced in Wheeler & Baddokwya (1981). Treated in many Chinese works as valid in *Arius*, but Wheeler & Baddokwya (1981) and Taylor (1986a) indicate that it is an unrecognizable siluriform from the description and/or illustration.

Silurus ichneumon Hermann, 1804: 309. Type locality: Cairo. No types known.

Pimelodes fossor Lichtenstein, 1823: 112. Type locality: Cape of Good Hope, South Africa. Type(s): at ZMB.

Pimelodus octocirrhus Cuvier, 1829: 294. Type locality: not stated. Available from footnote as, "Pim. octocirrhus, N., Seb., III, xxix, 1" [= Seba (1734–65: pl. 29, fig. 1)].

Pimelodus javus Valenciennes, in Cuvier & Valenciennes, 1840b: 187 (139 in Strasbourg deluxe edition). Type locality: Java. Holotype (4 ½ pouces): at MNHN. Status uncertain (Roberts, 1993: 47).

Pimelodus tachisurus Valenciennes, in Cuvier & Valenciennes, 1840b: 163 (121 of Strasbourg deluxe edition). Type locality: China. No types known; based on a Chinese painting. Unneeded new name for *Tachisurus chinensis* [sic, for *sinensis*] La Cepède.

Pimelodus Cantonensis Valenciennes, in Cuvier & Valenciennes, 1840b: 142 (106 of Strasbourg deluxe edition). Type locality: Canton [China]. No types known. Based solely on an illustration.

Bagrus chinta Valenciennes, in Cuvier & Valenciennes, 1840a: 445 (330 of Strasbourg deluxe edition). Type locality: [Vizagapatam]. No types known, based on account and illustration in Russell (1803: pl. 167).

Pimelodus pusillus Ranzani, 1842: 332, pl. 27. Type locality: Unknown [apparently Brazil, based on other species in publication]. Holotype: MZUB 933. Name appeared first in Ranzani (1841: 64) as a *nomen nudum*.

† *Pimelodus Sadleri* Heckel, 1849: 19. Type locality: Bihar Comitat, Hungary; Miocene. Holotype? (fin ray fragments). Illustrated and described in more detail in Heckel (1850: 213, pl. 16, fig. 3).

Silurus ferox Gronow, in Gray, 1854: 135. Type locality: not stated. No types known.

Rhamdia javanica Bleeker, 1858b: 139. Type locality: Java. Holotype: at MNHN. Unjustified emendation of *Pimelodus javus* Valenciennes, 1840.

- Laimumena barbonica* Sauvage, 1884a: 147. Type locality: Réunion. No types known. Not found in catalogs of MNHN (pers. obs.) and not seen on shelves.
- † *Silurus Serdicensis* Toula, 1889: 108, pl. 9. Type locality: im Westen von Sofia. Upper Tertiary. Syntypes: (fragmentary remains).
- † *Silurus gaudryi* Leriche, 1900: 181, pl. 1, figs. 7–12. Type locality: Cuis, Monthelon [Marne, Lower Eocene]. Types: at laboratoire de Géologie de l'Université de Lille (dorsal fin spines, pectoral fin spine). Originally *Silurus (?)gaudryi*. Remarks: As *Pimelodus gaudryi* in Leriche (1901:165).
- † *Silurus pliocaenicus* Leidenfrost, 1925, 121, text fig. 2 and pl. 6. Type locality: pannonisch-pontischen Tonen von Rákos bei Budapest [Hungary, Pliocene]. Holotype: at Samml. der Kön. ung. Geologischen Anstalt, Budapest, partial neurocranium. Remarks: Possibly a bagrid (S. L.), or a silurid (Gayet & Meunier, 2003: 503).
- † *Silurus stenocephalus* Leidenfrost, 1925, 122, pl. 7. Type locality: Pannonisch-pontischen Tonen von Rákos bei Budapest [Hungary, Pliocene]. Holotype: at Samml. der Kön. ung. Geologischen Anstalt, Budapest, partial neurocranium.
- † *Rhineastes grangeri* Hussakof, 1932: 17, fig. 26. Type locality: Tung Gur beds, 50 miles southeast of Iren Dabasu, Inner Mongolia; Pliocene. Holotype: at AMNH (right pectoral fin spine). Distribution: Known only from the type locality, in central Asia.
- Trachymochlus cupido* Hoedeman, 1961: 137, fig. 4. Type locality: Nickerie, entrance of Cupido creek into Maratakka River, Suriname. Holotype: ZMA 102236.
- † *Schmidelia graciliformis* Berry. [no other information; from Dolgopol de Saez (1945)].
- † *Clarias pliocaenicus* Sauvage (in Depéret, 1885). [no other information; from Gayet & Meunier (2003)]
- † *Ariopsis peyeria* Eocene, North Africa [no additional information, From Romer. Probably a mistake for *Ariopsis Peyer*]

Names wrongly treated as Siluriformes

- † *Arius lemoinei* of Eocene of France [No additional information] Remarks: Considered an acipenserid by Gayet & Meunier (2003).
- † *BRACHYSPONDYLUS* Marck, in Marck & Schläter, 1868: 283. Type species: † *Brachyspondylus cretaceus* Marck, 1868. Type by monotypy. Gender: Masculine. Remarks: Listed by Jordan (1923: 643) as a cyprinid with †*Brachyspondylus saropterix* Marck, 1876, as orthotype. *Brachyspondylus saropterix* has been considered to be a siluriform taxon, most recently by Gayet & Meunier (2003), but † *Brachyspondylus* is actually available from Marck & Schläter (1868: 283), with † *Brachyspondylus cretaceus* (283, pl. 43, fig. 2) as type by monotypy. This species is a scaly fish, probably a cyprinid, so that † *Brachyspondylus* is not a catfish name. See Sanders (1934: 30) for discussion of the taxonomic history of this name.
- † *BUCKLANDIUM* König, 1825
- † *Bucklandium* König, 1825: 4. Type species: † *Bucklandium diluvii* König, 1825. Type by monotypy. Gender: Neuter.
- † *Glyptocephalus* Agassiz, 1843 (in Agassiz, 1833–43): 264. Type species: † *Glyptocephalus radiatus* Agassiz, 1843. Type by monotypy. Gender: Masculine. Preoccupied by *Glyptocephalus* Gottsche, 1834, in Recent fishes (Pleuronectidae).
- † *Glyptocara* Gill, 1888: 926. Replacement for † *Glyptocephalus* Agassiz, 1844, preoccupied by *Glyptocephalus* Gottsche, 1834.
- Remarks: The name *Ephippus Owenii* appears in Agassiz (1843 (in Agassiz, 1833–43): 264) in the account of the newly proposed name *Glyptocephalus radiatus*. It is not treated as a valid name and is therefore not available from this usage. Gayet & Meunier (2003) place † *Bucklandium* as Siluriformes incertae sedis, possibly belonging within the Arioida, which appears to follow Woodward (1889b: 208), in which the name is treated as a siluriform fish near to *Auchenoglanis*. However, the illustration provided by Woodward is apparently not that of a

siluriform fish and after close examination of the type specimen, Ralf Britz (pers. commun.) concluded that the bones are definitely not those of a catfish.

† *Bucklandium diluvii* König, 1825

† *Bucklandium diluvii* König, 1825: 4, pl. 8, no. 91. Type locality: Isle of Sheppey, Lower Eocene, London Clay.

Holotype: BMNH P.9230 (cranium and pectoral arch). Redescribed and illustrated in Woodward (1889b: 208, pl. 22).

† *Glyptocephalus radiatus* Agassiz, 1843 (in Agassiz, 1833–43): 264. Type locality: [Argiles de Londres] Sheppy.

Holotype: BMNH P.9230 (cranium and pectoral arch).

Distribution: London Clay, Lower Eocene.

Remarks: † *Glyptocephalus radiatus* Agassiz, 1843, is an unneeded new name for † *Bucklandium diluvii* König, 1825, which Agassiz apparently did not associate with the examined specimen.

† *PROPYGIDIUM* Bocchino, 1964: 186. Type species: † *Propygidium primaevus* Bocchino, 1964. Type by original designation. Gender: Neuter.

Remarks: Although originally described as a member of the siluriform family Trichomycteridae, the material was reexamined and interpreted as a member of the Perciformes by Cione & Torno (1988).

† *Propygidium primaevus* Bocchino, 1964: 186, fig. on p. 189. Type locality: Serie Andesítica Lask Bayas, to the west of Rio Negro Province, Argentina; Eocene/Oligocene. [Corrected to: Cerro David, 0.5 km from the road from San Carlos de Bariloche to Chenqueniyeu, to the west of the Province of Rio Negro, from the middle levels of the Nirihuau Formation (Upper Oligocene-Lower Miocene), by Cione & Torno (1988)]. Holotype: División de Paleontología Vertebrados del Museo de La Plata 21-871, partial skeletal impression.

† *TREWAVASIA* White & Moy Thomas, 1941

† *Xenopholis* Davis, 1887: 548. Type species: † *Xenopholis carinatus* Davis, 1887, by monotypy. Gender: Feminine.

Preoccupied by *Xenopholis* Peters, 1869, in Reptiles; replaced by † *Trewavasia* White & Moy Thomas, 1941, and †*Xenopholoides* Fowler, 1958.

† *Trewavasia* White & Moy Thomas, 1941: 400. Type species: † *Xenopholis carinatus* Davis, 1887. Gender: Feminine. Type by being a replacement name. Replacement for † *Xenopholis* Davis, 1887; preoccupied by *Xenopholis* Peters, 1869, in Reptiles. Gender: Feminine.

† *Xenopholoides* Fowler, 1958: 13. Type species: † *Xenopholis carinatus* Davis, 1887. Gender: Feminine. Type by being a replacement name. Replacement for † *Xenopholis* Davis, 1887; preoccupied by *Xenopholis* Peters, 1869, in Reptiles.

Remarks: Described by Davis (1887) as probably showing a “closer connection with siluroids than with any other group” but clearly not a siluriform fish. Most likely an Acanthopterygian.

† *Trewavasia carinata* (Davis, 1887)

† *Xenopholis carinatus* Davis, 1887: 549, pl. 29, fig. 4. Type locality: Mt. Lebanon, Syria, Hard Chalk.

Appendix 1. Siluriform genera and species named from otoliths. Unless otherwise noted, all names were found in Weiler's (1968) catalogue of fish otoliths as belonging to catfishes

Generic names

† *Claibornichthys* Frizzel & Dante, 1965: 697. Type species: † *Claibornichthys troelli* Frizzel & Dante, 1965. Type by original designation. Gender: Masculine.

Remarks: † *Otolithus (Sciaenidarum) decipiens* Koken, 1888, tentatively included as a second species of this genus by Frizzel & Dante (1965).

† *Vorhisia* Frizzell, 1965: 179. Type species: † *Vorhisia vulpes* Frizzell, 1965. Type by original designation. Gender: Feminine.

Remarks: The family group name † Vorhisidae was proposed in the same publication.

Species names

(Names listed by date of publication)

† *Otolithus (incertae sedis) crassus* Koken, 1884: 559, pl. 12, fig. 13. Type locality: Heaton Hill, Isle of Wight. Holotype: at Roy. Mus. Nat. Hist. Berlin (otolith), but apparently lost (Stinton, 1977: 71).

Remarks: Placed in *Arius* by Weiler (1968). Treated as valid as † *Tachysurus crassus* by Stinton (1977: 71).

† *Otolithus (Sciaenidarum) decipiens* Koken, 1888: 285, pl. 19 (figs. 5-6). Type locality: Calyborne-Schichten. Syntypes (2): Whereabouts unknown..

Remarks: Not listed among siluriform otolith based names in Weiler (1968), but treated a possible second species of † *Claibornichthys* (as † *Otolithus (Arius) decipiens*) in Frizzel & Dante (1965).

† *Arius baroni* Newton, 1889: 207, pl. 21, fig. 7. Type locality: Ankoala, Madagascar; Eocene. Holotype: at BMNH, in the Baron collection (1 of 8 otoliths). Name available from figure caption, thus illustrated specimen is the holotype.

† *Raja similis* Woodward, 1899a: 86, pl. 4, figs. 4-5. Type locality: Hampshire, England; Eocene. (Otoliths).

Remarks: Name corrected to † *Arius similis* on plate caption. Treated as a synonym of † *Tachysurus crassus* (Koken) by Stinton (1977: 71).

† *Otolithus (Arius) danicus* Koken, 1891: 81, fig. 1. Type locality: Copenhagen; lower Eocene. Based on otolith identified as † *Otolithus cf. crassus* in Koken (1885: 116, pl. 5, fig. 29).

† *Otolithus (Arius) germanicus* Koken, 1891: 81, pl. 1, fig. 3, and pl. 6, fig. 8. Type locality: Lattorf, Westeregeln, and Osterweddingen, Lower Oligocene; Sollingern, Weinheim, Waldboeckelheim, Middle Oligocene. (otoliths).

† *Otolithus (Arius) vanigonis* Koken, 1891: 81, pl. 6, fig. 4. Type locality: Mitteloligöcan, Waldböeckelheim.

† *Otolithus (Arius) Lerichei* Priem, 1906: 277, figs. 46-47. Type locality: problemant du Thanétien ou Yprésien ? des environs de Reims, éocènes du bassin Parisien. Holotype (partial Lapillus): Collection Bourdot.

† *Otolithus (Siluridarum ?) incertus* Priem, 1906: 277, figs. 48-49. Type locality: problemant d'Hérouval (Yprésien supérieur), éocènes du bassin Parisien. Holotype: ? MNHN.

† *Otolithus (Arius ?) moravicus* Schubert, 1908: 106, abb. 3. Type locality: Marnes de Pausram (Lower Oligocene). Holotype: GBW 1908/01/4.

Remarks: Nolf (1981: 136) listed this name as a rejected species.

† *Otolithus (Arius) angelicus* Bassoli, 1909: 41, fig. 1. Type locality: Headon Member, Solent formation, Brockenhurst, Hampshire, Britain. Holotype: (Lapillus).

Remarks: Treated as valid as † *Galeichthys angelicus* by Stinton (1977: 73).

† *Otolithus (Arius) crassus bartonensis* Shepherd, 1916: 183, fig. 155 (5). Type locality: Ober- Eozän, England. Originally † *Otolithus (Arius) crassus* var. *bartonensis*.

Remarks: Treated as a synonym of † *Tachysurus crassus* (Koken) by Stinton (1977: 71).

† *Otolithus (Arius) danicus bartonensis* Shepherd, 1916: 180, fig. 157 (11). Type locality: Upper Eocene, England. Originally † *Otolithus (Arius) danicus* var. *bartonensis*.

Remarks: Treated as a synonym of † *Tachysurus planus* (Frost), as well † *Tachysurus crassus* (Koken) as by Stinton

(1977: 70).

† *Otolithus (Arius?) parvus* Schubert, 1916: 287, pl. 7 (fig. 24). Type locality: Barton Formation (Upper Eocene) at Barton Cliff, England. Syntypes: GBW 1916/01/25, 25a.

Remarks: Treated as a synonym of † *Tachysurus crassus* (Koken) by Stinton (1977: 71). Nolf (1981: 136) listed this name as a rejected species.

† *Otolithus (Arius) newtoni* Schubert, 1916: 286, pl. 7 (fig. 3). Type locality: Barton Formation (Upper Eocene) at Barton Cliff, England. Syntypes: GBW 1916/01/3 (or MGB 0427, see Stinton, 1977: 72).

Remarks: Treated as a synonym of † *Tachysurus crassus* (Koken) by Stinton (1977: 71). Nolf (1981: 136) treated this name as a synonym of † *Arius crassus* (Koken).

† *Otolithus (Arius) tenuis* Frost, 1925: 28, pl. 2 (fig. 37). Type locality: Hoofdsossiel-zone, Al. Geureugah (Nisam), Sumatra; upper Tertiary (Neogene). Holotype: (? Lapillus).

Remarks: Name spelled † *Otolithus (Arius) tenuis* on plate, which is treated here as an invalid name.

† *Otolithus (Arius) africanus* Frost, 1926: 84, pl. 18, figs. 7–8. Type locality: Ameki, Nigeria; Eocene. Syntypes (5): (Lapillus).

† *Otolithus (Arius) amekiensis* Frost, 1926: 84, pl. 18, fig. 10. Type locality: Ameki, Nigeria; Eocene. Holotype: (Lapillus).

† *Otolithus (Arius) angulatus* Frost, 1926: 84, pl. 18, fig. 9. Type locality: Ameki, Nigeria; Eocene. Syntypes (3): (Lapillus).

† *Otolithus (Arius?) glaber* Voigt, 1926: 177, pl. 2, figs. 23–24. Type locality: Bavaria, Cretaceous.

† *Otolithus (Arius) jaekeli* Richter 1928: 138, pl. 1, fig. 3. Type locality: Pomerania, Cretaceous.

† *Otolithus (Arius) aequus* Frost, 1934: 504, pl. 14 (fig. 15). Type locality: Barton Formation, Barton, Hampshire; Upper Eocene. Holotype: BMNH P 22732 (Lapillus).

Remarks: Treated as valid as † *Galeichthys aequus* by Stinton (1977: 71).

† *Otolithus (Arius) planus* Frost, 1934: 504, pl. 14 (fig. 14). Type locality: Barton Formation, Barton, Hampshire; Upper Eocene. Holotype: BMNH P 22730 (otolith).

Remarks: Treated as valid as † *Tachysurus planus* by Stinton (1977: 70).

† *Arius rutschi* Casier, 1958: 22, pl. 2 (figs. 12, 15). Type locality: l'Ile de la Barbade; formation de Scotland superieure. Holotype: TLSS 80. (Lapillus).

† *Arius cavatus* Stinton, 1962: 82, pl. 19 (fig. 6). Type locality: Borneo, Lower Pliocene. Holotype: (Lapillus).

† *Tachysurus oblongus* Stinton, 1962: 81, pl. 19 (fig. 3). Type locality: Borneo, Miocene–Pliocene. Type(s): Whereabouts unknown.

† *Netuma radiata* Stinton, 1962: 81, pl. 1 (fig. 17). Type locality: Borneo, Pliocene.

† *Netuma regularis* Stinton, 1962: 82, pl. 19 (fig. 5). Type locality: Borneo, Pliocene.

† *Otolithus (Arius) rotundatus* Roedel, 1930: 52, pl. 1, fig. 17. Type locality: Köthen und Frankfurt [Germany], Eocene. Holotype: (Lapillus).

† *Vorhisia vulpes* Frizzell, 1965: 180, figs. 2, 3. Type locality: Upper Cretaceous (Maestrichtian): along left bank of Grand River; Sec. 19, T20N, R20W; Corson County, South Dakota. Holotype: Frizzell personal collection, no number (otolith: left lapillus).

Distribution: Fox Hill Formation: Irish Creek lithofacies, Bullhead lithofacies, and Colgate lithofacies, South Dakota, USA; Upper Cretaceous (Frizzell & Koenig, 1973).

† *Claibornichthys troelli* Frizzell & Dante, 1965: 697, pl. 86 (figs. 1, 2, 8, 17, 18, 19). Type locality: Stone City Beds: Brazos River, west of Bryan Texas. Holotype: USNM 23371 (otolith).

† *Diplomystes rudis* Stinton, 1966: 423, pl. 66, fig. 12. Type locality: London Clay, England.

Distribution: London Clay, England, Lower Eocene (Weiler, 1968).

Appendix 2. Institutional codes

AFY	Personal collection of Agustín Fernández-Yépez; portions now at SCN and MBUCV, Venezuela.
AI	Asociación Ictiológica, La Plata, Argentina.
AMG	Albany Museum, Grahamstown, South Africa.
AMNH	American Museum of Natural History, New York, U.S.A.
AMS	Australian Museum, Sydney, N. S. W., Australia.
ANSP	Academy of Natural Sciences, Philadelphia, Pennsylvania, U.S.A.
ASIZB	Academia Sinica Institute of Zoology, Beijing, China.
BCUE	Department of Biology, Ch'ongju University of Education, Korea.
BKNU	Kunsan National University, Department of Biology, Korea.
BMNH	Natural History Museum, London. Formerly British Museum (Natural History), U.K.
BSMP	Department of Agriculture, Bureau of Science, Manila, Philippines. Evidently entire collection destroyed during World War II.
CAS	California Academy of Sciences, San Francisco, California, U.S.A.
CM	Carnegie Museum, Pittsburgh, Pennsylvania, U.S.A. Now at FMNH.
CPUC	Colecciones Paleontológicas, Departamento Ciencias de la Tierra, Universidad de Concepción, Concepción, Chile.
CSIRO	Commonwealth Science & Industrial Research Organization, Division of Marine Research, Hobart, Tasmania, Australia.
CU	Cornell University Museum of Vertebrates, Ithaca, New York, U.S.A.
DGM	Paleontological collection of the Setor de Paleontologia, Departamento Nacional de Produção Mineral, Universidade do Estado do Rio de Janeiro, Brazil.
DVZUT	Department of Vertebrate Zoology, University of Tông-Hop, Hanoi, Vietnam.
DZSASP	Departamento de Zoologia, Secretaria da Agricultura, São Paulo, Brazil. Formerly Museu Paulista.
ECO-SC	El Colegio de la Frontera Sur, San Cristóbal, Chiapas, Mexico.
EEBP	Estação Experimental de Biologia e Piscicultura de Pirassununga, Brazil.
FAKU	Kyoto University, Department of Fisheries, Faculty of Agriculture, Japan.
FFSUC	Faculty of Forestry Sciences, Universidad de Chile, Ichthyological Collection, Santiago, Chile.
F/GUZ	Department of Zoology, Gauhati University, Guwahati, Assam, India.
FML	Instituto Fundacion Miguel Lillo, Tucuman, Argentina.
FMNH	Field Museum of Natural History, Chicago, Illinois, U.S.A.
FRLM	Faculty of Fisheries, Mie University, Fisheries Research Laboratory, Mie-ken, Japan.
GBW	Collection du Geologische Bundesanstalt, Vienna, Austria.
GCM	Government College, Department of Zoology, Lahore, Pakistan.
GSJ	Geological Society of Japan.
ICNMHN	Instituto de Ciencias Naturales, Museo de Historia Natural, Universidad Nacional de Colombia, Bogotá, Colombia. .
IHASW	Institute Hydrobiology Academy Sinica, Wuhan, China.
ILPLA	Museo de La Plata, Instituto de Limnología, La Plata, Argentina.
IMCN	Natural Sciences Museum Federico Carlos Lehmann V.- INCIVA Cali, Colombia
INHS	Illinois Natural History Survey, Champaign, Illinois, U.S.A.
INPA	Instituto Nacional de Pesquisas da Amazonia, Manaus, Amazonas, Brazil.
INVEMAR	Instituto de Investigaciones Marinas de Punta de Betín, Santa Marta, Colombia.
IRSNB	Institut Royal des Sciences Naturelles de Belgique, Brussels, Belgium.
IU	Indiana University, Bloomington, Indiana, U.S.A. Now distributed among several institutions, primarily CAS, UMMZ and USNM; many IU types remain unaccounted for.
IUQ	Laboratorio de Ictiología, Departamento de Biología, Universidad del Quindío, Armenia, Colombia.
IZUA	Universidad Austral de Chile, Instituto de Zoológica, Valdivia, Chile.
JFBM	Bell Museum Fish Collection at the University of Minnesota.

JNU	Department of Biology, Ji Nan University, China.
KIZ	Kunming Institute of Zoology, Chinese Academy of Sciences, Kunming, Yunnan, China.
KU	University of Kansas, Museum of Natural History, Lawrence, Kansas, U.S.A.
KUMF	Kasetsart University Museum of Fisheries, Bangkok, Thailand.
LACM	Los Angeles County Museum of Natural History, Los Angeles, California, U.S.A.
LIVCM	World Museum Liverpool, U.K.
LUVP	Vertebrate Palaeontology Laboratory, Geology Department, Lucknow University, Lucknow, India.
MACN	Museo Argentino de Ciencias Naturales, Bernardino Rivadavia, Capital Federal, Argentina.
MB	Universidade de Lisboa, Museu Bocage, Lisboa, Portugal.
MBLUZ	Museo de Biología de la Universidad del Zulia, Venezuela.
MBUCV	Universidad Central de Venezuela, Museo de Biología, Caracas, Venezuela.
MCN.USB	Museo de Ciencias Naturales, Universidad Simón Bolívar, Caracas, Venezuela.
MCNG	Museo de Ciencias Naturales, Guanare, Venezuela.
MCP	Pontifícia Universidade Católica do Rio Grande do Sul, Museu de Ciências, Rio Grande do Sul, Porto Alegre, Brazil.
MCZ	Museum of Comparative Zoology, Harvard University, Cambridge, Massachusetts, U.S.A.
MD	Museo do Dundo, Dundo, Angola.
MEPN	Escuela Politécnica Nacional, Quito, Ecuador.
MHNC	Museo de Historia Natural de Cochabamba, Bolivia.
MHNG	Museum d'Histoire Naturelle, Genève, Switzerland.
MHNL	Musée d'Histoire Naturelle du Luxembourg.
MHNLR	Museum d'Histoire Naturelle, La Rochelle, France.
MHNLS	Museo de Historia Natural de La Salle, Caracas, Venezuela.
MHNM	Museo de Historia Natural de Montevideo, Montevideo, Uruguay.
MHNN	Museum d'Histoire Naturelle, Neuchâtel, Switzerland.
MHV	Musée de Haute Volta, Ouagadougou, Upper Volta.
MIKU	Marine Biological Institute, Kyoto University, Japan. Specimens now at FAKU.
MLP	Museo de La Plata, La Plata, Argentina.
MNHN	Muséum National d'Histoire Naturelle, Paris, France.
MNHNP	Museo Nacional de Historia Natural de Paraguay.
MNKP	Museo de Historia Natural Noel Kempff Mercado, Santa Cruz, Bolivia.
MNRJ	Universidade Federal do Rio de Janeiro, Museu Nacional, Rio de Janeiro, Brazil.
MNSB	Museum of Natural Sciences, Budapest, Hungary.
MPEG	Museu Paraense 'Emílio Goeldi', Pará, Brazil.
MRAC	Musée Royal de l'Afrique Centrale, Tervuren, Belgium.
MRCN	Museu Rio-Grandense de Ciências Naturais, Brazil.
MSINR	Museum Sichuan Institute of Natural Resources, China.
MSNG	Museo Civico di Storia Naturale di Genova 'Giacomo Doria', Genova, Italy.
MSNM	Museo Civico di Storia Naturale, Milano, Italy.
MSUM	Michigan State University, University Museum, East Lansing, Michigan, U.S.A.
MTD	Staatliches Museum für Tierkunde, Dresden, Germany.
MUMF	Department of Life Sciences, Manipur University Museum of Fishes, Canchipur, Manipur, India.
MUSM	Museo de Historia Natural de la Universidad Nacional Mayor de San Marcos, Lima, Peru.
MZB	Museum Zoologicum Bogoriense, Bogor, Indonesia.
MZUF	Università di Firenze, Museo Zoologico della Specola, Firenze, Italy.
MZS	Université de Strasbourg, Musée de Zoologie, Strasbourg, France.
MZUSP	Universidade de São Paulo, Museu de Zoologia, São Paulo, Brazil.
MZUT	Università di Torino, Museo Zoologico, Torino, Italy.
NIFI	National Inland Fisheries Institute, Fish Taxonomy Division, Bangkok, Thailand.
NMBA	Naturhistorisches Museum Basel, Basel, Switzerland.

NMBE	Naturhistorisches Museum Bern, Bern, Switzerland.
NMC	National Museums of Canada, Ottawa, Canada.
NMK	National Museum, Nairobi, Kenya.
NMSL	National Museum, Sir Marcus Fernando Mawatha, Colombo, Sri Lanka.
NMSZ	National Museums of Scotland, Edinburgh, Scotland.
NMV	National Museum of Victoria, Melbourne, Victoria, Australia.
NMW	Naturhistorisches Museum, Vienna, Austria.
NRM	Swedish Museum of Natural History (Naturhistoriska Riksmuseet), Stockholm, Sweden.
NSMT	National Science Museum, Department of Zoology, Tokyo, Japan.
NTM	Northern Territory Museum of Arts & Sciences, Darwin, Northern Territory, Australia.
OSUS	Oklahoma State University, Department of Zoology, Stillwater, Oklahoma, U.S.A.
PEM	Port Elizabeth Museum, Port Elizabeth, South Africa.
QM	Queensland Museum, Brisbane, Queensland, Australia.
QVMS	Queen Victoria Museum, Harare, Zimbabwe.
RMNH	Rijksmuseum van Natuurlijke Historie, Leiden, Netherlands.
ROM	Royal Ontario Museum, Toronto, Ontario, Canada.
SAIAB	South African Institute for Aquatic Biodiversity; formerly J. L. B. Smith Institute of Ichthyology, Grahamstown, South Africa (RUSI).
SAM	South African Museum, Cape Town, South Africa.
SAMA	South Australian Museum, Adelaide, South Australia, Australia.
SBM	Sabah Museum, Sabah, East Malaysia.
SCNU	South China Normal University, Guangzhou, China.
SIUC	Southern Illinois University (Carbondale), University Museum, Carbondale, Illinois, U.S.A.
SMF	Natur-Museum und Forschungs-Institut Senckenberg, Frankfurt-am-Main, Germany.
SMK	Sarawak Museum, Kuching, Sarawak, Malaysia.
SMNS	Staatliches Museum fur Naturkunde in Stuttgart, Ludwigsburg, Germany.
SNMB	Slovak National Museum, Bratislava, Czechoslovakia.
SPNRI	Sichuan Province, Natural Resources Institute, China.
SRS/ZSI	Southern Regional Station, Zoological Survey of India, Madras, India.
STRI	Smithsonian Tropical Research Institute, Panama.
SU	Stanford University, Palo Alto, California, U.S.A. Specimens on long term loan to CAS.
TM	Transvaal Museum, Pretoria, South Africa.
TU	Tulane University, Department of Zoology, New Orleans, Louisiana, U.S.A.
UAB	Universidad Autonoma de Barcelona, Departamento de Biología, Barcelona, Spain.
UBJTL	Universidad Bogotá Jorge Tadeo Lozano, Colombia.
UFRJ	Universidade Federal do Rio de Janeiro, Brazil.
UG/CSBD	University of Guyana, Center for the study of Biological Diversity, Georgetown, Guyana.
UMMP	University of Michigan Museum of Paleontology, Ann Arbor, Michigan, U.S.A.
UMMZ	University of Michigan Museum of Zoology, Ann Arbor, Michigan, U.S.A.
UNAM	Universidad Nacional Autonoma de Mexico, Departamento de Zoologia, Instituto de Biologia, Mexico, DF, Mexico.
UO	University of Oregon, Corvallis, Oregon, U.S.A.
USNM	National Museum of Natural History, Washington, D.C., U.S.A. Formerly United States National Museum.
UW	University of Washington, College of Fisheries, Seattle, Washington, U.S.A.
WAM	Western Australian Museum, Perth, Western Australia, Australia.
ZFMK	Zoologisches Forschungsinstitut und Museum Alexander Koenig, Bonn, Germany.
ZIN	Zoological Institute, Academy of Sciences, St. Petersburg, Russia.
ZISP	Zoological Institute, Academy of Sciences, Leningrad, Russia.
ZMA	Universiteit van Amsterdam, Zoologisch Museum, Amsterdam, The Netherlands.
ZMB	Universitat Humboldt, Museum fur Naturkunde, Berlin, Germany.

ZMFMIB	Zoological Museum Fan Memorial Institute of Biology, Tsing Hua University, Peiping, China.
ZMH	Universitat Hamburg, Zoologisches Institut und Museum, Hamburg, Germany.
ZMMU	Zoological Museum, Moscow University, Russia.
ZMUB	Universitetet i Bergen, Zoologisk Museum, Bergen, Norway.
ZMUC	Kobenhavns Universitet Zoologisk Museum (Zoological Museum, University of Copenhagen), Copenhagen, Denmark.
ZMUL	Universitetets Lund, Zoologiska Museet, Lund, Sweden.
ZMUO	Universitetets I Oslo, Zoologisk Museum, Oslo, Norway.
ZMUT	University of Tokyo, Department of Zoology, University Museum, Tokyo, Japan.
ZMUU	Zoologiska Museet, Uppsala Universitet, Uppsala, Sweden.
ZRC	Zoological Reference Collection, National University of Singapore, Singapore.
ZSI	Zoological Survey of India, Kolkata [Calcutta], India.
ZSIC	Zoological Survey of India, Chennai, India.
ZSI/NRS	Northern Regional Station, Zoological Survey of India, Dehradun, India.
ZSI/SRS	Southern Regional Station, Zoological Survey of India, Madras, India.
ZVC-P	Facultad de Ciencias, Universidad de la República, Montevideo, República Oriental del Uruguay
ZSM	Zoologische Museum Staatssammlung, München, Germany.
ZX	Zhuhai City Fisheries Science Research Institute, Guandong Province, China.

Appendix 3. Taxonomic publications on Siluriformes issued in 2006

The following publications were issued in 2006, after the cutoff date for this Checklist. The publications are listed in alphabetical order of authorship, with an abbreviated summary of the taxonomic and nomenclatural actions taken, to make the checklist as up to date as possible. New names and other names that were subject to nomenclatural action have been noted and are included in the index. However, new names and any other proposed taxonomic changes were not incorporated into the statistics provided in Table 1.

Alencar, A.R. & Costa, W.J.E.M. (2006) *Trichomycterus pauciradiatus*, a new catfish species from the upper rio Paraná basin, southeastern Brazil (Siluriformes: Trichomycteridae). *Zootaxa*, 1269, 43–49.

Trichomycterus pauciradiatus Alencar & Wilson, 2006: 44, fig. 1. Type locality: Brazil: Estado de Minas Gerais: Município de Carrancas, córrego Debaixo da Serra, stream tributary of córrego Água Limba, rio Paraná basin, 21°26'40"S, 44°36'09"W. Holotype: UFRJ 5831.

Distribution: Upper Paraná River basin, southeastern Brazil.

Almirón, A., Azpelicueta, M. de las M., Casciotta, J. & Litz, T. (2006) A new species of *Hisonotus* (Siluriformes, Loricariidae, Otothyrini) from the República Oriental del Uruguay. *Revue suisse de Zoologie, Annales de la Société zoologique suisse et du Muséum d'Histoire naturelle de Genève*, 113, 87–94.

Hisonotus charrua Almirón Azpelicueta, Casciotta & Litz, 2006: 88, figs. 1–3. Type locality: República Oriental del Uruguay, Departamento Tacuarembó, río Uruguay basin, Cañada de Los Peña (31°39.09"S, 56°12.32"W). Holotype: ZVC-P 5639.

Distribution: Uruguay River and La Plata River mouth, Uruguay.

Betancur-R., R. & Acero P., A. (2006) A new species of *Notarius* (Siluriformes: Ariidae) from the Colombian Pacific. *Zootaxa*, 1249, 47–59.

Notarius armbrusteri Betancur-R. & Acero P., 2006: 50, figs. 2–3. Type locality: fish market of Buenaventura, Valle del Cauca, Colombia. Holotype: INVEMAR-PEC 6677.

Distribution: Eastern Pacific coast, at Valle del Cauca, Colombia.

Key: Species of *Notarius* of Eastern Pacific on p. 58.

Casciotta, J., Azpelicueta, M. de las M., Almirón, A. & Litz, T. (2006) *Hisonotus candombe*, a new species from the río Uruguay basin in the República Oriental del Uruguay (Siluriformes, Loricariidae, Otothyrini). *Spixiana*, 26 (2), 147–152.

Hisonotus candombe Casciotta, Azpelicueta, Almirón & Litz, 2006: 147, figs. 1–3. Type locality: República Oriental del Uruguay, Departamento Salto, río Uruguay basin, arroyo Palomas (31°04'43"S – 57°37'26"W). Holotype: ZVC-P 5595.

Distribution: lower Uruguay River basin, Uruguay.

Covain, R., Le Bail, P.Y., Sagnes, P. & Fisch-Muller, S. (2006) Species of the genus *Harttia* (Siluriformes: Loricariidae) in French Guiana: Morphology, taxonomy and distribution. *Cybium*, 30, 3–18.

Day, J. J. & Wilkinson, M. (2006) On the origin of the *Synodontis* catfish species flock from Lake Tanganyika. *Biology Letters*, doi:10.1098/rsbl.2006.0532, 1–5.

Diogo, R. & Bills, R. (2006) Osteology and myology of the cephalic region and pectoral girdle of the South African catfish *Austroglanis gilli*, with comments on the autapomorphies and phylogenetic relationships of the Austroglanididae (Teleostei: Siluriformes). *Animal Biology*, 56, 39–62.

Diogo, R., Chardon, M. & Vandewalle, P. (2006) Osteology and myology of the cephalic region and pectoral girdle of the *Cetopsis coecutiens* Spix & Agassiz, 1829, comparison with other cetopsids, and comments on the synapomorphies and phylogenetic relationships of the Cetopsidae (Teleostei: Siluriformes). *Belgian Journal of Zoology*, 136, 3–13.

Diogo, R., Chardon, M. & Vandewalle, P. (2006) Osteology and myology of the cephalic region and pectoral girdle of the *Nematogenys inermis* Guichenot, 1848, with comments on the autapomorphies and phylogenetic relationships of the Nematogenyidae (Teleostei: Siluriformes). *Belgian Journal of Zoology*, 136, 15–24.

Do Nascimento, C. & Provenzano, F. (2006) The genus *Henonemus* (Siluriformes: Trichomycteridae) with a description of a new species from Venezuela. *Copeia*, 2006, 198–205.

Henonemus triacanthopomus Do Nascimento & Provenzano, 2006: 199, fig. 1. Type locality: Venezuela, Estado Delta Amacuro, beach at Caño Macareo, 8°49'16"N, 62°00'00"W. Holotype: MBUCV-V 29526
Distribution: Apure and Arauca Rivers and in Caño Macareo in the Orinoco Delta, Venezuela.

Egge, J.J.D. & Simons, A.M. (2006) The challenge of truly cryptic diversity: diagnosis and description of a new madtom catfish (Ictaluridae: *Noturus*). *Zoologica Scripta*, 35, 581–595.

Noturus maydeni Egge, 2006: 588, fig. 7. Type locality: Strawberry River at Simstown Public Access, 4 mi N of Evening Shade, 1.7 mi off U.S. Hwy 167 on Simstown Road (036°05'52"N, 091°36'27"W), Sharp Co., AR [USA]. Holotype: JFBM 39465.

Distribution: Black and St. Francis river basins, Mississippi River drainage, Arkansas and Missouri, USA (Egge & Simons, 2006).

Remarks: Not distinguishable morphologically from *Noturus albater*; diagnosis based on karyotypes, allozyme variation, and DNA sequences.

Fernández, L. & Osinaga, K. (2006) A new *Trichomycterus* (Siluriformes: Trichomycteridae) from Aguarague National Park of the Bolivian preandean region, with comments on the relationships within the genus. *Environmental Biology of Fishes*, 75, 385–393.

Trichomycterus aguarague Fernández & Osinaga, 2006: 386, fig. 1. Type locality: Bolivia, Tarija, Province Gran Chaco, National Park Aguarague, Quebrada Timboycito, a Río Caiguami tributary, elevation 700–800 m, 21°30'S, 63°60'W. Holotype: MNKP 4012.

Distribution: Caiguami River tributaries, Pilcomayo River basin, Paraná River system, Bolivia.

Friel, J.P. & Vigliotta, T.R., (2006) *Synodontis acanthoperca*, a new species from the Ogôoué River system, Gabon with comments on spiny ornamentation and sexual dimorphism in mochokid catfishes (Siluriformes: Mochokidae). *Zootaxa*, 1125, 45–56.

Synodontis acanthoperca Friel & Vigliotta, 2006: 47, figs. 1–3. Type locality: Gabon, Haut-Ogôoué Province, Ogôoué River at and below the Rapids of Massoukou (Masuku), 1°39'30"S 13°32'14"E. Holotype: CU 89005.

Distribution: Louétsi and Ogôoué Rivers, Ogôoué River basin, Gabon.

Jayaram, K.C. (2006) *Catfishes of India*, Narendra Publishing House, Delhi, xxii +323 p., 11 pl.

Pterocryptis barakensis Vishwanath & Sharma, in Jayaram, 2006: 99, pl. 5, fig. 2. Type locality: India, Manipur, Tamen-glong District, Barak River at Vanchengphai, village. Holotype: MUMF 4018.

Distribution: Barak River, Brahmaputra River basin, India.

Horabagrinae new subfamily [Schilbidae], Jayaram, 2006: 141. Type genus: *Horabagrus* Jayaram, 1955.

Horaglanidinae new subfamily [Clariidae], Jayaram, 2006: 309. Type genus: *Horaglanis* Menon, 1950.

Janssen, G., Devaere, S., Weekers, P. & Adriaens, D. (2006) Phylogenetic and biogeographical analysis of African air-breathing catfish (Siluriformes: Clariidae): inferred from ribosomal genes and spacers sequences with emphasize on anguilliformity. *Molecular Phylogeny and Evolution* 38: 65–78.

Jerep, F.C., Shibatta, O.A., Pereira, E.H.L. & Oyakawa, O.T. (2006) Two new species of *Isbrueckerichthys* Derijst, 1996 (Siluriformes: Loricariidae) from the rio Paranapanema basin, Brazil. *Zootaxa*, 1372, 53–68.

Isbrueckerichthys calvus Jerep, Shibatta, Pereira & Oyakawa, 2006: 60, fig. 2. Type locality: Brazil; Paraná State; Apucarana; rio Tibagi basin; córrego Juruba; 23°34'44.6"S/51°22'12.6"W. Holotype: MZUEL 3714. Distribution: Tributary streams of the Taquara River, Tibagi River basin, Paraná State, Brazil.

Isbrueckerichthys saxicola Jerep, Shibatta, Pereira & Oyakawa, 2006: 55, fig. 1. Type locality: Brazil; Paraná State; Londrina; rio Tibagi basin; ribeirão Jacutinga; 23°14'30"S/51°13'05"W. Holotype. MZUEL 3716. Distribution: Ribeirão Jacutinga headwaters, Tibagi River basin, Paraná State, Brazil.

Koblmüller, S., Sturmbauer C., Verheyen, E., Meyer, A. & Salzburger, W. (2006) Mitochondrial phylogeny and phylogeography of East African squeaker catfishes (Siluriformes: *Synodontis*). *BMC Evolutionary Biology*, 2006, 6 (49) (<http://www.biomedcentral.com/1471-2148/6/49>).

Mori, H. & Shibatta, O.A. (2006) A new species of *Microglanis* Eigenmann, 1912 (Siluriformes, Pseudopimelodidae) from rio São Francisco basin, Brazil. *Zootaxa*, 1302, 31–42.

Microglanis leptostriatus Mori & Shibatta, 2006: 33, fig. 1. Type locality: rio Verde Grande, 16° 39'S/46°11'57.8"W, Montes Claros, State of Minas Gerais, Brazil. Holotype: MZUSP 85985.

Distribution: São Francisco River basin, Brazil.

- Murdy, E.O. & Ferraris, C.J., Jr. (2006) A revision of the marine catfish genus *Euristhmus* (Teleostei: Siluriformes: Plostidae). *The Beagle, Records of the Museums and Art Galleries of the Northern Territory*, 22, 77–90.
- Euristhmus microphthalmus* Murdy & Ferraris, 2006: 84, figs. 3, 6. Type locality: AUSTRALIA. Northern Territory. Woods Inlet, Darwin Harbour, 12°30'S, 130°45'E. Holotype: NTM S. 11242-001.
 Distribution: Northern Territory, Australia and Irian Jaya, Indonesia, in nearshore habitats over soft bottoms.
- Euristhmus sandrae* Murdy & Ferraris, 2006: 88, figs 3, 9. Type locality: AUSTRALIA. Western Australia. Exmouth Gulf, 21°42'S, 114°48'E, 9 m. Holotype: WAM P.32730.001.
 Distribution: Western Australia from south of Rowley Shoals and Exmouth Gulf, in waters as deep as 80m over soft bottoms.
- Cnidoglanis lepturus* Günther, 1864, Lectotype: BMNH 1864.1.17.33, designated on p. 82.
 Key: species of *Euristhmus* on p. 80.
- Near, T. J. & Hardman, M. (2006) Phylogenetic relationships of *Noturus stanauli* and *N. crypticus* (Siluriformes: Ictaluridae), two imperiled freshwater fish species from the southeastern United States. *Copeia*, 2006, 378–383.
- Ng, H.H. (2006) A Phylogenetic Analysis of the Asian Catfish Family Sisoridae (Teleostei: Siluriformes), and the Evolution of Epidermal Characters in the Group. Unpublished Ph.D. dissertation, University of Michigan, Ann Arbor.
- Ng, H.H. (2006) *Erethistoides infuscatus*, a new species of catfish (Teleostei: Erethistidae) from South Asia. *Ichthyological Explorations of Freshwaters*, 17, 283–287.
- Erethistoides infuscatus* Ng, 2006: 284, fig. 1. Type locality: India: Meghalaya, East Khasi Hills, Umsing River. Holotype: UMMZ 245695.
 Distribution: Brahmaputra and Meghna River drainages, northeast India and Bangladesh.
- Ng, H.H. (2006) The identity of *Batasio tengana* (Hamilton, 1822), with the description of two new species of *Batasio* from north-eastern India (Teleostei: Bagridae). *Journal of Fish Biology*, 68 (suppl. A), 101–118.
- Batasio tengana* Hamilton, 1822: Neotype designated by Ng, 2006: 103, fig. 1. Type locality: India: West Bengal: Tista River at Tista barrage; 26°45'10"N; 88°34'11"E. Neotype: UMMZ 244796.
 Distribution: Ganges and Brahmaputra River basins.
- Batasio fasciolatus* Ng, 2006: 107, fig. 4. Type locality: India: West Bengal, market at Malbazar: 26°32'30"N; 88°44'17"E. Holotype: UMMZ 244798.
 Distribution: Tista River basin, Brahmaputra River system.
- Batasio spilurus* Ng, 2006: 110, fig. 5. Type locality: India: Assam, Dibrugarh district, 27°29'N: 94°54'E. Holotype: ZRC 49133.
 Distribution: Brahmaputra River basin (Ng, 2006).
- Ng, H.H. (2006) *Pseudecheneis suppaetula*, a new species of glyptosternine catfish (Teleostei: Sisoridae) from India. *Zootaxa*, 1267, 59–68.
- Pseudecheneis suppaetula* Ng, 2006: 60, fig. 1. Type locality: India: Himachal Pradesh, Ganges River drainage, upper reaches of Giri River, in Chhaila area (in the vicinity of Kotkhai), 31°6'15"N 77°25'56"E. Holotype: NRM 36977.
 Distribution: Ganges River basin, Himachal Pradesh, India.
- Ng, H.H. (2006) *Akysis longifilis*, a new species of catfish (Teleostei: Akysidae) from Myanmar. *Zootaxa*, 1150, 19–30.
Akysis longifilis Ng, 2006: 20, fig. 1. Type locality: Myanmar: Bago division, Pyu township, Pyu stream (tributary of Sittang River) ca. 229 km from Yangon, 18°29'N, 96°26'E. Holotype: UMMZ 246172.
 Distribution: Sittang River basin, Myanmar.
- Ng, H.H. (2006) The identity of *Pseudecheneis sulcata* (M'Clelland, 1842), with descriptions of two new species of rheophilic catfish (Teleostei: Sisoridae) from Nepal and China. *Zootaxa*, 1254, 45–68.
- Pseudecheneis eddsi* Ng, 2006: 51, fig. 4. Type locality: Nepal: Tanahun, Khairenitar, Seti River (Ganges River drainage), 28°2'0.0"N 84°4'0.0"E. Holotype: KU 36872.
 Distribution: Gandaki River basin, Nepal.
- Pseudecheneis stenura* Ng, 2006: 57, fig. 5. Type locality: China: Yunnan, Baoshan Prefecture, Longchuanjiang at Lianmengie bridge (Irrawaddy River drainage). Holotype: KIZ 199811999.
 Distribution: Irrawaddy River basin, Yunnan, China.
- Ng, H.H. (2006) *Pseudolaguvia ferula*, a new species of sisoroid catfish (Teleostei: Erethistidae) from India. *Zootaxa*, 1229, 59–68.
Pseudolaguvia ferula Ng, 2006: 60, fig. 1. Type locality: India: West Bengal, Tista River at Tista Barrage, 26°45'10"N

88°34'11"E. Holotype: UMMZ 245985.
Distribution: Tista River, West Bengal, India.

Ng, H.H. & Bailey, R.M. (2006) *Chiloglanis productus*, a new species of suckermouth catfish (Siluriformes: Mochokidae) from Zambia. *Occasional Papers of the University of Michigan Museum of Zoology*, 738, 1–13.

Chiloglanis productus Ng, & Bailey, 2006: 2, figs. 1–3. Type locality: Zambia: Lunzua stream, 11.3 km SSE of Mpulungu, about 1 km N of bridge on Mbala–Mpulungu road, 8°49'S 31°10'E. Holotype: UMMZ 199816.

Distribution: Lunzua River basin, Lake Tanganyika drainage, Zambia.

Ng, H.H. & Lim, K.K.P. (2006) Two new species of *Leiocassis* (Teleostei: Bagridae), riverine catfishes from northeast Borneo. *Ichthyological Exploration of Freshwaters*, 17, 165–172.

Leiocassis collinus Ng & Lim, 2006: 166, fig. 1. Type locality: Borneo: Sabah: Danum Valley, Segama River drainage, Sungai Palum Tambun, tributary of Sungai Segama, upstream of Danum Valley Field Center. Holotype: ZRC 46154. Distribution: Kalabakan and Segama River basins, Sabah, Borneo.

Leiocassis tenebris Ng & Lim, 2006: 166, fig. 1. Type locality: Borneo: Kalimantan Timur: Kayan River drainage, Sungai Nah, tributary to Kayan River ca. 20 minutes upstream of confluence with Iwan River, 1°57'43.2"N 115°6'35.4"E at 550 m asl;. Holotype: MZB 10718.

Distribution: Kayan River basin, northeastern Borneo.

Ortega-Lara, A. & Lehmann A., P. (2006) *Cruciglanis*, a new genus of pseudopimelodid catfish (Ostariophysi: Siluriformes) with description of a new species from the Colombian Pacific coast. *Neotropical Ichthyology*, 4, 147–156.

Cruciglanis Ortega-Lara & Lehmann, 2006: 149. Type species: *Cruciglanis pacifici* Ortega-Lara & Lehmann, 2006, by original designation. Gender: Masculine.

Cruciglanis pacifici Ortega-Lara & Lehmann, 2006: 150, fig. 1. Type locality: Colombia, Valle del Cauca Department, near Buenaventura city, San Cipriano River where it crosses San Cipriano village, confluence of the La Sardina stream, approximately 500 m before the confluence with Dagua River, Dagua River basin, 03°50.543'N 76°54.068'W, 84 m of altitude. Holotype: IMCN 2359.

Distribution: Dagua River basin and Anchicayá River basin, Pacific versant, Colombia.

Parisi, B.M., Lundberg, J.G. & DoNascimento, C. (2006) *Propimelodus caesius* a new species of long-finned pimelodid catfish (Teleostei: Siluriformes) from the Amazon Basin, South America. *Proceedings of the Academy Of Natural Sciences, Philadelphia*, 155, 67–78.

Propimelodus caesius Parisi, Lundberg & DoNascimento, 2006: 68, figs. 1–2. Type locality: Brazil, Pará State, Rio Amazonas above Rio Trombetas, 20 km above Obidos, collected with 3 m bottom trawl in channel 4–7 m deep, 1°56'07.3"S, 55°41'18.5"W. Holotype: MZUSP 88582.

Distribution: Amazon River main stem and lower reaches of many tributary rivers, Brazil and Peru.

Peng, Z.-G., Ho, S.-W., Zhang, Y.-G. & He, S.-P. (2006) Uplift of the Tibetan plateau: Evidence from divergence times of glyptosternoid catfishes. *Molecular Phylogenetics and Evolution*, 39, 568–572.

Provenzano, F. & Milani, N. (2006) *Cordylancistrus nephelion* (Siluriformes, Loricariidae), a new and endangered species of suckermouth armored catfish from the Tuy River, north-central Venezuela. *Zootaxa*, 1116, 29–41.

Cordylancistrus nephelion Provenzano & Milani, 2006: 31, figs. 1–2. Type locality: Venezuela, Caribbean Sea basin, Tuy River system, Mesia River, tributary of the Guare River, near Village Corocito, approximately 10°12'N, 67°05'W. Holotype: MBUCV-V-21800.

Distribution: Tuy River basin, Venezuela (Provenzano & Milani, 2006).

Key: species of *Cordylancistrus* on p. 38.

Reis, R.E. & Borges, T.A.K. (2006) The South American catfish genus *Entomocorus* (Ostariophysi: Siluriformes: Auchenipteridae), with the description of a new species from the Paraguay River basin. *Copeia*, 2006, 412–422.

Entomocorus radiosus Reis & Borges, 2006: 416, figs. 6–7. Type locality: Brazil, Mato Grosso, Rio Paraguay at neighborhood of Cáceres, approx. 16°03'S, 057°42'W. Holotype: MCP 35902.

Distribution: Paraguay River basin (Reis & Borges, 2006).

Key: species of *Entomocorus* p. 419.

Reis, R.E., Pereira, E.H.L. & Armbruster, J.A.W. (2006) Delturinae, a new loricariid catfish subfamily (Teleostei, Siluriformes), with revisions of *Delturus* and *Hemipsilichthys*. *Zoological Journal of the Linnean Society*, 147, 277–299.

Delturus brevis Reis & Pereira, 2006: 295, fig. 12. Type locality: Rio Araçuáí upstream of its mouth on Rio Jequitinhonha, Araçuáí, Minas Gerais, Brazil. Holotype: MZUSP 69858.

- Distribution: Jequitinhonha River basin, Minas Gerais, Brazil.
Delturinae new subfamily Armbruster, Reis & Pereira, 2006: 279. Type genus: *Delturus* Eigenmann & Eigenmann, 1889.
- Key: subfamilies of Loricariidae, p. 279.
Key: *Hemipsilichthys* on p. 281.
Delturus parahybae Lectotype: MCZ 7726, designated on p. 293.
Delturus angulicauda Lectotype: NMW 44069, designated on p. 288.
Hemipsilichthys papillatus holotype illustrated on p. 285, fig. 6.
Hemipsilichthys nimius holotype illustrated on p. 287, fig. 8.
- Retzer, M. E. (2006) A new species of *Farlowella* Eigenmann and Eigenmann (Siluriformes: Loricariidae), a stick catfish from Bolivia. *Zootaxa*, 1282, 59–68.
Farlowella altocorpus Retzer, 2006: 60, figs. 1, 2. Type locality: Bolivia: La Paz State: Río Beni Basin: Río Coroico, Caranavi. Holotype: INHS 99773.
Distribution: Coroico River, Beni River basin, Bolivia.
- Salcedo, N.J. (2006) New species of *Chaetostoma* (Siluriformes: Loricariidae) from central Peru. *Copeia*, 2006, 60–67.
Chaetostoma changae, Salcedo, 2006: 61, fig. 1. Type locality: Peru, Departamento de Huánuco, vicinity of Tingo María, back-water near Puerto Nuevo, flowing into Río Tullamayo, 09°18'S, 75°59'W, 649 m. Holotype: ANSP 179125.
Distribution: Huallaga River basin at Tingo María, Peru.
- Salcedo, N.J. (2006) Two new species of *Chaetostoma* (Siluriformes: Loricariidae) from the Huallaga River in central Peru. *Ichthyological Exploration of Freshwaters*, 17, 207–220 [not seen].
Chaetostoma daidalmatos Salcedo, 2006
Chaetostoma stroumpoulos Salcedo, 2006
- Sarmento-Soares, L.M., Martins-Pinheiro, R.F., Aranda, A.T. & Chamon, C.C. (2006) *Microglanis pataxo*, a new catfish from southern Bahia coastal rivers, northeastern Brazil (Siluriformes: Pseudopimelodidae). *Neotropical Ichthyology*, 4, 157–166.
Microglanis pataxo Sarmento-Soares, Martins-Pinheiro, Aranda & Chamon, 2006: 158, Fig. 1. Type locality: Brazil, Bahia: Itamarajú, Jundiar creek on road BR-101 after joint with road to Jucuruçu, in the neighbourhoods of the city of Itamaraju (17°01'35"S 39°35'57"W). Holotype: MNRJ 28397.
Distribution: Peruipe, Jucuruçu and Cahy Rivers, northeastern Brazil.
- Sarmento-Soares, L.M., Martins-Pinheiro, R.F., Aranda A.T. & Chamon, C.C. (2006) *Ituglanis cahyensis*, a new catfish from Bahia, Brazil (Siluriformes: Trichomycteridae). *Neotropical Ichthyology*, 4, 309–318.
Ituglanis cahyensis Sarmento-Soares, Martins-Pinheiro, Aranda & Chamon, 2006: 310, fig. 1. Type locality: Brazil, Bahia: Prado, rio Palmares on road Guarany- Corumbau, in direction to Corumbau after the joint with the road to the mouth of rio Cahy, (16°57'48"S, 39°16'33"W). Holotype: MNRJ 28404.
Distribution: Palmares River, Cahy River basin, Southeastern Bahia State, Brazil.
- Sullivan J.P., Lundberg, J.G. & Hardman, M. (2006) A phylogenetic analysis of the major groups of catfishes (Teleostei: Siluriformes) using *rag1* and *rag2* nuclear gene sequences. *Molecular Phylogenetics and Evolution*, 41, 636–662.
- Tamang, L., Chaudhury, S. & Chauhury, D. (2006) On a new record of freshwater fish, *Pseudolaguvia shawi* (Hora) from Arunachal Pradesh, India (Teleostomi, Erethistidae). *Zoos' Print Journal*, 21 (11), 2443–2446.
- Thomson, A.W. & Page, L.M. (2006) Genera of the Asian catfish families Sisoridae and Erethistidae (Teleostei: Siluriformes). *Zootaxa*, 1345, 1–90.
Diagnoses and review of named taxa above species level.
Synonymy of *Hara* and *Erethistes*.
- Vari, R. P. & Ferraris, C.J., Jr. (2006) The Catfish genus *Tetranematicichthys* (Auchenipteridae). *Copeia*, 2006, 168–180.
Tetranematicichthys wallacei Vari & Ferraris, 2006: 169, figs. 1a, 2, 4. Type locality: Brazil, Amazonas, upper Rio Negro, São Pedro, mouth of Igarapé do Ibará, approximately 0°15'S, 66°46'W. Holotype: MZUSP 31096.
Distribution: Orinoco, Negro Amazon and Tocantins River basins.
- Villa-Verde, L. & Costa, W.J.E.M. (2006) A new glanapterygine catfish of the genus *Listrura* (Siluriformes: Trichomycteridae) from the southeastern Brazilian coastal plains. *Zootaxa*, 1142, 43–50.

Listrura picinguabae Villa-Verde & Costa, 2006: 44, figs. 1–3. Type locality: Brazil: Estado de São Paulo: Município de Ubatuba, serra do Mar, Picinguaba, small stream tributary to rio da Fazenda, on small road near Km 11 of the road BR-101, Parque Estadual da Serra do Mar, about 23°20'S 44°45'W. Holotype: UFRJ 6111, 48.6 mm SL.
Distribution: da Fazenda River tributaries, São Paulo State, southeastern Brazil.

Vishwanath, W. & Darshan, A. (2006) A new species of the genus *Batasio* Blyth (Teleostei: Bagridae) from Manipur, India. *Zoos' Print Journal*, 21, 2160–2163.

Batasio niger Vishwanath & Darshan, 2006: 2160, fig. 1. Type locality: Khujailok stream, Chandel district, Manipur, India. Holotype: MUMF 9028.

Distribution: Chindwin River basin, Manipur, India.

Remarks: Publication includes illustration of tooth plates only; illustration of holotype available only on web version of paper.

Wright, J.J. & Page, L.M. (2006) Taxonomic revision of Lake Tanganyikan *Synodontis* (Siluriformes: Mochokidae). *Florida Museum of Natural History Bulletin*, 46, 99–154.

Synodontis grandiops Wright & Page, 2006: 109, figs. 3c, 8, 9, 10. Type locality: Mwakizega coastline, L. Tanganyika. Holotype: BMNH 1982.4.13.4785.

Distribution: Lake Tanganyika.

Synodontis ilebrevis Wright & Page, 2006: 117, figs. 5d, 13, 14, 15. Type locality: Lake Tanganyika at Chaitika, Zambia. Holotype: UF 160942.

Distribution. Lake Tanganyika, only from the Cape Chaitika area, Zambia.

Synodontis lucipinnis Wright & Page, 2006: 126, figs. 4a, 17, 18. Type locality: Zambia, Mpulungu, Musende Rocks, 08°46'00"S, 031°51'00"E. Holotype: SAIAB 39577.

Distribution: Lake Tanganyika, only from the Musende Rocks area, Zambia.

Key: *Synodontis* species of Lake Tanganyika.

References

- Abbott, C.C. (1861) Descriptions of two new species of *Pimelodus* from Kansas. *Proceedings of the Academy of Natural Sciences, Philadelphia*, 12, 568–569.
- Abu Gideiri, Y.B. (1967) A new species of genus *Synodontis* (Siluroidea, Teleostei). *Revue de Zoologie et de Botanique Africaines*, 76, 132–136.
- Acero-P., A. (2002) Ariidae. In: Carpenter, K.E. (Ed.), *FAO Species Identification Guides for Fishery Purposes: The living Marine Resources of the Western Central Atlantic*, FAO, Rome, pp. 831–852.
- Acero-P., A. & Betancur-R., R. (2002a) *Arius cookei*, a new species of ariid catfish from the tropical American Pacific. *Aqua, Journal of Ichthyology and Aquatic Biology*, 5, 133–138.
- Acero-P., A. & Betancur-R., R. (2002b) Description of *Arius neogranatensis*, a new species of sea catfish from Colombia, with an identification key for Caribbean ariid fishes. *Aqua, Journal of Ichthyology and Aquatic Biology*, 6, 133–138.
- Acero-P., A., Tavera, J.J. & Reyes, J. (2005) Systematics of the genus *Bagre* (Siluriformes: Ariidae), a morphometric approach. *Cybium (3e série)*, 29, 127–133.
- Adams, A., Baikie, W.B. & Barron, C. (1854) *A Manual of Natural History for the Use of Travellers; Being a Description of the Families of the Animal and Vegetable Kingdoms: with Remarks on the Practical Study of Geology and Meteorology*, John Van Voorst, Paternoster Row, London, viii + 749 p.
- Adépo-Gourène, B., Teugels, G.G., Risch, L.M., Hanssens, M. & Agnèse, J.-F. (1997) Morphological and genetic differentiation of 11 populations of the African catfish *Chrysichthys nigrodigitatus* (Siluroidei; Claroteidae), with consideration of their biogeography. *Canadian Journal of Zoology*, 75, 102–109.
- Agassiz, L. (1844) *Recherches sur les Poissons Fossiles, Part 4*, Petitpierre, Neuchâtel and Soleure, xlix + 188 p.; atlas, 11 pls.
- Agassiz, L. (1846) *Nomenclatoris Zoologici. Index Universalis, Continens Nomina Systematica Classium, Ordinum, Familiarum et Generum Animalium Omnim, tam Viventium quam Fossilium*, Jent et Gassmann, Soloduri, viii + 393 p.
- Agassiz, L. (1850) *Lake Superior: its Physical Character, Vegetation, and Animals, Compared with Those of Other Similar Regions, by Louis Agassiz; with a Narrative of the Tour, by J. Elliott Cabot; and Contributions by Other Scientific gentlemen*, Gould, Kendall and Lincoln, Boston, 428 p., 8 pl.

- Agassiz, L. (1857) [The identity of the *Glanis* of Aristotle]. *Proceedings of the American Academy of Arts and Sciences*, 3, 325–334.
- Agnèse, J.F. & Teugels, G.G. (2001a) The *Bathyclarias* — *Clarias* species flock. A new model to understand rapid speciation in African Great lakes. *Comptes Rendus de l'Academie des Sciences, Serie III: Sciences de la Vie*, 324, 683–688.
- Agnèse, J.F. & Teugels, G.G. (2001b) Genetic evidence for monophyly of the genus *Heterobranchus* and paraphyly of the genus *Clarias* (Siluriformes, Clariidae). *Copeia*, 2001, 548–552.
- Agnèse, J.F. & Teugels, G.G. (2005) Insights into the phylogeny of African Clariidae (Teleostei: Siluriformes): implications for their body shape evolution, biogeography, and taxonomy. *Molecular Phylogenetics and Evolution*, 36, 546–553.
- Ahl, E. (1925) Neue südamerikanische Fische aus dem Zool. Museum Berlin. *Sitzungsberichte der Gesellschaft Naturforschender Freunde zu Berlin*, 1925, 106–109.
- Ahl, E. (1931) Neue Süßwasserfische aus dem Stromgebiet des Amazonenstromes. *Sitzungsberichte der Gesellschaft Naturforschender Freunde zu Berlin*, 1931, 206–211.
- Ahl, E. (1934) Weitere Fische aus dem Toba-See in Sumatra. *Sitzungsberichte der Gesellschaft Naturforschender Freunde zu Berlin*, 1934, 235–238.
- Ahl, E. (1936) Beschreibungen dreier neuer Welse aus Brasilien. *Zoologischer Anzeiger*, 116, 109–111.
- Ahl, E. (1937a) Ueber eine kleine Fischsammlung aus Sierra Leone. *Mitteilungen aus dem Zoologischen Museum in Berlin*, 22, 239–243.
- Ahl, E. (1937b) Zwei neue Süßwasserfische aus Südamerika. *Sitzungsberichte der Gesellschaft Naturforschender Freunde zu Berlin*, 1937, 445–447.
- Akama, A. & Ferraris, C.J., Jr. (2003) *Entomocorus melaphareus*, a new species of auchenipterid catfish (Osteichthyes: Siluriformes) from the lower and middle reaches of the Amazon River. *Neotropical Ichthyology*, 1, 77–82.
- Alencar, A.R. & Costa, W.J.E.M. (2004) Description of two new species of the catfish genus *Trichomycterus* from south-eastern Brazil (Siluriformes: Trichomycteridae). *Zootaxa*, 744, 1–8.
- Alexander, R.M. (1965) Structure and function in the catfish. *Journal of Zoology (London)*, 148, 88–152.
- Alfred, E.R. (1966) A new catfish of the genus *Akysis* from Malaya. *Copeia*, 1966, 467–470.
- Allen, G.R. (1985) Descriptions of two new species of freshwater catfishes (Plotosidae) from Papua New Guinea. *Records of the Western Australian Museum*, 12, 247–256.
- Allen, G.R. (1989) *Die Süßwasserfische Neuguineas. Eine Kommentierte Checkliste. (Freshwater Fishes of New Guinea. An Annotated Checklist)*, 35 p. [not seen]
- Allen, G.R. (1991) *Field Guide to the Freshwater Fishes of New Guinea*, Christensen Research Institute, Madang, Papua New Guinea, 268 p.
- Allen, G.R. (1996) Freshwater fishes of Irian Jaya. In: Kitchener, D.J. & Suyanto, A. (Eds.), *Proceedings of the first international conference on eastern Indonesian-Australian vertebrate fauna*, Manado, Indonesia, pp. 15–21. November 22–26, 1994
- Allen, G.R. (1997) *Marine Fishes of Tropical Australia and South-east Asia*, Western Australian Museum, Perth, 292 p.
- Allen, G.R. (1998) A review of the marine catfish genus *Paraplotosus* (Plotosidae) with the description of a new species from north-western Australia. *The Raffles Bulletin of Zoology*, 46, 123–124.
- Allen, G.R. & Coates, D. (1990) An ichthyological survey of the Sepik River, Papua New Guinea. *Records of the Western Australian Museum, Supplement*, 34, 31–116.
- Allen, G.R. & Feinberg, M.N. (1998) Descriptions of a new genus and four new species of freshwater catfishes (Plotosidae) from Australia. *Aqua, Journal of Ichthyology and Aquatic Biology*, 3, 9–18.
- Allen, G.R., Parenti, L.R. & Coates, D. (1992) Fishes of the Ramu River, Papua New Guinea. *Ichthyological Exploration of Freshwaters*, 3, 289–304.
- Almirón, A.E., Azpelicueta, M. & Casciotta, J.R. (2004) A new species of *Epauctionotus* (Siluriformes: Loricariidae: Otothyrini) from the río Iguazú basin, Argentina. *Zoologische Abhandlungen; Staatliches Museums für Tierkunde in Dresden*, 54, 137–144.
- Alonso de Arámburu, A. & Arámburu, R.H. (1962) Una nueva especie de *Xyliphius* de la Argentina (Siluriformes, Bunocephalidae). *Physis (Buenos Aires)*, 23, 219–222.
- Anderson, J. (1879) Pisces. In: Anderson, J. (Ed.), *Anatomical and Zoological Researches; Comprising an Account of the Zoological Results of the Two Expeditions to Western Yunnan in 1868 and 1875, and a Monograph of the Two Cetacean Genera, Platanista and Orcella*, Bernard Quaritch, London, pp. 863–869, pl. 79.
- Anseaume, L. & Teugels, G.G. (1999) On the rehabilitation of the clariid catfish genus *Bathyclarias* endemic to the east

- African rift Lake Malawi. *Journal of Fish Biology*, 55, 405–419.
- Anuradha, S. (1986) Contributions to the study of bagrid fishes, 19.— Systematic position of *Macrones halepensis colvillii* Hora & Misra, 1943, with description of a new species (Siluriformes, Bagridae). *Revue suisse de Zoologie, Annales de la Société zoologique suisse et du Muséum d'Histoire naturelle de Genève*, 93, 291–296.
- Anuradha, S. & Jayaram, K.C. (1985) Contributions to the study of bagrid fishes. 18.— Redescription of *Mystus pelusius* Solander, the type species of the genus *Mystus* Scopoli. *Bulletin of the Zoological Survey of India*, 7, 107–111.
- Aquino, A.E. (1996) Redescripción de *Otocinclus flexilis* Cope, 1894 (Siluriformes, Loricariidae, Hypoptopomatinae) con un nuevo sinónimo. *Iheringia, Série Zoología*, 81, 13–22.
- Aquino, A.E. (1997) Las especies de Hypoptopomatinae (Pisces, Siluriformes, Loricariidae) en la Argentina. *Revista de Ictiología*, 5, 5–21.
- Aquino, A.E. & Miquelarena, A.M. (2001) Redescription of *Hypoptopoma inexspectata* (Holmberg, 1883), with notes on its anatomy (Siluriformes: Loricariidae). *Physis (Buenos Aires), Sección B*, 58, for 2000, 1–18.
- Aquino, A.E. & Schaefer, S.A. (2002) Revision of *Oxyropsis* Eigenmann and Eigenmann, 1889 (Siluriformes, Loricariidae). *Copeia*, 2002, 374–390.
- Aquino, A.E., Schaefer, S.A. & Miquelarena, A.M. (2001) A new species of *Hisonotus* (Siluriformes, Loricariidae) of the upper Río Uruguay basin. *American Museum Novitates*, 3333, 1–12.
- Ardila Rodríguez, C.A. (2005) *Callichthys oibaensis*, una nueva especie de pez para el departamento de Santander – Colombia (Siluriformes: Callichthyidae). *Memoires del VIII Simposio Colombiano de Ictiología*, 31–43. [not seen]
- Armbruster, J.W. (1998a) Phylogenetic relationships of the suckermouth armored catfishes of the *Rhinelepis* group (Loricariidae: Hypostominae). *Copeia*, 1998, 620–636.
- Armbruster, J.W. (1998b) Review of the loricariid catfish genus *Aphanotorulus* and redescription of *A. unicolor* (Teleostei: Siluriformes). *Ichthyological Exploration of Freshwaters*, 8, 253–262.
- Armbruster, J.W. (2002) *Hypancistrus inspector*: a new species of suckermouth armored catfish (Loricariidae: Ancistrinae). *Copeia*, 2002, 86–92.
- Armbruster, J.W. (2003a) The species of the *Hypostomus cochliodon* group (Siluriformes: Loricariidae). *Zootaxa*, 249, 1–60.
- Armbruster, J.W. (2003b) *Peckoltia sabaji*, a new species from the Guyana Shield (Siluriformes: Loricariidae). *Zootaxa*, 344, 1–12.
- Armbruster, J.W. (2004a) Phylogenetic relationships of the suckermouth armoured catfishes (Loricariidae) with emphasis on the Hypostominae and the Ancistrinae. *Zoological Journal of the Linnean Society*, 141, 1–80.
- Armbruster, J.W. (2004b) *Pseudancistrus sidereus*, a new species from southern Venezuela (Siluriformes: Loricariidae) with a redescription of *Pseudancistrus*. *Zootaxa*, 628, 1–15.
- Armbruster, J.W. (2005) The loricariid catfish genus *Lasiancistrus* with description of two new species. *Neotropical Ichthyology*, 3, 549–569.
- Armbruster, J.W. & Hardman, M. (1999) Redescription of *Pseudorinelepis genibarbis* (Loricariidae: Hypostominae) with comments on behavior as it relates to air-holding. *Ichthyological Exploration of Freshwaters*, 10, 53–61.
- Armbruster, J.W. & Page, L.M. (1996) Redescription of *Aphanotorulus* (Teleostei: Loricariidae) with description of one new species, *A. ammophilus*, from the Río Orinoco Basin. *Copeia*, 1996, 379–389.
- Armbruster, J.W. & Page, L.M. (1997) Generic reassignment of the loricariid species *Monistancistrus carachama* Fowler 1940, *Plecostomus lacerta* Nichols, 1919, and *Rhinelepis levius* Pearson, 1924 (Teleostei: Siluriformes). *Copeia*, 1997, 227–232.
- Armbruster, J.W. & Provenzano, F. (2000) Four new species of the suckermouth armored catfish genus *Lasiancistrus* (Loricariidae: Ancistrinae). *Ichthyological Exploration of Freshwaters*, 11, 241–254.
- Armbruster, J.W., Sabaj, M.H., Hardman, M., Page, L.M. & Knouft, J.H. (2000) Catfish genus *Corymbophanes* (Loricariidae: Hypostominae) with description of one new species: *Corymbophanes kaiei*. *Copeia*, 2000, 997–1006.
- Armbruster, J.W. & de Souza, L.S. (2005) *Hypostomus macushi*, a new species of the *Hypostomus cochliodon* group (Siluriformes: Loricariidae) from Guyana. *Zootaxa*, 920, 1–12.
- Armbruster, J.W. & Werneke, D.C. (2005) *Peckoltia cavatica*, a new loricariid catfish from Guyana and a redescription of *P. braueri* (Siluriformes). *Zootaxa*, 882, 1–14.
- Arratia, G. (1983a) Preferencias de habitat de peces Siluriformes de aguas continentales de Chile (Fam. Diplomystidae y Trichomycteridae). *Studies on Neotropical Fauna and Environment*, 18, 217–237.
- Arratia, G. (1983b) *Trichomycterus chungaraensis* n. sp. and *Trichomycterus laucaensis* n. sp. (Pisces, Siluriformes, Trichomycteridae) from the high Andean range. *Studies on Neotropical Fauna and Environment*, 18, 65–87.
- Arratia, G. (1987) Description of the primitive family Diplomystidae (Siluriformes, Teleostei, Pisces): Morphology, tax-

- onomy and phylogenetic implications. *Bonner Zoologische Monographien*, 24, 1–120.
- Arratia, G. (1990) The South American Trichomycterinae (Teleostei: Siluriformes), a problematic group. In: Peters, G. & Hutterer, R. (Eds.), *Vertebrates in the Tropics: Proceedings of the International Symposium on Vertebrate Biogeography and Systematics in the Tropics, Bonn, June 5–8, 1989*, Alexander Koenig Zoological Research Institute and Zoological Museum, Bonn, pp. 395–403.
- Arratia, G. (1992) Development and variation of the suspensorium of primitive catfishes (Teleostei: Ostariophysi) and their phylogenetic relationships. *Bonner Zoologische Monographien*, 32, 1–149.
- Arratia, G. (1998) *Silvinichthys*, a new genus of trichomycterid catfishes from the Argentinian Andes, with redescription of *Trichomycterus nigricans*. *Ichthyological Exploration of Freshwaters*, 9, 347–370.
- Arratia, G. & Chang, A. (1975) Osteocráneo de *Nematogenys inermis* Guichenot, 1848, y consideraciones acerca de la primitividad del género (Pisces, Siluriformes, Trichomycteridae). *Museo Nacional de Historia Natural, Santiago de Chile, Publicación Ocasional*, 19, 3–7.
- Arratia, G., Chang, A., G., Menu-Marque, S. & Rojas, G. M. (1978) About *Bullockia* gen. nov., *Trichomycterus mendozensis* n. sp. and revision of the family Trichomycteridae (Pisces, Siluriformes). *Studies on Neotropical Fauna and Environment*, 13, 157–194.
- Arratia, G. & Cione, A. (1996) The record of fossil fishes of southern South America. *Münchner Geowissenschaftliche Abhandlungen, Reihe A –Geologie und Paläontologie*, 30, 9–72.
- Arratia, G. & Gayet, M. (1995) Sensory canals and related bones of Tertiary siluriform crania from Bolivia and North America and comparison with Recent forms. *Journal of Vertebrate Paleontology*, 15, 482–505.
- Arratia, G. & Huaquin, L. (1995) Morphology of the lateral line system and of the skin of diplomystid and certain primitive loricarioid catfishes and systematic and ecological considerations. *Bonner Zoologische Monographien*, 36, 1–110.
- Arratia, G., Kapoor, B.G., Chardon, M. & Diogo, R. (Eds.) (2003) *Catfishes*, Science Publishers, Enfield, NH, USA, 812 p.
- Arratia, G. & Menu-Marque, S. (1981) Revision of the freshwater catfishes of the genus *Hatcheria* (Siluriformes, Trichomycteridae) with commentaries on ecology and biogeography. *Zoologischer Anzeiger*, 207, 88–111.
- Arratia, G. & Menu-Marque, S. (1984) New catfishes of the genus *Trichomycterus* from the high Andes of South America (Pisces, Siluriformes) with remarks on distribution and ecology. *Zoologische Jahrbücher: Abteilung für Systematik, Ökologie und Geographie der Tiere*, 111, 493–520.
- Arunkumar, L. (2000) *Laguvia manipurensis*, a new species of sisorid cat fish (Pisces: Sisoridae) from the Yu River system of Manipur. *Indian Journal of Fisheries*, 47, 193–200.
- Arunkumar, L. & Tombi Singh, H. (1997) *Silurus morehensis*, a new species of silurid cat fish (Order: Siluriformes, Family: Siluridae) from Manipur, India. *Journal of Freshwater Biology*, 9, 72–76.
- Ataur Rahman, A.K. (1976) A new species of the genus *Gagata* Bleeker from river of Bangladesh. *Bangladesh Journal of Biological Sciences*, 5, 4–8.
- Axelrod, H.R. (1970) *Synodontis davidii*, David's upside-down catfish. *Tropical Fish Hobbyist*, 18, 33–35.
- Axelrod, H.R. (1987) Two new species of catfishes (Siluriformes, Callichthyidae and Pimelodidae) from the Rio Unini, Amazonas, Brazil. *Tropical Fish Hobbyist*, 35, 22–25.
- Axenrot, T.E. & Kullander, S.O. (2003) *Corydoras diphyes* (Siluriformes: Callichthyidae) and *Otocinclus mimulus* (Siluriformes: Loricariidae), two new species of catfishes from Paraguay, a case of mimetic association. *Ichthyological Exploration of Freshwaters*, 14, 249–272.
- Azpelicueta, M.M. (1990) *Tridentopsis cahuali* n. sp. (Siluriformes, Trichomycteridae), a new miniature tridentine from Paraguay System, in Argentina. *Revue suisse de Zoologie, Annales de la Société zoologique suisse et du Muséum d'Histoire naturelle de Genève*, 97, 981–988.
- Azpelicueta, M.M. (1994a) Los diplomistidos en Argentina (Siluriformes, Diplomystidae). *Fauna de Agua Dulce de la República Argentina*, 40, Pisces, 5–27.
- Azpelicueta, M.M. (1994b) Three east-Andean species of *Diplomystes* (Siluriformes: Diplomystidae). *Ichthyological Exploration of Freshwaters*, 5, 223–240.
- Azpelicueta, M.M. (1995) *Pimelodus absconditus*, a new species of pimelodid catfish from the la Plata basin (Siluriformes: Pimelodidae). *Ichthyological Exploration of Freshwaters*, 6, 71–76.
- Azpelicueta, M.M. (1998) A new species of *Pimelodus* (Siluriformes: Pimelodidae) from the Paraguay and lower Paraná rivers. *Neotropica*, 44, 87–94.
- Azpelicueta, M.M. (2001) A new species of *Pimelodus* (Siluriformes: Pimelodidae) from the upper Paraná basin, Brazil. *Ichthyological Exploration of Freshwaters*, 12, 193–200.

- Azpelicueta, M.M., Casciotta, J., Almirón, A. & Körber, S. (2004) A new species of Otothyridini (Siluriformes: Loricariidae: Hypoptopomatinae) from the Río Uruguay basin, Argentina. — Eine neue Art des Stammes Otothyridini (Siluriformes: Loricariidae: Hypoptopomatinae) aus dem System des Rio Uruguay, Argentinien. *Verhandlungen der Gesellschaft für Ichthyologie*, 4, 81–90.
- Azpelicueta, M.M. & Gosztonyi, A.E. (1998) Redescription of *Diplomystes mesembrinus* (Siluriformes, Diplomystidae). *Revue suisse de Zoologie, Annales de la Société zoologique suisse et du Muséum d'Histoire naturelle de Genève*, 105, 901–910.
- Azpelicueta, M.M. & Rubilar, A. (1998) A Miocene *Nematogenys* (Teleostei: Siluriformes: Nematogenyidae) from South-Central Chile. *Journal of Vertebrate Paleontology*, 18, 475–483.
- Bailey, R.G. (1968) A new species of *Synodontis* (Mochokidae) from Tanzania, East Africa. *Revue de Zoologie et de Botanique Africaines*, 77, 346–348.
- Bailey, R.M. & Baskin, J.N. (1976) *Scolopax dicra*, a new armored catfish from the Bolivian Amazon. *Occasional Papers of the Museum of Zoology, University of Michigan*, 674, 1–14.
- Bailey, R.M. & Robins, C.R. (1988) *Ameiurus* Rafinesque, 1820 (Osteichthyes, Siluriformes): proposed designation of *Silurus lividus* Rafinesque, 1820 (= *Pimelodus natalis* Lesueur, 1819) as the type species. *Bulletin of Zoological Nomenclature*, 45, 135–137.
- Bailey, R.M. & Stewart, D.J. (1983) *Bagrus* Bosc, 1816 (Pisces, Siluriformes): Proposal to place on the Official List. Z.N.(S.) 2371. *Bulletin of Zoological Nomenclature*, 40, 167–172.
- Bailey, R.M. & Stewart, D.J. (1984) Bagrid catfishes from Lake Tanganyika, with a key and descriptions of new taxa. *Miscellaneous Publications, Museum of Zoology, University of Michigan*, 168, i–iv + 1–41.
- Bailey, R.M. & Taylor, W.R. (1950) *Schilbeodes hildebrandi*, a new ameiurid catfish from Mississippi. *Copeia*, 1950, 31–38, pls. 1–2.
- Baird, S.F. & Girard, C.F. (1854) Descriptions of new species of fishes collected in Texas, New Mexico and Sonora, by Mr. John H. Clark, on the U. S. and Mexican Boundary Survey, and in Texas by Capt. Stewart Van Vliet, U. S. A. *Proceedings of the Academy of Natural Sciences, Philadelphia*, 7, 24–29.
- Balon, E.K. & Hensel, K. (1970) Notes on a small collection of fishes from Afghanistan with a description of *Glyptothorax jalalensis*, sp. n. (Pisces, Sisoridae). *Vestnik Ceskoslovenské Společnosti Zoologické*, 34, 159–163.
- Barbosa, M.A. & Costa, W.J.E.M. (2003a) Validade, relações filogenéticas e redescrição de *Eremophilus candidus* Ribeiro, 1949 (Teleostei, Siluriformes, Trichomycteridae). *Arquivos do Museu Nacional do Rio de Janeiro*, 61, 179–188.
- Barbosa, M.A. & Costa, W.J.E.M. (2003b) *Trichomycterus potschi* (Siluriformes: Loricarioidei): a new trichomycterid catfish from coastal streams of southeastern Brazil. *Ichthyological Exploration of Freshwaters*, 14, 281–288.
- Barbour, T. & Cole, L.J. (1906) Vertebrata from Yucatan: Reptilia, Amphibia, and Pisces. *Bulletin of the Museum of Comparative Zoology*, 50, 146–159, pls. 1–2.
- Bardack, J. (1961) New Tertiary teleosts from Argentina. *American Museum Novitates*, 2041, 1–27.
- Barnard, K.H. (1943) Revision of the indigenous freshwater fishes of the S. W. Cape region. *Annals of the South African Museum*, 36, 101–262.
- Bashir, K.A. & Mirza, M.R. (1975) Fishes of the river Sutlej in Lahore District, Pakistan, with the description of a new subspecies. *Bulletin of Hydrobiological Research, series 1*, 9, 91–104.
- Basilewsky, S. (1855) Ichthyographia Chiae Borealis. *Nouveaux Mémoires de la Société Impériale des Naturalistes de Moscou*, 10, 215–263, pls. 1–9.
- Baskin, J.N. (1973) *Structure and relationships of the Trichomycteridae*. Unpublished Ph.D. dissertation, City University of New York, New York.
- Baumgartner, J.V. (1982) A new fossil ictalurid catfish from the Miocene middle member of the Truckee Formation, Nevada. *Copeia*, 1982, 38–46.
- Bean, T.H. (1880a) Description of a new species of *Amiurus* (*A. ponderosus*) from the Mississippi River. *Proceedings of the United States National Museum*, 2, 286–290.
- Bean, T.H. (1880b) Descriptions of two species of fishes collected by Prof. A. Dugès in central Mexico. *Proceedings of the United States National Museum*, 2, 302–305.
- Beaufort, L.F. de. (1931) Pisces, Reptilia and Aves. In: Onse palaeontologische kennis van Nederlandsch Oost-Indië 1930. *Leidsche geologische Mededeelingen*, 5, 461–470.
- Beaufort, L.F. de. (1933) On some new or rare species of Ostariophysi from the Malay Peninsula and a new species of *Betta* from Borneo. *Bulletin of the Raffles Museum*, 8, 31–36.
- Behre, E.H. (1928) A list of the fresh water fishes of western Panama between 81°45' and 83°15'W. *Annals of the Carn-*

- egie Museum, 18, 305–328, pl. 1.
- Bell-Cross, G. (1973) The fish fauna of the Buzi River system in Rhodesia and Moçambique. *Arnoldia (Rhodesia)*, 6, 1–14.
- Bellotti, C. (1892) Un nuovo siluroide giapponese. *Atti della Società Italiana di Scienze Naturali, Milano*, 34, 99–101.
- Benech, V., Teugels, G.G. & Gourene, G. (1993) Critère pratique pour distinguer deux poissons-chats Africains, *Clarias anguillaris* et *C. gariepinus* (Siluriformes; Clariidae). *Cybium (3e série)*, 17, 83–85.
- Bennett, E.T. (1830) Class Pisces. In: *Catalogue of Zoological Specimens*, in: S. Raffles, *Memoirs of the Life and Public Services of Sir Thomas Stamford Raffles [...], Particularly in the Government of Java, 1811–1816, and of Bencoolen and its Dependencies, 1817–1824; with Details of the Commerce and Resources of the Eastern Archipelago, and Selections from his Correspondence, by his Widow*, Murray, London, pp. 686–694. [Authorship of Class Pisces indicated on p. [xi] and 629–630]
- Berg, C. (1895) Sobre peces de agua dulce nuevos ó poco conocidos de la República Argentina. *Anales del Museo Nacional de Buenos Aires, secunda Serie*, 4, 121–165, pls. 2–3.
- Berg, C. (1897) Contribuciones al conocimiento de los peces Sudamericanos, especialmente de los de la República Argentina. *Anales del Museo Nacional de Buenos Aires, secunda Serie*, 5, 263–302.
- Berg, C. (1901) Comunicaciones ictiológicas, IV. *Comunicaciones del Museo Nacional de Buenos Aires*, 1, 293–311.
- Berg, L.S. (1907) Beschreibungen einiger neuer Fische aus dem Stromgebiete des Amur. *Ezhegodnik Zoologicheskii Muzei Akademii Nauk*, 12, 418–423.
- Berg, L.S. (1918) [On a new siluroid fish, *Glyptosternum armeniacum* n. sp., from the Upper Euphrates]. *Bulletin du Musée du Caucase, Tiflis*, 11, 145–148. [In Russian, with English title and summary]
- Berg, L.S. (1931) Description of a new siluroid fish, *Glyptosternum kurdistanicum*, from the basin of the Tigris River. *Izvestiia Akademii Nauk SSSR. Otdelenie Matematicheskikh i Estestvennykh Nauk*, 1931, 1267–1270, pl. 1.
- Bertaco, V.A. & Cardoso, A.R. (2005) A new species of *Microglanis* (Siluriformes: Pseudopimelodidae) from the rio Uruguay drainage, Brazil. *Neotropical Ichthyology*, 3, 61–67.
- Bertin, L. & Estève, R. (1950) *Catalogue des Types de Poissons du Muséum National d'Histoire Naturelle. 5^e partie.—Ostariophysaires (Siluriformes)*, Imp. Nationale, Paris, 85 p.
- Bertoletti, J.J. (1967) Nova espécie de "Rhamdella" Eigenmann & Eigenmann (Actinopterygii, Siluriformes, Pimelodidae). *Revista Brasileira de Biologia*, 27, 77–83.
- Bertoletti, J.J., Pereira, E.H.L. & da Silva, J.F.P. (1992) Nota sobre o gênero *Trachelyopterus* Valenciennes, 1840, no estado do Rio Grande do Sul, Brasil (Siluriformes, Auchenipteridae). *Comunicações do Museu de Ciências da PUCRS, Série Zoologia, Porto Alegre*, 5, 169–177.
- Bertoletti, J.J., Pezzi da Silva, J.F. & Pereira, E.H.L. (1995) A new species of the catfish genus *Trachelyopterus* (Siluriformes, Auchenipteridae) from southern Brazil. *Revue Française d'Aquariologie et Herpetologie*, 22, 71–74.
- Bessednov, L.N. (1967) [Sea stinging catfishes of the genus *Plotosus* Lacépède from the Tonkin Bay]. *Zoologicheskii Zhurnal*, 46, 443–447. [In Russian, with English abstract.]
- Betancur-R., R. (2003) *Filogenia de los Bagres Marinos (Siluriformes: Ariidae) del Nuevo Mundo*, M.Sc.Thesis, Universidad Nacional de Colombia, Bogotá.
- Betancur-R., R. & Acero P., A. (2004) Description of *Notarius biffi* n. sp. and redescription of *N. insculptus* (Jordan and Gilbert) (Siluriformes: Ariidae) from the eastern Pacific, with evidence of monophyly and limits of *Notarius*. *Zootaxa*, 703, 1–20.
- Betancur-R., R. & Acero P., A. (2005) Description of *Cathorops mapale*, a new species of sea catfish (Siluriformes: Ariidae) from the Colombian Caribbean, based on morphological and mitochondrial evidence. *Zootaxa*, 1045, 45–60.
- Betancur-R., R., Acero-P., A. & Mejia-Ladino, L.M. (2004) Análisis filogenético preliminar de algunos bagres marinos (Siluriformes: Ariidae) neotropicales. Preliminar [Preliminary] phylogenetic analysis of some neotropical marine catfishes (Siluriformes: Ariidae). *Memoria de la Fundación la Salle de Ciencias Naturales*, 62, 61–85.
- Bhatt, A. & Jayaram, K.C. (2004) A new species of the genus *Batasio* Blyth (Siluriformes: Bagridae) from Sharavati River, Uttara Kannada, Karnataka. *Zoos'Print Journal*, 19, 1339–1342.
- Bichuette, M.E. & Trajano, E. (2004) Three new subterranean species of *Ituglanis* from Central Brazil (Siluriformes: Trichomycteridae). *Ichthyological Exploration of Freshwaters*, 25, 243–256.
- Bichuette, M.E. & Trajano, E. (2005) A new cave species of *Rhamdia* Bleeker, 1858 (Siluriformes: Heptapteridae) from Serra do Ramalho, northeastern Brazil, middle São Francisco River basin, with notes on ecology and behavior. *Neotropical Ichthyology*, 3, 587–595.
- Bizerril, C.R.S.F. (1991) Descrição de uma nova espécie de *Brachyglanis* Eigenmann, 1912 da Bacia do rio Trombetas (Siluroidei, Pimelodidae). *Boletim do Museu Nacional do Rio de Janeiro, Nova Série, zoologia*, 347, 1–8.

- Bizerril, C.R.S.F. (1994) Descrição de uma nova espécie de *Trichomycterus* (Siluroidei, Trichomycteridae) do Estado de Santa Catarina, com uma sinopse da composição da família Trichomycteridae no leste Brasileiro. *Arquivos de Biologia e Tecnologia* (Curitiba), 37, 617–628.
- Bizerril, C.R.S.F. (1995a) Description of new species of *Hemipsilichthys* (Loricariidae, Hypostominae) from the State of Santa Catarina, Brazil. *Acta Biologica Leopoldensia*, 17, 115–122.
- Bizerril, C.R.S.F. (1995b) Descrição de uma nova espécie de *Neoplecostomus* (Loricariidae, Neoplecostominae) com uma sinopse da composição taxonômica dos Loricariidae no leste Brasileiro. *Arquivos de Biologia e Tecnologia* (Curitiba), 38, 693–704.
- Bizerril, C.R.S.F. & Borba, I.M.M. (1994) Status taxonômico de *Rhamdella schultzi* Ribeiro, 1964 (Siluriformes, Pimelodidae). *Boletim do Museu Nacional do Rio de Janeiro, Nova Série, zoologia*, 357, 1–6.
- Bizerril, C.R.S.F. & Perez-Neto, P.R. (1992) Description of a new species of *Microglanis* (Siluroidei, Pimelodidae) from eastern Brazil. *Revue Française d'Aquariologie et Herpetologie*, 18, for 1991, 97–100.
- Blache, J. (1964) Les poissons du bassin du Tchad et du bassin adjacent du Mayo Kebbi. Étude systématique et biologique. *Mémoires ORSTOM*, 4, 1–483.
- Black, B. (1987) A new species of *Corydoras*, *Corydoras davidsandsi* (Siluriformes, Callichthyidae) from the Rio Unini, Amazonas, Brazil. *Practical Fishkeeping*, 1987, 74–75.
- Bleeker, P. (1846a) Overzigt der siluroïden, welke te Batavia voorkomen. *Natuur-en Geneeskundig Archif voor Nederlandsch Indië*, 3, 135–184.
- Bleeker, P. (1846b) Siluroideorum bataviensium species nuperime detectae. *Natuur-en Geneeskundig Archif voor Nederlandsch Indië*, 3, 284–293.
- Bleeker, P. (1847a) Pharyngognathorum Siluroideorumque species novae Javanenses. *Natuur-en Geneeskundig Archif voor Nederlandsch Indië*, 4, 155–169.
- Bleeker, P. (1847b) Nieuwe bijdrage tot de kennis der Siluroïden van Java. *Verhandelingen van het Bataviaasch Genootschap van Kunsten en Wetenschappen*, 21, 1–12.
- Bleeker, P. (1847c) Siluroideorum bataviensium conspectus diagnosticus. *Verhandelingen van het Bataviaasch Genootschap van Kunsten en Wetenschappen*, 21, 1–60.
- Bleeker, P. (1849) Bijdrage tot de kennis der ichthyologische fauna van het eiland Madura, met beschrijving van eenige nieuwe soorten. *Verhandelingen van het Bataviaasch Genootschap van Kunsten en Wetenschappen*, 22, 1–16.
- Bleeker, P. (1851a) Bijdrage tot de kennis der ichthyologische fauna van Borneo, met beschrijving van 16 nieuwe soorten van zoetwatervisschen. *Natuurkundig Tijdschrift voor Nederlandsch Indië*, 1, 1–16.
- Bleeker, P. (1851b) Nieuwe bijdrage tot de kennis der ichthyologische fauna van Borneo met beschrijving van eenige nieuwe soorten van zoetwatervisschen. *Natuurkundig Tijdschrift voor Nederlandsch Indië*, 1, 259–275.
- Bleeker, P. (1851c) Derde bijdrage tot de kennis der ichthyologische fauna van Borneo, met beschrijving van eenige nieuwe soorten van zoetwatervisschen. *Natuurkundig Tijdschrift voor Nederlandsch Indië*, 2, 57–70.
- Bleeker, P. (1851d) Vierde bijdrage tot de kennis der ichthyologische fauna van Borneo, met beschrijving van eenige nieuwe soorten van zoetwatervisschen. *Natuurkundig Tijdschrift voor Nederlandsch Indië*, 2, 193–208.
- Bleeker, P. (1851e) Vijfde bijdrage tot de kennis der ichthyologische fauna van Borneo, met beschrijving van eenige nieuwe soorten van zoetwatervisschen. *Natuurkundig Tijdschrift voor Nederlandsch Indië*, 2, 415–442.
- Bleeker, P. (1851f) Bijdrage tot de kennis der ichthyologische fauna van Riouw. *Natuurkundig Tijdschrift voor Nederlandsch Indië*, 2, 469–497.
- Bleeker, P. (1852a) Bijdrage tot de kennis der ichthyologische fauna van Blitong (Billiton), met beschrijving van eenige nieuwe soorten van zoetwatervisschen. *Natuurkundig Tijdschrift voor Nederlandsch Indië*, 3, 87–100.
- Bleeker, P. (1852b) Zesde bijdrage tot de kennis der ichthyologische fauna van Borneo. Vissen van Pamangkat, Bandjermassing, Praboekarta en Sampit. *Natuurkundig Tijdschrift voor Nederlandsch Indië*, 3, 407–442.
- Bleeker, P. (1852c) Bijdrage tot de kennis der ichthyologische fauna van het eiland Banka. *Natuurkundig Tijdschrift voor Nederlandsch Indië*, 3, 443–460.
- Bleeker, P. (1852d) Diagnostische beschrijvingen van nieuwe of weinig bekende vischsoorten van Sumatra. Tiental I – IV. *Natuurkundig Tijdschrift voor Nederlandsch Indië*, 3, 569–608.
- Bleeker, P. (1852e) Nieuwe bijdrage tot de kennis der ichthyologische fauna van het eiland Banka. *Natuurkundig Tijdschrift voor Nederlandsch Indië*, 3, 715–738.
- Bleeker, P. (1853a) Nalezingen op de ichthyologische fauna van het eiland Banka. *Natuurkundig Tijdschrift voor Nederlandsch Indië*, 5, 175–193.
- Bleeker, P. (1853b) Nieuwe tientallen diagnostische beschrijvingen van nieuwe of weinig bekende vischsoorten van Sumatra. *Natuurkundig Tijdschrift voor Nederlandsch Indië*, 5, 495–534.

- Bleeker, P. (1854a) Overzigt der ichthyologische fauna van Sumatra, met beschrijving van eenige nieuwe soorten. *Natuurkundig Tijdschrift voor Nederlandsch Indië*, 7, 49–108.
- Bleeker, P. (1854b) Specierum piscium javanensium novarum vel minus cognitarum diagnoses adumbratae. *Natuurkundig Tijdschrift voor Nederlandsch Indië*, 7, 415–448.
- Bleeker, P. (1854c) Nalezingen op de ichthyologische fauna van Bengalen en Hindostan. *Verhandelingen van het Bataviaansch Genootschap van Kunsten en Wetenschappen*, 25, 1–164, pls. 1–6.
- Bleeker, P. (1855a) Nalezingen op de vischfauna van Sumatra. Visschen van Lahat en Sibogha. *Natuurkundig Tijdschrift voor Nederlandsch Indië*, 9, 257–280.
- Bleeker, P. (1855b) Negende bijdrage tot de kennis der ichthyologische fauna van Borneo. Zoetwatervissen van Pontianak en Bandjermasin. *Natuurkundig Tijdschrift voor Nederlandsch Indië*, 9, 415–430.
- Bleeker, P. (1856) Verslag van eenige verzamelingen van zee- en zoetwatervissen van het eiland Banka. *Natuurkundig Tijdschrift voor Nederlandsch Indië*, 11, 415–420.
- Bleeker, P. (1857a) Descriptionum specierum piscium Javanensium novarum vel minus cognitarum diagnosticae. *Natuurkundig Tijdschrift voor Nederlandsch Indië*, 13, 323–368.
- Bleeker, P. (1857b) Index: Descriptionum specierum piscium Bleekerianarum in voluminibus I ad XIV diarii Societatis Scientiarum Indo-Batavae. *Natuurkundig Tijdschrift voor Nederlandsch Indië*, 14, 447–486.
- Bleeker, P. (1858a) Zesde bijdrage tot de kennis der vischfauna van Sumatra. Visschen van Padang, Troessan, Priaman, Sibogha en Palembang. *Verhandelingen der Koninklijke Natuurkundige Vereeniging in Nederlandsch Indië [= Acta Societatis Regiae Scientiarum Indo-Neerlandicae]*, 3, 1–50.
- Bleeker, P. (1858b) De visschen van den Indischen Archipel. Beschreven en toegelicht. Siluri. *Verhandelingen der Koninklijke Natuurkundige Vereeniging in Nederlandsch Indië [= Acta Societatis Regiae Scientiarum Indo-Neerlandicae]*, 4, i–xii + 1–370. [Also: Ichthyologiae Archipelagi Indici Prodromus, Vol 1. Siluri. Lange & Co., Batavia. xii + 370.]
- Bleeker, P. (1858c) Twaalfde bijdrage tot de kennis der vischfauna van Borneo. Visschen van Sinkawang. *Verhandelingen der Koninklijke Natuurkundige Vereeniging in Nederlandsch Indië [= Acta Societatis Regiae Scientiarum Indo-Neerlandicae]*, 5, 1–10.
- Bleeker, P. (1858d) Vierde bijdrage tot de kennis der vischfauna van Biliton. *Natuurkundig Tijdschrift voor Nederlandsch Indië*, 15, 219–240.
- Bleeker, P. (1858e) Enumeratio specierum piscium javanensium hucusque cognitarum. *Natuurkundig Tijdschrift voor Nederlandsch Indië*, 15, 359–456.
- Bleeker, P. (1859) Enumeratio specierum piscium hucusque in Archipelago indicō observatarum, adjectis habitationibus citationibusque, ubi descriptiones earum recentiores reperiuntur, nec non speciebus Musei Bleekeriani Bengalensisibus, Japonicis, Capensisbus Tasmanicisque. *Verhandelingen der Koninklijke Natuurkundige Vereeniging in Nederlandsch Indië [= Acta Societatis Regiae Scientiarum Indo-Neerlandicae]*, 6, i–xxxvi, 1–276.
- Bleeker, P. (1860) Negende bijdrage tot de kennis der vischfauna van Sumatra (Visschen uit de Lematang-Enim en van Benkoelen). *Verhandelingen der Koninklijke Natuurkundige Vereeniging in Nederlandsch Indië [= Acta Societatis Regiae Scientiarum Indo-Neerlandicae]*, 8, 1–12.
- Bleeker, P. (1861) Iets over de vischfauna van het eiland Pinang. *Verslagen en Mededeelingen der Koninklijke Akademie van Wetenschappen (Afdeeling Natuurkunde)*, Amsterdam, 12, 64–80.
- Bleeker, P. (1862–1863) *Atlas Ichthyologique des Indes Orientales Néerlandaises, Publié sous les Auspices du Gouvernement Colonial Néerlandais. Tome II.— Siluroïdes, Chacoïdes et Hétérobranchoïdes*, Amsterdam, 112 p., pls. 49–101. [Issued in installments. Text pages 1–32 and plates 49–72 date to 1862; pages 33–112 and plates 73–101 date to 1863. See Boeseman (1963: 4) for details.]
- Bleeker, P. (1862a) Descriptions de quelques espèces nouvelles de Silures de Suriname. *Verslagen en Mededeelingen der Koninklijke Akademie van Wetenschappen (Afdeeling Natuurkunde)*, Amsterdam, 14, 371–389.
- Bleeker, P. (1862b) Notice sur les genres *Parasilurus*, *Eutropiichthys*, *Pseudeutropius*, et *Pseudopangasius*. *Verslagen en Mededeelingen der Koninklijke Akademie van Wetenschappen (Afdeeling Natuurkunde)*, Amsterdam, 14, 390–399.
- Bleeker, P. (1862c) Notice sur les genres *Trachelyopterichthys*, *Hemicetopsis* et *Pseudocetopsis*. *Verslagen en Mededeelingen der Koninklijke Akademie van Wetenschappen (Afdeeling Natuurkunde)*, Amsterdam, 14, 400–403.
- Bleeker, P. (1863a) Mémoire sur les poissons de la côte de Guinée. *Natuurkundige Verhandelingen van de Hollandsche Maatschappij der Wetenschappen te Haarlem (Ser. 2)*, 18, 1–136, col. pls. 1–28.
- Bleeker, P. (1863b) Sur quelques genres nouveaux du groupe des *Doras*. *Nederlandsch Tijdschrift voor de Dierkunde*, 1, 10–18.
- Bleeker, P. (1863c) Systema Silorum revisum. *Nederlandsch Tijdschrift voor de Dierkunde*, 1, 77–122.

- Bleeker, P. (1863d) Description de quelques espèces de poissons, nouvelles ou peu connues de Chine, envoyées au Musée de Leide par M.- G. Schlegel. *Nederlandsch Tijdschrift voor de Dierkunde*, 1, 135–150.
- Bleeker, P. (1863e) Description de trois espèces nouvelles de Siluroïdes de l'Inde archipelagique. *Nederlandsch Tijdschrift voor de Dierkunde*, 1, 368–371.
- Bleeker, P. (1863f) Description de trois espèces nouvelles de Siluroïdes de l'Inde archipelagique. *Verslagen en Mededeelingen der Koninklijke Akademie van Wetenschappen (Afdeeling Natuurkunde)*, Amsterdam, 15, 70–76.
- Bleeker, P. (1864a) Description des espèces de Silures de Suriname, conservées aux Musées de Leide et d'Amsterdam. *Natuurkundige Verhandelingen van de Hollandsche Maatschappij der Wetenschappen te Haarlem (Ser. 2)*, 20, 1–104, pls. 1–16.
- Bleeker, P. (1864b) Notice sur la faune ichthyologique de Siam. *Verslagen en Mededeelingen der Koninklijke Akademie van Wetenschappen (Afdeeling Natuurkunde)*, Amsterdam, 16, 352–358, 1 pl. [Also published in 1865 in *Nederlandsch Tijdschrift voor de Dierkunde*, 2, 96–99.]
- Bleeker, P. (1864c) *Rhinobagrus* et *Pelteobagrus* deux genres nouveaux de Siluroïdes de Chine. *Nederlandsch Tijdschrift voor de Dierkunde*, 2, 7–10.
- Bleeker, P. (1870) Description et figure d'une espèce inédite de *Hemibagrus* de Chine. *Verslagen en Mededeelingen der Koninklijke Akademie van Wetenschappen (Afdeeling Natuurkunde)*, Amsterdam (Ser. 2), 4, 257–258, 1 pl.
- Bleeker, P. (1872) Mémoire sur la faune ichthyologique de Chine. *Nederlandsch Tijdschrift voor de Dierkunde*, 4, 113–154.
- Bloch, M.E. (1794) *Naturgeschichte der Ausländischen Fische*, Vol. 8, Berlin, iv + 174 p., pls. 361–396.
- Bloch, M.E. (1797) *Ichtyologie, ou Histoire Naturelle des Poissons*, Vol. 2, Berlin.
- Bloch, M.E. & Schneider, J.G. (1801) *M. E. Blochii, Systema Ichthyologiae Iconibus ex Illustratum. Post Obitum Auctoris Opus Inchoatum Absolvit, Correxit, Interpolavit Jo. Gottlob Schneider; Saxo, Sumtibus Austoris Impressum et Bibliopolio Sanderiano Commissum*, Berolini, lx + 584 p., 110 pl.
- Blyth, E. (1858) Report of Curator, Zoological Department, for May, 1858. *Journal of the Asiatic Society of Bengal*, 27, 267–290.
- Blyth, E. (1860) Report on some fishes received chiefly from the Sitang River and its tributary streams, Tenasserim Provinces. *Journal of the Asiatic Society of Bengal*, 29, 138–174.
- Bocchino, A. (1964) Sobre un Pygidiidae (Pisces, Siluriformes) del Eoceno de Río Negro. *Ameghiniana*, 3, 185–189.
- Bockmann, F.A. (1994) Description of *Mastiglanis asopos*, a new pimelodid catfish from northern Brazil, with comments on phylogenetic relationships inside the subfamily Rhamdiinae (Siluriformes: Pimelodidae). *Proceedings of the Biological Society of Washington*, 107, 760–777.
- Bockmann, F.A. (1998) *Análise Filogenética da Família Heptapteridae (Teleostei, Ostariophysi, Siluriformes) e Redefinição de seus Gêneros*, Unpublished Ph.D. thesis, Universidade de São Paulo, São Paulo.
- Bockmann, F.A., Casatti, L. & Pinna, M.C.C., de. (2004) A new species of trichomycterid catfish from the Rio Paranapanema basin, southeastern Brazil (Teleostei: Siluriformes), with comments on the phylogeny of the family. *Ichthyological Exploration of Freshwaters*, 25, 225–242.
- Bockmann, F.A. & Ferraris, C.J., Jr. (2005) Systematics of the Neotropical catfish genera *Nemuroglanis* Eigenmann and Eigenmann 1889, *Imparales* Schultz 1944, and *Medemicthys* Dahl 1961 (Siluriformes: Heptapteridae). *Copeia*, 2005, 124–137.
- Bockmann, F.A. & Guazzelli, G.M. (2003) Heptapteridae. In: Reis, R.E., Kullander, S.O. & Ferraris, C.J., Jr. (Eds.), *Check list of the Freshwater Fishes of South and Central America*. Edipucrs, Porto Alegre, Brazil, pp. 406–431.
- Bockmann, F.A. & Pinna, M.C.C., de. (2004) *Heptapterus colletti* Steindachner, 1881: a member of the Asian bagrid genus *Olyra* erroneously assigned to the Neotropical fauna (Siluriformes: Ostariophysi). *Copeia*, 2004, 665–675.
- Bockmann, F.A. & Ribeiro, A.C. (2003) Description of a new suckermouth armoured catfish of the genus *Pareiorhina* (Siluriformes: Loricariidae), from southeastern Brazil. *Ichthyological Exploration of Freshwaters*, 14, 231–242.
- Bocourt, F. (1866) Poissons. In: Notes sur les reptiles, les batraciens et les poissons recueillis pendant un voyage dans le royaume de Siam. *Nouvelles Archives du Muséum d'Histoire Naturelle*, Paris, 2, 11–20, pl. 1.
- Boeseman, M. (1947) Revision of the fishes collected by Burger and Von Siebold in Japan. *Zoologische Mededelingen (Leiden)*, 28, i–vii + 1–242, pls. 1–5. [Also issued as a book, with the same title, by E. J. Brill, Leiden.]
- Boeseman, M. (1953) Scientific results of the Surinam Expedition 1948–1949. Part II.— Zoology, no. 2: The Fishes (I). *Zoologische Mededelingen (Leiden)*, 32, 1–24.
- Boeseman, M. (1954) On *Sciadeichthys (Selenaspis) walrechti*, a new South American catfish. *Zoologische Mededelingen (Leiden)*, 33, 59–62, pl. 3.
- Boeseman, M. (1957) On a collection of fishes from Stanley Pool (Belgian Congo). *Zoologische Mededelingen (Leiden)*,

- 35, 139–151, pl. 5.
- Boeseman, M. (1963) An annotated list of fishes from the Niger Delta. *Zoologische Verhandelingen* (Leiden), 61, 1–48, pls. 1–6.
- Boeseman, M. (1966) A new sisorid catfish from Java, *Sundagagata robusta* gen. et spec. nov. *Proceedings of the Koninklijke Nederlandse Akademie van Wetenschappen (Series C)*, 69, 242–247 + table.
- Boeseman, M. (1968) The genus *Hypostomus* Lacépède, 1803, and its Surinam representatives (Siluriformes, Loricariidae). *Zoologische Verhandelingen* (Leiden), 99, 1–89, pls. 1–18.
- Boeseman, M. (1969) Additional new species of *Hypostomus* La Cepède, 1803, from Surinam; with remarks on the apparent "gymnorhynchus-complex" (Siluriformes, Loricariidae). *Beaufortia*, 16, 119–136.
- Boeseman, M. (1971) The "comb-toothed" Loricariinae of Surinam, with reflections on the phylogenetic tendencies within the family Loricariidae (Siluriformes, Siluroidei). *Zoologische Verhandelingen* (Leiden), 116, 1–56, pls. 1–8.
- Boeseman, M. (1972) Notes on South American catfishes, including remarks on Valenciennes and Bleeker types in the Leiden Museum. *Zoologische Mededelingen* (Leiden), 47, 293–320, pls. 1–2.
- Boeseman, M. (1974) On two Surinam species of Hypoptopomatinae, both new to science (Loricariidae, Siluriformes, Ostariophysi). *Proceedings of the Koninklijke Nederlandse Akademie van Wetenschappen (Series C)*, 77, 251–271.
- Boeseman, M. (1976) A short review of the Surinam Loricariinae; with additional information on Surinam Harttiinae, including the description of a new species (Loricariidae, Siluriformes). *Zoologische Mededelingen* (Leiden), 50, 153–177, pls. 1–8.
- Boeseman, M. (1982) The South American mailed catfish genus *Lithoxus* Eigenmann, 1910, with the description of three new species from Surinam and French Guyana and records of related species (Siluriformes, Loricariidae). *Proceedings of the Koninklijke Nederlandse Akademie van Wetenschappen (Series C)*, 85, 41–58, pls. 1–5.
- Boeseman, M. (1983a) Introduction. In: P. Bleeker; *Atlas Ichthyologique des Indes Orientales Néerlandaises; Plates Originally Prepared for Planned Tomes XI–XIV*, Smithsonian Institution Press, Washington, pp. 1–12.
- Boeseman, M. (1983b) Some remarks on the South American pimelodid catfish usually known by the name of *Phractocephalus hemiliopterus* (Bloch & Schneider) (Pimelodidae, Siluriformes). *Zoologische Mededelingen* (Leiden), 57, 105–114.
- Bogačew. (1924) Nowie materiali k istorii tretitschnich slonow w Jugo-Wostotschnej Rossii. *Izwestij Aserbaidjanskogo Universiteta*, Baku, 3, for 1923/24, [Not seen, from Weiler, 1956: 187.]
- Böhlke, E.B. (1984) Catalog of type specimens in the ichthyological collection of the Academy of Natural Sciences of Philadelphia. *Academy of Natural Sciences, Philadelphia, Special Publication*, 14, i–viii + 1–246.
- Böhlke, J.E. (1950) A new catfish of the genus *Corydoras* from the Peruvian Amazon. *The Fish Culturist*, 30, 26–27.
- Böhlke, J.E. (1951a) A new miniature catfish of the genus *Corydoras* from the Rio Orinoco in Venezuela. *Annals and Magazine of Natural History (Ser. 12)*, 4, 824–827.
- Böhlke, J.E. (1951b) Description of a new achenipterid catfish of the genus *Pseudopapetus* from the Amazon Basin. *Stanford Ichthyological Bulletin*, 4, 38–40.
- Böhlke, J.E. (1953) A catalogue of the type specimens of Recent fishes in the Natural History Museum of Stanford University. *Stanford Ichthyological Bulletin*, 5, 1–168.
- Böhlke, J.E. (1959) Results of the Catherwood Foundation Peruvian Amazon Expedition. *Petacara*, a new genus for the bunocephalid catfish, *Bunocephalus dolichurus* Delsman. *Notulae Naturae (Philadelphia)*, 318, 1–6.
- Böhlke, J.E. (1970) A new species of the doradid catfish genus *Leptodoras*, with comments on related forms. *Proceedings of the California Academy of Sciences, Series 2*, 38, 53–61.
- Böhlke, J.E. (1980) *Gelanoglanis stroudi*: a new catfish from the Rio Meta system in Colombia (Siluriformes, Doradidae, Auchenipterinae). *Proceedings of the Academy of Natural Sciences, Philadelphia*, 132, 150–155.
- Bonaparte, C.L. (1838) Synopsis vertebratorum systematis. *Nuovi Annali delle Scienze Naturali (Bologna)*, 2, 105–133.
- Bonaparte, C.L. (1846) *Catalogo Metodico dei Pesci Europei*. Stamperia e Cartieri del Fibreno, Napoli, 98 p.
- Bonnaterre, J.P. (1788) *Tableau Encyclopédique et Méthodique des Trois Règnes de la Nature*.— *Ichthyologie*, Chez Panckoucke, Paris, lvi + 215 p., 102 pls.
- Bornbusch, A.H. (1991a) Redescription and reclassification of the silurid catfish *Apodoglanis furnessi* Fowler (Siluriformes: Siluridae), with diagnoses of three intrafamilial silurid subgroups. *Copeia*, 1991, 1070–1084.
- Bornbusch, A.H. (1991b) Monophyly of the catfish family Siluridae (Teleostei: Siluriformes), with a critique of previous hypotheses of the family's relationships. *Zoological Journal of the Linnean Society*, 101, 105–120.
- Bornbusch, A.H. (1995) Phylogenetic relationships within the Eurasian catfish family Siluridae (Pisces: Siluriformes), with comments on generic validities and biogeography. *Zoological Journal of the Linnean Society*, 115, 1–46.
- Bornbusch, A.H. & Lundberg, J.G. (1989) A new species of *Hemisilurus* (Siluriformes, Siluridae) from the Mekong

- River, with comments on its relationships and historical biogeography. *Copeia*, 1989, 434–444.
- Borodin, N.A. (1927a) A new blind catfish from Brazil. *American Museum Novitates*, 263, 1–5.
- Borodin, N.A. (1927b) Some new catfishes from Brazil. *American Museum Novitates*, 266, 1–7.
- Borodin, N.A. (1927c) *Pimelodus platicirris*, new species, and other notes on Brazilian catfishes. *American Museum Novitates*, 271, 1–4.
- Borodin, N.A. (1934) *Netuma hassleriana*, a new catfish from Panama. *Copeia*, 1934, 33–34.
- Borodin, N.A. (1936) On a collection of freshwater fishes from lakes Nyasa, Tanganyika and Viktoria in Central Afrika. *Zoologische Jahrbücher. Abteilung für Systematik, Ökologie und Geographie der Tiere*, 68, 1–34, pl. 1.
- Bosc, L.A.G. (1816) [Pisces accounts] Vol. 3. *Nouveau Dictionnaire d'Histoire Naturelle...*, Nouv. éd., Chez Deterville, Paris.
- Boulenger, G.A. (1887a) On new fishes from the lower Congo. *Annals and Magazine of Natural History (Ser. 5)*, 19, 148–149.
- Boulenger, G.A. (1887b) On new siluroid fishes from the Andes of Columbia. *Annals and Magazine of Natural History (Ser. 5)*, 19, 348–350.
- Boulenger, G.A. (1887c) An account of the fishes collected by Mr. C. Buckley in eastern Ecuador. *Proceedings of the Zoological Society of London*, 1887, 274–283, pls. 20–24.
- Boulenger, G.A. (1889) Descriptions of a new snake and two new fishes obtained by Dr. H. von Ihering in Brazil. *Annals and Magazine of Natural History (Ser. 6)*, 4, 265–267.
- Boulenger, G.A. (1890a) List of the reptiles, batrachians and freshwater fishes collected by Professor Moesch and Mr. Iversen in the district of Deli, Sumatra. *Proceedings of the Zoological Society of London*, 1890, 31–40.
- Boulenger, G.A. (1890b) Descriptions of two new species of the siluroid genus *Arges*. *Proceedings of the Zoological Society of London*, 1890, 450–452, pl. 41.
- Boulenger, G.A. (1891) An account of the siluroid fishes obtained by Dr. H. von Ihering and Herr Sebastian Wolff in the Province Rio Grande do Sul, Brazil. *Proceedings of the Zoological Society of London*, 1891, 231–235, pls. 25–26.
- Boulenger, G.A. (1892a) Description of a new siluroid fish from China. *Annals and Magazine of Natural History (Ser. 6)*, 9, 247.
- Boulenger, G.A. (1892b) On some new or little-known fishes obtained by Dr. J. W. Evans and Mr. Spencer Moore during their recent expedition to the Province of Matto Grosso, Brazil. *Annals and Magazine of Natural History (Ser. 6)*, 10, 9–12, pls. 1–2.
- Boulenger, G.A. (1894a) Descriptions of new freshwater fishes from Borneo. *Annals and Magazine of Natural History (Ser. 6)*, 13, 245–251.
- Boulenger, G.A. (1894b) Description of a new siluroid fish from Burma. *Annals and Magazine of Natural History (Ser. 6)*, 14, 196.
- Boulenger, G.A. (1895a) Descriptions of two new fishes obtained by Mr. C. Hose in Sarawak. *Annals and Magazine of Natural History (Ser. 6)*, 15, 247.
- Boulenger, G.A. (1895b) [Abstract of a report on a large collection of fishes formed by Dr. C. Ternetz in Matto Grosso and Paraguay, with descriptions of new species]. *Proceedings of the Zoological Society of London*, 1895, 523–529.
- Boulenger, G.A. (1896a) Descriptions of new fishes from the upper Congo. *Annals and Magazine of Natural History (Ser. 6)*, 17, 309–311.
- Boulenger, G.A. (1896b) Description of a new siluroid fish from the Organ Mountains, Brazil. *Annals and Magazine of Natural History (Ser. 6)*, 18, 154.
- Boulenger, G.A. (1896c) On a collection of fishes from the Rio Paraguay. *Transactions of the Zoological Society of London*, 24, 25–39, pls. 1–6.
- Boulenger, G.A. (1897a) On a collection of fishes from the island of Marajo, Brazil. *Annals and Magazine of Natural History (Ser. 6)*, 20, 294–299.
- Boulenger, G.A. (1897b) Descriptions of new fishes from the upper Congo.— II. *Annals and Magazine of Natural History (Ser. 6)*, 20, 422–427.
- Boulenger, G.A. (1897c) Viaggio del Dott. Alfredo Borelli nel Chaco boliviano e nella Repubblica Argentina. III.— Poissons. *Bollettino dei Musei di Zoologia ed Anatomia Comparata della Università di Torino*, 12, 1–4.
- Boulenger, G.A. (1898a) Description of a new siluroid fish from West Africa. *Annals and Magazine of Natural History (Ser. 7)*, 2, 415.
- Boulenger, G.A. (1898b) Descriptions of two new siluroid fishes from Brazil. *Annals and Magazine of Natural History (Ser. 7)*, 2, 477–478.
- Boulenger, G.A. (1898c) Viaggio del Dr. Enrico Festa nell' Equateur e regioni vicine. Poissons de l'Équateur.— Part I.

- Bollettino dei Musei di Zoologia ed Anatomia Comparata della Università di Torino*, 13, 1–13.
- Boulenger, G.A. (1898d) Report on the fishes recently obtained by Mr. J. E. S. Moore in Lake Tanganyika. *Proceedings of the Zoological Society of London*, 1898, 494–497.
- Boulenger, G.A. (1898e) On a collection of fishes from the Rio Jurua, Brazil. *Transactions of the Zoological Society of London*, 14, 421–428, pls. 39–42.
- Boulenger, G.A. (1898f) Report on the collection of fishes made by Mr. J. E. S. Moore in Lake Tanganyika during his expedition, 1895–96. *Transactions of the Zoological Society of London*, 15, 1–30, pls. 1–8.
- Boulenger, G.A. (1899a) Matériaux pour la faune du Congo. Poissons nouveaux du Congo. Troisième Partie. Silures, Acanthoptérygiens, Mastacembles, Plectognathes. *Annales du Musée du Congo, Zoologie*, 1, 39–58, pls. 20–29.
- Boulenger, G.A. (1899b) Matériaux pour la faune du Congo. Poissons nouveaux du Congo. Cinquième Partie. Cyprins, Silures, Cyprinodontes, Acanthoptérygiens. *Annales du Musée du Congo, Zoologie*, 1, 97–128, pls. 40–47.
- Boulenger, G.A. (1900a) Matériaux pour la faune du Congo. Poissons nouveaux du Congo. Sixième Partie. Mormyres, Characins, Cyprins, Silures, Acanthoptérygiens, Dipneustes. *Annales du Musée du Congo, Zoologie*, 1, 129–164, pls. 48–56.
- Boulenger, G.A. (1900b) Descriptions of three new species of siluroid fishes from southern Brazil. *Annals and Magazine of Natural History (Ser. 7)*, 5, 165–166.
- Boulenger, G.A. (1900c) Diagnoses of new fishes discovered by Mr. J. E. S. Moore in Lake Tanganyika. I.—*Cyprinidae, Siluridae*. *Annals and Magazine of Natural History (Ser. 7)*, 6, 478–481.
- Boulenger, G.A. (1900d) On some little-known African silurid fishes of the subfamily Doradinae. *Annals and Magazine of Natural History (Ser. 7)*, 6, 520–529.
- Boulenger, G.A. (1900e) List of the fishes collected by Mr. J. S. Budgett in the river Gambia. *Proceedings of the Zoological Society of London*, 1900, 511–516.
- Boulenger, G.A. (1901a) *Les Poissons du Bassin du Congo*. Publication de l'État Indépendant du Congo, Brussels, lxii + 532 p., 25 pls., 1 map.
- Boulenger, G.A. (1901b) Descriptions of four new African freshwater fishes. *Annals and Magazine of Natural History (Ser. 7)*, 7, 80–82.
- Boulenger, G.A. (1901c) Descriptions of three new siluroid fishes of the genus *Synodontis* discovered by Mr. W. L. S. Loat in the White Nile. *Annals and Magazine of Natural History (Ser. 7)*, 8, 10–12.
- Boulenger, G.A. (1901d) Diagnoses of four new fishes discovered by Mr. J. E. S. Moore in Lakes Albert and Albert Edward. *Annals and Magazine of Natural History (Ser. 7)*, 8, 12–14.
- Boulenger, G.A. (1901e) Diagnoses of new fishes discovered by Mr. W. L. S. Loat in the Nile. *Annals and Magazine of Natural History (Ser. 7)*, 8, 444–446.
- Boulenger, G.A. (1901f) Description of a new silurid fish of the genus *Anoplopterus*, from Cameroon. *Annals and Magazine of Natural History (Ser. 7)*, 8, 447–448.
- Boulenger, G.A. (1901g) Description of a new silurid fish of the genus *Gephyroglanis*, from South Africa. *Annals of the South African Museum*, 2, 227–228, pl. 13.
- Boulenger, G.A. (1902a) Descriptions of two new fishes of the genus *Loricaria* from north-western Ecuador. *Annals and Magazine of Natural History (Ser. 7)*, 9, 69–71.
- Boulenger, G.A. (1902b) Descriptions of new fishes and reptiles discovered by Dr. F. Silvestri in South America. *Annals and Magazine of Natural History (Ser. 7)*, 9, 284–288.
- Boulenger, G.A. (1902c) Descriptions of new fishes from the collection made by Mr. E. Degen in Abyssinia. *Annals and Magazine of Natural History (Ser. 7)*, 10, 421–439.
- Boulenger, G.A. (1902d) Additions à la faune ichthyologique de bassin du Congo. Matériaux pour la faune du Congo. *Annales du Musée du Congo, Zoologie*, 2, 19–57, pl. 7–16.
- Boulenger, G.A. (1902e) Contributions to the ichthyology of the Congo.—I. On some new fishes from the French Congo. *Proceedings of the Zoological Society of London*, 1902, pt. 1, 234–237, pls. 22–24.
- Boulenger, G.A. (1902f) On the fishes collected by Mr. S. L. Hinde in the Kenya District, East Africa, with descriptions of four new species. *Proceedings of the Zoological Society of London*, 1902, pt. 1, 221–224, pls. 16–17.
- Boulenger, G.A. (1902g) Contributions to the ichthyology of the Congo—II. On a collection of fishes from the Lindi River. *Proceedings of the Zoological Society of London*, 1902, pt. 1, 265–271, pl. 28–30.
- Boulenger, G.A. (1902h) Descriptions of two new fishes discovered by Dr. W. J. Ansorge in southern Nigeria. *Proceedings of the Zoological Society of London*, 1902, pt. 2, 623–624, pl. 37.
- Boulenger, G.A. (1903a) Description of a new fish of the genus *Arges* from Venezuela. *Annals and Magazine of Natural History (Ser. 7)*, 11, 601–602.

- Boulenger, G.A. (1903b) Description of a new silurid fish of the genus *Clarias* from British Central Africa. *Annals and Magazine of Natural History* (Ser. 7), 12, 362–363.
- Boulenger, G.A. (1903c) Descriptions of new freshwater fishes from southern Cameroon. *Annals and Magazine of Natural History* (Ser. 7), 12, 435–441.
- Boulenger, G.A. (1903d) Second account of the fishes collected by Dr. W. J. Ansorge in the Niger Delta. *Proceedings of the Zoological Society of London*, 1902, pt. 2, 324–330, pls. 28–29.
- Boulenger, G.A. (1903e) On the fishes collected by Mr. G. L. Bates in southern Cameroon. *Proceedings of the Zoological Society of London*, 1903, pt. 1, 21–29, pls. 1–5.
- Boulenger, G.A. (1904a) Descriptions of new West-African freshwater fishes. *Annals and Magazine of Natural History* (Ser. 7), 14, 16–20.
- Boulenger, G.A. (1904b) Report on the fishes collected by Mr. Oscar Neumann and Baron Carlo von Erlanger in Gallaland and southern Ethiopia. *Proceedings of the Zoological Society of London*, 1903, pt. 2, 328–334, pls. 29–31.
- Boulenger, G.A. (1904c) Descriptions of three new fishes discovered by the late Mr. J. S. Budgett in the Niger. *Proceedings of the Zoological Society of London*, 1904, pt. 1, 197–199, pls. 7–8.
- Boulenger, G.A. (1905a) On a small collection of fishes from the Kasai river (Congo). *Annals and Magazine of Natural History* (Ser. 7), 16, 640–642.
- Boulenger, G.A. (1905b) On a collection of fishes from Lake Bangwelo. *Annals and Magazine of Natural History* (Ser. 7), 16, 642–647.
- Boulenger, G.A. (1905c) On a second collection of fishes made by Mr. S. L. Hinde in the Kenya District, East Africa. *Proceedings of the Zoological Society of London*, 1905, pt. 1, 62–64, pl. 7.
- Boulenger, G.A. (1906a) On some fishes from the Kwango River (Congo System) in Angola, collected by Dr. W. J. Ansorge. *Annals and Magazine of Natural History* (Ser. 7), 17, 110–112.
- Boulenger, G.A. (1906b) Descriptions of new fishes discovered by Mr. E. Degen in Lake Victoria. *Annals and Magazine of Natural History* (Ser. 7), 17, 433–452.
- Boulenger, G.A. (1906c) Description of a new fish of the genus *Clarias* from Uganda. *Annals and Magazine of Natural History* (Ser. 7), 17, 569.
- Boulenger, G.A. (1906d) On a second species of the silurid genus *Mochocetus*. *Annals and Magazine of Natural History* (Ser. 7), 18, 147–148.
- Boulenger, G.A. (1906e) Description of a new silurid fish of the genus *Doumea*, Sauvage, from Angola. *Annals and Magazine of Natural History* (Ser. 7), 18, 347–348.
- Boulenger, G.A. (1906f) Fourth contribution to the ichthyology of Lake Tanganyika.—Report on the collection of fishes made by Dr. W. A. Cunnington during the Third Tanganyika Expedition, 1904–1905. *Transactions of the Zoological Society of London*, 17, 537–619, pls. 30–41.
- Boulenger, G.A. (1907a) *Zoology of Egypt: The Fishes of the Nile*, Hugh Rees, Ltd., London, li + 578 p., 97 pls.
- Boulenger, G.A. (1907b) Descriptions of three new freshwater fishes discovered by Mr. G. L. Bates in South Cameroon. *Annals and Magazine of Natural History* (Ser. 7), 20, 50–52.
- Boulenger, G.A. (1907c) Descriptions of three new fishes from Central Africa. *Annals and Magazine of Natural History* (Ser. 7), 20, 487–489.
- Boulenger, G.A. (1908a) Description of a new silurid fish of the genus *Synodontis* from South Cameroon. *Annals and Magazine of Natural History* (Ser. 8), 2, 30–31.
- Boulenger, G.A. (1908b) A revision of the African silurid fishes of the subfamily Clariinae. *Proceedings of the Zoological Society of London*, 1907, 1062–1097.
- Boulenger, G.A. (1909) Descriptions of new freshwater fishes discovered by Mr. G. L. Bates in South Cameroon. *Annals and Magazine of Natural History* (Ser. 8), 4, 186–188.
- Boulenger, G.A. (1910) On a large collection of fishes made by Dr. W. J. Ansorge in the Quanza and Bengo rivers, Angola. *Annals and Magazine of Natural History* (Ser. 8), 6, 537–561.
- Boulenger, G.A. (1911a) *Catalogue of the Fresh-water Fishes of Africa in the British Museum (Natural History)*, Vol. 2, British Museum (Natural History) Trustees, London, xii + 529 p.
- Boulenger, G.A. (1911b) Descriptions of new freshwater fishes discovered by Dr. W. J. Ansorge in Portuguese Guinea. *Annals and Magazine of Natural History* (Ser. 8), 7, 373–376.
- Boulenger, G.A. (1911c) Description of a new African fish of the genus *Clarias* from Lake Rukwa. *Annals and Magazine of Natural History* (Ser. 8), 8, 54.
- Boulenger, G.A. (1911d) Further descriptions of new freshwater fishes discovered by Dr. W. J. Ansorge in Portuguese Guinea. *Annals and Magazine of Natural History* (Ser. 8), 8, 56–57.

- Boulenger, G.A. (1911e) Descriptions of three new freshwater fishes discovered by Mr. G. L. Bates in South Cameroon. *Annals and Magazine of Natural History (Ser. 8)*, 8, 372–373.
- Boulenger, G.A. (1911f) Liste des poissons représentés dans une nouvelle collection de la rivière Sankuru (Kasai) reçue de M. E. Luja. *Bulletins mensuels. Société des naturalistes luxembourgeois [= Monatsberichte / Gesellschaften Luxemburger Naturfreunde]*, n. s., 5, 218–225.
- Boulenger, G.A. (1912a) Descriptions of two new fishes from the Nile system. *Annals and Magazine of Natural History (Ser. 8)*, 10, 601–602.
- Boulenger, G.A. (1912b) Poissons recueillis dans la Région du Bas-Congo par M. le Dr W. J. Ansorge. — Poissons, Batraciens et Reptiles. Matériaux pour la Faune du Congo. *Annales du Musée du Congo Belge, Zoologie (Ser. 1)*, 2, 1–28, pls. 17–22.
- Boulenger, G.A. (1912c) On a collection of fishes made by Mr. A. Blayney Percival in British East Africa to the East of Lake Baringo. *Proceedings of the Zoological Society of London*, 1912, 672–676, pls. 78–80.
- Boulenger, G.A. (1913) Descriptions of four new fishes discovered by Mr. G. L. Bates in the Nyong River, S. Cameroon. *Annals and Magazine of Natural History (Ser. 8)*, 12, 67–70.
- Boulenger, G.A. (1915) Diagnoses de poissons nouveaux. II.— Mormyrides, Kneriides, Characinae, Cyprinidae, Siluridae. (Mission Stappers au Tanganyika-Moero.). *Revue de Zoologie Africaine*, 4, 162–171.
- Boulenger, G.A. (1916) *Catalogue of the Fresh-water Fishes of Africa in the British Museum (Natural History)*, Vol. 4, Trustees, London, xxvii + 392 p.
- Boulenger, G.A. (1917a) Description of a new silurid fish from Natal. *Annals of the Durban Museum*, 1, 432.
- Boulenger, G.A. (1917b) Descriptions of new fishes from Lake Tanganyika forming part of the collection made by the late Dr. L. Stappers for the Belgian Government. *Annals and Magazine of Natural History (Ser. 8)*, 20, 363–368.
- Boulenger, G.A. (1918) On some fishes from the Shari River, with descriptions of two new species. *Annals and Magazine of Natural History (Ser. 9)*, 2, 426–427.
- Boulenger, G.A. (1919) Descriptions de poissons nouveaux du Congo. *Revue de Zoologie Africaine*, 7, 158–161.
- Boulenger, G.A. (1920a) Poissons recueillis au Congo Belge par l'expédition du Dr. C. Christy. Zoologie. — Série I. Matériaux pour la Faune du Congo. *Annales du Musée du Congo Belge, Zoologie (Ser. 1)*, 2, 1–38, pls. 23–25.
- Boulenger, G.A. (1920b) Descriptions de deux poissons nouveaux de l'Ituri. *Revue de Zoologie Africaine*, 8, 148–150.
- Boulenger, G.A. (1920c) On some new fishes from near the west coast of Lake Tanganyika. *Proceedings of the Zoological Society of London*, 1919, 399–404.
- Boulenger, G.A. (1923) Descriptions of three new fresh-water fishes from northern Rhodesia. *Annals of the South African Museum*, 13, 437–438.
- Bowdich, S.L. (1825) Fishes of Madeira. In: T. E. Bowdich, *Excursions in Madeira and Porto Santo During the Autumn of 1823, While on His Third Voyage to Africa*. G. B. Whittaker, London, pp. 121–125, 233–238.
- Breder, C.M., Jr. (1925) New loricariate, characin and poeciliid fishes from the Rio Chucunaque, Panama. *American Museum Novitates*, 180, 1–9.
- Breder, C.M. (1927) The fishes of the Rio Chucunaque drainage, Eastern Panama. *Bulletin of the American Museum of Natural History*, 57, 91–176, 6 pls.
- Briggs, J.C. (1961) Emended generic names in Berg's classification of fishes. *Copeia*, 1961, 161–166.
- Britski, H.A. (1969) Lista dos tipos de peixes das coleções do Departamento de Zoologia da Secretaria da Agricultura de São Paulo. *Papeis Avulsos de Zoologia, São Paulo*, 22, 197–215.
- Britski, H.A. (1972) *Sistemática e Evolução dos Auchenipteridae e Ageneiosidae (Teleostei, Siluriformes)*, Doctoral Dissertation, Universidade de São Paulo, São Paulo, Brazil.
- Britski, H.A. (1981) Sobre um novo gênero e espécie de Sorubiminae da Amazônia (Pisces, Siluriformes). *Papeis Avulsos de Zoologia, São Paulo*, 34, 109–114.
- Britski, H.A. (1993) Uma nova espécie de *Phenacorhamdia* da bacia do alto Paraná (Pisces, Siluriformes). *Comunicações do Museu de Ciências da PUCRS, Série Zoologia, Porto Alegre*, 6, 41–50.
- Britski, H.A. (1997) Descrição de um novo gênero de Hypoptopomatinae, com duas espécies novas (Siluriformes, Loricariidae). *Papeis Avulsos de Zoologia, São Paulo*, 40, 231–255.
- Britski, H.A., de Silimon, K.Z. de S. & Lopes, B.S. (1999) *Peixes do Pantanal: Manual de Identificação*, Embrapa, Brasília, 184 p.
- Britski, H.A. & Garavello, J.C. (1984) Two new southeastern Brazilian genera of Hypoptopomatinae and a redescription of *Pseudotocinclus* Nichols, 1919 (Ostariophysi, Loricariidae). *Papeis Avulsos de Zoologia, São Paulo*, 35, 225–41.
- Britski, H.A. & Garavello, J.C. (2002) *Parotocinclus jumbo*, a new species of the subfamily Hypoptopomatinae from northeastern Brazil (Ostariophysi: Loricariidae). *Ichthyological Exploration of Freshwaters*, 13, 279–288.

- Britski, H.A. & Garavello, J.C. (2003) *Hisonotus insperatus*: new species, from the upper Rio Paraná basin (Pisces: Ostariophysi: Loricariidae). *Copeia*, 2003, 588–593.
- Britski, H.A. & Langeani, F. (1988) *Pimelodus paranaensis* sp. n., um novo Pimelodidae (Pisces, Siluriformes) do Alto Paraná, Brasil. *Revista Brasileira de Zoologia*, 5, 409–417.
- Britski, H.A. & Ortega, H. (1983) *Trichogenes longipinnis*, novo gênero e espécie de Trichomycterinae do sudeste do Brazil (Pisces, Siluriformes). *Revista Brasileira de Zoologia*, 1, 211–216.
- Britski, H.A., Sato, Y. & Rosa, A.B.S. (1986) *Manual de Identificação de Peixes da Região de Três Marias (com Chaves de Identificação para os Peixes da Bacia do São Francisco)*. CODEVASF, Brasília, 115 p.
- Britto, M.R. (1998) Two new species of the genus *Aspidoras* (Siluriformes: Callichthyidae) from central Brazil. *Ichthyological Exploration of Freshwaters*, 8, 359–368.
- Britto, M.R. (2000) *Aspidoras depinnai* (Siluriformes: Callichthyidae): A new species from northeastern Brazil. *Copeia*, 2000, 1048–1055.
- Britto, M.R. (2003) Phylogeny of the subfamily Cordoradinae Hoedeman, 1952 (Siluriformes: Callichthyidae), with a definition of its genera. *Proceedings of the Academy of Natural Sciences, Philadelphia*, 153, 119–154.
- Britto, M.R. & Castro, R.M.C. (2002) New corydorine catfish (Siluriformes: Callichthyidae) from the upper Paraná and São Francisco: the sister group of *Brochis* and most of *Corydoras* species. *Copeia*, 2002, 1006–1015.
- Britto, M.R. & Lima, F.C.T. (2003) *Corydoras tukano*, a new species of corydoradine catfish from the rio Tiquié, upper rio Negro basin, Brazil (Ostariophysi: Siluriformes: Callichthyidae). *Neotropical Ichthyology*, 1, 83–91.
- Britto, M.R., Lima, F.C.T. & Moreira, C.R. (2002) *Aspidoras velites*, a new catfish from the upper Rio Araguaia basin, Brazil (Teleostei: Siluriformes: Callichthyidae). *Proceedings of the Biological Society of Washington*, 115, 727–736.
- Britto, M.R., Lima, F.C.T. & Santos, A.C.A. (2005) A new *Aspidoras* (Siluriformes: Callichthyidae) from rio Paraguaçu basin, Chapada Diamantina, Bahia, Brazil. *Neotropical Ichthyology*, 3, 473–479.
- Britto, M.R. & Moreira, C.R. (2002) *Otocinclus tapirape*: a new hypoptopomatine catfish from central Brazil (Siluriformes: Loricariidae). *Copeia*, 2002, 1063–1069.
- Britto, M.R. & Reis, R.E. (2005) A new *Scleromystax* species (Siluriformes: Callichthyidae) from coastal rivers of Southern Brazil. *Neotropical Ichthyology*, 3, 481–488.
- Britz, R. & Ferraris, C.J., Jr. (2003) A new species of the Asian catfish genus *Pseudolaguvia* from Myanmar (Teleostei: Ostariophysi: Siluriformes: Erethistidae). *Zootaxa*, 388, 1–8.
- Brown, B.A. & Ferraris, C.J., Jr. (1988) Comparative osteology of the Asian catfish family Chacidae, with the description of a new species from Burma. *American Museum Novitates*, 2907, 1–16.
- Brunet, M. & Franco-Thaciene, M.P. (2000) Chad: discovery of a vertebrate fauna close to the Mio-Pliocene border. *Journal of Vertebrate Paleontology*, 20, 205–209.
- Bruwer, E.E. & van der Bank, F.H. (2003) A morphological key for the identification of southern African *Synodontis* species Cuvier, 1816 (Teleostei: Mochokidae). *African Journal of Aquatic Science*, 28, 183–186.
- Buckup, P.A. (1981) *Microlepidogaster taimensis* sp. n., novo Hypoptopomatinae da Estação Ecológica do Taim, Rio Grande do Sul, Brasil (Ostariophysi, Loricariidae). *Iheringia, Série Zoologia*, 60, 19–31.
- Buckup, P.A. (1988) The genus *Heptapterus* (Teleostei, Pimelodidae) in southern Brazil and Uruguay, with the description of a new species. *Copeia*, 1988, 641–653.
- Buckup, P.A. & Malabarba, L.R. (1990) Sobre as localidades-tipo de *Characidium rachovii* e *Corydoras macropterus* (Teleostei: Ostariophysi). *Comunicações do Museu de Ciências da PUCRS, Série Zoologia, Porto Alegre*, 23, 103–109.
- Buen, F. de (1946a) Investigaciones sobre ictiología Mexicana. III.— La ictiofauna del Lago de Chapala, con descripción de una nueva especie (*Haustor ochoterenai* de Buen). *Anales del Instituto de Biología de la Universidad Nacional Autónoma de México*, 17, 261–281.
- Buen, F. de (1946b) Ictiogeografía continental mexicana (I, II, y III). *Revista de la Sociedad Mexicana de Historia Natural*, 7, 87–138.
- Burchell, W.J. (1822) *Travels in the Interior of Southern Africa*, 2 Vols, Longman, Hurst, Rees, Orme, and Brown, London.
- Burgess, W.E. (1982) *Corydoras adolfoi*, a new species of catfish (Siluriformes, Callichthyidae) from the upper Rio Negro, Brazil, near São Gabriel da Cachoeira. *Tropical Fish Hobbyist*, 30, 15–16.
- Burgess, W.E. (1983) *Corydoras robineae*, a new species of callichthyid catfish from Brazil. *Tropical Fish Hobbyist*, 31, 42–43.
- Burgess, W.E. (1989) *An Atlas of Freshwater and Marine Catfishes. A Preliminary Survey of the Siluriformes*, T.F.H. Publications, Neptune City, New Jersey, 784 p.

- Burgess, W.E. (1993) Three new species of catfishes of the genus *Corydoras* (Callichthyidae: Siluriformes). *Tropical Fish Hobbyist*, 41, 152–158.
- Burgess, W.E. (1994) *Scobinancistrus aureatus*, a new species of loricariid catfish from the Rio Xingu (Loricariidae: Ancistrinae). *Tropical Fish Hobbyist*, 43, 236–242.
- Burgess, W.E. (1997) *Corydoras coriatae*, a new species of callichthyid catfish related to *Corydoras fowleri*. *Tropical Fish Hobbyist*, 45, 138–147.
- Burr, B.M., Eisenhour, D.J. & Grady, J.M. (2005) Two new species of *Noturus* (Siluriformes: Ictaluridae) from the Tennessee River drainage: description, distribution, and conservation status. *Copeia*, 2005, 783–802.
- Bussing, W.A. (1970) Two new species of catfishes of the genera *Nannorhamdia* and *Imparales* (family Pimelodidae) from Central America. *Contributions in Science, Los Angeles County Museum*, 196, 1–11.
- Bussing, W.A. (1987) *Peces de las Aguas Continentales de Costa Rica*, Editorial de la Universidad de Costa Rica, San José, 271 p.
- Bussing, W.A. (1998) *Peces de las Aguas Continentales de Costa Rica.—Freshwater Fishes of Costa Rica, Second Edition*, Editorial de la Universidad de Costa Rica, San José, 468 p. [Also, Revista de Biología Tropical, 46 (supplement 2): 1–468.]
- Cagnolaro, L. & Violani, C. (1988) Introduction to the anastatic reprint of "Vertebratorum Synopsis..." by E. Cornalia (1849). *Atti della Società Italiana di Scienze Naturali, Milano*, 129, 433–434 + 16 pp. and 1 pl.
- Cala, P. (1981) Catalogo de los ejemplares tipo en la colección de peces del Instituto de Ciencias Naturales — Museo de Historia Natural de la Universidad Nacional de Colombia. *Lozania (Acta Zoologica Colombiana)*, 34, 1–5.
- Campanario, C.M. & Pinna, M.C.C., de. (2000) A new species of the primitive trichomycterid subfamily Copionodontinae from northeastern Brazil (Teleostei: Trichomycteridae). *Ichthyological Exploration of Freshwaters*, 11, 369–375.
- Cantor, T.E. (1842) General features of Chusan, with remarks on the flora and fauna of that island. *Annals and Magazine of Natural History (n. s.)*, 9, 265–278, 361–370, 481–493.
- Cantor, T.E. (1849) Catalogue of Malayan fishes. *Journal of the Asiatic Society of Bengal*, 18, i–xii + 981–1443, pls. 1–14. [Also issued in 1850, as a book with the same title, by J. Thomas, Calcutta.]
- Capello, F.B. (1870) Noticia ácerca de um peixe pouco conhecido proveniente do Brasil. *Jornal do Sciências Mathemáticas, Physicas e Naturaes, Lisboa*, 2, 64–69, pl. 7.
- Caramaschi, E.P. & Caramaschi, U. (1991) Taxonomic status of the trichomycterid catfish *Trichomycterus itatiayae*. *Copeia*, 1991, 222–224.
- Cardona, L. & Guerao, G. (1994) *Astroblepus riberae*, una nueva especie de siluriforme cavernícola del Perú (Osteichthyes: Astroblepidae). *Mémoires de Biospéologie*, 21–24.
- Cardoso, A.R. (2004) *Hemiancistrus megalopteryx*, a new species of loricariid catfish from the rio Tubarão drainage, Santa Catarina State, Brazil (Teleostei: Siluriformes: Trichomycteridae). *Ichthyological Exploration of Freshwaters*, 15, 173–178.
- Cardoso, A.R. & Lucinda, P.H.F. (2003) Three new species of *Hemiancistrus* (Teleostei: Siluriformes: Loricariidae) from the rio Tocantins basin with comments on the genus. *Ichthyological Exploration of Freshwaters*, 14, 73–84.
- Cardoso, A.R. & Malabarba, L.R. (1999) Description of three new species of *Hemiancistrus* Bleeker, 1862, from southern Brazil (Teleostei: Siluriformes: Loricariidae). *Comunicações do Museu de Ciências da PUCRS, Série Zoologia, Porto Alegre*, 12, 141–161.
- Cardoso, A.R. & Pezzi da Silva, J.F. (2004) Two new species of the genus *Hemiancistrus* Bleeker (Teleostei: Siluriformes: Loricariidae) from the upper rio Uruguai basin. *Neotropical Ichthyology*, 2, 1–8.
- Carranza, J. (1954) Descripción del primer bagre anoftalmo y depigmentado encontrado en aguas mexicanas. *Ciencia (Mexico City)*, 14, 129–136, pl. 1.
- Carvalho, A.L. (1967) Novos dados para o conhecimento de *Phreatobius cisternarum* Goeldi (Pisces, Pygidiidae, Phreatobiinae). *Atlas do Simpósio sobre a Biota Amazonica 3 (Limnologia)*, 83–88.
- Casciotta, J.R. & Almirón, A.E. (1996) *Scleronema minutum* (Boulenger) y *Ochmacanthus batrachostoma* (M. Ribeiro) (Siluriformes: Trichomycteridae), dos citas nuevas para la cuenca del Plata en Argentina. *Neotropica*, 42, 51–54.
- Casier, E. (1958) Contribution à l'étude des poissons fossiles des Antilles. *Mémoires Suisses de Paléontologie*, 74, 1–95, pls. 1–3.
- Casier, E. (1960) Note sur la collection des poissons Paléocènes et Eocènes de l'Enclave de Cabinda (Congo): recueillis par J. Bequaert au cours de sa mission in 1913. *Annales du Musée Royal du Congo Belge, A — Minérologie, Géologie, Paléontologie, Sér. 3*, 1, 1–47, 2 pl.
- Castaño, M.L., Mojica C., J.I. & Royero, R. (2005) Revisión taxonómica del género *Pimelodella*, Eigenmann &

- Eigenmann, 1888 (Pisces, Siluriformes: Heptapteridae) de la región transandina de Colombia. *Acta Biologia Colombia*, 19, 79.
- Castelnau, F.L. (1855) Poissons. In: *Animaux Nouveaux or Rares Recueillis Pendant l'Expédition dans les Parties Centrales de l'Amérique du Sud, de Rio de Janeiro à Lima, et de Lima au Para; Exécutée par Ordre du Gouvernement Français Pendant les Années 1843 a 1847*, P. Bertrand, Paris, pp. i–xii + 1–112, 50 pl.
- Castelnau, F.L. (1861) *Mémoire sur les Poissons de l'Afrique Australe*. J. Ballière et fils, Paris, vii + 78 p.
- Castelnau, F.L. (1873) Contribution to the ichthyology of Australia.— No. VIII. *Proceedings of the Zoological and Acclimatisation Society of Victoria*, 2, 123–149.
- Castelnau, F.L. (1875) Researches on the fishes of Australia. *Intercolonial Exhibition Essays*, 1875–6, 1–52.
- Castelnau, F.L. (1878a) Australian fishes. New or little known species. *Proceedings of the Linnean Society of New South Wales*, 2, 225–248, pls. 2–3.
- Castelnau, F.L. (1878b) On several new Australian (chiefly) fresh-water-fishes. *Proceedings of the Linnean Society of New South Wales*, 3, 140–144.
- Castillo, G.O. & Brull, G.O. (1989) *Ageneiosus magoi* una nueva especie de bagre ageneiosido (Teleostei, Siluriformes) para Venezuela y algunas notas sobre su historia natural. *Acta Biologica Venezolica*, 12, 72–87.
- Castro, D.M. (1986a) Los bagres de la subfamilia Sorubiminae de la Orinoquía y Amazonía Colombiana (Siluriformes – Pimelodidae). *Boletín Ecotrópica*, 13, 1–40.
- Castro, D.M. (1986b) *Corydoras gomezi* a new species from Colombia (Pisces, Siluriformes, Callichthyidae). *Boletín Ecotrópica*, 15, 33–38.
- Castro, D.M. (1987) The fresh-water fishes of the genus *Corydoras* from Colombia, including two new species (Pisces, Siluriformes, Callichthyidae). *Boletín Ecotrópica*, 16, 23–57, pls. 1–11.
- Catesby, M. (1731–1743) *The Natural History of Carolina, Florida and the Bahama Islands; Containing the Figures of Birds, Beasts, Fishes, Serpents, Insects and Plants: Together with their Descriptions in English and French etc.*, 2 vols., Privately printed, London.
- Catesby, M. (1771) *The Natural History of Carolina, Florida and the Bahama Islands; Containing the Figures of Birds, Beasts, Fishes, Serpents, Insects and Plants: ... Together with their Descriptions in English and French. ..., Revised by Mr. Edwards*, 3rd ed. 2 vols, Printed for B. White, London.
- Ceas, P.A. & Page, L.M. (1996) *Chaetostoma yurubiense* (Teleostei: Siluriformes), a new species of loricariid catfish from the Aroa, Urama, and Yaracuy River systems in Venezuela. *Copeia*, 1996, 671–677.
- Chabanaud, P. (1934) Mission Jean Thomas en Afrique équatoriale française (septembre 1929 à mai 1930). *Faune des Colonies Françaises*, 5, 195–231.
- Chakrabarty, P. & Ng, H.H. (2005) The identity of catfishes identified as *Mystus cavasius* (Hamilton, 1822) (Teleostei: Bagridae), with a description of a new species from Myanmar. *Zootaxa*, 1093, 1–24.
- Chamon, C.C., Aranda, A.T. & Buckup, P.A. (2005) *Pareiorhina brachyrhyncha* (Loricariidae: Siluriformes): a new species of fish from the Paraíba do Sul slope of Serra da Mantiqueira, southeastern Brazil. *Copeia*, 2005 550–558.
- Chang, F. (1999) New species of *Myoglanis* (Siluriformes, Pimelodidae) from the Río Amazonas, Peru. *Copeia*, 1999, 434–438.
- Chang, F. & Castro, E. (1999) *Crossoloricaria bahuaja*, a new loricariid fish from Madre de Dios, southeastern Peru. *Ichthyological Exploration of Freshwaters*, 10, 81–88.
- Chang, M.-M. & Zhou, J.-J. (1993) A brief survey of the Chinese Eocene ichthyofauna. *Kaupia Darmstaedler Beiträge zur Naturgeschichte*, 2, 157–162.
- Chang, Y.-W. & Wu, C.-T. (1965) [A new pangasid cat-fish, *Sinopangasius semicultratus* gen. et sp. nov., found in China]. *Acta Zootaxonomica Sinica*, 2, 11–14. [In Chinese and English.]
- Chardon, M. (1968) Anatomie comparée de l'appareil de Weber et structures connexes chez les Siluriformes. *Annales du Musée Royal de l'Afrique Centrale, Series in 8°, Sciences Zoologiques*, 169, 1–277.
- Chaudhuri, B.L. (1911) Contributions to the fauna of Yunnan based on collections made by J. Coggin Brown, B.Sc., 1909–1910. Part II.— Fishes. *Records of the Indian Museum*, 6, 13–24, pl. 1.
- Chaudhuri, B.L. (1912) Descriptions of some new species of freshwater fishes from north India. *Records of the Indian Museum*, 7, 437–444, pls. 38–41.
- Chaudhuri, B.L. (1913) Zoological results of the Abor Expedition, 1911–12. XVIII.— Fish. *Records of the Indian Museum*, 8, 243–257, pls. 7–9.
- Chaudhuri, B.L. (1916) Fauna of the Chilka Lake. Fish.— Part I. *Memoirs of the Indian Museum*, 5, 403–439.
- Chaudhuri, B.L. (1919) Report on a small collection of fish from Putao (Hkamti Long) on the northern frontier of Burma. *Records of the Indian Museum*, 16, 271–287, pl. 22.

- Chaux, J. & Fang, P.W. (1949a) Catalogue des Siluroidei d'Indochine de la collection du Laboratoire des Pêches Coloniales au Muséum, avec la description de six espèces nouvelles. *Bulletin du Muséum National d'Histoire Naturelle* (2^e Série), 21, 194–201.
- Chaux, J. & Fang, P.W. (1949b) Catalogue des Siluroidei d'Indochine de la collection du Laboratoire des Pêches Coloniales au Muséum, avec la description de six espèces nouvelles. (Suite et fin). *Bulletin du Muséum National d'Histoire Naturelle* (2^e Série), 21, 342–346.
- Chen, H.-L. (1977) [A review of the Chinese Siluridae]. *Acta Hydrobiologica Sinica*, 6, 197–218, pls. 1–2. [In Chinese, with English abstract.]
- Chen, X.-P. & Lundberg, J.G. (1995) *Xiurenbagrus*, a new genus of amblycipitid catfishes (Teleostei: Siluriformes), and phylogenetic relationships among the genera of Amblycipitidae. *Copeia*, 1995, 780–800.
- Chen, X.-Y., Ferraris, C.J., Jr. & Yang, J.-X. (2005) A new species of catfish of the genus *Clariasoma* (Siluriformes: Schilbidae) from the Salween River, Yunnan, China. *Copeia*, 2005 566–570.
- Chevey, P. (1931) Sur un nouveau silure géant du Bassin du Mékong *Pangasianodon gigas* nov. g., nov. sp. *Bulletin de la Société Zoologique de France*, 55, for 1930, 536–542, pl. 1.
- Chevey, P. (1932) *Poissons des Campagnes du "de Lanessan"* (1925–1929). 1^{re} Partie. *Travaux de l'Institut Océanographique de l'Indochine*, 4^o Mémoire. Gouvernement General de l'Indochine, Saigon, 155 p., 50 pls.
- Chockley, B.R. & Armbruster, J.W. (2002) *Panaque changae*, a new species of catfish (Siluriformes: Loricariidae) from eastern Peru. *Ichthyological Exploration of Freshwaters*, 13, 81–90.
- Choo, I.-Y. (1987) *Systematic Studies on the Torrent Catfish, Family Amblycipitidae (Pisces: Siluriformes)* from Korea. Department of Biology, Choong Ang University.
- Chu, X.-L. (1979) [Systematics and evolutionary pedigree of the glyptosternoid fishes (family Sisoridae)]. *Acta Zootaxonomica Sinica*, 4, 72–82. [In Chinese.]
- Chu, X.-L. (1981) [Taxonomic revision of the genera *Pareuchiloglanis* and *Euchiloglanis*]. *Zoological Research*, 2, 25–31. [In Chinese.]
- Chu, X.-L. (1982) [Phylogeny of the genus *Pseudecheneis* (Siluriformes: Sisoridae), with descriptions of two new species]. *Acta Zootaxonomica Sinica*, 7, 428–437. [In Chinese.]
- Chu, X.-L. (1986) [Glyptosternoid fishes in the Himalaya-Hengduan Mountain region]. *Freshwater Fishes*, 12, 39–44. [In Japanese.]
- Chu, X.-L. & Chen, Y.-R. (Eds.) (1990) *The Fishes of Yunnan, China. Part II — Cyprinidae [sic, non-Cyprinidae]*, Science Press, Beijing.
- Chu, X.-L. & Kuang, P.R. (1990) Sisoridae (Siluriformes). In: Chu, X.-L. & Chen, Y.-R. (Eds.), *The Fishes of Yunnan, China. Part II — Cyprinidae [sic, non-Cyprinidae]*, Science Press, Beijing, pp. 170–225. [In Chinese.]
- Chu, X.-L. & Mo, T.-P. (1999) Sisoridae. In: Chu, X.-L., Zheng, B.-S., Dai, D.-Y. & al. (Eds.), *Fauna Sinica: Osteichthyes: Siluriformes*, Science Press, Beijing, pp. 114–181. [In Chinese.]
- Chu, X.-L., Zheng, B.-S. & Dai, D.-Y. (1999) *Fauna Sinica. Osteichthyes: Siluriformes*, Science Press, Beijing, vii + 230 p. [In Chinese.]
- Cione, A.L. (1982) Peces del Pleistoceno tardío de la provincia de Buenos Aires. Consideraciones biogeográficas. *Circular Informativa de la Asociación Paleontológica Argentina*, 8, 12.
- Cione, A.L. (1986) Los peces continentales del Cenozoico de Argentina. Su significación paleoambiental y paleobiogeográfica. *Congreso Argentino de Paleontología y Bioestratigrafía*, No. 4, Actas. Vol. 2, Mendoza, pp. 101–106.
- Cione, A.L., Azpelicueta, M.M. & Caille, G. (1996) An arid catfish (Osteichthyes: Siluriformes) from marine middle Miocene beds of Patagonia; Recent arid biogeography in southern South America. *Cienc. Nat. Bernardino Rivadavia, Paleontología [? Revista del Museo Argentino de Ciencias Naturales "Bernardino Rivadavia" e Instituto Nacional de Investigación de las Ciencias Naturales. Paleontología]*, 3, 111–120.
- Cione, A.L., Pereira, S.M., Alonso, R. & Arias, J. (1985) Los bagres (Osteichthyes, Siluriformes) de la Formación Yacoraite (Cretácico tardío) del Noroeste Argentino: Consideraciones biogeográficas y bioestratigráficas. *Ameghiniana*, 21, 294–304.
- Cione, A.L. & Prashad, G.V.R. (2002) The oldest known catfish (Teleostei: Siluriformes) from Asia (India, Late Cretaceous). *Journal of Paleontology*, 76, 190–193.
- Cione, A.L. & Torno, A.E. (1988) Assignment of the bony fish "*Propygodium primaevus*"(a supposed siluriform from the Tertiary of Patagonia) to the Order Perciformes. *Journal of Paleontology*, 62, 656–657.
- Cloquet, H. (1816) [Pisces accounts. vol. 4, supplement]. *Dictionnaire des Sciences Naturelles*, Imprint Le Normant, Paris, pp. 52–53. [Not seen, citation from Bailey & Stewart, 1983]
- Coad, B.W. (1981a) *Glyptothorax silviae*, a new species of sisorid catfish from southwestern Iran. *Japanese Journal of*

- Ichthyology*, 27, 291–295.
- Coad, B.W. (1981b) Fishes of Afghanistan, an annotated checklist. *Publications in Zoology, National Museum of Natural Sciences (Canada)*, 14, i–v + 1–26.
- Coad, B.W. (1995) Freshwater fishes of Iran. *Acta Scientiarum Naturalium Academiae Scientiarum Bohemicae, Brno*, 29, 1–64.
- Coad, B.W. & Delmastro, G.B. (1985) Notes on a sisorid catfish from the Black Sea drainage of Turkey. *Cybium (3e série)*, 9, 221–224.
- Coad, B.W. & Holcik, J. (2000) On *Silurus* species from Iran (Actinopterygii: Siluridae). *Folia Zoologica*, 49, 139–148.
- Cockerell, T. (1925) A fossil fish of the family Callichthyidae. *Science (n. s.)*, 62, 397–398.
- Collette, B.B. (1962) *Astroblepus pholeti*, a new species of cave-dwelling catfish from eastern Ecuador. *Proceedings of the Biological Society of Washington*, 75, 311–314.
- Cope, E.D. (1864) On a blind silurid, from Pennsylvania. *Proceedings of the Academy of Natural Sciences, Philadelphia*, 16, 231–233.
- Cope, E.D. (1867) Supplement on some new species of American and African fishes. *Transactions of the American Philosophical Society*, 13, 400–407.
- Cope, E.D. (1868) On the distribution of fresh-water fishes in the Allegheny region of southwestern Virginia. *Journal of the Academy of Natural Sciences of Philadelphia (Ser. 2)*, 6, 207–247, pls. 26–28 [listed as 22–24].
- Cope, E.D. (1870a) Partial synopsis of the fishes of the fresh waters of North Carolina. *Proceedings of the American Philosophical Society*, 11, 448–495.
- Cope, E.D. (1870b) Contribution to the ichthyology of the Marañon. *Proceedings of the American Philosophical Society*, 11, 559–570.
- Cope, E.D. (1871a) [Fishes from the Amazon above the mouth of the Rio Negro]. *Proceedings of the Academy of Natural Sciences, Philadelphia*, 23, 55.
- Cope, E.D. (1871b) [Some anatomical points of importance in the classification of the siluroids of the Amazon]. *Proceedings of the Academy of Natural Sciences, Philadelphia*, 23, 112–113.
- Cope, E.D. (1872a) On the fishes of the Ambyiacu River. *Proceedings of the Academy of Natural Sciences, Philadelphia*, 23, for 1871, 250–294, pls 3–16.
- Cope, E.D. (1872b) Notices of new Vertebrata from the upper waters of Bitter Creek, Wyoming Territory. *Proceedings of the American Philosophical Society*, 7, 483–486.
- Cope, E.D. (1873) On the extinct Vertebrata of the Eocene of Wyoming observed by the expedition of 1872, with notes on the geology. Sixth Report. *U. S. Geological and Geographic Survey of the Territories*, 546–649.
- Cope, E.D. (1874a) Supplementary notices of fishes from the freshwater Tertiaries of the Rocky Mountains. *Bulletin of the U. S. Geological and Geographic Survey of the Territories*, 1, 49–51.
- Cope, E.D. (1874b) On some Batrachia and Nematognathi brought from the upper Amazon by Prof. Orton. *Proceedings of the Academy of Natural Sciences, Philadelphia*, 26, 120–137.
- Cope, E.D. (1877) Synopsis of the cold blooded Vertebrata, procured by Prof. James Orton during his exploration of Peru in 1876–77. *Proceedings of the American Philosophical Society*, 17, 33–49.
- Cope, E.D. (1878) Synopsis of the fishes of the Peruvian Amazon, obtained by Professor Orton during his expeditions of 1873 and 1877. *Proceedings of the American Philosophical Society*, 17, 673–701.
- Cope, E.D. (1880) On the zoological position of Texas. *Bulletin of the U. S. National Museum*, 17, 1–51.
- Cope, E.D. (1884a) The fishes of the Batsto River, N. J. *Proceedings of the Academy of Natural Sciences, Philadelphia*, 35, 132–133.
- Cope, E.D. (1884b) The Vertebrata of the Tertiary formations of the West. Book 1. *Report, U. S. Geological and Geographic Survey of the Territories*, 3, 1–1009, 76 pl.
- Cope, E.D. (1891) On Vertebrata from the Tertiary and Cretaceous rocks of the Northwest Territory. I.— The species from the Oligocene of lower Miocene beds of the Cypress Hills. *Contributions to Canadian Palaeontology*, 3, 1–25.
- Cope, E.D. (1894) On the fishes obtained by the Naturalist Expedition in Rio Grande do Sul. *Proceedings of the American Philosophical Society*, 33, 84–108, pls. 4–9.
- Copley, H. (1941) A short account of the freshwater fishes of Kenya. *Journal of the East Africa and Uganda Natural History Society*, 16, 1–24.
- Cornalia, A. (1849) *Vertebratorum Synopsis in Museo Mediolanense Extantium quae per Novum Orbem Cajetanus Osculati Collegit ... (1849)*, 16 p., 1 pl. [Not seen]
- Costa, W.J.E.M. (1987) Feeding habits of a fish community in a tropical coastal stream, rio Mato Grosso, Brazil. *Studies on Neotropical Fauna and Environment*, 22, 145–153.

- Costa, W.J.E.M. (1992) Description de huit nouvelles espèces du genre *Trichomycterus* (Siluriformes: Trichomycteridae), du Brésil oriental. *Revue Française d'Aquariologie et Herpetologie*, 18, for 1991, 101–110.
- Costa, W.J.E.M. (1994) A new genus and species of Sarcoglanidinae (Siluriformes: Trichomycteridae) from the Arauá basin, central Brazil, with notes on subfamilial phylogeny. *Ichthyological Exploration of Freshwaters*, 5, 207–216.
- Costa, W.J.E.M. & Bockmann, F.A. (1993) Un nouveau genre néotropical de la famille des Trichomycteridae (Siluriformes: Loricarioidei). *Revue Française d'Aquariologie et Herpetologie*, 20, 43–46.
- Costa, W.J.E.M. & Bockmann, F.A. (1994a) A new genus and species of Sarcoglanidinae (Siluriformes: Trichomycteridae) from southeastern Brazil, with a re-examination of subfamilial phylogeny. *Journal of Natural History*, 28, 715–730.
- Costa, W.J.E.M. & Bockmann, F.A. (1994b) *Typhlobelus macromycterus*, a new blind glanapterygine fish (Siluriformes: Trichomycteridae) from the Rio Tocantins, Brazil. *Tropical Zoology*, 7, 67–72.
- Costa, W.J.E.M., Lima, S.M.Q. & Bizerril, C.R.S.F. (2004) *Microcambeva ribeirae* sp. n. (Teleostei: Siluriformes: Trichomycteridae): a new sarcoglanidine catfish from the Rio Ribeira do Iguape basin, southeastern Brazil. *Zootaxa*, 563, 1–10.
- Crass, R.S. (1960) Notes on the freshwater fishes of Natal with descriptions of four new species. *Annals of the Natal Museum*, 14, 405–458.
- Cui, G.-H. (1990) Bagridae (Siluriformes). In: Chu, X.-L. & Chen, Y.-R. (Eds.), *The Fishes of Yunnan, China. Part II—Cyprinidae [sic, non-Cyprinidae]*, Science Press, Beijing, pp. 145–162. [In Chinese, with diagnoses of new taxa in English.]
- Curran, D.J. (1989) Phylogenetic relationships among the catfish genera of the family Auchenipteridae (Teleostei: Siluroidea). *Copeia*, 1989, 408–419.
- Cuvier, G. (1816) *Le Règne Animal Distribué d'après son Organisation pour Servir de Base à l'Histoire Naturelle des Animaux et d'Introduction à l'Anatomie Comparée. Tome 2. Les Reptiles, les Poissons, les Mollusques et les Annélides*, Deterville, Paris, xviii + 532 p. [pls. 9–10, in vol. 4]
- Cuvier, G. (1829) *Le Règne Animal Distribué d'après son Organisation pour Servir de Base à l'Histoire Naturelle des Animaux et d'Introduction à l'Anatomie Comparée. Nouvelle Éd., rev. et augm., Tome 2—Les Reptiles et les Poissons*, Deterville, Paris, xviii + 532 p.
- Cuvier, G. (1836) *Le Règne Animal Distribué d'après son Organisation pour Servir de Base à l'Histoire Naturelle des Animaux et d'Introduction à l'Anatomie Comparée, Troisième Édition, Tome Premier*, Louis Hauman et Comp e., Bruxelles, xxii + 626 p., 14 pls.
- Cuvier, G. & Valenciennes, A. (1840a) *Histoire Naturelle des Poissons. Tome Quatorzième. Suite du Livre Seizième. Labroïdes. Livre Dix-septième. Des Malacoptérygiens*, Ch. Pitois & V^e Levrault, Paris; Strasbourg, xxii + 2 + 464 + 4 p., pls. 389–420. [Vol. dated 1839]
- Cuvier, G. & Valenciennes, A. (1840b) *Histoire Naturelle des Poissons. Tome Quinzième. Suite du Livre Dix-septième. Siluroïdes*, Ch. Pitois & V^e Levrault, Paris & Strasbourg, xxxi + 540 p., pls. 421–455.
- Cuvier, G. & Valenciennes, A. (1846) *Histoire Naturelle des Poissons. Tome Dix-huitième. Suite du Livre Dix-huitième. Cyprinoïdes. Livre Dix-neuvième. Des Ésoces ou Lucioïdes*, Ch. Pitois, & V^e Levrault, Paris & Strasbourg, xix + 2 + 505 + 2 p., pls. 520–553.
- Dabry de Thiersant, P. (1872) Nouvelles espèces de poissons de Chine. In: *La pisciculture et la pêche en Chine*. Masson, Paris, pp. 178–192, pls. 36–50.
- Daget, J. (1948) La collection des poissons d'eau douce de l'I.F.A.N. *Catalogues et Documents (Institut Français d'Afrique Noire)*, 3, 1–59.
- Daget, J. (1954) Les poissons du Niger Supérieur. *Memoirs de l'Institut Français d'Afrique Noire*, 36, 1–391.
- Daget, J. (1959) Description de trois poissons nouveaux de la Gambie: *Barbus niokoloensis* n. sp., *Chrysichthys johnelsi* n. sp. et *Amphilophus rheophilus* n. sp. *Bulletin de l'Institut Français d'Afrique Noire (Série A, Sciences naturelles)*, 21, 689–694.
- Daget, J. (1962) Les poissons du Fouta Dialon et de la basse Guinée. *Memoirs de l'Institut Français d'Afrique Noire*, 65, 1–210, pl. 1–13.
- Daget, J. (1963) La Réserve naturelle intégrale du Mont Nimba. XXVII.— Poissons (deuxième note). *Memoirs de l'Institut Français d'Afrique Noire*, 66, 573–600.
- Daget, J. (1965) Espèces nouvelles ou rares de *Synodontis* récoltées en Côte d'Ivoire. (Pisces, Siluriformes). *Bulletin du Muséum National d'Histoire Naturelle (2^e Série)*, 36, for 1964, 472–476.
- Daget, J. (1979) Description de *Platylagnis depierrei* n. gen., n. sp. (Pisces, Bagridae) du Sanaga (Sud Cameroun). *Bulletin du Muséum National d'Histoire Naturelle (2^e Série)*, 36, for 1979, 1–10.

- tin du Muséum National d'Histoire Naturelle (Série 4, section A)*, 1, for 1978, 821–825.
- Daget, J. (1992) Ariidae. In: Lévêque, C., Paugy, D. & Teugels, G.G. (Eds.), *Faune des Poissons d'Eaux Douces et Saumâtres de l'Afrique de l'Ouest*, Vol. 2, Musée Royal de l'Afrique Central, Tervuren, and Editions de l'ORSTOM, Paris, pp. 564–568.
- Daget, J., Gosse, J.-P. & Thys van den Audenaerde, D.F.E. (Eds.) (1986) *Check-list of the Freshwater Fishes of Africa*, Vol. 2, ISBN, Bruxelles; MRAC, Tervuren; ORSTOM, Paris.
- Daget, J. & Lévêque, C. (1981) Description d'un poisson nouveau de Côte d'Ivoire *Synodontis comoensis* n. sp. (Pisces, Mochocidae). *Cybium (3e série)*, 5 (2), 45–52.
- Daget, J. & Planquette, P. (1967) Sur quelques poissons de Côte d'Ivoire avec la description d'une espèce nouvelle, *Clarrias lamottei* n. sp. (Pisces, Siluriformes, Clariidae). *Bulletin du Muséum National d'Histoire Naturelle (2e Série)*, 39, 278–281.
- Daget, J. & Stauch, A. (1963) Poissons de la partie camerounaise du Bassin de la Bénoué. *Memoirs de l'Institut Français d'Afrique Noire*, 68, 85–107.
- Dahl, G. (1942) Three new fishes of the family Loricariidae from the Magdalena system. *Förhandlingar, Kungl. Fysiografiska Sällskapet i Lund*, 11, 80–86.
- Dahl, G. (1955) An ichthyological reconnaissance of the Sinu River. *Revista Linneana*, 1, 11–19.
- Dahl, G. (1960a) New fresh-water fishes from western Colombia. *Caldasia*, 8, 451–484.
- Dahl, G. (1960b) Nematognathous fishes collected during the Macarena Expedition 1959.— Part I. *Novedades Colombianas*, 1, 302–317.
- Dahl, G. (1961) Nematognathous fishes collected during the Macarena Expedition 1959.— Part II: Pimelodidae, Callophysidae. *Novedades Colombianas*, 1, 483–514.
- Dahl, G. & Medem, F. (1964) *Informe Sobre la Fauna Acuática del Río Sinú. I Parte. Los Peces y la Pesca del Río Sinú*, Corporacion Autonoma Regional de los Valles del Magdalena y del Sinu -CVM-. Departamento de Investigaciones Ictiologicas y Faunisticas, 109 p.
- Dahl, G., Medem, F. & Ramos Henao, A. (1964) "Bocachico," Contribución al Estudio de su Biología y de su Ambiente, El Departamento de Pesca de la Corporación Autónoma Regional de los Valles del Magdalena y del Sinú – C.V.M., 144 p.
- Datovo, A. & Landim, M.I. (2005) *Ituglanis macunaima*, a new catfish from rio Araguaia basin, Brazil (Siluriformes, Trichomycteridae). *Neotropical Ichthyology*, 3, 455–464.
- Datta, A.K., Barman, R.P. & Jayaram, K.C. (1987) On a new species of *Kryptopterus* (Pisces: Siluroidea, family: Siluridae) from Namdapha Wildlife Sanctuary, Arunachal Pradesh, India. *Bulletin of the Zoological Survey of India*, 8, 29–31.
- Datta, A.K. & Karmakar, A.K. (1980) *Clarias bastari* sp. nov. (Pisces: Schilbeidae) from Bastar, Madhya Pradesh, India. *Bulletin of the Zoological Survey of India*, 2, 193–196.
- Datta Munshi, J.S. & Srivastava, M.P. (1988) *Natural History of Fishes and Systematics of Freshwater Fishes of India*, Narendra Publ. House, Delhi, xviii + 403 p.
- David, A. (1962) Brief taxonomic account of the gangetic *Pangasius pangasius* (Hamilton) with a description of a new subspecies from the Godavary. *Proceedings of the Indian Academy of Sciences, Section B*, 56, 136–156.
- David, L. (1935) Die Entwicklung der Clariiden und ihre Verbreitung. Eine anatomisch-systematische Untersuchung. *Revue de Zoologie et de Botanique Africaines*, 28, 77–147 + 5 foldout tables.
- David, L. (1936a) *Uegitglanis*, silure aveugle de la Somalie italienne: Chainon entre Bagrides et Clariides. *Revue de Zoologie et de Botanique Africaines*, 28, 416–418.
- David, L. (1936b) Deux espèces de *Synodontis* du Moyen-Congo. *Revue de Zoologie et de Botanique Africaines*, 28, 416–418.
- David, L. & Poll, M. (1937) Contribution à la faune ichthyologique du Congo Belge: Collections du Dr. H. Schouteden (1924–1926) et d'autres récolteurs. *Annales du Musée du Congo Belge, Zoologie (Ser. I)*, 3, 189–294, pl. 12.
- Davis, J.W. (1887) The fossil fishes of the chalk of Mount Lebanon, in Syria. *Transactions of the Royal Dublin Society, series 2*, 3, 457–636, pls. 14–38.
- Day, F. (1865a) *The Fishes of Malabar*; Bernard Quaritch, London, xxxii + 293 p., 20 pls.
- Day, F. (1865b) On the fishes of Cochin, on the Malabar Coast of India. Part II.— *Anacanthini*. *Proceedings of the Zoological Society of London*, 1865, 286–318.
- Day, F. (1867) On some new or imperfectly known fishes of Madras. *Proceedings of the Zoological Society of London*, 1867, 558–565.
- Day, F. (1868) On some new or imperfectly known fishes of India. *Proceedings of the Zoological Society of London*,

- 1868, 149–156.
- Day, F. (1869) On the fishes of Orissa.— Part I. *Proceedings of the Zoological Society of London*, 1869, 296–310.
- Day, F. (1870a) Notes on the genus *Hara*. *Journal of the Asiatic Society of Bengal*, 39, 37–40, pl. 4.
- Day, F. (1870b) Remarks on some of the Fishes in the Calcutta Museum.— Part I. *Proceedings of the Zoological Society of London*, 1869, 511–527.
- Day, F. (1870c) On the freshwater fishes of Burma.— Part I. *Proceedings of the Zoological Society of London*, 1869, 614–623.
- Day, F. (1871) On the fishes of the Andaman Islands. *Proceedings of the Zoological Society of London*, 1870, 677–705.
- Day, F. (1872) On the freshwater siluroids of India and Burmah. *Proceedings of the Zoological Society of London*, 1871, 703–721.
- Day, F. (1873a) *Report on the Fresh Water Fish and Fisheries of India and Burma*, Office of the Superintendent of Government Printing, Calcutta, 118 + cccvii p.
- Day, F. (1873b) On some new fishes of India. *The Journal of the Linnean Society of London. Zoology*, 11, 524–530.
- Day, F. (1873c) On new or imperfectly known fishes of India. *Proceedings of the Zoological Society of London*, 1873, 236–240.
- Day, F. (1873d) Extracts from the late Dr. Buchanan's 'Fishes of Bengal,' with remarks. *Proceedings of the Zoological Society of London*, 1873, 743–748.
- Day, F. (1875–78) *The Fishes of India; being a Natural History of the Fishes Known to Inhabit the Seas and Fresh Waters of India, Burma, and Ceylon*, Bernard Quaritch, London. [Issued in 5 parts, plus supplement (see Day, 1888). Dates of issuance of parts follow Menon & Rama Rao (1974); all catfish names date to 1877.]
- Day, F. (1876) On some of the fishes of the Deccan. *The Journal of the Linnean Society of London. Zoology*, 12, 565–578.
- Day, F. (1877a) Geographical distribution of Indian freshwater fishes. Part II.— the Siluridae. *The Journal of the Linnean Society of London. Zoology*, 13, 338–352.
- Day, F. (1877b) On the fishes of Yarkand. *Proceedings of the Zoological Society of London*, 1876, 781–807.
- Day, F. (1888) *The Fishes of India; being a Natural History of the Fishes Known to Inhabit the Seas and Fresh Waters of India, Burma, and Ceylon, Supplement*, Williams and Norgate, London and Edinburgh, pp. 779–816.
- De Filippi, F. (1853) Nouvelles espèces de poissons. *Revue et Magasin de Zoologie (Ser. 2)*, 5, 164–171.
- De Kay, J.E. (1842) Fishes. In: *Zoology of New-York; or the New-York Fauna; Comprising Detailed Descriptions of All the Animals Hitherto Observed Within the State of New-York, with Brief Notices of Those Occasionally Found Near its Borders, and Accompanied by Appropriate Illustrations*, part 4, W. & A. White, and J. Visscher, Albany, pp. 1–415, pls. 1–79. [In 6 volumes, 1842–1844]
- De Vis, C.W. (1884) Fishes from South Sea islands. *Proceedings of the Linnean Society of New South Wales*, 8, 445–457.
- De Vos, L. (1981) Description of *Eutropius nyongensis* spec. nov. from the Cameroons (Pisces, Schilbeidae). *Revue de Zoologie Africaine*, 95, 968–974.
- De Vos, L. (1983) Note on the synonyms of *Eutropius mandibularis* Günther, 1867, and *Eutropius multitaeniatus* Pellegrin, 1913 (Pisces, Schilbeidae). *Revue de Zoologie Africaine*, 97, 268–287.
- De Vos, L. (1984a) Note on the species of the genus *Eutropius* (Pisces, Schilbeidae) from the Quanza and Bengo rivers (Angola) with description of *Eutropius angolensis* spec. nov. *Cybium (3e série)*, 8 (2), 3–18.
- De Vos, L. (1984b) Preliminary data of a systematic revision of the African species of the family Schilbeidae (Pisces: Siluriformes). *Revue de Zoologie Africaine*, 98, 424–433.
- De Vos, L. (1986) Schilbeidae. In: Daget, J., Gosse, J.-P. & Thys van den Audenaerde, D.F.E. (Eds.), *Check-list of the Freshwater Fishes of Africa*, Vol. 2, ISBN, Bruxelles; MRAC, Tervuren; ORSTOM, Paris, pp. 36–53.
- De Vos, L. (1992) Schilbeidae. In: Lévêque, C., Paugy, D. & Teugels, G.G. (Eds.), *Faune des Poissons d'Eaux Douces et Saumâtres de l'Afrique de l'Ouest*, Vol. 2, Musée Royal de l'Afrique Central, Tervuren, and Editions de l'ORSTOM, Paris, pp. 432–450.
- De Vos, L. (1993) Le genre *Chiloglanis* (Siluriformes, Mochokidae) dans le bassin de la Ruzizi: description de deux nouvelles espèces. *Journal of African Zoology*, 107, 153–168.
- De Vos, L. (1995) A systematic revision of the African Schilbeidae (Teleostei, Siluriformes), with an annotated bibliography. *Musée Royal de l'Afrique Centrale, Annales, Sciences Zoologiques*, 271, 1–450.
- De Vos, L. (2001a) *Synodontis manni* (Teleostei: Mochokidae), a new catfish from the Lower Tana River, Kenya. *Ichthyological Exploration of Freshwaters*, 12, 41–50.
- De Vos, L. (2001b) Rediscovery of the giant catfish *Pardiglanis tarabinii* (Siluriformes: Claroteidae). *Ichthyological Exploration of Freshwaters*, 12, 212–214.

- De Vos, L. & Lévéque, C. (1983) Etude systématique et morphologique du genre *Eutropius* (Pisces, Schilbeidae) en Afrique de l'Ouest. *Revue de Zoologie Africaine*, 97, 469–532.
- De Vos, L. & Skelton, P. (1990) Name changes for two common African catfishes. Rehabilitation of *Schilbe intermedius* Rüppell, 1832 (Siluriformes, Schilbeidae). *Cybium (3e série)*, 14, 323–326.
- De Vos, L. & Thys van den Audenaerde, D.F.E. (1998) Le statut taxonomique de *Synodontis serratus tanganyicae* Borodin, 1936 du lac Tanganyika (Teleostei; Mochokidae): synonymie avec *S. lacustricolus* Poll, 1953. *Journal of African Zoology*, 112, 147–156.
- Dean, B. (1916) *A Bibliography of Fishes; Vol. 1, Publications Grouped Under the Names of Authors: A–K*, American Museum of Natural History, New York.
- Debouche, C., Marquet, J.-P. & Teugels, G. (1979) Détermination de poissons du genre *Clarias* par une méthode généralisable aux poissons non écailleux. *Bulletin de l'Institut fondamental d'Afrique noire. Série A, Sciences naturelles*, 41, 844–862.
- Delsman, N.C. (1941) Résultats scientifiques des croisières du Navire-École Belge "Mercator", vol. III, No. 3. Pisces. *Mémoires du Musée Royal d'Histoire Naturelle de Belgique (Ser. 2)*, 21, 47–82.
- Depéret, C. (1885) Description géologique du bassin tertiaire du Roussillon et description des vertébrés fossiles du terrain pliocène du Roussillon. *Ann. Sci. géol. (Paris)* 16, 1–272, pls. I–V, map. [not seen]
- Deraniyagala, P.E.P. (1952) *A Colored Atlas of Some Vertebrates from Ceylon*, Vol. 1—Fishes, The Ceylon Government Press, Colombo, 149 p., 34 pls.
- Deraniyagala, P.E.P. (1953) A new race of *Wallago attu* from Ceylon. *Spolia Zeylanica*, 27, 45.
- Deraniyagala, P.E.P. (1958) Three new cyprinoids, a new cat fish and variation among some cyprinoids and an anabantoid of Ceylon. *Spolia Zeylanica*, 28, 129–138, pls. 1–2.
- Derijst, E. (1991) Kataloog der ichthyologische publikaties en nieuwe taxa beschreven door: Dr. Max Poll. *Aquarium Wereld*, Special Publication 2, i–vii + 1–87. [Not seen, from Catalog of Fishes]
- Derijst, E. (1996) Note on the type species of the mailed catfish genus *Pareiorhaphis* Miranda-Ribeiro, 1918 (Pisces: Siluriformes: Loricariidae), with the introduction of *Isbrueckerichthys* nom. nov. *Aquarium Wereld*, 49, 62–64.
- Desoutter, M. (1975) Étude de quelques Bagridae (Siluriformes, Pisces) du Cambodge. Description d'une espèce nouvelle: *Mystus aubentoni*. *Bulletin du Muséum National d'Histoire Naturelle, Zoologie (Sér. 3)*, 206, 441–462.
- Desoutter, M. (1977) Révision du genre *Hemipimelodus* Bleeker, 1858 (Tachysuridae, Siluriformes, Pisces). *Cybium (3e série)*, 1, 9–36.
- Devaere, S., Adriaens, D., Teugels, G.G. & Verraes, W. (2005) Morphology and spatial constraints in a dorso-ventrally flattened skull, with a revised species description of *Platyallabes tihoni* (Poll, 1944). *Journal of Natural History*, 39, 1653–1673.
- Devaere, S., Adriaens, D., Teugels, G.G., Verraes, W., De Clerq, N. & Postnov, A. (2005) Holotype skeletal morphology of *Gymnallabes nops* (Roberts & Stewart, 1976), using micro CT-scanning. *Cybium (3e série)*, 29, 281–293.
- Devaere, S., Teugels, G.G., Adriaens, D., Huysentruyt, F. & Verraes, W. (2004) Redescription of *Dolichallabes microphthalmus* (Poll, 1942) (Siluriformes, Clariidae). *Copeia*, 2004, 108–115.
- Devincenzi, G.J. (1925) El primer ensayo sobre Ictiología del Uruguay. La clase "Peces" de la zoología de don Dámaso A. Larrañaga. *Anales del Museo Nacional de Historia Natural de Montevideo (Ser. 2)*, 6, 295–323.
- Devincenzi, G.J. (1933) Peces del Uruguay. Notas complementarias, II. *Anales del Museo Nacional de Historia Natural de Montevideo (Ser. 2)*, 4, 1–11, pl. 1.
- Devincenzi, G.J. (1943) El género *Loricaria* en la cuenca Platense; descripción de una nueva especie, *Loricaria commersonoides* n. sp. *Comunicaciones Zoológicas del Museo de Historia Natural de Montevideo*, 1, 1–6, 1 plate.
- Devincenzi, G.J. & Teague, G.W. (1942) Ictiofauna del Rio Uruguay medio. *Anales del Museo Nacional de Historia Natural de Montevideo (Ser. 2)*, 5, 1–100 + index + i–viii, pls. 1–6.
- Devincenzi, G.J. & Vaz-Ferreira, R. (1939) Nota preliminar sobre un pygidido hematófago del Rio Uruguay. *Archivos de la Sociedad de Biología de Montevideo*, 9, 165–178.
- Di Capriacco, L. (1935) Spedizione Nello Beccari nella Guiana Britannica. *Monitore Zoologico Italiano*, 46, 55–70.
- Di Capriacco, L. (1948) Miscellanea ichthyologica. *Bollettino di Pesca, Piscicoltura e Idrobiologia*, 23 [= N.S., vol. 2], 193–205.
- Ding, R.-H. (2003) [Pareuchiloglanis in western China and neighbouring countries]. *Sichuan Journal of Zoology*, 22, 27–28. [In Chinese.]
- Ding, R.-H. & Fang, S.-G. (1997) [Studies on the DNA fingerprinting in three species of the genus *Pareuchiloglanis* from China, with description of a new species.] *Transactions of the Chinese Ichthyological Society*, 6, 15–21. [In Chinese, with English abstract.]

- Ding, R.-H., Fu, T.-Y. & Ye, M.-R. (1991) [Two new species of the genus *Pareuchiloglanis* from China (Siluriformes: Sisoridae)]. *Acta Zootaxonomica Sinica*, 16, 369–374. [In Chinese, with English abstract.]
- Dinkelmeyer, J. (1995) Zwei neue Arten von Panzerwelsen der Gattung *Corydoras* Lacépède, 1803 aus Brasilien (Pisces, Siluriformes, Callichthyidae). *Aquaristik Aktuell*, 1995, 60–61.
- Dinkelmeyer, J. (1996) *Corydoras seussi* n. sp., ein neuer Panzerwels aus Brasilien (Pisces, Siluriformes, Callichthyidae). *Aquaristik Aktuell*, 1996, 25–26.
- Diogo, R. (2003a) Higher-level phylogeny of Siluriformes — an overview. In: Arratia, G., Kapoor, B.G., Chardon, M. & Diogo, R. (Eds.), *Catfishes*, Science Publishers, Enfield, NH, USA, pp. 353–384.
- Diogo, R. (2003b) Anatomy, phylogeny, and taxonomy of Amphiliidae. In: Arratia, G., Kapoor, B.G., Chardon, M. & Diogo, R. (Eds.), *Catfishes*, Science Publishers, Enfield, NH, USA, pp. 401–438.
- Diogo, R. (2004a) Phylogeny, origin and biogeography of catfishes: support for a Pangean origin of ‘modern teleosts’ and reexamination of some Mesozoic Pangean connections between the Gondwanan and Laurasian supercontinents. *Animal Biology*, 54, 331–351.
- Diogo, R. (2004b) *Morphological Evolution, Aptations, Homoplasies, Constraints, And Evolutionary Trends: Catfishes As A Case Study On General Phylogeny And Macroevolution*, Science Publishers, Inc. Enfield, NH, USA, 487 p.
- Diogo, R. & Chardon, M. (2000) The structures associated with catfish (Teleostei: Siluriformes) mandibular barbels: origin, anatomy, function, taxonomic distribution, nomenclature and synonymy. *Netherlands Journal of Zoology*, 50, 455–478.
- Diogo, R., Chardon, M. & Vandewalle, P. (2001) Osteology and myology of the cephalic region and pectoral girdle of *Bunocephalus knerii*, and a discussion on the phylogenetic relationships of the Aspredinidae (Teleostei: Siluriformes). *Netherlands Journal of Zoology*, 51, 457–481.
- Diogo, R., Chardon, M. & Vandewalle, P. (2002) Osteology and myology of the cephalic region and pectoral girdle of *Glyptothorax fukiensis*, comparison with other sisorids, and discussion on the synapomorphies and phylogenetic relationships of the Sisoridae (Teleostei: Siluriformes). *Belgian Journal of Zoology*, 132, 95–103.
- Diogo, R., Chardon, M. & Vandewalle, P. (2003) Osteology and myology of the cephalic region and pectoral girdle of *Centromochlus heckelii*, comparison with other achenipterids, and comments on the synapomorphies and phylogenetic relationships of the Achenipteridae (Teleostei: Siluriformes). *Animal Biology*, 53, 397–416.
- Diogo, R., Chardon, M. & Vandewalle, P. (2003a) Osteology and myology of the cephalic region and pectoral girdle of *Heteropneustes fossilis* (Teleostei: Siluriformes), with comments on the phylogenetic relationships between *Heteropneustes* and the clariid catfishes. *Animal Biology*, 53, 379–396.
- Diogo, R., Chardon, M. & Vandewalle, P. (2003b) Osteology and myology of the cephalic region and pectoral girdle of *Erethistes pusillus*, comparison with other erethistids, and comments on the synapomorphies and phylogenetic relationships of the Erethistidae (Teleostei: Siluriformes). *Journal of Fish Biology*, 63, 1160–1176.
- Diogo, R., Chardon, M. & Vandewalle, P. (2004) Osteology and myology of the cephalic region and pectoral girdle of *Schilbe mystus*, comparison with other schilbids, and discussion on the monophyly and phylogenetic relationships of the Schilbidae (Teleostei: Siluriformes). *Animal Biology*, 54, 91–110.
- Diogo, R., Oliveira, C. & Chardon, M. (2000) The origin and transformation of catfish palatine-maxillary system: an example of macroevolution. *Netherlands Journal of Zoology*, 50, 373–389.
- Diogo, R., Oliveira, C. & Chardon, M. (2001) On the homologies of the skeletal components of catfish (Teleostei: Siluriformes) suspensorium. *Belgian Journal of Zoology*, 131, 93–109.
- Diogo, R., Vandewalle, P. & Chardon, M. (1999) Morphological description of the cephalic region of *Bagrus docmac*, with a reflection on Bagridae (Teleostei: Siluriformes) autapomorphies. *Netherlands Journal of Zoology*, 49, 207–232.
- Dixon, F. & Egerton, P. de M.G. (1850) Description of fishes (from the Eocene of Bracklesham Bay and Selsey). In: Dixon, F. (Ed.), *The Geology and Fossils of the Tertiary and Cretaceous Formations of Sussex*, Brighton, pp. 195–205, pls. 11–12.
- Dmitrenko, E.M. (1974) [*Arius* of Day— *Arius dayi* sp. n. (Cypriniformes, Ariidae) from the Arabian Sea]. *Vestnik Zoologii*, 1974, 37–41. [In Russian.]
- Do Nascimento, C. & Lundberg, J.G. (2005) *Myoglanis aspredinoides* (Siluriformes: Heptapteridae), a new catfish from the Río Ventuari, Venezuela. *Zootaxa*, 1009, 37–49.
- Do Nascimento, C., Provenzano, F. & Lundberg, J.G. (2004) *Rhamdia guasarensis* (Siluriformes, Heptapteridae) a new species of cave catfish from the Sierra de Perijá, northwestern Venezuela. *Proceedings of the Biological Society of Washington*, 117, 564–574.
- Do Nascimento, C., Villarreal, O. & Provenzano, F. (2001) Descripción de una nueva especie de bagre anoftalmo del

- género *Trichomycterus* (Siluriformes, Trichomycteridae), de una cueva de la Sierra de Perijá, Venezuela. *Boletín de la Sociedad Venezolana de Espeleología*, 35, 20–26.
- Dolgopol de Saez, M. (1945) Noticias sobre peces fósiles Argentinos: Siluroideos terciarios del Chabut. *Notes del Museo de la Plata*, 6, 451–457.
- Douglas, N.H. (1972) *Noturus taylori*, a new species of madtom (Pisces, Ictaluridae) from the Caddo River, southwest Arkansas. *Copeia*, 1972, 785–789.
- Driver, C.S. (1919) On the Luciopimelodinae, a new subfamily of the South American Siluridae. *Proceedings of the American Philosophical Society*, 58, 448–456, pls. 2–3.
- Duméril, A.M.C. (1856) Ichthyologie analytique, ou classification des poissons, suivant la méthode naturelle, à l'aide de tableaux synoptiques. *Mémoires de l'Académie des Sciences de l'Institut impérial de France*, 27, 1–507.
- Duncker, G. (1904) Die Fische der malayischen Halbinsel. *Mitteilungen aus dem Naturhistorischen Museum in Hamburg*, 21, 133–207, pls. 1–2.
- Durand, J. (1940) Notes sur quelques poissons d'espèces nouvelles ou peu connues des eaux douces cambodgiennes. *Station Maritime de Cauda; Institut Océanographique de l'Indochine*, 34, 1–40, table, pls. 1–8.
- Durand, J. (1968) Étude des poissons récoltés dans la grotte de Umayalanta (Bolivie), *Trichomycterus chaberti* sp. n. *Annales de Spéléologie*, 23, 343–353.
- Dutt, S. & Sharma, S.V. (1979) Provisional key to the common freshwater catfishes of central coastal Andhra Pradesh. *Mem. Soc. Zool. Guntur*, 1, 109–119. [Not seen]
- Dutt, S., Sharma, S.V. & Desoutter, M. (1982) On the taxonomic position of *Mystus cavasius* (Hamilton-Buchanan) vis-à-vis *M. nigriceps* (Valenciennes, 1839) and *M. keletius* (Valenciennes, 1839). *Cybium (3e série)*, 6 (4), 27–30.
- Dybowski, B.I. (1872) Zur Kenntniss der Fischfauna des Amurgebietes. *Verhandlungen der Kaiserlich Zoologisch-Botanischen Gesellschaft in Wien*, 22, 209–222.
- Eastman, C.R. (1917) Fossil fishes in the collection of the United States National Museum. *Proceedings of the United States National Museum*, 52, 235–304.
- Eigenmann, C.H. (1890) The evolution of catfishes. *Zoe*, 1, 10–15.
- Eigenmann, C.H. (1905) The mailed catfishes of South America. *Science (n. s.)*, 21, 792–795.
- Eigenmann, C.H. (1907) On a collection of fishes from Buenos Aires. *Proceedings of the Washington Academy of Science*, 8, 449–458, pls. 21–23.
- Eigenmann, C.H. (1909a) Reports on the expedition to British Guiana of the Indiana University and the Carnegie Museum, 1908. Report no. 1.— Some new genera and species of fishes from British Guiana. *Annals of the Carnegie Museum*, 6, 4–54.
- Eigenmann, C.H. (1909b) The fresh-water fishes of Patagonia and an examination of the Archiplata-Archipelagic theory. *Reports of the Princeton University expeditions to Patagonia 1896–1899, Zoology*, vol. 3, pt. 1, Princeton University, pp. 225–374, pls. 30–37.
- Eigenmann, C.H. (1910) Catalogue of the fresh-water fishes of tropical and south temperate America. *Reports of the Princeton University Expeditions to Patagonia 1896–1899, Zoology*, vol. 3, pt. 2, Princeton University pp. 375–511.
- Eigenmann, C.H. (1911) Description of a new species of *Pygidium*. *Annals of the Carnegie Museum*, 7, 214, pl. 32.
- Eigenmann, C.H. (1912a) Some results from an ichthyological reconnaissance of Colombia, South America. Part I. *Indiana University Studies*, 16, [sic. 8], 1–27.
- Eigenmann, C.H. (1912b) The freshwater fishes of British Guiana, including a study of the ecological grouping of species, and the relation of the fauna of the plateau to that of the lowlands. *Memoirs of the Carnegie Museum*, 5, i–xxii + 1–578, pls. 1–103.
- Eigenmann, C.H. (1913) On two new species of fishes collected by Miss Lola Vance in Peru. *Annals of the Carnegie Museum*, 8, 421–422.
- Eigenmann, C.H. (1914a) Some results from studies of South American fishes. IV.— New genera and species of South American fishes. *Indiana University Studies*, 20, 44–48.
- Eigenmann, C.H. (1914b) On new species of fishes from the Rio Meta Basin of eastern Colombia and on albino or blind fishes from near Bogotá. *Indiana University Studies*, 23, 229–230.
- Eigenmann, C.H. (1916) New and rare fishes from South American rivers. *Annals of the Carnegie Museum*, 10, 77–86, pls. 13–16.
- Eigenmann, C.H. (1917a) New and rare species of South American Siluridae in the Carnegie Museum. *Annals of the Carnegie Museum*, 11, 398–404, pls. 39–41.
- Eigenmann, C.H. (1917b) *Pimelodella* and *Typhlobagrus*. *Memoirs of the Carnegie Museum*, 7, 229–258, pls. 29–35.
- Eigenmann, C.H. (1918a) The Pygidiidae, a family of South American catfishes. *Memoirs of the Carnegie Museum*, 7,

- 259–398, pls. 36–56.
- Eigenmann, C.H. (1918b) Eighteen new species of fishes from northwestern South America. *Proceedings of the American Philosophical Society*, 56, for 1917, 673–689.
- Eigenmann, C.H. (1918c) Descriptions of sixteen new species of Pygidiidae. *Proceedings of the American Philosophical Society*, 56, for 1917, 690–703.
- Eigenmann, C.H. (1919a) Peces Colombianos de las cordilleras y de los llanos al oriente de Bogotá. *Boletín de la Sociedad Colombiana de Ciencias Naturales*, 7, 126–136.
- Eigenmann, C.H. (1919b) *Trogloglanis pattersoni* a new blind fish from San Antonio, Texas. *Proceedings of the American Philosophical Society*, 58, 397–400.
- Eigenmann, C.H. (1920a) The fishes of Lake Valencia, Caracas, and of the Rio Tuy at El Concejo, Venezuela. *Indiana University Studies*, 7, 1–13.
- Eigenmann, C.H. (1920b) South America West of the Maracaibo, Orinoco, Amazon, and Titicaca basins, and the horizontal distribution of its fresh-water fishes. *Indiana University Studies*, 7, 1–24.
- Eigenmann, C.H. (1920c) On a new species of *Hatcheria* and a new species of *Pygidium*. *Revista Chilena de Historia Natural, Valparaíso*, 23, 53–54.
- Eigenmann, C.H. (1920d) Limits of the genera *Vandellia* and *Urinophilus*. *Science (n. s.)*, 51 (no. 1322), 441.
- Eigenmann, C.H. (1922a) On a new genus and two new species of Pygidiidae, a family of South American nematognaths. *Bijdragen tot de Dierkunde*, 22, 113–114, pls. 3–4.
- Eigenmann, C.H. (1922b) The fishes of western South America, Part I.—The fresh-water fishes of northwestern South America, including Colombia, Panama, and the Pacific slopes of Ecuador and Peru, together with an appendix upon the fishes of the Rio Meta in Colombia. *Memoirs of the Carnegie Museum*, 9, 1–346, pls. 1–38.
- Eigenmann, C.H. (1925) A review of the Doradidae, a family of South American Nematognathi, or catfishes. *Transactions of the American Philosophical Society (N. S.)*, 22, 280–365, pls. 1–27.
- Eigenmann, C.H. (1927) The fresh-water fishes of Chile. *Memoirs of the National Academy of Sciences*, 22, 1–63, pls. 1–16.
- Eigenmann, C.H. & Allen, W.R. (1942) *Fishes of Western South America. I.—The Intercordilleran and Amazonian Lowlands of Peru. II.—The High Pampas of Peru, Bolivia, and Northern Chile; with a Revision of the Peruvian Gymnotidae, and of the Genus Orestias*, University of Kentucky, xv + 494 p., 22 pl.
- Eigenmann, C.H. & Bean, B.A. (1907) An account of Amazon River fishes collected by J. B. Steere; with a note on *Pimelodus clarias*. *Proceedings of the United States National Museum*, 31, 659–668.
- Eigenmann, C.H. & Eigenmann, R.S. (1888a) American Nematognathi. *American Naturalist*, 22, 647–649.
- Eigenmann, C.H. & Eigenmann, R.S. (1888b) Preliminary notes on South American Nematognathi, I. *Proceedings of the California Academy of Sciences, Series 2*, 1, 119–172.
- Eigenmann, C.H. & Eigenmann, R.S. (1889a) Description of new nematognathoid fishes from Brazil. *West American Scientist*, 6, 8–10.
- Eigenmann, C.H. & Eigenmann, R.S. (1889b) Preliminary notes on South American Nematognathi, II. *Proceedings of the California Academy of Sciences, Series 2*, 2, 28–56.
- Eigenmann, C.H. & Eigenmann, R.S. (1890) A revision of the South American Nematognathi or cat-fishes. *Occasional Papers of the California Academy of Sciences*, 1, 1–508 + errata and map.
- Eigenmann, C.H. & Eigenmann, R.S. (1891) A catalogue of the fresh-water fishes of South America. *Proceedings of the United States National Museum*, 14, 1–81.
- Eigenmann, C.H. & Eigenmann, R.S. (1919) *Steindachneridion*. *Science (n. s.)*, 50, 525–526.
- Eigenmann, C.H. & Fisher, H.G. (1917) On some species of *Rhamdia*, a genus of South American Siluridae, in the Carnegie Museum. *Annals of the Carnegie Museum*, 11, 394–397, pl. 38.
- Eigenmann, C.H., Henn, A.W. & Wilson, C. (1914) New fishes from western Colombia, Ecuador, and Peru. *Indiana University Studies*, 19, 1–15.
- Eigenmann, C.H. & Kennedy, C.H. (1903) On a collection of fishes from Paraguay, with a synopsis of the American genera of cichlids. *Proceedings of the Academy of Natural Sciences, Philadelphia*, 55, 497–537.
- Eigenmann, C.H., McAtee, W.L. & Ward, D.P. (1907) On further collections of fishes from Paraguay. *Annals of the Carnegie Museum*, 4, 110–157, pls. 31–45.
- Eigenmann, C.H. & Norris, A.A. (1900) Sobre alguns peixes de S. Paulo, Brazil. *Revista do Museu Paulista*, 4, 349–362.
- Eigenmann, C.H. & Norris, A.A. (1901) *Bergiaria*. *Comunicaciones del Museo Nacional de Buenos Aires*, 1, 272.
- Eigenmann, C.H. & Vance, L. (1917) Some species of *Farlowella*. *Annals of the Carnegie Museum*, 11, 297–303, pls. 29–31.

- Ellis, M.D. (1913) The plated nematognaths. *Annals of the Carnegie Museum*, 8, 384–413, pls. 25–31.
- Elwin, M.G. (1939) *Corydoras arcuatus* sp. n., an Amazonian catfish. *Annals and Magazine of Natural History (Ser. 11)*, 3, 126–128, pl. 3.
- Emmens, C.W. & Axelrod, H.R. (1968) *Catfish*. TFH Publications, Neptune City, NJ, 96 p.
- Eschmeyer, W.N. (1990) *Catalog of the Genera of Recent Fishes*, California Academy of Sciences, San Francisco, v + 697 p.
- Eschmeyer, W.N. (Ed.) (1998) *Catalog of Fishes*, California Academy of Sciences, San Francisco, 2905 p.
- Eschmeyer, W.N., Ferraris, C.J., Jr., Hoang, M. & Long, D. (1998) Species of fishes. In: Eschmeyer, W.N. (Ed.), *Catalog of Fishes*, California Academy of Sciences, San Francisco, pp. 25–1820.
- Espinosa Pérez, H., Gaspar Dillanes, M.T. & Fuentes Mata, P. (1993) *Listados Faunísticos de México. III.—Los Peces Dulceacuícolas Mexicanos*. Universidad Nacional Autónoma de México, 98 p.
- Etnier, D.A. & Jenkins, R.E. (1980) *Noturus stanauli*, a new madtom catfish (Ictaluridae) from the Clinch and Duck rivers, Tennessee. *Bulletin of the Alabama Museum of Natural History*, 5, 17–22.
- Evermann, B.W. & Goldsborough, E.L. (1902) A report on fishes collected in Mexico and Central America, with notes and descriptions of five new species. *Bulletin of the U. S. Fish Commission*, 21, for 1901, 137–159.
- Evermann, B.W. & Kendall, W.C. (1894) The fishes of Texas and the Rio Grande basin, considered chiefly with reference to their geographic distribution. *Bulletin of the U. S. Fish Commission*, 12, for 1892, 57–126, pls. 10–50.
- Evermann, B.W. & Kendall, W.C. (1898) Descriptions of new or little-known genera and species of fishes from the United States. *Bulletin of the U. S. Fish Commission*, 17, for 1897, 125–133, pls. 6–9.
- Fang, S.-M., Xu, T.-Q. & Cui, G.-H. (1984) [A new species of the catfish genus *Pareuchiloglanis* (Pisces: Sisoridae) from China]. *Acta Zootaxonomica Sinica*, 9, 209–211. [In Chinese, with English abstract.]
- Fernández, L.A. (1999) *Revisión Taxonómica del Género Trichomycterus (Pisces: Trichomycteridae) de la República Argentina*. Unpublished Ph.D. Dissertation, Universidad Tucumán, Argentina.
- Fernández, L.A. (2000) A new species of *Trichomycterus* from northwestern Argentina (Ostariophysi: Trichomycteridae). *Ichthyological Exploration of Freshwaters*, 11, 349–354.
- Fernández, L.A. (2000) Redescription of the teleost *Trichomycterus barbouri* (Eigenmann, 1911), occurrence in Argentina and comparison with related species (Ostariophysi: Siluriformes: Trichomycteridae). *Studies on Neotropical Fauna and Environment*, 35 27–33.
- Fernández, L.A. & Bichuette, M.E. (2002) A new cave dwelling species of *Ituglanis* from the São Domingos karst, central Brazil (Siluriformes: Trichomycteridae). *Ichthyological Exploration of Freshwaters*, 13, 273–278.
- Fernández, L.A. & Pinna, M.C.C., de. (2005) Phreatic catfish of the genus *Silvinichthys* from southern South America (Teleostei, Siluriformes, Trichomycteridae). *Copeia*, 2005, 100–108.
- Fernández, L.A. & Schaefer, S.A. (2003) *Trichomycterus yuska*, a new species from high elevations of Argentina (Siluriformes: Trichomycteridae). *Ichthyological Exploration of Freshwaters*, 14, 353–360.
- Fernández, L.A. & Schaefer, S.A. (2005) New *Trichomycterus* (Siluriformes: Trichomycteridae) from an offshore island of Colombia. *Copeia*, 2005, 68–76.
- Fernández, L.A. & Vari, R.P. (2000) New species of *Trichomycterus* (Teleostei: Siluriformes: Trichomycteridae) lacking a pelvic fin and girdle from the Andes of Argentina. *Copeia*, 2000, 990–996.
- Fernández, L.A. & Vari, R.P. (2002) New species of *Trichomycterus* from the Andes of Argentina with a redescription of *Trichomycterus alterus* (Siluriformes: Trichomycteridae). *Copeia*, 2002, 739–747.
- Fernández, L.A. & Vari, R.P. (2004) New species of *Trichomycterus* from midelevation localities of northwestern Argentina (Siluriformes: Trichomycteridae). *Copeia*, 2004, 876–882.
- Fernández-Yépez, A. (1945) Un nuevo loricarido para Venezuela. Descripción de un ejemplar de Loricariidae, colectado en el Río Encantado, afluente del Río Tuy por medio del Río Grande. *Memoria, Sociedad de Ciencias Naturales La Salle*, 5, 27–34.
- Fernández-Yépez, A. (1950a) Algunos peces del Río Autana. *Novedades Científicas; Contribuciones ocasionales del Museo de Historia Natural La Salle (Serie Zoológica)*, 2, 1–18, pls. 1–3.
- Fernández-Yépez, A. (1950b) Notas sobre la fauna ictiológica de Venezuela. *Memoria, Sociedad de Ciencias Naturales La Salle*, 10, 113–118.
- Fernández-Yépez, A. (1950c) Un nuevo pez de la familia Doradidae. *Memoria, Sociedad de Ciencias Naturales La Salle*, 10, 195–198.
- Fernández-Yépez, A. (1951) *Ginesia cunaguaro*, nuevo pez para la Ciencia colectado en el Río Apure, Venezuela. *Evenias*, [1–4].
- Fernández-Yépez, A. (1953) Algunas notas sobre los peces Asprediformes con descripción de *Ernstichthys anduzei*,

- nuevo e interesante bunocephalido. *Novedades Científicas; Contribuciones ocasionales del Museo de Historia Natural La Salle (Serie Zoológica)*, 11, 1–6, 1 pl.
- Fernández-Yépez, A. (1965) Contribución al conocimiento de los peces de Venezuela. Dieciséis especies nuevas para Venezuela. *Evencias*, 1–12.
- Fernández-Yépez, A. (1967) Resultados zoologicos de la expedicion de la Universidad Central de Venezuela a la region del Auyantepui en la Guayana Venezolana, abril de 1956. 6.— Primera contribucion al conocimiento de los peces, con descripción de dos especies y una subespecie nuevas. *Acta Biologica Venezolica*, 5, 159–177.
- Fernández-Yépez, A. (1968) Contribución al conocimiento de la familia Doradidae en Venezuela. Boletín del Instituto Oceanográfico. *Universidad de Oriente, Cumaná*, 7, 7–72.
- Fernández-Yépez, A. (1970) *Análisis Ictiológico del Complejo Hidrográfico (07) Río Unare*, Dirección de Obras Hidráulicas, Ministerio de Obras Públicas, República de Venezuela, 20 p., 41 pl.
- Fernández-Yépez, A. (1972a) *Análisis Ictiológico del Complejo Hidrográfico (04) Río Yaracuy*, Dirección de Obras Hidráulicas, Ministerio de Obras Públicas, República de Venezuela, 25 pp., 41 pl.
- Fernández-Yépez, A. (1972b) El género *Hemicetopis* Bleeker [sic, Bleeker], 1863 (Cetopsidae) en Venezuela. *Lagena*, 30, 19–21.
- Fernández-Yépez, A. (1973) Contribución al conocimiento de Auchenipteridae. *Evencias*, 29, [1–7].
- Fernández-Yépez, A. & Martín Salazar, F. (1952) Notas sobre la fauna ictiológica de la region Baruta-El Hatillo. *Memoria, Sociedad de Ciencias Naturales La Salle*, 12, 31–45.
- Fernández-Yépez, A. & Martín Salazar, F. (1953) Apuntes sobre la ictiología de Perija. *Memoria, Sociedad de Ciencias Naturales La Salle*, 13, 227–243.
- Fernholm, B. & Wheeler, A. (1983) Linnaean fish specimens in the Swedish Museum of Natural History, Stockholm. *Zoological Journal of the Linnean Society*, 78, 199–286.
- Ferraris, C.J., Jr. (1988) *The Auchenipteridae: Putative Monophyly and Systematics, with a Classification of the Neotropical Doradoid Catfishes (Ostariophysi: Siluriformes)* Doctoral Dissertation, City University of New York, New York.
- Ferraris, C.J., Jr. (1988) Relationships of the Neotropical catfish genus *Nemuroglanis*, with a description of a new species (Osteichthyes: Siluriformes: Pimelodidae). *Proceedings of the Biological Society of Washington*, 101, 509–516.
- Ferraris, C.J., Jr. (1991) On the type species of *Bunocephalus* (Siluriformes: Aspredinidae). *Copeia*, 1991, 224–225.
- Ferraris, C.J., Jr. (1996) *Denticetopsis*, a new genus of South American whale catfish (Siluriformes: Cetopsidae, Cetopsinae), with two new species. *Proceedings of the California Academy of Sciences, Series 4*, 49, 161–170.
- Ferraris, C.J., Jr. (1999a) Plotosidae. In: Carpenter, K.E. & Niem, V.H. (Eds.), *FAO Species Identification Guides for Fishery Purposes: The living Marine Resources of the Western Central Pacific* Vol. 3: Batoid fishes, chimeras and bony fishes, part 1 (Elopidae to Linophrynidae), FAO, Rome, pp. 1880–1883.
- Ferraris, C.J., Jr. (1999b) *Rita sacerdotum*, a valid species of catfish from Myanmar (Pisces, Bagridae). *Bulletin of the Natural History Museum, London, (Zoology)*, 65, 15–21.
- Ferraris, C.J., Jr. (2003a) Diplomystidae. In: Reis, R.E., Kullander, S.O. & Ferraris, C.J., Jr. (Eds.), *Check list of the Freshwater Fishes of South and Central America* Edipuers, Porto Alegre, Brazil, pp. 255–256.
- Ferraris, C.J., Jr. (2003b) Neoplecostominae. In: Reis, R.E., Kullander, S.O. & Ferraris, C.J., Jr. (Eds.), *Check list of the Freshwater Fishes of South and Central America*. Edipuers, Porto Alegre, Brazil, pp. 319–320.
- Ferraris, C.J., Jr. (2003c) Loricariinae. In: Reis, R.E., Kullander, S.O. & Ferraris, C.J., Jr. (Eds.), *Check list of the Freshwater Fishes of South and Central America* Edipuers, Porto Alegre, Brazil, pp. 330–350.
- Ferraris, C.J., Jr. (2003d) Auchenipteridae. In: Reis, R.E., Kullander, S.O. & Ferraris, C.J., Jr. (Eds.), *Check list of the Freshwater Fishes of South and Central America* Edipuers, Porto Alegre, Brazil, pp. 470–482.
- Ferraris, C.J., Jr. (2004) A new species of the Asian schilbid catfish genus *Clarias* from Myanmar, with a redescription of *Clarias prateri* Hora (Osteichthyes: Siluriformes: Schilbidae). *Zootaxa*, 439, 1–10.
- Ferraris, C.J., Jr. & Britz, R. (2005) A diminutive new species of *Glyptothorax* (Siluriformes: Sisoridae) from the upper Irrawaddy River basin, Myanmar, with comments on sisorid and erethistid phylogenetic relationships. *Ichthyological Exploration of Freshwaters*, 16, 375–383.
- Ferraris, C.J., Jr. & Brown, B.A. (1991) A new species of *Pseudocetopsis* from the Río Negro drainage of Venezuela (Siluriformes: Cetopsidae). *Copeia*, 1991, 161–165.
- Ferraris, C.J., Jr. & Fernandez, J. (1987) *Trachelyopterichthys anduzei*, a new species of achenipterid catfish from the upper Río Orinoco of Venezuela with notes on *T. taeniatus* (Kner). *Proceedings of the Biological Society of Washington*, 100, 257–261.
- Ferraris, C.J., Jr., Isbrücker, I.J.H. & Nijssen, H. (1986) *Neblinichthys pilosus*, a new genus and species of mailed catfish

- from the Rio Baria system, southern Venezuela (Pisces, Siluriformes, Loricariidae). *Revue Française d'Aquariologie et Herpetologie*, 13, 69–72.
- Ferraris, C.J., Jr. & Mago-Leccia, F. (1989) A new genus and species of pimelodid catfish from the Río Negro and Río Orinoco drainages of Venezuela (Siluriformes: Pimelodidae). *Copeia*, 1989, 166–171.
- Ferraris, C.J., Jr. & Pinna, M.C.C., de. (1999) Higher level names for catfishes (Ostariophysi: Siluriformes). *Proceedings of the California Academy of Sciences, Series 4*, 51, 1–17.
- Ferraris, C.J., Jr. & Runge, K.E. (1999) Revision of the south Asian bagrid catfish genus *Sperata*, with the description of a new species from Myanmar. *Proceedings of the California Academy of Sciences, Series 4*, 51, 397–424.
- Ferraris, C.J., Jr. & Vari, R.P. (1992) Catalog of type specimens of Recent fishes in the National Museum of Natural History, Smithsonian Institution, 4: Gonorynchiformes, Gymnotiformes, and Siluriformes (Teleostei: Ostariophysi). *Smithsonian Contributions to Zoology*, 535, 1–52.
- Ferraris, C.J., Jr. & Vari, R.P. (1999) The South American catfish genus *Auchenipterus* Valenciennes, 1840 (Ostariophysi: Siluriformes: Auchenipteridae): monophyly and relationships, with a revisionary study. *Zoological Journal of the Linnean Society*, 126, 387–450.
- Ferraris, C.J., Jr. & Vari, R.P. (2000) The deep-water South American catfish genus *Pseudepapterus* (Ostariophysi: Auchenipteridae). *Ichthyological Exploration of Freshwaters*, 11, 97–112.
- Ferraris, C.J., Jr., Vari, R.P. & Raredon, S.J. (2005) Catfishes of the genus *Auchenipterichthys* (Osteichthyes: Siluriformes: Auchenipteridae), a revisionary study. *Neotropical Ichthyology*, 3, 81–98.
- Figueiredo, F.J. & Costa-Carvalho, B.C.M. (1999) *Steindachneridion silvasantosi* sp. nov. (Teleostei, Siluriformes, Pimelodidae) from the Tertiary of Taubaté Basin, São Paulo, Brazil. *Anais da Academia Brasileira de Ciências*, 71, 683–695.
- Fisch-Muller, S. (1999) *Systématique du Genre Ancistrus Kner (Teleostei, Loricariidae): Approches Morphologique et Génétique*, Thèse de Doctorat, Université de Genève.
- Fisch-Muller, S. (2003) Ancistrinae. In: Reis, R.E., Kullander, S.O. & Ferraris, C.J., Jr. (Eds.), *Check list of the Freshwater Fishes of South and Central America*. Edipucrs, Porto Alegre, Brazil, pp. 373–400.
- Fisch-Muller, S., Cardoso, A.R., da Silva, J.F.P. & Bertaco, V.A. (2005a) Two new Amazonian species of armored catfishes (Siluriformes: Loricariidae): *Ancistrus verecundus* and *Ancistrus parecis*. *Neotropical Ichthyology*, 3, 525–532.
- Fisch-Muller, S., Cardoso, A.R., da Silva, J.F.P. & Bertaco, V.A. (2005b) Three new species of *Ancistrus* Kner (Teleostei: Siluriformes: Loricariidae) from the upper Tapajós and Tocantins rivers. *Revue suisse de Zoologie, Annales de la Société zoologique suisse et du Muséum d'Histoire naturelle de Genève*, 112, 559–572.
- Fisch-Muller, S., Mazzoni, R. & Weber, C. (2001) Genetic and morphological evidences for two new sibling species of *Ancistrus* (Siluriformes: Loricariidae) in upper rio Tocantins drainage, Brazil. *Ichthyological Exploration of Freshwaters*, 12, 289–304.
- Fisher, H.G. (1917) A list of the Hypophthalmidae, the Diplomystidae and of some unrecorded species of Siluridae in the collections of the Carnegie Museum. *Annals of the Carnegie Museum*, 11, 405–427, pl. 42.
- Forey, P.L. & Young, S.V.T. (1999) Late Miocene fishes of the Emirate of Abu Dhabi, United Arab Emirates. In: Whybrow, P.J. & Hill, A. (Eds.), *Fossil Vertebrates of Arabia*, Yale University Press, London and New Haven, pp. 120–135.
- Forsskål, P. (1775) *Descriptiones Animalium Avium, Amphibiorum, Piscium, Insectorum, Vermium; quae in Itinere Orientali Observavit... Post Mortem Auctoris Edidit Carsten Niebuhr*, Hauniae, 20 + xxxiv + 164 p., map.
- Fowler, H.W. (1904) A collection of fishes from Sumatra. *Journal of the Academy of Natural Sciences of Philadelphia (Ser. 2)*, 12, 495–560, pls. 7–28.
- Fowler, H.W. (1905) Some fishes from Borneo. *Proceedings of the Academy of Natural Sciences, Philadelphia*, 57, 455–523.
- Fowler, H.W. (1911) Some fishes from Venezuela. *Proceedings of the Academy of Natural Sciences, Philadelphia*, 63, 419–437.
- Fowler, H.W. (1913) Fishes from the Madeira River, Brazil. *Proceedings of the Academy of Natural Sciences, Philadelphia*, 65, 517–579.
- Fowler, H.W. (1914) Fishes from the Rupununi River, British Guiana. *Proceedings of the Academy of Natural Sciences, Philadelphia*, 66, 229–284.
- Fowler, H.W. (1915a) Notes on nematognathous fishes. *Proceedings of the Academy of Natural Sciences, Philadelphia*, 67, 203–243.
- Fowler, H.W. (1915b) Cold-blooded vertebrates from Florida, the West Indies, Costa Rica, and eastern Brazil. *Proceed-*

- ings of the Academy of Natural Sciences, Philadelphia*, 67, 244–269.
- Fowler, H.W. (1919a) A new siluroid fish of the genus *Cyclopium* from Colombia. *Proceedings of the Academy of Natural Sciences, Philadelphia*, 71, 125–127, pl. 8.
- Fowler, H.W. (1919b) The fishes of the United States Eclipse Expedition to West Africa. *Proceedings of the United States National Museum*, 56, 195–292.
- Fowler, H.W. (1930) The freshwater fishes obtained by the Gray African Expedition — 1929. With notes on other species in the Academy collection. *Proceedings of the Academy of Natural Sciences, Philadelphia*, 82, 27–83.
- Fowler, H.W. (1931) Fishes obtained by the Barber Asphalt Company in Trinidad and Venezuela in 1930. *Proceedings of the Academy of Natural Sciences, Philadelphia*, 83, 391–410.
- Fowler, H.W. (1932a) Zoological results of the Matto Grosso Expedition to Brazil in 1931, I.— Fresh water fishes. *Proceedings of the Academy of Natural Sciences, Philadelphia*, 84, 343–377.
- Fowler, H.W. (1932b) Notes on fresh water fishes from Central America. *Proceedings of the Academy of Natural Sciences, Philadelphia*, 84, 379–385.
- Fowler, H.W. (1934a) Zoological results of the third De Schauensee Siamese Expedition, Part I.— Fishes. *Proceedings of the Academy of Natural Sciences, Philadelphia*, 86, 67–163, pl. 12.
- Fowler, H.W. (1934b) Zoological results of the third De Schauensee Siamese Expedition, Part V.— Additional fishes. *Proceedings of the Academy of Natural Sciences, Philadelphia*, 86, 335–352.
- Fowler, H.W. (1935a) Scientific results of the Vernay-Lang Kalahari expedition, March to September, 1930. Fresh-water fishes. *Annals of the Transvaal Museum*, 16, 251–293, pls. 6–9.
- Fowler, H.W. (1935b) Zoological results of the third De Schauensee Siamese Expedition, Part VI.— Fishes obtained in 1934. *Proceedings of the Academy of Natural Sciences, Philadelphia*, 87, 89–163.
- Fowler, H.W. (1936a) Fresh-water fishes obtained in Guatemala by Mr. Rodolphe Meyer de Schauensee in 1935. *Proceedings of the Academy of Natural Sciences, Philadelphia*, 87, for 1935, 515–531.
- Fowler, H.W. (1936b) Zoological results of the George Vanderbilt African Expedition of 1934. Part III.— The fresh water fishes. *Proceedings of the Academy of Natural Sciences, Philadelphia*, 88, 243–335.
- Fowler, H.W. (1937) Zoological results of the third De Schauensee Siamese Expedition. Part VIII.— Fishes obtained in 1936. *Proceedings of the Academy of Natural Sciences, Philadelphia*, 89, 125–264.
- Fowler, H.W. (1939) Zoological results of the third De Schauensee Siamese Expedition. Part IX.— Additional fishes obtained in 1936. *Proceedings of the Academy of Natural Sciences, Philadelphia*, 91, 39–76.
- Fowler, H.W. (1940a) A collection of fishes obtained by Mr. William C. Morrow in the Ucayali River Basin, Peru. *Proceedings of the Academy of Natural Sciences, Philadelphia*, 91, for 1939, 219–289.
- Fowler, H.W. (1940b) Fishes obtained in Chile by Mr. D. S. Bullock. *Proceedings of the Academy of Natural Sciences, Philadelphia*, 92, 171–190.
- Fowler, H.W. (1940c) Zoological results of the second Bolivian expedition for the Academy of Natural Sciences of Philadelphia, 1936–1937. Part I.— The fishes. *Proceedings of the Academy of Natural Sciences, Philadelphia*, 92, 43–103.
- Fowler, H.W. (1941a) A collection of fresh-water fishes obtained in eastern Brazil by Dr. Rodolpho von Ihering. *Proceedings of the Academy of Natural Sciences, Philadelphia*, 93, 123–199.
- Fowler, H.W. (1941b) Los peces del Peru. Catálogo sistemático de los peces que habitan en aguas peruanas (Continuación). *Boletín del Museo de Historia Natural "Javier Prado"*, 5, 466–487.
- Fowler, H.W. (1941c) Notes on Colombian fresh-water fishes with descriptions of four new species. *Notulae Naturae (Philadelphia)*, 73, 1–10.
- Fowler, H.W. (1942) A new cat-fish from Brazil (*Plecostomus gomesi*, new species). *The Fish Culturist*, 21, unpaginated.
- Fowler, H.W. (1943a) A collection of fresh-water fishes from Colombia, obtained chiefly by Brother Nicéforo María. *Proceedings of the Academy of Natural Sciences, Philadelphia*, 95, 223–266.
- Fowler, H.W. (1943b) Notes and descriptions of new or little known fishes from Uruguay. *Proceedings of the Academy of Natural Sciences, Philadelphia*, 95, 311–334.
- Fowler, H.W. (1943c) Zoological results of the second Bolivian expedition for the Academy of Natural Sciences of Philadelphia, 1936–1937. Part II.— Additional new fishes. *Notulae Naturae (Philadelphia)*, 120, 1–7.
- Fowler, H.W. (1944a) A new glass catfish from Borneo. *The Fish Culturist*, 24, 1–2.
- Fowler, H.W. (1944b) Fresh-water fishes from northwestern Colombia. *Proceedings of the Academy of Natural Sciences, Philadelphia*, 96, 227–248.
- Fowler, H.W. (1944c) Results of the fifth George Vanderbilt expedition (1941) (Bahamas, Caribbean Sea, Panama,

- Galápagos Archipelago and Mexican Pacific islands).— The Fishes. *Monograph. Academy of Natural Sciences, Philadelphia*, 6, 57–529, pls. 1–20.
- Fowler, H.W. (1945a) Colombian zoological survey. Pt. I.— The freshwater fishes obtained in 1945. *Proceedings of the Academy of Natural Sciences, Philadelphia*, 97, 93–135.
- Fowler, H.W. (1945b) Descriptions of two new fresh-water fishes from Colombia. *Notulae Naturae (Philadelphia)*, 158, 1–11.
- Fowler, H.W. (1945c) Descriptions of seven new fresh-water fishes from Peru. *Notulae Naturae (Philadelphia)*, 159, 1–11.
- Fowler, H.W. (1945d) A study of the fishes of the southern Piedmont and coastal plain. *Monograph. Academy of Natural Sciences, Philadelphia*, 7, i–vi + 1–408, 313 figs. on 73 unnum. pls.
- Fowler, H.W. (1946) Notes on a collection of fishes from Trinidad. *Notulae Naturae (Philadelphia)*, 165, 1–11.
- Fowler, H.W. (1949) Results of the two Carpenter African expeditions, 1946–1948. Pt. II.— The fishes. *Proceedings of the Academy of Natural Sciences, Philadelphia*, 101, 233–275.
- Fowler, H.W. (1951a) Os peixes de água doce do Brasil, 3.^a entrega. *Arquivos de Zoologia do Estado de São Paulo*, 6, 405–624.
- Fowler, H.W. (1951b) Some new or emended names of fish-like vertebrates. *The Fish Culturist*, 39, 1–4.
- Fowler, H.W. (1954) Os peixes de água doce do Brasil. 4.^a entrega. *Arquivos de Zoologia do Estado de São Paulo*, 9, 1–400.
- Fowler, H.W. (1958) Some new taxonomic names of fishlike vertebrates. *Notulae Naturae (Philadelphia)*, 310, 1–16.
- França, C. (1922) Doutor Alexandre Rodrigues Ferreira. História de uma missão científica ao Brasil no século XVIII. *Boletim da Sociedade Broteriana*, 1, 65–123, figs.
- Franz, D. (2001) Schwielenwelse. Morphologische Untersuchung der Gattung *Hoplosternum* Gill, 1858 und Beschreibung der Gattung *Diasternum* gen. nov. (Actinopterygii: Callichthyidae). *Das Aquarium*, 389, 17–23.
- Fraser-Brunner, A. (1938) Debutantes of the month. *Aquarist and Pondkeeper*, 8, 207–208.
- Fraser-Brunner, A. (1947) New fishes of the genus *Corydoras*. *Proceedings of the Zoological Society of London*, 117, 241–246, pl. 1.
- Freycinet, L. (Ed.) (1824–1825) *Voyage Autour du Monde, Enterpris par Ordre du Roi, sous le Ministère et Conformément aux Instructions de S. Exc. M. le Vicomte du Bouchage, Secrétaire d'État au Département de la Marine, Exécuté sur les Corvettes de S. M. l'Uranie et la Physicienne, Pendant les Années 1817, 1818, 1819 et 1820*, Pillet Ainé, Imprimeur-Libraire, Paris, [p.1–328 in 1824; 329–616 in 1825].
- Friel, J.P. (1994) *A Phylogenetic Study of the Neotropical Banjo Catfishes (Teleostei: Siluriformes: Aspredinidae)*, Unpublished Ph.D dissertation, Duke University, Durham, NC.
- Friel, J.P. (1995) *Acanthobunocephalus nicoi*, a new genus and species of miniature banjo-catfish from the upper Orinoco and Casiquiare rivers, Venezuela (Siluriformes: Aspredinidae). *Ichthyological Exploration of Freshwaters*, 6, 89–95.
- Friel, J.P. (2003) Aspredinidae. In: Reis, R.E., Kullander, S.O. & Ferraris, C.J., Jr. (Eds.), *Check list of the Freshwater Fishes of South and Central America*, Edipuress, Porto Alegre, Brazil, pp. 261–267.
- Friel, J.P. & Lundberg, J.G. (1996) *Micromyzon akamai*, gen. et sp. nov., a small and eyeless banjo catfish (Siluriformes: Aspredinidae) from the river channels of the lower Amazon basin. *Copeia*, 1996, 641–648.
- Frizzell, D.L. (1965) Otoliths of new fish (*Vorhisia vulpes*, n. gen., n. sp. Siluroidei?) from Upper Cretaceous of South Dakota. *Copeia*, 1965, 178–181.
- Frizzell, D.L. & Dante, J.H. (1965) Otoliths of some early Cenozoic fishes of the Gulf Coast. *Journal of Paleontology*, 39, 687–718.
- Frizzell, D.L. & Koenig, J.W. (1973) Upper Cretaceous ostariophysine (*Vorhisia*) redescribed from unique association of utricular and lagenal otoliths (lapillus and asteriscus). *Copeia*, 1973, 692–698.
- Frost, G.A. (1925) Description of fish otoliths from the Tertiary formations of Atcheen, northern Sumatra. *Wetenschappelijke Mededeelingen // Dienst van den Mijnbouw in Nederlandsch-Oost Indië*, 2, 1–28. [Not seen, Journal title not verified]
- Frost, G.A. (1926) Appendix.— Teleost otoliths. Pp. 83–86, pl. 18, figs. 7–12, In: E.I. White, Eocene fishes from Nigeria. *Bulletin, Geological Survey of Nigeria*, 10, 1–86, pls. 1–18.
- Frost, G.A. (1934) Otoliths of fishes from the lower Tertiary formations of southern England. V.— Anacanthini, Heterosomatidae, Ostariophysidae. *Annals and Magazine of Natural History (Ser. 10)*, 14, 500–505, pl. 14.
- Fu, T.-S. (1935) Study of the fishes of Sin-Yang. *Bulletin of the Fan Memorial Institute of Biology, Peiping (Zoological series)*, 6, 178–181.

- Fuhrmann, O. (1906) *Scleropages formosum* und über *Phreatobius cisternarum*. *Verhandlungen der Schweizerischen Naturforschenden Gesellschaft in Luzern*, 88, 50–51.
- Fumihiito, A. (1989) Morphological comparison of the Mekong giant catfish, *Pangasianodon gigas*, with other pangasiid species. *Japanese Journal of Ichthyology*, 36, 113–119.
- Galvis, G., Mojica, J.I. & Camargo, M. (1997) *Peces del Catatumbo*, Asociación Cravo Norte, Ecopetrol, DXY & Shell, Bogotá, 118 p.
- Ganguly, D.N. & Datta, N.C. (1975) A new cat fish of the genus *Mystus* Scopoli (family: Bagridae) from the vicinity of the Hundru Falls, Bihar, India with comment on the genus *Mystus*. In: Tiwari, K.K. & Srivastava, C.B. (Eds.), *Dr. B.S. Chauhan Commemoration Volume*, Zoological Society of India, Orissa, pp. 293–298.
- Ganguly, D.N., Datta, N.C. & Sen, S. (1972) Two new catfishes of the genus *Glyptothorax* Blyth (family: Sisoridae) from Subarnarekha River, Bihar, India. *Copeia*, 1972, 340–344.
- Garavello, J.C. (1976) Systematics and geographical distribution of the genus *Parotocinclus* Eigenmann & Eigenmann, 1889 (Ostariophysi, Loricariidae). *Arquivos de Zoologia (São Paulo)*, 28, 1–37.
- Garavello, J.C. (1988) Three new species of *Parotocinclus* Eigenmann & Eigenmann, 1889 with comments on their geographical distribution (Pisces, Loricariidae). *Naturalia (São Paulo)*, 13, 117–128.
- Garavello, J.C. (2005) Revision of genus *Steindachneridion* Eigenmann & Eigenmann, 1919 (Ostariophysi, Siluriformes, Pimelodidae). *Neotropical Ichthyology*, 3, 607–623.
- Garavello, J.C. & Britski, H.A. (2003) *Parotocinclus planicauda*, a new species of the subfamily Hypoptopomatinae from southeastern Brazil (Ostariophysi: Loricariidae). *Brazilian Journal of Biology*, 63, 253–260.
- Garavello, J.C., Britski, H.A. & Schaefer, S.A. (1998) Systematics of the genus *Otothyris* Myers 1927, with comments on geographic distribution (Siluriformes: Loricariidae: Hypoptopomatinae). *American Museum Novitates*, 3222, 1–19.
- Garman, S. (1890) *Silurus (Parasilurus) aristotelis*. *Bulletin of the Essex Institute*, 22, 56–59.
- Garman, S. (1912) Pisces. In: Some Chinese vertebrates. *Memoirs of the Museum of Comparative Zoology*, 40, 111–123.
- Gauger, M.F.W. & Buckup, P.A. (2005) Two new species of Hypoptopomatinae from the rio Paraíba do Sul basin, with comments on the monophyly of *Parotocinclus* and the Otothyriini (Siluriformes: Loricariidae). *Neotropical Ichthyology*, 3, 509–518.
- Gayet, M. (1988) Le plus ancien crane de siluriforme: *Andinichthys bolivianensis* nov. gen., nov. sp. (Andinichthyidae nov. fam.) du Maastrichtien de Tiupampa (Bolivie). *Comptes Rendus de l'Academie des Sciences, Serie II: Mécanique, Physique-Chimie Sciences de l'Univers Sciences de la Terre*, 307, 833–836.
- Gayet, M. (1990) Nouveaux Siluriformes du Maastrichtien de Tiupampa (Bolivie). *Comptes Rendus de l'Academie des Sciences, Serie II: Mécanique, Physique-Chimie Sciences de l'Univers Sciences de la Terre*, 310, 867–872.
- Gayet, M. (1991) Holostean and teleostean fossils from Bolivia. In: R. Suárez-Sourco (Ed.), *Fósiles y Facies de Bolivia. Revista Tech. YPFB Cochabamba*, 12, 453–494.
- Gayet, M. & Meunier, F.J. (2003) Palaeontology and palaeobiogeography of catfishes. In: Arratia, G., Kapoor, B.G., Chardon, M. & Diogo, R. (Eds.), *Catfishes*, Science Publishers, Inc., Enfield, NH, USA, pp. 491–522.
- Geerinckx, T., Adriaens, D., Teugels, G.G. & Verraes, W. (2003) Taxonomic evaluation and redescription of *Anaspis-doglanis akiri* (Risch, 1987) (Siluriformes: Claroteidae). *Cybium (3e série)*, 27, 17–25.
- Geerinckx, T., Adriaens, D., Teugels, G.G. & Verraes, W. (2004) A systematic revision of the African catfish genus *Parauchenoglanis* Boulenger, 1911 (Siluriformes: Claroteidae). *Journal of Natural History*, 38, 775–803.
- Geisler, R. (1969) *Corydoras baderi*, ein neuer Panzerwels, und sein Lebensraum im Grenzgefiet Brasilien–Surinam (Pisces, Teleostei, Callichthyidae). *Senckenbergiana Biologica*, 50, 353–357.
- Geoffroy Saint-Hilaire, E. (1809) Poissons du Nil, de la mer Rouge et de la Méditerranée. In: Savigny, J. (Ed.), *Description de l'Egypte... . Histoire Naturelle, vol. 1, part 1*. Paris, pp. 1–52, poissons pls. 1–17.
- Geoffroy Saint-Hilaire, I. (1827) Suite de l'histoire des poissons du Nil. In: Savigny, J. (Ed.), *Description de l'Egypte... . Histoire Naturelle, vol. 1, part 1*. Paris, pp. 265–310.
- Ghazzi, M.S. (2005) *Sturisoma kneri*, new species, a name for an old yet poorly-known catfish (Siluriformes: Loricariidae). *Copeia*, 2005, 559–565.
- Gianferrari, L. (1923) *Uegitglanis zammaronoi* un nuovo siluride cieco africano. *Atti della Società Italiana di Scienze Naturali, Milano*, 62, 1–3, pl. 1.
- Gianferrari, L. (1932) Pesci raccolti in Africa dalla spedizione Baragiola-Durini. *Atti della Società Italiana di Scienze Naturali, Milano*, 71, 138–144.
- Giebel, C.G.A. (1871) *Trachypoma marmoratum*, ein neuer Wels aus dem Amazonenstrom. *Zeitschrift für Gesammten Naturwissenschaften, Berlin*, 37 [= n.f., 3], 97.

- Gilbert, C.H. (1884) Notes on the fishes from Kansas. *Bulletin of the Washburn Laboratory of Natural History*, 1, 10–16.
- Gilbert, C.H. (1885) Description of three new fishes from Kansas. *Proceedings of the United States National Museum*, 7, 512–514.
- Gilbert, C.H. (1891) Report of explorations made in Alabama during 1889, with notes on the fishes of the Tennessee, Alabama, and Escambia rivers. *Bulletin of the U. S. Fish Commission*, 9, for 1889, 143–159, pl. 43.
- Gilbert, C.H. (1897) Descriptions of twenty-two new species of fishes collected by the steamer Albatross, of the United States Fish Commission. *Proceedings of the United States National Museum*, 19, 437–457, pls. 49–55.
- Gilbert, C.H. (1904) Notes on fishes from the Pacific coast of North America. *Proceedings of the California Academy of Sciences, Series 3*, 3, 255–271, pls. 25–29.
- Gilbert, C.H. & Starks, E.C. (1904) The fishes of Panama Bay. *Memoirs of the California Academy of Sciences*, 4, 1–304, pls.
- Gilbert, C.R. (1998) Type catalog of Recent and fossil North American Freshwater fishes: families Cyprinidae, Catostomidae, Ictaluridae, Centrarchidae and Elassomatidae. *Florida Museum of Natural History, Special Publication*, 1, ii + 284 p.
- Gilchrist, J.D.F. & Thompson, W.W. (1913) The freshwater fishes of South Africa. *Annals of the South African Museum*, 11, 321–463.
- Gilchrist, J.D.F. & Thompson, W.W. (1916) Description of four new S. African fishes. *Mar. Biol. Rep. So. Afr.*, 1914–1918, 56–61.
- Gilchrist, J.D.F. & Thompson, W.W. (1917) The freshwater fishes of South Africa (Continued). *Annals of the South African Museum*, 11, 465–575, pl. 3.
- Gill, T.N. (1858) Synopsis of the fresh water fishes of the western portion of the island of Trinidad, W. I. *Annals of the Lyceum of Natural History, New York*, 6, 363–430.
- Gill, T.N. (1859a) Description of a new genus of Pimelodinae from Canada. *Annals of the Lyceum of Natural History, New York*, 7, 39–42.
- Gill, T.N. (1859b) Description of new South American type of siluroids, allied to *Callophysus*. *Proceedings of the Academy of Natural Sciences, Philadelphia*, 11, 196–197.
- Gill, T.N. (1861a) Catalogue of the fishes of the eastern coast of North America, from Greenland to Georgia. *Proceedings of the Academy of Natural Sciences, Philadelphia*, 13, Supplement, 1–63.
- Gill, T.N. (1861b) Descriptions of new species of Pimelodinae (abridged from the forthcoming report of Captain J. H. Simpson). *Proceedings of the Boston Society of Natural History*, 8, for 1861–1862, 42–46.
- Gill, T.N. (1861c) Synopsis of the genera of the sub-family of Pimelodinae. *Proceedings of the Boston Society of Natural History*, 8, for 1861–1862, 46–55.
- Gill, T.N. (1862) On the West African genus *Hemicromis* and descriptions of new species in the museums of the Academy and Smithsonian Institution. *Proceedings of the Academy of Natural Sciences, Philadelphia*, 14, 134–139.
- Gill, T.N. (1863) Descriptive enumeration of a collection of fishes from the western coast of Central America, presented to the Smithsonian Institution by Captain John M. Dow. *Proceedings of the Academy of Natural Sciences, Philadelphia*, 15, 162–174.
- Gill, T.N. (1870) On some new species of fishes obtained by Prof. Orton from the Maranon, or Upper Amazon, and Napo Rivers. *Proceedings of the Academy of Natural Sciences, Philadelphia*, 22, 92–96.
- Gill, T.N. (1872) Arrangement of the families of fishes, or classes Pisces, Marsipobranchii, and Leptocardii. *Smithsonian Miscellaneous Collection*, 247, 1–49.
- Gill, T.N. (1877) Notes on fishes from the Isthmus of Panama, collected by Dr. J. F. Bransford, U. S. N. *Proceedings of the Academy of Natural Sciences, Philadelphia*, 28, for 1876, 335–339.
- Gill, T.N. (1888) *Glyptocephalus* not identical with *Bucklandium*. *American Naturalist*, 22, 925–926.
- Gill, T.N. (1891a) Note on the Aspredinidae. *Proceedings of the United States National Museum*, 13, 347–352.
- Gill, T.N. (1891b) Note on the genus *Felichthys* of Swainson. *Proceedings of the United States National Museum*, 13, 353–354.
- Gill, T.N. (1893) Families and subfamilies of fishes. *Memoirs of the National Academy of Sciences, Washington*, 6, 125–138.
- Giltay, L. (1930a) Notes ichthyologiques, IV.— Description d'une espèce nouvelle de Bagridae (*Auchenoglanis Wittei*, nov. sp.). *Revue de Zoologie et de Botanique Africaines*, 19, 91–94.
- Giltay, L. (1930b) Notes ichthyologiques, V.— Description d'une sous-espèce nouvelle de *Ansorgia vitatta* Blgr., provenant du Bas-Uélé. VI.— *Ichthyoborus besse congolensis*, nov. subsp., de Nyonga (Congo Belge). *Revue de Zoologie*

- et de Botanique Africaines*, 19, 393–396.
- Giltay, L. (1935) Notes ichthyologiques, X.— Description d'une espèce nouvelle de Trichomycteridae. *Bulletin du Musée Royal d'Histoire Naturelle de Belgique*, 11, 1–3.
- Giltay, L. (1936) Notes Ichthyologiques, XI.— Revision du genre *Hemipsilichthys* (Loricariidae). *Bulletin du Musée Royal d'Histoire Naturelle de Belgique*, 12, 1–7.
- Girard, C.F. (1855) *Contributions to the Fauna of Chile. Report to Lieut. James M. Gilliss, U. S. N., Upon the Fishes Collected by the U. S. Naval Astronomical Expedition to the Southern Hemisphere During the Years 1849-50-51-52*, Washington.
- Girard, C.F. (1858a) Notes upon various new genera and new species of fishes, in the museum of the Smithsonian Institution, and collected in connection with the United States and Mexican boundary survey: Major William Emory, Commissioner. *Proceedings of the Academy of Natural Sciences, Philadelphia*, 10, 167–171.
- Girard, C.F. (1858b) Fishes. In: *General Report upon Zoology of the Several Pacific Railroad Routes, 1857*, U. S. Senate, Miscellaneous Document, no. 78 (33rd. Congress, 2nd. Session), pp. i–xiv + 1–400, 21 pls.
- Girard, C.F. (1859a) Ichthyology of the boundary. In: *United States and Mexican Boundary Survey, Under the Order of Lieut. Col. W. H. Emory, Major First Cavalry and United States Commissioner. Vol. 2.— Zoology of the Boundary, Department of the Interior*, Washington, pp. 1–85, fishes pls. 1–41.
- Girard, C.F. (1859b) Ichthyological notices. *Proceedings of the Academy of Natural Sciences, Philadelphia*, 11, 157–161.
- Gistel, J. (1848) *Naturgeschichte des Thierreichs, für Höhere Schulen*. Stuttgart, xvi + 216 p., 32 pl.
- Glaw, F. & Vences, M. (1994) *A Field Guide to the Amphibians and Reptiles of Madagascar. Second Edition, Including Mammals and Freshwater Fish*, Privately published, Köln, Germany, 480 pp.
- Glodek, G.S. (1976) *Rhynchodoras woodsi*, a new catfish from eastern Ecuador (Siluriformes: Doradidae) with a redefinition of *Rhynchodoras*. *Copeia*, 1976, 43–46.
- Glodek, G.S. & Carter, H.J. (1978) A new helogeneid catfish from eastern Ecuador (Pisces, Siluriformes, Helogeneidae). *Fieldiana, Zoology*, 72, 75–82.
- Glodek, G.S., Whitmire, G.L. & Orcés, G. (1976) *Rhinodoras boehlkei*, a new catfish from eastern Ecuador (Osteichthyes, Siluroidei, Doradidae). *Fieldiana, Zoology*, 70, 1–11.
- Gmelin, J.F. (1788–1793) *Caroli a Linné... Systema Naturae per Regna Tria Naturae, Secundum Classes, Ordines, Genera, Species; cum Characteribus, Differentiis, Synonymis, Locis. Editio Decimo Tertia, Aucta, Reformata*, Lipsiae [Fishes in vol. 1, pt 3 (dated [1789]), pp 1033–1516.]
- Godoy, M.P. (1969) Nova espécie de "Plecostomus" Gronovius, 1763 (Pisces, Loricariidae, Nematognathi). *Revista Brasileira de Biologia*, 29, 175–180.
- Goeldi, E.A. (1898) Primeira contribuição para o conhecimento dos peixes do valle do Amazonas e das Guyanas. Estudos ichthyologicos dos annos 1894–1898. *Boletim do Museu Paraense de Historia Natural e Ethnographia, Pará*, 2, 443–488, 1 pl.
- Goeldi, E.A. (1905) Nova zoologica aus der Amazonas-region. Neue Wirbeltiere. In: Bedot, M. (Ed.), *Compte Rendu des Séances du Sixième Congrès International de Zoologie*. Imprimerie W. Kündig & Fils, Genève, pp. 542–549.
- Golani, D. (2002) The Indo-Pacific striped eel catfish, *Plotosus lineatus* (Thunberg, 1787), (Osteichthyes: Siluriformes) a new record from the Mediterranean. *Scientia Marina*, 66, 321–323.
- Gomes, A.L. (1946) A review of *Microglanis*, a genus of South American catfishes, with notes on related genera. *Occasional Papers of the Museum of Zoology, University of Michigan*, 494, 1–19, pl. 1.
- Gomes, A.L. (1955) Descrição de *Pseudotocinclus ribeiroi*, nova espécie de cascudinho (Loricariidae, Hypoptopomatinae) da bacia do rio Itanhaém, São Paulo. *Arquivos do Museu Nacional do Rio de Janeiro*, 42, 221–227, pls. 1–2.
- Gomes, A.L. (1956) Descrição de uma nova espécie de "Luciopimelodinae" do Rio Mogi Guaçu, Estado de São Paulo, (Pisces, Nematognathi, Pimelodidae). *Revista Brasileira de Biologia*, 16, 403–413.
- Gomes, A.L. & Schubart, O. (1958) Descrição de "*Chasmocranus brachynema*" sp. n., novo "Luciopimelodinae" da Bacia do Rio Mogi Guaçu, estado de São Paulo (Pisces, Nematognathi, Pimelodidae). *Revista Brasileira de Biologia*, 18, 413–416.
- Gomez, S.E., Lopez, H.L. & Toresani, N.I. (1990) *Hypostomus derbyi* (Haseman) e *Hypostomus myersi* (Gosline), descripción complementaria y primeros registros para Argentina (Pisces, Loricariidae). *Studies on Neotropical Fauna and Environment*, 25, 139–152.
- Gomon, J.R. & Taylor, W.R. (1982) *Plotosus nkunga*, a new species of catfish from South Africa, with a redescription of *Plotosus limbatus* Valenciennes and key to the species of *Plotosus* (Siluriformes: Plotosidae). *J. L. B. Smith Institute of Ichthyology, Special Publication*, 22, 1–16.

- Gosline, W.A. (1940a) Rediscovery and redescription of *Pariolius armillatus*, a genus and species of pimelodid catfishes described by E. D. Cope from the Peruvian Amazon in 1872. *Copeia*, 1940, 78–80.
- Gosline, W.A. (1940b) A revision of the neotropical catfishes of the family Callichthyidae. *Stanford Ichthyological Bulletin*, 2, 1–29.
- Gosline, W.A. (1941) Synopsis of the genera of pimelodid catfishes without a free orbital rim. *Stanford Ichthyological Bulletin*, 2, 83–88.
- Gosline, W.A. (1942) Notes on South American catfishes (Nematognathi). *Copeia*, 1942, 39–41.
- Gosline, W.A. (1945) Catálogo dos nematognatos de água-doce da América do Sul e Central. *Boletim do Museu Nacional, Nova série, Zoologia*, 33, 1–138.
- Gosline, W.A. (1947) Contributions to the classification of the loricariid catfishes. *Arquivos do Museu Nacional do Rio de Janeiro*, 41, 79–134, pls. 1–9.
- Gosse, J.-P. (1982) Mutanda ichthyologica: *Synodontis polli* nom. nov. et *Synodontis ornatissimus* nom. nov. *Cybium (3e série)*, 6, 48.
- Gosse, J.-P. (1986a) Malapteruridae. In: Daget, J., Gosse, J.-P. & Thys van den Audenaerde, D.F.E. (Eds.), *Check-list of the Freshwater Fishes of Africa*, Vol. 2, ISBN Bruxelles, MRAC Tervuren, ORSTOM, Paris, pp. 102–104.
- Gosse, J.-P. (1986b) Mochokidae. In: Daget, J., Gosse, J.-P. & Thys van den Audenaerde, D.F.E. (Eds.), *Check-list of the Freshwater Fishes of Africa*, Vol. 2, ISBN Bruxelles, MRAC Tervuren, ORSTOM, Paris, pp. 105–152.
- Grady, J.M. (1987) *Biochemical Systematics and Evolution of the Ictalurid Catfish genus Noturus (Pisces, Siluriformes)*, Unpublished Ph.D. Dissertation, Southern Illinois University at Carbondale, Carbondale, IL.
- Grady, J.M. & LeGrande, W.H. (1992) Phylogenetic relationships, modes of speciation, and historical biogeography of the madtom catfishes, genus *Noturus* Rafinesque (Siluriformes: Ictaluridae). In: Mayden, R.L. (Ed.), *Systematics, Historical Ecology, & North American Freshwater Fishes*, Stanford University Press, Stanford, California, pp. 747–777.
- Grande, L. (1987) Redescription of †*Hypsidoris farsonensis* (Teleostei: Siluriformes), with a reassessment of its phylogenetic relationships. *Journal of Vertebrate Paleontology*, 7, 24–54.
- Grande, L. & Lundberg, J.G. (1988) Revision and redescription of the genus †*Astephus* (Siluriformes: Ictaluridae) with a discussion of its phylogenetic relationships. *Journal of Vertebrate Paleontology*, 8, 139–171.
- Grande, L. & Pinna, M., de. (1998) Description of a second species of the catfish †*Hypsidoris* and a reevaluation of the genus and the family †Hypsidoridae. *Journal of Vertebrate Paleontology*, 18, 451–474.
- Grant, S. (1997) Descriptions of two new species of *Corydoras*, Lacepede, 1803 (Pisces, Siluriformes, Callichthyidae). *Aquarist and Pondkeeper*, 62, 41–45.
- Grant, S. (1999) A replacement name (nomen novum) and neotype designation for *Hara malabarica* Day, 1865, with notes on related species (Siluriformes). *Aqua, Journal of Ichthyology and Aquatic Biology*, 3, 169–174.
- Gras, R. (1961) Contribution à l'étude des poissons du Bas-Dahomey. Description de quatre espèces nouvelles. *Bulletin du Muséum National d'Histoire Naturelle (2e Série)*, 32, for 1960, 401–410.
- Gratzianov, V.J. (1907) [A synoptic essay of the fishes of the Russian Empire]. *Trudy Otdela Ichtiol. Russ. Obshch. Akklimat. Zhiv. Moskva*, 4, i–xxx + 1–567. [In Russian.]
- Gray, J.E. (1830–1835) *Illustrations of Indian Zoology; Chiefly Selected from the Collection of Major-General Hardwicke*, Treuttel, Jun. and Richter, Paris; Grande Rue, Strasburg; Parbury Allen and Co., London. 20 parts in 2 vols., 202 pls. [Catfish illustrated in Vol. 1, plates 84 and 85, and dated 1830, following Sawyer (1953). See Eschmeyer (1998: 2627) for more information.]
- Gray, J.E. (1831) Description of twelve new genera of fish, discovered by Gen. Hardwicke, in India, the greater part in the British Museum. *Zoological Miscellany*, 1831, 7–9.
- Gray, J.E. (1854) *Catalogue of Fish Collected and Described by Laurence Theodore Gronow, now in the British Museum*, British Museum, London, vii + 196 p.
- Greenfield, D.W. & Glodek, G.S. (1977) *Trachelyichthys exilis*, a new species of catfish (Pisces: Auchenipteridae) from Peru. *Fieldiana, Zoology*, 72, 47–58.
- Greenfield, D.W., Greenfield, T.A. & Woods, R.L. (1982) A new subspecies of cave-dwelling pimelodid catfish, *Rhamdia laticauda typhla* from Belize, Central America. *Brenesia*, 19/20, 563–576.
- Greenfield, D.W. & Thomerson, J.E. (1997) *Fishes of the Continental Waters of Belize*, University of Florida Press, Gainesville, xxii + 311 p.
- Greenwood, P.H. (1956) A new species of *Clariallabes* (Pisces, Clariidae), from the Nile. *Proceedings of the Zoological Society of London*, 127, 555–564.
- Greenwood, P.H. (1958) A new genus and species of cat-fish (Pisces, Clariidae) from the deeper waters of Lake Victoria.

- Annals and Magazine of Natural History* (Ser. 13), 1, 321–325.
- Greenwood, P.H. (1961) A revision of the genus *Dinotopterus* Blgr. (Pisces, Clariidae) with notes on the comparative anatomy of the suprabranchial organs in the Clariidae. *Bulletin of the British Museum (Natural History)*, Zoology, 7, 215–241.
- Greenwood, P.H. (1963) A collection of fishes from the Aswa river drainage system, Uganda. *Proceedings of the Zoological Society of London*, 140, 61–74.
- Gromov, I.A. (1970) [A new species of bagrid catfish (*Mystus mica* Gromov, sp. n. (Pisces, Bagridae)) in the Amur Basin]. *Voprosy Ikhtiologii*, 10, 400–405. [In Russian; translated into English in *Journal of Ichthyology*, 10 (3), 277–281.]
- Gronovius, L.T. (1754) *Museum Ichthyologicum, Sistens Piscium Indigenorum & Nonnullorum Exoticorum, quorum Maximum pars in Museo Laurenti Theodori Gronovii, J. U. D. Adservantar, Descriptiones, Ordine Systematico; Accedunt Nonnullorum Exoticorum Piscium Icones, Aeri Incisae*, Theodorum Haak, Lugduni-Batavorum, 70 p. 4 pls.
- Gronovius, L.T. (1756) *Museum Ichthyologicum, Tomus Secundus, Sistens Piscium Indigenorum & Nonnullorum Exoticorum, quorum Maximum pars in Museo Laurenti Theodori Gronovii, J. U. D. Adservantar, Descriptiones, Ordine Systematico; Accedunt Nonnullorum Exoticorum Piscium Icones, Aeri Incisae*, Theodorum Haak, Lugduni-Batavorum, 88 p., pls. 5–7.
- Gronovius, L.T. (1763) *Zoophylacii Gronoviani, Fasciculus Primus, Exhibens Animalia Quadrupeda, Amphibia atque Pisces, quae in Museo suo Adservat, Rite Examinavit, Systematice Disposuit, Descripsit atque Iconibus Illustravit Laur. Theod. Gronovius, J.U.D. Theodorum Haak et Socium, et Samuelem et Johannem Luchtmans, Lugduni Batavorum*, 136 p., 14 pls.
- Guazzelli, G.M. (1997) *Revisão das Espécies de Pimelodella Eigenmann & Eigenmann, 1888 (Teleostei: Siluriformes: Pimelodidae) dos Sistemas Costeiros do Sul e Sudeste Brasileiro*, Unpublished Master Dissertation, Pontifícia Universidade Católica do Rio Grande do Sul, Porto Alegre, 150 p.
- Guichenot, A. (1848) Fauna Chilena. Peces. In: Gay, C. (Ed.), *Historia Física y Política de Chile, Zoología*. Vol. 2, En casa del Autor, Paris; Museo de Historia Natural de Santiago, Chile, pp. 137–370.
- Guichenot, A. (1860) Notice sur un nouveau poisson du genre des Trichomyctères. *Revue et Magasin de Zoologie* (Ser. 2), 12, 525–527.
- Guimarães, A.R.P. (1884a) Description d'un nouveau poisson de l'intérieur d'Angola. *Jornal do Sciências Mathemáticas, Physicas e Naturaes, Lisboa*, 9, for 1882, 85–87, 1 pl. [apparently not published until 1884.]
- Guimarães, A.R.P. (1884b) Diagnoses de trois nouveaux poissons d'Angola. *Jornal do Sciências Mathemáticas, Physicas e Naturaes, Lisboa*, 10, 1–10, pls. 1–2.
- Guimarães, J.R.A. (1935) Contribuição para o conhecimento de uma nova espécie de peixe hematófago, ectoparasita de "Characidae," encontrado em São Paulo (Rio Tietê). *Vandellia hematophaga* sp. n. *Revista de Industria Animal*, 2, 300–304.
- Güntert, H. (1938) Beschreibung einiger unbestimmter Clariiden mit drei neuen Arten aus dem Naturhistorischen Museum in Wien. *Zoologischer Anzeiger*, 124, 289–303.
- Güntert, H. (1939) Ein neuer Clariide aus Kamerun. *Anzeiger der Mathematisch- Naturwissenschaftlichen Classe der Kaiserlichen Akademie der Wissenschaften in Wien*, 75, 25–26.
- Güntert, H. (1942) Beschreibung einiger zum Teil noch unbekannter südamerikanischer Siluriden aus dem Naturhistorischen Museum in Basel. *Zoologischer Anzeiger*, 138, 27–40.
- Günther, A. (1859) List of the cold-blooded vertebrates collected by Mr. Fraser in the Andes of western Ecuador. *Proceedings of the Zoological Society of London*, 27, 89–93.
- Günther, A. (1860) Third list of cold-blooded Vertebrates collected by Mr. Fraser in Ecuador. *Proceedings of the Zoological Society of London*, 28, 233–240, pl. 10.
- Günther, A. (1863) On new species of fishes from the Essequibo. *Annals and Magazine of Natural History* (Ser. 3), 12, 441–443.
- Günther, A. (1864) *Catalogue of the Fishes in the British Museum, vol. 5.—Catalogue of the Physostomi, Containing the Families Siluridae, Characinidae, Haplochitonidae, Sternopychidae, Scopelidae, Stomiataidae in the Collection of the British Museum*, Trustees, London, xxii + 455 p.
- Günther, A. (1865a) A contribution to the ichthyology of West Africa. *Annals and Magazine of Natural History* (Ser. 3), 15, 452–453.
- Günther, A. (1865b) Pisces. *Zoological Record*, for 1864, 133–188.
- Günther, A. (1866) Description of a new siluroid fish from Ceylon. *Annals and Magazine of Natural History* (Ser. 3), 18,

- 473–474, pl. 15.
- Günther, A. (1867a) Additions to the knowledge of Australian reptiles and fishes. *Annals and Magazine of Natural History* (Ser. 3), 20, 45–68.
- Günther, A. (1867b) New fishes from the Gaboon and Gold Coast. *Annals and Magazine of Natural History* (Ser. 3), 20, 110–117, pls. 2–3.
- Günther, A. (1867c) On the fishes of the states of Central America, founded upon specimens collected in fresh and marine waters of various parts of that country by Messrs. Salvin and Godman and Capt. J. M. Dow. *Proceedings of the Zoological Society of London*, 1866, 600–604.
- Günther, A. (1867d) Description of some new or little-known species of fishes in the collection of the British Museum. *Proceedings of the Zoological Society of London*, 1867, 99–104, pl. 10.
- Günther, A. (1868a) Diagnoses of some new freshwater fishes from Surinam and Brazil, in the collection of the British Museum. *Annals and Magazine of Natural History* (Ser. 4), 1, 475–481.
- Günther, A. (1868b) Descriptions of freshwater fishes from Surinam and Brazil. *Proceedings of the Zoological Society of London*, 1868, 229–247, pls. 20–22.
- Günther, A. (1868c) An account of the fishes of the states of Central America, based on collections made by Capt. J. M. Dow, F. Godman, Esq. and O. Salvin, Esq. *Transactions of the Zoological Society of London*, 6, 377–494, pls. 63–87.
- Günther, A. (1869) Descriptions of some species of fishes from the Peruvian Amazons. *Proceedings of the Zoological Society of London*, 1869, 423–429.
- Günther, A. (1872) Notice of a large siluroid from the upper Amazons. *Annals and Magazine of Natural History* (Ser. 4), 10, 449–450.
- Günther, A. (1873a) New fishes from Angola. *Annals and Magazine of Natural History* (Ser. 4), 12, 142–144.
- Günther, A. (1873b) Report on a collection of fishes from China. *Annals and Magazine of Natural History* (Ser. 4), 12, 239–250.
- Günther, A. (1874) A contribution to the fauna of the river Tigris. *Annals and Magazine of Natural History* (Ser. 4), 14, 36–38, pls. 8–9.
- Günther, A. (1876) Contributions to our knowledge of the fish-fauna of the Tertiary deposits of the highlands of Padang, Sumatra. *The Geological Magazine, Series 2*, 3, 443–440. [Also in 1878: Jaarb. Mijnw. Ned. Oost-Indië, 7: 171–184.]
- Günther, A. (1880a) A contribution to the knowledge of the fish fauna of the Rio de la Plata. *Annals and Magazine of Natural History* (Ser. 5), 6, 7–13, pl. 2.
- Günther, A. (1880b) Report on the shore fishes procured during the voyage of H. M. S. Challenger in the years 1873–1876. *Report on the Scientific Results of the Voyage of H. M. S. Challenger During the Years 1873–76. Zoology* Vol. 1, pt. 6, pp. 1–82, pls. 1–32.
- Günther, A. (1883) Notes on some Indian fishes in the collection of the British Museum. *Annals and Magazine of Natural History* (Ser. 5), 11, 137–140.
- Günther, A. (1889) On some fishes from Kilima-Njaro District. *Proceedings of the Zoological Society of London*, 1889, 70–72, pl. 8.
- Günther, A. (1892) List of the species of reptiles and fishes collected by Mr. A. E. Pratt on the upper Yang-tze-kiang and in the province Sze-Chuen, with description of the new species. Appendix II. In: Pratt, A.E. (Ed.), *To the Snows of Tibet Through China*, Longmans, Green and Co., London and New York, pp. 238–250, pls. 1–4.
- Günther, A. (1894) Second report on the reptiles, batrachians, and fishes transmitted by Mr. H. H. Johnston, C. B., from British Central Africa. *Proceedings of the Zoological Society of London*, 1893, 616–628, pls. 53–57.
- Günther, A. (1896a) Report on a collection of reptiles and fishes made by Miss. M. H. Kingsley during her travels on the Ogowe River and in Old Calabar. *Annals and Magazine of Natural History* (Ser. 6), 17, 261–285, pls. 13–15.
- Günther, A. (1896b) Report on a collection of fishes made by Dr. A. Donaldson Smith during his expedition to Lake Rudolf. *Proceedings of the Zoological Society of London*, 1896, 217–224, pl. 9.
- Günther, A. (1899) An account of a collection of fishes made by Mr. R. B. N. Walker, C. M. Z. S., on the Gold Coast. *Proceedings of the Zoological Society of London*, 1899, 716–729, pls. 41–45.
- Günther, A. (1903) Last account of fishes collected by Mr. R. B. N. Walker, C. M. Z. S., on the Gold Coast. *Proceedings of the Zoological Society of London*, 1902, pt 2, 330–339, pls. 30–33.
- Guo, X.-G., He, S.-P. & Zhang, Y.-G. (2005) Phylogeny and biogeography of Chinese sisorid catfishes re-examined using mitochondrial cytochrome b and 16S rRNA gene sequences. *Molecular Phylogenetics and Evolution*, 35, 344–362.
- Gupta, S.K. (1982) Redescription and distribution of a sisorid catfish *Laguvia ribeiroi kapuri* Tilak and Hussain, from

- Banda district (UP). *Indian Journal of Physical and Natural Sciences, Section A*, 2, 36–39.
- Gupta, S.K., Jayaram, K.C. & Hajela, K.P. (1981) On a new silurid cat-fish from Uttar Pradesh, India. *Journal of the Bombay Natural History Society*, 77, 290–291.
- Gustiano, R., Teugels, G.G. & Pouyaud, L. (2003) Revision of the *Pangasius kunyit* catfish complex, with description of two new species from South-east Asia (Siluriformes: Pangasiidae). *Journal of Natural History*, 37, 357–376.
- Gustiano, R., Teugels, G.G. & Pouyaud, L. (2004) *Pangasius bedado* Roberts, 1999: a junior synonym of *Pangasius djambal* Bleeker, 1846 (Siluriformes, Pangasiidae). *Cybium (3e série)*, 28, 13–18.
- Haig, J. (1952) Studies on the classification of the catfishes of the Oriental and Palaearctic family Siluridae. *Records of the Indian Museum*, 48, for 1950, 59–116.
- Hamilton, F. (1822) *An Account of the Fishes Found in the River Ganges and its Branches*, Edinburgh & London, vii + 405 p., 39 pl.
- Hancock, J. (1828) Notes on some species of fishes and reptiles, from Demerara, presented to the Zoological Society by John Hancock, Esq., corr. memb. Zool. Soc. In a letter addressed to the secretary of the Society. *Zoological Journal, London*, 4, 240–247.
- Hardenberg, J.D.F. (1936a) On a collection of fishes from the estuary and the lower and middle course of the River Kapuas (W. Borneo). *Treubia*, 15, 225–254.
- Hardenberg, J.D.F. (1936b) Some new or rare fishes of the Indo-Australian Archipelago. — V. *Treubia*, 15, 367–378.
- Hardenberg, J.D.F. (1938) Some new or rare fishes of the Indo-Australian Archipelago. — VI. *Treubia*, 16, 311–320.
- Hardenberg, J.D.F. (1941) Fishes of New Guinea. *Treubia*, 18, 217–231.
- Hardenberg, J.D.F. (1948) Some new or rare fishes of the Indo-Australian Archipelago. — 8. *Treubia*, 19, 407–415.
- Hardman, M. (2002) *Phylogenetic Relationships Among Species of Ictaluridae (Otophysi: Siluriformes) and of the Family to Other Catfishes*, Unpublished Ph.D. dissertation, University of Illinois, Urbana-Champaign.
- Hardman, M. (2004) The phylogenetic relationships among *Noturus* catfishes (Siluriformes: Ictaluridae) as inferred from mitochondrial gene cytochrome *b* and nuclear recombination activating gene 2. *Molecular Phylogenetics and Evolution*, 30, 395–408.
- Hardman, M. (2005) The phylogenetic relationships among non-diplomystid catfishes as inferred from mitochondrial cytochrome *b* sequences; the search for the ictalurid sister taxon (Otophysi: Siluriformes). *Molecular Phylogenetics and Evolution*, 37, 700–720.
- Hardman, M. & Page, L.M. (2003) Phylogenetic relationships among bullhead catfishes of the genus *Ameiurus* (Siluriformes: Ictaluridae). *Copeia*, 2003, 20–33.
- Hardy, M.E., Grady, J.M. & Routman, E.J. (2002) Intraspecific phylogeography of the slender madtom: The complex evolutionary history of the Central Highlands of the United States. *Molecular Ecology*, 11, 2393–2403.
- Harka, A. & Pinter, K. (1990) Systematic status of Hungarian bullhead pout: *Ictalurus nebulosus pannonicus* ssp. n. *Tiszaia (Szeged)*, 25, 65–73.
- Harry, R.R. (1947) *Platysomatos*, a neglected name for a genus of aspredinid catfishes. *Copeia*, 1947, 194.
- Harry, R.R. (1953) A contribution to the classification of the African catfishes of the family Amphiliidae, with description of collections from Cameroon. *Revue de Zoologie et de Botanique Africaines*, 47, 177–200; 201–232.
- Hase, A. (1914) Die Fische der Deutschen Grenzexpedition 1910 in das Kaiser-Wilhelms-Land, Neu Guinea. *Jenaische Zeitschrift für Naturwissenschaft*, 51, 525–548.
- Haseman, J.D. (1911a) Descriptions of some new species of fishes and miscellaneous notes on others obtained during the expedition of the Carnegie Museum to central South America. *Annals of the Carnegie Museum*, 7, 315–328, pls. 46–52.
- Haseman, J.D. (1911b) Some new species of fishes from the Rio Iguassú. *Annals of the Carnegie Museum*, 7, 374–387, pls. 50, 58, 73–83.
- Hay, O.P. (1924) Description of some fossil vertebrates from the Upper Miocene of Texas. *Proceedings of the Biological Society of Washington*, 37, 1–19, pls. i–vi.
- He, M.-J. & Chen, Y.-R. (1981) [Two new catfishes of the genus *Akysis* from China]. *Zoological Research*, 2, 209–214. [In Chinese]
- He, S.-P. (1995) The analysis of historical biogeography for the glyptosternoid fishes (Teleostei: Siluriformes: Sisoridae). *Biogeographica*, 71, 145–160.
- He, S.-P. (1996a) [A new species of the genus *Gagata* (Pisces: Sisoridae)]. *Acta Zootaxonomica Sinica*, 21, 380–382. [In Chinese and English]
- He, S.-P. (1996b) The phylogeny of the glyptosternoid fishes (Teleostei: Siluriformes, Sisoridae). *Cybium (3e série)*, 20, 115–159.

- He, S.-P., Gayet, M. & Meunier, F.J. (1999) Phylogeny of the Amphiliidae (Teleostei: Siluriformes). *Annales des sciences naturelles. Zoologie et biologie animale* (Sér. 13), 20, 117–146.
- He, S.-P. & Meunier, F. (1998) The biogeography of Amphiliidae and Sisoridae (Teleostei: Siluriformes). *Acta Zootaxonomica Sinica*, 23 (Supplement), 1–9.
- Heckel, J.J. (1838) *Fische aus Caschmir Gesammelt und Herausgegeben von Carl Freiherrn v. Hügel, Beschrieben von Joh. Jacob Heckel*, Gedruckt bei den P. P. Mechitaristen, Wien, 112 p., 13 pls.
- Heckel, J.J. (1843) Ichthyologie [von Syrien]. In: J. von Russegger, 1841–1849, *Reisen in Europa, Asien und Afrika, mit besonderer Rücksicht auf die naturwissenschaftlichen Verhältnisse der betreffenden Länder, unternommen in den Jahren 1835 bis 1841*, Vol. 1, part 2, E. Schweizerbart, Stuttgart, pp. 990–1099.
- Heckel, J.J. (1849) Über eine neue fossile Fishgattung, *Chirocentrites*, und die ersten Ueberreste eines Siluroiden aus der Vorwelt. *Sitzungsberichte der Kaiserlichen Akademie der Wissenschaften, Mathematisch- Naturwissenschaftlichen Classe*, Wien, 2, 16–19.
- Heckel, J.J. (1850) Beiträge zur Kenntniss der fossilen Fische Oesterreichs. *Denkschriften der Mathematisch- Naturwissenschaftlichen Classe der Kaiserlichen Akademie der Wissenschaften in Wien*, 1, 201–242, 13 pls.
- Heilprin, A. (1887) Explorations on the west coast of Florida and in the Okeechobee wilderness, with special reference to the geology and zoology of the Floridian peninsula. A narrative of researches undertaken under the auspices of the Wagner Free Institute of Science. *Transactions of the Wagner Free Institute of Science of Philadelphia*, 1, 1–134, pls. 1–19.
- Heitmans, W.R.B., Nijssen, H. & Isbrücker, I.J.H. (1983) The mailed catfish genus *Lasiancistrus* Regan, 1904, from French Guiana and Surinam, with descriptions of two new species (Pisces, Siluriformes, Loricariidae). *Bijdragen tot de Dierkunde*, 53, 33–48.
- Hemming, F. (Ed.) (1953) *Copenhagen Decisions on Zoological Nomenclature: Additions to, and Modifications of, the Règles Internationales de la Nomenclature Zoologique. Approved and Adopted by the Fourteenth International Congress of Zoology, Copenhagen, August, 1953*, International Trust for Zoological Nomenclature, London.
- Henn, A.W. (1928) List of types of fishes in the collection of the Carnegie Museum on September 1, 1928. *Annals of the Carnegie Museum*, 19, 51–99.
- Hensel, R.F. (1868) Beiträge zur Kenntniss der Wirbelthiere Südbrasiliens. *Archiv für Naturgeschichte*, 34, 323–375.
- Hensel, R.F. (1870) Beiträge zur Kenntniss der Wirbelthiere Südbrasiliens (Fortsetzung). *Archiv für Naturgeschichte*, 36, 50–91.
- Hermann, J. (1804) *Observationes Zoologicae, Quibus Novae Complures, Aliaeque Animalium Species Describuntur et Illustrantur. Opus Posthumum Edidit Fridericus Ludovicus Hammer*, Argentorati, viii + 332 p. [Pisces on pp. 290–328]
- Herre, A.W.C.T. (1924a) Distribution of the true fresh-water fishes in the Philippines, II.— The Philippine Labyrinthici, Clariidae, and Siluridae. *Philippine Journal of Science*, 24, 683–709, pls. 1–2.
- Herre, A.W.C.T. (1924b) The distribution of true fresh-water fishes in the Philippines and its significance. *Proceedings of the Pan-Pacific Science Congress, 2 (Australia)*. pp. 1561–1570.
- Herre, A.W.C.T. (1926) A summary of the Philippine catfishes, order Nematognathi. *Philippine Journal of Science*, 31, 385–411, pl. 1.
- Herre, A.W.C.T. (1932) Fishes from Kwangtung Province and Hainan Island, China. *Lingnan Science Journal*, 11, 423–443.
- Herre, A.W.C.T. (1933) *Herklotella anomala*.— A new fresh water cat-fish from Hong Kong. *Hong Kong Naturalist*, 4, 179–180.
- Herre, A.W.C.T. (1934) Notes on new or little known fishes from southeastern China. *Lingnan Science Journal*, 13, 285–296.
- Herre, A.W.C.T. (1935) New fishes obtained by the Crane Pacific expedition. *Publication, Zoological Series, Field Museum of Natural History*, 18, 383–438.
- Herre, A.W.C.T. (1940) New species of fishes from the Malay Peninsula and Borneo. *Bulletin of the Raffles Museum*, 16, 5–26, pls. 1–20.
- Herre, A.W.C.T. (1942) *Glyptothorax housei*, a new sisorid catfish from south India. *Stanford Ichthyological Bulletin*, 2, 117–119.
- Herre, A.W.C.T. & Myers, G.S. (1937) A contribution to the ichthyology of the Malay Peninsula. *Bulletin of the Raffles Museum*, 13, 5–75, pls. 1–7.
- Herzenstein, S.M. (1889a) Über einen russischen Wels (*Exostoma Oschanini* Herz.). *Mélanges Biologiques, tirés du Bulletin physico-mathématique de l'Académie Impériale des Sciences de St. Pétersbourg*, 13, 69–73.

- Herzenstein, S.M. (1889b) Über einen russischen Wels (*Exostoma Oschanini* Herz.). *Bulletin de l'Académie Impériale des Sciences de St. Pétersbourg (n. s.)*, 1, 119–123.
- Hieronimus, H. (1991) *Corydoras similis* spec. nov., ein neuer Panzerwels aus Brasilien (Pisces: Siluriformes: Callichthyidae). *Zeitschrift für Fischkunde*, 1, 39–46.
- Hieronimus, H. (1995) *Corydoras lacerdai* spec. nov., ein neuer Panzerwels aus Brasilien (Teleostei: Siluriformes: Callichthyidae). *Zeitschrift für Fischkunde*, 3, 107–116.
- Higuchi, H. (1992) *A Phylogeny of the South American Thorny Catfishes (Osteichthyes; Siluriformes, Doradidae)*. Unpublished Ph.D. dissertation, Harvard University, Cambridge, Massachusetts.
- Higuchi, H., Britski, H.A. & Garavello, J.C. (1990) *Kalyptodoras bahiensis*, a new genus and species of thorny catfish from northeastern Brazil (Siluriformes: Doradidae). *Ichthyological Exploration of Freshwaters*, 1, 219–225.
- Higuchi, H., Reis, E.G. & Araújo, F.G. (1982) Uma nova espécie de bagre marinho do litoral do Rio Grande do sul e considerações sobre o gênero nominal *Netuma* Bleeker, 1858 no Atlântico sul ocidental (Siluriformes, Ariidae). *Atlantica, Rio Grande*, 5, 1–15.
- Hildebrand, S.F. (1925) Fishes of the Republic of El Salvador, Central America. *Bulletin of the Bureau of Fisheries*, 41, 237–287.
- Hildebrand, S.F. (1938) A new catalogue of the fresh-water fishes of Panama. *Zoological Series, Field Museum of Natural History*, 22, 217–259.
- Hilgendorf, F.M. (1878) Einige neue japanische Fischgattungen. *Sitzungsberichte der Gesellschaft Naturforschender Freunde zu Berlin*, 1878, 404–457.
- Hilgendorf, F.M. (1888) Fische aus dem Victoria-Nyanza (Ukerewe-See), gesammelt von dem verstorbenen Dr. G. A. Fischer. *Sitzungsberichte der Gesellschaft Naturforschender Freunde zu Berlin*, 1888, 75–79.
- Hilgendorf, F.M. (1905) Fische von Deutsch und Englisch Ost-Africa. *Zoologische Jahrbücher, Abteilung für Systematik, Geographie und Biologie der Tiere (Jena)*, 22, 405–420.
- Hilgendorf, F.M. & Pappenheim, P. (1903) Über die Fischfauna des Rukwa-Sees. *Sitzungsberichte der Gesellschaft Naturforschender Freunde zu Berlin*, 1903, 259–271.
- Hoedeman, J.J. (1949) Onderorde Siluroidea "Meervallen". *Encyclopaedie voor de aquariumhouder*: pp. X.30.X.1–6.
- Hoedeman, J.J. (1952a) *Encyclopaedie voor de Aquariumhouder*. De Regenboog, Amsterdam, 17 p.
- Hoedeman, J.J. (1952b) Notes on the ichthyology of Surinam (Dutch Guiana). The catfish genera *Hoplosternum* and *Callichthys*, with key to the genera and groups of the family Callichthyidae. *Beaufortia*, 12, 1–11.
- Hoedeman, J.J. (1961) Notes on the ichthyology of Surinam and other Guianas. 8.— Additional records of siluriform fishes (2). *Bulletin of Aquatic Biology*, 2, 129–139.
- Hoedeman, J.J. (1965) *Elseviers Pocketboek voor de Aquariumliefhebber*, Elsevierpocket (A) 108, Elsevier, Amsterdam/Brussels, 176 p.
- Holbrook, J.E. (1855) An account of several species of fish observed in Florida, Georgia, etc. *Journal of the Academy of Natural Sciences of Philadelphia (Ser. 2)*, 3, 47–58, pls. 5–6.
- Hollanda Carvalho, P. & Weber, C. (2004) Five new species of the *Hypostomus cochliodon* group (Siluriformes: Loricariidae) from the middle and lower Amazon System. *Revue suisse de Zoologie, Annales de la Société zoologique suisse et du Muséum d'Histoire naturelle de Genève*, 111, 953–978.
- Holly, M. (1926) Zwei neue Siluriden und ein neuer Charcinide aus Kamerun. *Anzeiger der Mathematisch- Naturwissenschaftlichen Classe der Kaiserlichen Akademie der Wissenschaften in Wien*, 62, 157–159.
- Holly, M. (1927a) Eine neue Siluriden-Subspezies aus Kamerun. *Anzeiger der Mathematisch- Naturwissenschaftlichen Classe der Kaiserlichen Akademie der Wissenschaften in Wien*, 64, 8–9.
- Holly, M. (1927b) Siluriden, Cyprinodontiden, Acanthopterygien und Mastacembeliden aus Kamerun. *Sitzungsberichte der Kaiserlichen Akademie der Wissenschaften, Mathematisch- Naturwissenschaftlichen Classe, Wien, Abt. 1, Botanik, Zoologie, Anatomie, Geologie und Paläontologie*, 136, 195–230.
- Holly, M. (1927c) Weitere Fische aus Kamerun. *Sitzungsberichte der Kaiserlichen Akademie der Wissenschaften, Mathematisch- Naturwissenschaftlichen Classe, Wien, Abt. 1, Botanik, Zoologie, Anatomie, Geologie und Paläontologie*, 136, 421–427.
- Holly, M. (1929) Einige neue Fischformen aus Brasilien. *Anzeiger der Mathematisch- Naturwissenschaftlichen Classe der Kaiserlichen Akademie der Wissenschaften in Wien*, 66, 117–120.
- Holly, M. (1930) Synopsis der Süßwasserfische Kameruns. *Sitzungsberichte der Kaiserlichen Akademie der Wissenschaften, Mathematisch- Naturwissenschaftlichen Classe, Wien, Abt. 1, Botanik, Zoologie, Anatomie, Geologie und Paläontologie*, 139, 195–281, pls. 1–2.
- Holly, M. (1939) Zur Nomenklatur der Siluridengattung *Macrones*, C. Duméril. *Zoologischer Anzeiger*, 125, 143.

- Holly, M. (1940) Vier noch nicht beschriebene *Corydoras* Arten. *Anzeiger der Mathematisch- Naturwissenschaftlichen Classe der Kaiserlichen Akademie der Wissenschaften in Wien*, 77, 107–112.
- Holmberg, E.L. (1893a) Nombres vulgares de peces Argentinos con sus equivalencias científicas. *Revista del Jardín Zoológico de Buenos Ayres*, 1, 85–96.
- Holmberg, E.L. (1893b) Dos peces argentinos. *Revista del Jardín Zoológico de Buenos Ayres*, 1, 353–354.
- Holota, K. (1990) Pflege und Zucht von *Ancistrus* cf. *hoplogenys*. *Die Aquarien- und Terrarien- Zeitschrift*, 43, 147–148.
- Hora, S.L. (1921a) Fish and fisheries of Manipur with some observations on those of the Naga Hills. *Records of the Indian Museum*, 22, 165–214, pls. 9–12.
- Hora, S.L. (1921b) On some new or rare species of fish from the eastern Himalayas. *Records of the Indian Museum*, 22, 731–744, pl. 29.
- Hora, S.L. (1923a) On a collection of fish from Siam. *Journal of the Natural History Society of Siam*, 6, 143–184, pls. 10–12.
- Hora, S.L. (1923b) Notes on fishes in the Indian Museum, V.— On the composite genus *Glyptosternon* M'Clelland. *Records of the Indian Museum*, 25, 1–44, pls. 1–4.
- Hora, S.L. (1929) An aid to the study of Hamilton Buchanan's "Gangetic Fishes". *Memoirs of the Indian Museum*, 9, 169–192, pl. 13–23.
- Hora, S.L. (1931) On two small collections of fishes from Burma. *Records of the Indian Museum*, 33, 1–2.
- Hora, S.L. (1932) *Glyptosternum reticulatum*, a siluroid fish from Afghanistan. *Annals and Magazine of Natural History (Ser. 10)*, 10, 176–179.
- Hora, S.L. (1933) Siluroid fishes of India, Burma and Ceylon. I.— Loach-like fishes of the genus *Amblyceps* Blyth. *Records of the Indian Museum*, 35, 607–621.
- Hora, S.L. (1934) The fish of Chitral. *Records of the Indian Museum*, 36, 279–319, 2 pls.
- Hora, S.L. (1936a) Siluroid fishes of India, Burma and Ceylon. II.— Fishes of the genus *Akysis* Bleeker. III.— Fishes of the genus *Olyra* M'Clelland. IV.— On the use of the generic name *Wallago* Bleeker. V.— Fishes of the genus *Heteropneustes* Müller. *Records of the Indian Museum*, 38, 199–209.
- Hora, S.L. (1936b) Siluroid fishes of India, Burma and Ceylon. VI.— Fishes of the genus *Clarias* Gronovius. VII.— Fishes of the genus *Silurus* Linnaeus. VIII.— Fishes of the genus *Callichrous* Hamilton. *Records of the Indian Museum*, 38, 347–361.
- Hora, S.L. (1937a) Geographical distribution of Indian freshwater fishes and its bearing on the probable land connections between India and the adjacent countries. *Current Science*, 5, 351–356.
- Hora, S.L. (1937b) The game fishes of India. III.— Garua bachcha or Gaurchcha. *Journal of the Bombay Natural History Society*, 39, 659–678, pl. 1.
- Hora, S.L. (1937c) A new genus of Siamese catfishes. *Journal of the Siam Society, Natural History Supplement*, 11, 39–46, pl. 2.
- Hora, S.L. (1937d) Notes on fishes in the Indian Museum, XXX.— On the systematic position of *Cyprinus cosuatis* Hamilton. XXXI.— On a small collection of fish from Sandoway, lower Burma. XXXII.— On a small collection of fish from the upper Chindwin drainage. XXXIII.— On a collection of fish from the Kumaon Himalayas. XXXIV.— On a new catfish from Kwangsi, China. XXXV.— A further note on Hamilton's *Cyprinus* (*Garra*) *lamta*. XXXVI.— On a new genus of Chinese catfishes allied to *Pseudecheneis*. *Records of the Indian Museum*, 39, 321–350.
- Hora, S.L. (1937e) The game fishes of India. II.— The Bachhwa or Butchwa. *Journal of the Bombay Natural History Society*, 39, 431–446, 1 plate.
- Hora, S.L. (1938a) The game fishes of India. IV.— The silonid catfish, *Silonia silondia*. *Journal of the Bombay Natural History Society*, 40, 137–147, plate.
- Hora, S.L. (1938b) The game fishes of India. V.— The pungas catfish, *Pangasius pangasius* (Hamilton). *Journal of the Bombay Natural History Society*, 40, 355–366, plate.
- Hora, S.L. (1938c) A new name for *Silurus sinensis* Hora. *Records of the Indian Museum*, 40, 243.
- Hora, S.L. (1938d) Notes on fishes in the Indian Museum, XXXVIII.— On the systematic position of *Bagrus lonah* Sykes, with descriptions of and remarks on other glyptosternoid fishes from the Deccan. *Records of the Indian Museum*, 40, 363–375, pl. 7.
- Hora, S.L. (1939a) The game fishes of India. VI.— The goonch, *Bagarius bagarius* (Hamilton). *Journal of the Bombay Natural History Society*, 40, 583–593, plate.
- Hora, S.L. (1939b) The game fishes of India. VII.— The mulley of boali, *Wallagonia attu* (Bloch and Schneider). *Journal of the Bombay Natural History Society*, 41, 64–71, plate.

- Hora, S.L. (1941) Siluroid fishes of India, Burma and Ceylon. XI.— Fishes of the schilbeid genera *Silonopangasius* Hora, *Pseudeutropius* Bleeker, *Proeutropiichthys* Hora, and *Ailia* Gray. XII.— A further note on fishes of the genus *Clarias* Gronovius. *Records of the Indian Museum*, 43, 97–115.
- Hora, S.L. (1949a) Systematic position of three glyptosternoid fishes described by Hamilton. *Records of the Indian Museum*, 46, for 1948, 55–62, pl. 2.
- Hora, S.L. (1949b) Systematic position of siluroids in Hamilton's Gangetic fishes. *Records of the Indian Museum*, 46, for 1948, 63–72.
- Hora, S.L. (1950) Siluroid fishes of India, Burma and Ceylon. XIII.— Fishes of the genera *Erethistes* Müller and Troschel, *Hara* Blyth and of two new allied genera. *Records of the Indian Museum*, 47, 183–202, pls. 1–2.
- Hora, S.L. (1952) Parallel evolution of *Pseudecheneis* Blyth and similar fishes of south east Asia. *Journal of the Asiatic Society. Science*, 18, 183–202.
- Hora, S.L. & Chabanaud, P. (1930) The siluroid fish *Pseudecheneis* and an allied new genus. *Records of the Indian Museum*, 32, 215–221.
- Hora, S.L. & Gupta, J.C. (1941) Notes on Malayan fishes in the collection of the Raffles Museum, Singapore. I.— Catfishes of the families Siluridae, Bagridae, Amblycepidae, Akysidae, Sisoridae, Chacidae, Schilbeidae, and Clariidae. *Bulletin of the Raffles Museum*, 17, 12–43, pls. 2–4.
- Hora, S.L. & Law, N.C. (1941) Siluroid fishes of India, Burma and Ceylon. IX.— Fishes of the genera *Gagata* Bleeker and *Nangra* Day. X.— Fishes of the genus *Batasio* Blyth. *Records of the Indian Museum (Calcutta)*, 43, 9–42, 2 pls.
- Hora, S.L. & Silas, E.G. (1952a) Notes on fishes in the Indian Museum, XLVII.— Revision of the glyptosternoid fishes of the family Sisoridae, with descriptions of new genera and species. *Records of the Indian Museum*, 49, for 1951, 5–29, pl. 1.
- Hora, S.L. & Silas, E.G. (1952b) Evolution and distribution of glyptosternoid fishes of the family Sisoridae (Order: Siluroidea). *Proceedings of the National Institute of Sciences of India*, 18, 309–322.
- Hough, F.B. (1852) Catalogue of reptiles and fishes. Fishes. In: *Fifth annual report of the Regents of the University, on the condition of the state cabinet of natural history, and the historical and antiquarian collection annexed thereto, Report no. 122*, University of the State of New York, Albany, 24–28.
- Houttuyn, M. (1782) Beschrijving van eenige Japanse visschen en andere zee-schepzelen. *Verhandelingen der Hollandsche Maatschappij der Wetenschappen, Haarlem*, 20, 311–350.
- Howes, G.J. (1980) A new catfish from Sierra Leone. *Bulletin of the British Museum (Natural History), Zoology*, 38, 165–170.
- Howes, G.J. (1983a) Problems in catfish anatomy and phylogeny exemplified by the Neotropical Hypophthalmidae (Teleostei: Siluroidei). *Bulletin of the British Museum (Natural History), Zoology*, 45, 1–39.
- Howes, G.J. (1983b) The cranial muscles of the loricarioid catfishes, their homologies and value as taxonomic characters (Teleostei: Siluroidei). *Bulletin of the British Museum (Natural History), Zoology*, 45, 309–345.
- Howes, G.J. (1985) The phylogenetic relationships of the electric catfish family Malapteruridae (Teleostei: Siluroidei). *Journal of Natural History*, 19, 37–67.
- Howes, G.J. & Fumihito, A. (1991) Cranial anatomy and phylogeny of the south east Asian catfish genus *Belodontichthys*. *Bulletin of the British Museum (Natural History), Zoology*, 57, 133–160.
- Hu, X.-Y., Lan, J.-H. & Zhang, C.-G. (2004) A new species of the genus *Duanensis* [sic, *Silurus*] (Siluriformes, Siluridae) from Guangxi, China. *Acta Zootaxonomica Sinica*, 29, 586–590.
- Huang, S.-Y. (1981) On two new species of the catfish genus *Platytopius* Hora from Yunnan, China. *Acta Zootaxonomica Sinica*, 6, 437–440. [In Chinese, with new species description repeated in English.]
- Hubbs, C.L. (1935) Fresh-water fishes collected in British Honduras and Guatemala. *Miscellaneous Publications, Museum of Zoology, University of Michigan*, 28, 1–22, pls. 1–4.
- Hubbs, C.L. (1936) Fishes of the Yucatan Peninsula. *Carnegie Institution of Washington Publication*, 157–287, pls. 1–15.
- Hubbs, C.L. & Bailey, R.M. (1947) Blind catfishes from artesian waters of Texas. *Occasional Papers of the Museum of Zoology, University of Michigan*, 499, 1–15, pl. 1.
- Hubbs, C.L. & Hibbard, C.W. (1951) *Ictalurus lambda*, a new catfish, based on a pectoral spine from the lower Pliocene of Kansas. *Copeia*, 1951, 8–14.
- Hubbs, C.L. & Miller, R.R. (1960) *Potamarius*, a new genus of ariid catfishes from the fresh waters of Middle America. *Copeia*, 1960, 101–112, pl. 1.
- Hubbs, C.L. & Raney, E.C. (1944) Systematic notes on North American siluroid fishes of the genus *Schilbeodes*. *Occasional Papers of the Museum of Zoology, University of Michigan*, 487, 1–36, 1 pl.

- Hubrecht, A.A.W. (1881) On a collection of fishes from the St. Paul's River, Liberia, with description of three new species. *Notes from the Leyden Museum*, 3, 66–71.
- Humboldt, F.H.A.von. (1805a) Mémoire sur l'*Eremophilus* et *Astroblepus*, deux nouveaux genres de l'ordre des apodes. In: *Voyage aux Régions Équinoxiales du Nouveau Continent fait en 1799 et 1800, par A. de Humboldt et A. Bonpland. Deuxième Partie: Observations de Zoologie et d'Anatomie Comparée*, Paris, pp. 17–20, pls. 6–7.
- Humboldt, F.H.A.von. (1805b) Mémoire sur une nouvelle espèce de pimelode, jetée par les volcans du Royaume de Quito. In: *Voyage aux Régions Équinoxiales du Nouveau Continent fait en 1799 et 1800, par A. de Humboldt et A. Bonpland. Deuxième Partie: Observations de Zoologie et d'Anatomie Comparée*, Paris, pp. 21–25, pl. 7.
- Humboldt, F.H.A.von & Valenciennes, A. (1821) Recherches sur les poissons fluviatiles de l'Amérique Équinoxiale. In: *Voyage aux Régions Équinoxiales du Nouveau Continent fait en 1799 et 1800, par A. de Humboldt et A. Bonpland. Deuxième Partie: Observations de Zoologie et d'Anatomie Comparée*, Paris, pp. 145–216, pls. 45–52.
- Husain, A. & Tilak, R. (1978) On the redescription and distribution of *Hara jerdoni* Day (Sisoridae: Siluriformes). *Journal of the Inland Fishery Society of India*, 9, 92–97.
- Hussakof, L. (1932) The fossil fishes collected by the Central Asiatic Expedition. *American Museum Novitates*, 553, 1–19.
- Hyrtl, C.J. (1859) Anatomische Untersuchung des *Clarotes (Gonocephalus) heuglini* Kner. Mit einer Abbildung und einer osteologischen Tabelle der Siluroiden. *Denkschriften der Mathematisch-Naturwissenschaftlichen Classe der Kaiserlichen Akademie der Wissenschaften in Wien*, 16, 1–18, 1 pl.
- Ihering, H. von. (1898) Description of a new fish from São Paulo. Pp. 108–109, appended to: Contributions to the herpetology of São Paulo.— 1. *Proceedings of the Academy of Natural Sciences, Philadelphia*, 50, 101–109.
- Ihering, R. von. (1905) Descriptions of four new loricariid fishes of the genus *Plecostomus* from Brazil. *Annals and Magazine of Natural History (Ser. 7)*, 15, 558–561.
- Ihering, R. von. (1907) Diversas especies novas de peixes nemathognathos do Brasil. (Several new species of Brazilian nematognath fishes.) Notas preliminares. *Revista do Museu Paulista (N. S.)*, 1, 13–39.
- Ihering, R. von. (1911) Algunas especies novas de peixes d'água doce (Nematognatha) (*Corydoras*, *Plecostomus*, *Hemipsilichthys*). *Revista do Museu Paulista*, 8, for 1910, 380–404.
- Ihering, R. von. (1928a) Uma nova especie de *Otocinclus* (Pisces. Nematognatha) "cascudinho" de S. Paulo. *Boletim Biológico, São Paulo*, 11, 1–3.
- Ihering, R. von. (1928b) *Glanidium cesarpintoi* n. sp. de Peixe de couro (fam. Siluridae sub-fam. Auchenipterinae). *Boletim Biológico, São Paulo*, 12, 46–49.
- Ihering, R. von. (1930) Notas ecológicas referentes a peixes d'água doce do Estado de S. Paulo e descrição de 4 espécies novas. *Archivos do Instituto Biológico, São Paulo*, 3, 93–103, pl. 13.
- Ihering, R. von. (1968) *Dicionário dos Animais do Brasil*. Editora da Universidade de Brasília, São Paulo, Brazil, 790 p.
- Inger, R.F. (1956) Notes on a collection of fishes from southeastern Venezuela. *Fieldiana, Zoology*, 34, 425–440.
- Inger, R.F. & Chin, P.K. (1959) New species of fresh-water catfishes from North Borneo. *Fieldiana, Zoology*, 39, 279–296.
- Innes, W.T. & Myers, G.S. (1950) The "Imitator catfish," which mimics a *Corydoras*. *The Aquarium*, 19, 222–223.
- International Commission on Zoological Nomenclature [ICZN] (1961) *International Code of Zoological Nomenclature, Second Edition*, International Trust for Zoological Nomenclature, London and Berkeley.
- International Commission on Zoological Nomenclature [ICZN] (1985) *International Code of Zoological Nomenclature, Third Edition*, International Trust for Zoological Nomenclature, London and Berkeley.
- International Commission on Zoological Nomenclature [ICZN] (1999) *International Code of Zoological Nomenclature, Fourth Edition*, International Trust for Zoological Nomenclature, London, 306 p.
- International Commission on Zoological Nomenclature [ICZN] (2002) Opinion 2011. *Hemibagrus* Bleeker, 1862 (Osteichthyes, Siluriformes), *Bagrus nemurus* Valenciennes in Cuvier & Valenciennes, 1840, *B. planiceps* Valenciennes, 1840, *B. flavus* Bleeker, 1846 and *B. sieboldii* Bleeker, 1846: Previous fixations of type specimens not to be set aside. *Bulletin of Zoological Nomenclature*, 59, 220–221.
- Isbrücker, I.J.H. (1969) Over de namen van enkele *Corydoras*-soorten. *Het Aquarium*, 39, 202–203.
- Isbrücker, I.J.H. (1971a) A redescription of the South American catfish *Loricariichthys maculatus* (Bloch, 1794), with designation of the lectotype and restriction of its type locality (Pisces, Siluriformes, Loricariidae). *Bijdragen tot de Dierkunde*, 41, 10–18.
- Isbrücker, I.J.H. (1971b) *Pseudohemiodon (Planiloricaria) cryptodon*, a new species and subgenus from Peru (Pisces, Siluriformes, Loricariidae). *Bonner Zoologische Beiträge*, 21, for 1970, 274–283, 7 pls.
- Isbrücker, I.J.H. (1972) The identity of the South American catfish *Loricaria cataphracta* Linnaeus, 1758, with descrip-

- tions of the original type specimens of four other nominal *Loricaria* species (Pisces, Siluriformes, Loricariidae). *Beaufortia*, 19, 163–191.
- Isbrücker, I.J.H. (1973) Status of the primary homonymous South American catfish *Loricaria cirrhosa* Perugia, 1897, with remarks on some other loricariids (Pisces, Siluriformes, Loricariidae). *Annali del Museo Civico de Storia Naturale di Genova*, 79, 172–191.
- Isbrücker, I.J.H. (1975a) *Pseudohemiodon thorectes*, a new species of mailed catfish from the Rio Mamoré system, Bolivia (Pisces, Siluriformes, Loricariidae). *Beaufortia*, 23, 85–92.
- Isbrücker, I.J.H. (1975b) *Metaloricaria paucidens*, a new species and genus of mailed catfish from French Guiana (Pisces, Siluriformes, Loricariidae). *Bulletin, Institut royal des Sciences naturelles de Belgique, Biologie*, 50, 1–9, pls. 1–3.
- Isbrücker, I.J.H. (1979a) Description préliminaire de nouveaux taxa de la famille des Loricariidae, poissons-chats cuirassés néotropicaux, avec un catalogue critique de la sous-famille nominale (Pisces, Siluriformes). *Revue Française d'Aquariologie et Herpetologie*, 5, for 1978, 86–116.
- Isbrücker, I.J.H. (1979b) Les poissons de la famille des Loricariidés ou poissons-chats cuirassés. *Revue Française d'Aquariologie et Herpetologie*, 6, 109–124.
- Isbrücker, I.J.H. (1980) Classification and catalogue of the mailed Loricariidae (Pisces, Siluriformes). *Verslagen en Technische Gegevens, Instituut voor Taxonomische Zoölogie, Universiteit van Amsterdam*, 22, 1–181.
- Isbrücker, I.J.H. (1981) Revision of *Loricaria* Linnaeus, 1758 (Pisces, Siluriformes, Loricariidae). *Beaufortia*, 31, 51–96.
- Isbrücker, I.J.H. (1986) Trichomycteridae, mysterieuze meervallen. *Het Aquarium*, 56, 274–279.
- Isbrücker, I.J.H. (1992) Überblick über die güttigen (Unter-) Gattungsnamen der Harnischwelse (Loricariidae) und ihre Synonyme. *Die Aquarien- und Terrarien-Zeitschrift*, 1992, 71–72.
- Isbrücker, I.J.H. (1992) Ein fish findet nach 138 Jahren sein Zuhause. Der verborgene Fundort von *Hemiancistrus medians* (Kner, 1854). In: Stawikowski, R. (Ed.), *Harnischwelse*, Die Aquarien- und Terrarien- Zeitschrift, Sonderheft, Eugen Ulmer, Stuttgart, pp. 56–57.
- Isbrücker, I.J.H. (1999) *Corydoras geoffroy* und *Cataphractus punctatus*. Revision der beiden zuerst entdeckten Arten der Unterfamilie Corydoradinae (Actinopterygii, Siluriformes, Callichthyidae). Teil 1. *Die Aquarien- und Terrarien-Zeitschrift*, 52, 38–43.
- Isbrücker, I.J.H. (1999b) *Corydoras bolivianus* und *Corydoras geryi*. *Die Aquarien- und Terrarien-Zeitschrift*, 52, 58–59.
- Isbrücker, I.J.H. (2000) *Corydoras geoffroy* und *Cataphractus punctatus*. History of both earliest discovered species of the subfamily Corydoradinae (Actinopterygii, Siluriformes, Callichthyidae). *Cat Chat*, 1, 11–18.
- Isbrücker, I.J.H. (2001a) Nomenklator der Gattungen und Arten der Harnischwelse, Familie Loricariidae Rafinesque, 1815 (Teleostei, Ostariophysi). In: Stawikowski, R. (Ed.), *Harnischwelse 2*, Die Aquarien- und Terrarien-Zeitschrift, Eugen Ulmer, Stuttgart, pp. 25–32.
- Isbrücker, I.J.H. (2001b) Catalogue of genera and species of Corydoradinae Hoedeman, 1952 (Teleostei, Ostariophysi, Callichthyidae), including type localities, type specimens, and etymology. In: Fuller, I.A.M. (Ed.), *Breeding Corydoradine Catfishes*, Ian Fuller Enterprises, Kidderminster, pp. 213–247.
- Isbrücker, I.J.H., Britski, H.A., Nijssen, H. & Ortega, H. (1983) *Aposturisoma myriodon*, une espèce et un genre nouveaux de poisson-chat cuirassé, tribu Farlowellini Fowler, 1958 du bassin du Rio Ucayali, Pérou (Pisces, Siluriformes, Loricariidae). *Revue Française d'Aquariologie et Herpetologie*, 10, 33–42.
- Isbrücker, I.J.H. & Nijssen, H. (1973) Two new species of the callichthyid catfish genus *Corydoras* from Brazil (Pisces, Siluriformes, Callichthyidae). *Beaufortia*, 21, 1–7.
- Isbrücker, I.J.H. & Nijssen, H. (1974a) *Rhadinoloricaria* gen. nov. and *Planiloricaria*, two genera of South American mailed catfishes (Pisces, Siluriformes, Loricariidae). *Beaufortia*, 22, 67–81.
- Isbrücker, I.J.H. & Nijssen, H. (1974b) On *Hemiodontichthys acipenserinus* and *Reganella depressa*, two remarkable mailed catfishes from South America (Pisces, Siluriformes, Loricariidae). *Beaufortia*, 22, 193–222.
- Isbrücker, I.J.H. & Nijssen, H. (1976a) The South American mailed catfishes of the genus *Pseudoloricaria* Bleeker, 1862 (Pisces, Siluriformes, Loricariidae). *Beaufortia*, 25, 107–129.
- Isbrücker, I.J.H. & Nijssen, H. (1976b) *Rineloricaria heteroptera*, a new species of mailed catfish from Rio Amazonas near Manaus, Brazil (Pisces, Siluriformes, Loricariidae). *Zoologischer Anzeiger*, 196, 109–124.
- Isbrücker, I.J.H. & Nijssen, H. (1978a) Two new species and a new genus of neotropical mailed catfishes of the subfamily Loricariinae Swainson, 1838 (Pisces, Siluriformes, Loricariidae). *Beaufortia*, 27, 177–206.
- Isbrücker, I.J.H. & Nijssen, H. (1978b) The neotropical mailed catfishes of the genera *Lamontichthys* P. de Miranda-

- Ribeiro, 1939 and *Pterosturisoma* n. gen., including the description of *Lamontichthys stibaros* n. sp. from Ecuador (Pisces, Siluriformes, Loricariidae). *Bijdragen tot de Dierkunde*, 48, 57–80.
- Isbrücker, I.J.H. & Nijssen, H. (1979) Three new South American mailed catfishes of the genera *Rineloricaria* and *Loricariichthys* (Pisces, Siluriformes, Loricariidae). *Bijdragen tot de Dierkunde*, 48, 191–211.
- Isbrücker, I.J.H. & Nijssen, H. (1982) New data on *Metaloricaria paucidens* from French Guiana and Surinam (Pisces, Siluriformes, Loricariidae). *Bijdragen tot de Dierkunde*, 52, 155–168.
- Isbrücker, I.J.H. & Nijssen, H. (1983a) *Aphanotorulus frankei*, une espèce et un genre nouveaux de poissons-chats cuirassés du bassin du Rio Ucayali au Pérou (Pisces, Siluriformes, Loricariidae). *Revue Française d'Aquariologie et Herpetologie*, 9, for 1982, 105–110.
- Isbrücker, I.J.H. & Nijssen, H. (1983b) *Crossoloricaria rhami* n. sp., un nouveau poisson-chat cuirassé du Rio Huacamayo, Pérou (Pisces, Siluriformes, Loricariidae). *Revue Française d'Aquariologie et Herpetologie*, 10, 9–12.
- Isbrücker, I.J.H. & Nijssen, H. (1984a) *Rineloricaria castroi*, a new species of mailed catfish from Rio Trombetas, Brazil (Pisces, Siluriformes, Loricariidae). *Beaufortia*, 34, 93–99.
- Isbrücker, I.J.H. & Nijssen, H. (1984b) *Pyxiloricaria menezesi*, a new genus and species of mailed catfish from Rio Miranda and Rio Cuiabá, Brazil (Pisces, Siluriformes, Loricariidae). *Bijdragen tot de Dierkunde*, 54, 163–168.
- Isbrücker, I.J.H. & Nijssen, H. (1984c) *Hypostomus nematopterus*, a new species of mailed catfish from the Oyapock River system, French Guiana (Pisces, Siluriformes, Loricariidae). *Bulletin Zoologisch Museum, Universiteit van Amsterdam*, 10, 9–13.
- Isbrücker, I.J.H. & Nijssen, H. (1985) *Exastilithoxus hoedemani*, a new species of mailed catfish from Rio Marauiá, Est. Amazonas, Brazil (Pisces, Siluriformes, Loricariidae). *Spixiana*, 8, 221–229.
- Isbrücker, I.J.H. & Nijssen, H. (1986a) New records of the mailed catfish *Planiloricaria cryptodon* from the Upper Amazon in Peru, Brazil and Bolivia, with a key to the genera of the *Planiloricariina*. *Bijdragen tot de Dierkunde*, 56, 39–46.
- Isbrücker, I.J.H. & Nijssen, H. (1986b) *Aristoloricaria condei*, nouveau genre et nouvelle espèce de poisson-chat cuirassé, tribu Loricariini Bonaparte, 1831, du bassin du Rio Napo, haute Amazone, Equateur (Pisces, Siluriformes, Loricariidae). *Revue Française d'Aquariologie et Herpetologie*, 12, for 1985, 103–108.
- Isbrücker, I.J.H. & Nijssen, H. (1988) *Acanthicus adonis*, ein neuer Harnischwels aus dem Rio Tocantins, Brasilien (Pisces, Siluriformes, Loricariidae). *Die Aquarien- und Terrarien- Zeitschrift*, 41, 164–167.
- Isbrücker, I.J.H. & Nijssen, H. (1989) Diagnose dreier neuer Harnischwelsgattungen mit fünf neuen Arten aus Brasilien (Pisces, Siluriformes, Loricariidae). *Die Aquarien- und Terrarien- Zeitschrift*, 42, 541–547.
- Isbrücker, I.J.H. & Nijssen, H. (1991) *Hypancistrus zebra*, a new genus and species of uniquely pigmented ancistrine loricariid fish from the Rio Xingu, Brazil (Pisces: Siluriformes: Loricariidae). *Ichthyological Exploration of Freshwaters*, 1, 345–350.
- Isbrücker, I.J.H. & Nijssen, H. (1992) *Corydoras breei*, a new species of callichthyid catfish from the Corantijn River basin in Surinam (Pisces, Siluriformes, Callichthyidae). *Beaufortia*, 43, 9–14.
- Isbrücker, I.J.H. & Nijssen, H. (1992) Sexualdimorphismus bei Harnischwelsen (Loricariidae). Odontoden, Zähne, Lippen, Tentakel, Genitalpapillen und Flossen. In: Stawikowski, R. (Ed.), *Harnischwelse*. Die Aquarien- und Terrarien- Zeitschrift Sonderheft, pp. 19–33.
- Isbrücker, I.J.H., Nijssen, H. & Cala, P. (1988) *Lithoxancistrus orinoco*, nouveau genre et espèce de poisson-chat cuirassé du Rio Orinoco en Colombie (Pisces, Siluriformes, Loricariidae). *Revue Française d'Aquariologie et Herpetologie*, 15, 13–16.
- Isbrücker, I.J.H., Nijssen, H. & Nico, L.G. (1992) Ein neuer Rüsselzahnwels aus oberen Orinoco-Zuflüssen in Venezuela und Kolumbien: *Leporacanthicus triactis* n. sp. (Pisces, Siluriformes, Loricariidae). *Die Aquarien- und Terrarien- Zeitschrift*, 46, 1–5.
- Isbrücker, I.J.H., Seidel, I., Michels, J.P., Schraml, E. & Werner, A. (2001) Diagnose vierzehn neuer Gattungen der Familie Loricariidae Rafinesque, 1815 (Teleostei, Ostariophysi). In: Stawikowski, R. (Ed.), *Harnischwelse 2*, Die Aquarien- und Terrarien- Zeitschrift Sonderheft, Eugen Ulmer, Stuttgart, pp. 17–24.
- Jackson, P.B.N. (1955) A new fish of the genus *Clarias* Gronov. from Lake Nyasa, with notes on the distribution of the Clariidae and other catfishes in the lake. *Proceedings of the Zoological Society of London*, 125, 681–684.
- Jackson, P.B.N. (1959) Revision of the clariid catfishes of Nyasaland, with a description of a new genus and seven new species. *Proceedings of the Zoological Society of London*, 132, 109–128.
- Jacquemont, V. (1835–1844) *Voyage dans l'Inde Pendant les Années 1828 à 1832, Publié sous les Auspices de M. Guizot, Ministre de l'instruction Publique. Journal. 4 vols. Text, 2 vols, atlas*, Paris. [Fish plates in vol. 2: pls. 13–18; dated to 1840.]

- Jansen, G., Devaere, S., Weekers, P.H.H. & Adriaens, D. (2005) Phylogenetic relationships and divergence time estimate of African anguilliform catfish (Siluriformes: Clariidae) inferred from ribosomal gene and spacer sequences. *Molecular Phylogenetics and Evolution*, 38, for 2006, 65–78.
- Jayaram, K.C. (1952) Taxonomic notes on the fish *Pseudobagrus chryseus* Day, 1865. *Annals and Magazine of Natural History (Ser. 12)*, 5, 980–983.
- Jayaram, K.C. (1954) Siluroid fishes of India, Burma and Ceylon: XIV.— Fishes of the genus *Mystus* Scopoli. *Records of the Indian Museum*, 51, for 1953, 527–558, pl. 19.
- Jayaram, K.C. (1955a) A preliminary review of the genera of the family Bagridae (Pisces: Siluroidea). *Proceedings of the National Institute of Sciences of India*, 21 B, 120–128.
- Jayaram, K.C. (1955b) The Palaearctic element in the fish fauna of peninsular India. *Bulletin of the National Institute of Sciences of India*, 7, 260–263.
- Jayaram, K.C. (1956a) Nomenclatural status of the names *Bagre* Cuvier (Oken), *Bagrus* Valenciennes, and *Porcus*, Geoffroy St. Hilaire. *Copeia*, 1956, 248–249.
- Jayaram, K.C. (1956b) Taxonomic status of the Chinese catfish family Cranoglanididae Myers, 1931. *Proceedings of the National Institute of Sciences of India*, 21B, for 1955, 256–263, pl. 18.
- Jayaram, K.C. (1959) Systematic position of fishes described under *Bagrus* by Valenciennes, 1839. *Records of the Indian Museum*, 54, for 1956, 53–59.
- Jayaram, K.C. (1960) Racial analysis of *Rita chrysea* Day inhabiting the Mahanadi River. *Journal of the Zoological Society of India*, 12, for 1959, 85–103.
- Jayaram, K.C. (1962) The nomenclatural status of *Mystus*, *Macrones*, *Aoria* and other names for a genus of Asiatic siluroid fishes. *Proceedings of the First All India Congress of Zoology, 1959. Part 2—Scientific papers*. Jabalpur, pp. 632–635.
- Jayaram, K.C. (1963) Systematic status of *Ailichthys punctata* bay [sic, Day] and its relationship with *Ailia coila* (Hamilton) (Siluroidea: Schilbeidae). *Journal of the Zoological Society of India*, 14, for 1962, 244–248.
- Jayaram, K.C. (1965a) On a new species of fish of the family Bagridae. *Bulletin of Systematic Zoology, Calcutta*, 1, 9–10.
- Jayaram, K.C. (1965b) Systematic position of the genus *Liobagrus* Hilgendorf (Pisces: Siluroidea). *Science and Culture*, 31, 436–437.
- Jayaram, K.C. (1966a) Contributions to the study of bagrid fishes (Siluroidea: Bagridae): 5.— On a new species of the genus *Coreobagrus* Mori. *Bulletin of the Biogeographical Society of Japan*, 24, 29–32.
- Jayaram, K.C. (1966b) Contributions to the study of the fishes of the family Bagridae: 2.— A systematic account of the African genera with a new classification of the family. *Bulletin de l'Institut fondamental d'Afrique noire. Série A, Sciences naturelles*, 28, 1064–1139.
- Jayaram, K.C. (1966c) Contributions to the study of bagrid fishes (Siluroidea: Bagridae): 1.— A systematic account of the genera *Rita* Bleeker, *Rama* Bleeker, *Mystus* Scopoli, and *Horabagrus* Jayaram. *Internationale Revue der Gesamten Hydrobiologie*, 51, 433–450.
- Jayaram, K.C. (1966d) A new species of sisorid fish from the Kameng Frontier Division, N.E.F.A. *Journal of the Zoological Society of India*, 15, for 1963, 85–87.
- Jayaram, K.C. (1968a) Contributions to the study of bagrid fishes. 6.— Nomenclatural status and systematic position of the fishes of the subgenus *Osteobagrus* Jayaram (Siluroidea: Bagridae). *Proceedings of the Indian Sci. Congress*, 3, 370.
- Jayaram, K.C. (1968b) Contributions to the study of bagrid fishes (Siluroidea: Bagridae): 3.— A systematic account of the Japanese, Chinese, Malayan and Indonesian genera. *Treubia, Museum Zoologicum Borgoriense*, 27, 287–386.
- Jayaram, K.C. (1971b) Contributions to the study of bagrid fishes: 4.— Nomenclatural status and systematic position of the fishes of the subgenus *Osteobagrus* Jayaram (Siluroidea: Bagridae). *Records of the Zoological Survey of India*, 63, 185–194.
- Jayaram, K.C. (1972a) Siluroid fishes of India, Burma and Ceylon. 18.— Resurrection of the genus *Nangra* Day and its systematic position. *Journal of the Zoological Society of India*, 23, for 1971, 171–174.
- Jayaram, K.C. (1972b) Siluroid fishes of India, Burma and Ceylon. 19.— Fishes of the genus *Ketengus* Bleeker (Family Ariidae). *Proceedings of the Zoological Society of Calcutta*, 25, 135–139.
- Jayaram, K.C. (1973a) Contributions to the study of bagrid fishes (Siluroidea: Bagridae): 10.— Systematic position of *Pimelodus chandramara* Hamilton with description of a new genus. *Internationale Revue der Gesamten Hydrobiologie*, 57, for 1972, 815–820.
- Jayaram, K.C. (1973b) Contributions to the study of bagrid fishes: 9.— Generic status of *Aorichthys* Wu (Siluroidea:

- Bagridae). *Proceedings of the Zoological Society of Calcutta*, 24, for 1971, 149–156.
- Jayaram, K.C. (1973c) Siluroid fishes of India, Burma and Ceylon. 19.— Systematic position of the genus *Laguvia* Hora and its relationships. *Records of the Zoological Survey of India*, 67, 385–389.
- Jayaram, K.C. (1974) Contributions to the study of bagrid fishes: 11.— Designation of a lectotype for *Macrones argenteivittatus* Regan with notes on some bagrid species in the collections of the Natural History Museum, Geneva. *Revue suisse de Zoologie, Annales de la Société zoologique suisse et du Muséum d'Histoire naturelle de Genève*, 81, 785–790.
- Jayaram, K.C. (1976) Contributions to the study of bagrid fishes: 13.— Interrelationships of Indo-African catfishes of the family Bagridae. *Matsya*, 2, 47–53.
- Jayaram, K.C. (1977a) Year of publication and type-locality of *Ompok bimaculatus* (Bloch) (Pisces: Siluroidea). *Newsletter of the Zoological Survey of India*, 3, 50–51.
- Jayaram, K.C. (1977b) Contributions to the study of bagrid fishes: 12.— The correct family position of *Batasio* Blyth (Siluroidea). *Newsletter of the Zoological Survey of India*, 3, 242–245.
- Jayaram, K.C. (1977c) Aid to identification of siluroid fishes of India, Burma, Sri Lanka, Pakistan and Bangladesh: 1.— Bagridae. *Records of the Zoological Survey of India, Miscellaneous Publication, Occasional Paper*, 8, 1–41.
- Jayaram, K.C. (1977d) Aid to identification of siluroid fishes of India, Burma, Sri Lanka, Pakistan and Bangladesh: 2.— Siluridae, Schilbeidae, Pangasiidae, Amblycipitidae, Akysidae. *Records of the Zoological Survey of India, Miscellaneous Publication, Occasional Paper*, 10, 1–33.
- Jayaram, K.C. (1978a) The catfishes of India. *Zoologiana (Calcutta)*, 1, 9–17.
- Jayaram, K.C. (1978b) Contributions to the study of bagrid fishes: 14.— The systematic position of the species of *Mystus* Scopoli known from China. *Proceedings of the Indian Academy of Science*, 87B, 221–228.
- Jayaram, K.C. (1979) Aid to identification of siluroid fishes of India, Burma, Sri Lanka, Pakistan and Bangladesh: 3.— Sisoridae. *Records of the Zoological Survey of India, Miscellaneous Publication, Occasional Paper*, 11, 1–62.
- Jayaram, K.C. (1980) Aid to identification of siluroid fishes of India, Burma, Sri Lanka, Pakistan and Bangladesh: 4.— Clariidae, Heteropneustidae, Chacidae, and Olyridae. *Records of the Zoological Survey of India, Miscellaneous Publication, Occasional Paper*, 23, 1–23.
- Jayaram, K.C. (1981) *The Freshwater Fishes of India, Pakistan, Bangladesh, Burma, and Sri Lanka — a Handbook*, Zoological Survey of India, Calcutta, 475 p., 12 pl.
- Jayaram, K.C. (1982) Aid to identification of siluroid fishes of India, Burma, Sri Lanka, Pakistan and Bangladesh: 5.— Ariidae and Plotosidae. *Records of the Zoological Survey of India, Miscellaneous Publication, Occasional Paper*, 37, 1–41.
- Jayaram, K.C. (1983) Ariidae. *FAO Species Identification Sheets for Western Indian Ocean, Fishing Area 51*, FAO, Rome, [48] p.
- Jayaram, K.C. (1999) *The Freshwater Fishes of the Indian Region*, Narendra Publishing House, Delhi, xxvii + 551 p., 18 pl.
- Jayaram, K.C. & Anuradha, C. (1984) Contributions to the study of bagrid fishes: 17.— The history and usage of the name “*Mystus*”. *Bulletin of the Zoological Survey of India*, 6, 289–293.
- Jayaram, K.C. & Anuradha, S. (2003) A taxonomic revision of the fishes of the genus *Mystus* Scopoli (Family: Bagridae). *Records of the Zoological Survey of India, Miscellaneous Publication, Occasional Paper*, 207, 1–136, pls. 1–5.
- Jayaram, K.C. & Boeseman, M. (1976) The systematic position of the Chinese fish *Macrones sinensis* Bleeker (Siluroidea). *Zoologische Mededelingen (Leiden)*, 50, 117–119, pl. 1.
- Jayaram, K.C. & Dhanze, J.R. (1978) Siluroid fishes of India, Burma, and Ceylon: 21.— A note on the systematic position of *Tachysurus serratus* (Day) (Ariidae). *Bulletin of the Zoological Survey of India*, 1, 203–205.
- Jayaram, K.C. & Dhanze, J.R. (1979) Siluroid fishes of India, Burma and Ceylon: 22.— A preliminary review of the genera of the family Ariidae (Pisces: Siluroidea). *Matsya*, 4, for 1978, 42–51.
- Jayaram, K.C. & Dhanze, J.R. (1981a) Siluroid fishes of India, Burma, and Ceylon: 23.— The specific status of *Tachysurus malabaricus* (Day) (Ariidae: Siluriformes). *Bulletin of the Zoological Survey of India*, 4, 121–123.
- Jayaram, K.C. & Dhanze, J.R. (1981b) Siluroid fishes of India, Burma, and Ceylon 24.— The systematic status of *Arius satparanus* Chaudhuri (Ariidae: Siluriformes). *Bulletin of the Zoological Survey of India*, 4, 395–398.
- Jayaram, K.C. & Dhanze, J.R. (1985) Evolution and biogeography of the Indian genera of the family Ariidae. *Proceedings of the Indian Academy of Sciences (Animal Sciences)*, 95, 279–288.
- Jayaram, K.C. & Majumdar, N. (1964) Siluroid fishes of India, Burma and Ceylon. 16.— Fishes of the genus *Chaca* Gray, 1831. *Proceedings of the Zoological Society of Calcutta*, 17, 177–181.

- Jayaram, K.C. & Singh, R. (1982) Contributions to the study of bagrid fishes: 15.—A comparative account of the cranial musculature in four bagrid genera with a note on their phylogeny. *Records of the Zoological Survey of India*, 80, 231–250.
- Jenkins, J.T. (1910) Notes on fish from India and Persia, with descriptions of new species. *Records of the Indian Museum*, 5, 123–140, pl. 6.
- Jennet, A. (1928) Les poissons fossiles originaux conservés à l'institut de Géologie de l'université de Neuchâtel. *Bulletin de la société Neuchateloise des Sciences Naturelles, nouvelle série*, 1, for 1927, 102–124.
- Jenyns, L. (1840–1842) Fish. In: *The Zoology of the Voyage of H. M. S. Beagle, Under the Command of Captain FitzRoy, R. N., During the years 1832 to 1836 [part 4: xvi + 172 p, 29 pl.]*, Smith, Elder, and Co, Cornhill.
- Jerdon, T.C. (1849) On the fresh-water fishes of southern India. (Continued from p. 149). *Madras Journal of Literature and Science*, 15, 302–346.
- Joannis, L. de. (1835) Observations sur les poissons du Nil, et description de plusieurs espèces nouvelles. *Magasin de zoologie*, 5, [1–53], pls. 1–15. [Each species account with a unique header code that links text to a plate]
- Johnsen, S. (1926) On a collection of fishes from the White Nile and the Yei River. (With notes on the skull of *Alla-benchelys* and *Clarias*). *Bergens Museums Aarbok*, 10, 1–22.
- Jordan, D.S. (1877a) A partial synopsis of the fishes of upper Georgia; with supplementary papers on fishes of Tennessee, Kentucky, and Indiana. *Annals of the Lyceum of Natural History, New York*, 11, 307–377.
- Jordan, D.S. (1877b) Contributions to North American Ichthyology based primarily on the collections of the United States National Museum. I.—Review of Rafinesque's memoirs on North American fishes. *Bulletin of the U. S. National Museum*, 9, 1–53.
- Jordan, D.S. (1877c) Contributions to North American ichthyology based primarily on the collections of the United States National Museum. No. 2. B.—Synopsis of the *Siluridae* of the fresh waters of North America. *Bulletin of the U. S. National Museum*, 10, 69–103, pls. 1–45.
- Jordan, D.S. (1885) A catalogue of the fishes known to inhabit the waters of North America, north of the Tropic of Cancer, with notes on species discovered in 1883 and 1884. *Report of the U. S. Fish Commission*, 13, for 1885, 789–973.
- Jordan, D.S. (1889) Descriptions of fourteen species of fresh-water fishes collected by the U. S. Fish Commission in the summer of 1888. *Proceedings of the United States National Museum*, 11, 351–362, pls. 43–45.
- Jordan, D.S. (1895) The fishes of Sinaloa. *Proceedings of the California Academy of Sciences, Series 2*, 5, 377–514, pls. 26–55.
- Jordan, D.S. (1917) The genera of fishes, from Linnaeus to Cuvier, 1758–1833, seventy-five years, with the accepted type of each. A contribution to the stability of scientific nomenclature. (Assisted by Barton Warren Evermann.). *Leland Stanford Jr. University Publications, University Series*, 27, 1–161.
- Jordan, D.S. (1919a) The genera of fishes, part II, from Agassiz to Bleeker, 1833–1858, twenty-six years, with the accepted type of each. A contribution to the stability of scientific nomenclature. *Leland Stanford Jr. University Publications, University Series*, 36, i–ix, 155–284, i–xiii.
- Jordan, D.S. (1919b) The genera of fishes, part III, from Guenther to Gill, 1859–1880, twenty-two years, with the accepted type of each. A contribution to the stability of scientific nomenclature. *Leland Stanford Jr. University Publications, University Series*, 39, 285–410, i–xv.
- Jordan, D.S. (1920) The genera of fishes, part IV, from 1881 to 1920, thirty-nine years, with the accepted type of each. A contribution to the stability of scientific nomenclature. *Leland Stanford Jr. University Publications, University Series*, 43, 411–576 + i–xviii.
- Jordan, D.S. (1923) A classification of fishes including families and genera as far as known. *Stanford University Publications, University Series, Biological Sciences*, 3, 77–243 + i–x.
- Jordan, D.S. & Evermann, B.W. (1896) The fishes of North and Middle America: a descriptive catalogue of the species of fish-like vertebrates found in the waters of North America, north of the Isthmus of Panama. Part I. *Bulletin of the U. S. National Museum*, 47, i–lx + 1–1240.
- Jordan, D.S. & Evermann, B.W. (1898) The fishes of North and Middle America: a descriptive catalogue of the species of fish-like vertebrates found in the waters of North America north of the Isthmus of Panama. Part III. *Bulletin of the U. S. National Museum*, 47, i–xxiv + 2183a–3136.
- Jordan, D.S. & Evermann, B.W. (1900) The fishes of North and Middle America: a descriptive catalogue of the species of fish-like vertebrates found in the waters of North America, north of the Isthmus of Panama. Part IV. *Bulletin of the U. S. National Museum*, 47, i–ci + 3137–3313, pls. 1–392.
- Jordan, D.S. & Evermann, B.W. (1917) The genera of fishes, from Linnaeus to Cuvier, 1758–1833, seventy-five years, with the accepted type of each. A contribution to the stability of scientific nomenclature. *Leland Stanford Jr. University Publications, University Series*, 27, 1–161.

- sity Publications, University Series, 27, 1–161.
- Jordan, D.S. & Fowler, H.W. (1903) A review of the siluroid fishes or catfishes of Japan. *Proceedings of the United States National Museum*, 26, 897–911.
- Jordan, D.S. & Gilbert, C.H. (1877) On the genera of North American freshwater fishes. *Proceedings of the Academy of Natural Sciences, Philadelphia*, 29, 83–104.
- Jordan, D.S. & Gilbert, C.H. (1882) A review of the siluroid fishes found on the Pacific coast of tropical America, with descriptions of three new species. *Bulletin of the U. S. Fish Commission*, 2, 34–54. [Bulletin issued in dated signatures; much of Vol. 2 issued in 1883, but this article issued in September and October, 1882.]
- Jordan, D.S. & Gilbert, C.H. (1886) List of fishes collected in Arkansas, Indian Territory, and Texas, in September, 1884, with notes and descriptions. *Proceedings of the U. S. National Museum*, 9, 1–25.
- Jordan, D.S. & Hubbs, C.L. (1925) Record of fishes obtained by David Starr Jordan in Japan, 1922. *Memoirs of the Carnegie Museum*, 10, 93–346, pls. 5–12.
- Jordan, D.S. & Snyder, J.O. (1899) Notes on a collection of fishes from the rivers of Mexico, with description of twenty new species. *Bulletin of the U.S. Fish Commission*, 19, 115–147.
- Jubb, R.A. (1965) A new species of *Clariallabes* (Pisces, Clariidae) from the Upper Zambesi River. *Annals and Magazine of Natural History (Ser. 13)*, 7, for 1964, 393–395.
- Jubb, R.A. & Le Roux, P. (1969) Revision of the *Chiloglanis* (Pisces: Mochokidae) of southern Africa and descriptions of two new species. *Annals of the Cape Provincial Museums. Natural History*, 8, 13–23.
- Kailola, P.J. (1983) *Arius graeffei* and *Arius armiger*: Valid names for two common species of Australo-Papuan fork-tailed catfishes (Pisces: Ariidae). *Transactions of the Royal Society of South Australia*, 107, 187–196.
- Kailola, P.J. (1986a) Ariidae systematics: Comparison of the giant sea catfishes *Arius thalassinus* and *A. bilineatus* of the Indo-Pacific. In: Uyeno, T., Arai, R., Taniuchi, T. & Matsuura, K. (Eds.), *Indo-Pacific Fish Biology. Proceedings of the Second International Conference on Indo-Pacific Fishes*, Ichthyological Society of Japan, pp. 540–549.
- Kailola, P.J. (1986b) *Arius peronii*: A distinct species of fork-tailed catfish (Pisces: Ariidae). *Journal of Zoology (London)*, 210, 243–250.
- Kailola, P.J. (1990) A review of the freshwater fork-tailed catfishes (Pisces: Ariidae) of northern New Guinea, with descriptions of two new species. *Records of the Western Australian Museum, Supplement*, 34, 1–30.
- Kailola, P.J. (1999) Ariidae. In: Carpenter, K.E. & Niem, V.H. (Eds.), *FAO Species Identification Guides for Fishery Purposes: The Living Marine Resources of the Western Central Pacific. Vol. 3: Batoid fishes, chimeras and bony fishes, part 1 (Elopidae to Linophrynidae)*, FAO, Rome, pp. 1827–1879.
- Kailola, P.J. (2000) Six new species of fork-tailed catfishes (Pisces, Teleostei, Ariidae) from Australia and New Guinea. *The Beagle, Records of the Museums and Art Galleries of the Northern Territory*, 16, 127–144.
- Kailola, P.J. (2004) A phylogenetic exploration of the catfish family Ariidae (Otophysi; Siluriformes). *The Beagle, Records of the Museums and Art Galleries of the Northern Territory*, 20, 87–166.
- Kailola, P.J. & Bussing, W.A. (1995) Ariidae. In: Fischer, W., Krupp, F., Schneider, W., Sommer, C., Carpenter, K.E. & Niem, V.H. (Eds.), *Guía FAO para la Identificación para los Fines de la Pesca. Pacífico Centro-oriental*. Vol. II. Vertebrados — Parte 1, FAO, Rome, pp. 860–886.
- Kailola, P.J. & Pierce, B.E. (1988) A new freshwater catfish (Pisces: Ariidae) from northern Australia. *Records of the Western Australian Museum*, 14, 73–89.
- Kanazawa, R.H. (1958) A new species of catfish, family Loricariidae, from Ecuador. *Copeia*, 1958, 327–328.
- Kastenberger, G. (1977) Der trommelapparat der Doradidae (Siluriformes, Pisces). *Zoologische Jahrbücher. Abteilung für Allgemeine Zoologie und Physiologie der Tiere*, 81, 281–309.
- Keilhack, L. (1908) Bemerkungen zur Fishfauna des nördlichen Njassa-Gebietes: einige neue Arten aus den Gattungen *Barbus* und *Synodontis* und Beiträge zur Systematik der Gattung *Clarias*. *Sitzungsberichte der Gesellschaft Naturforschender Freunde zu Berlin*, 1908, 164–169.
- Keilhack, L. (1910) Über einige von Herrn Dr. H. Monke in Duala (Kamerun) gesammelte Fische. *Mitteilungen aus dem Zoologischen Museum in Berlin*, 5, 117–124.
- Keith, P. & Merona, B. de (2000) Présence de *Lithodoras dorsalis* (Valenciennes in Cuvier et Valenciennes, 1840) (Siluriformes, Doradidae) en Guyane française. *Cybium (3e série)*, 24, 101–103.
- Kessler, K.T. (1872) [Ichthyological Fauna of Turkestan]. *Izvestia Imperatorskago Obshchestva Liubitelei Estestvoznanija, Antropologii i Etnografii*, 10, 47–76, pls. 6–12. [In Russian.]
- Khin-Thant. (1966) *Silurus burmanensis*, a new species of fish from the Inlé Lake, Southern Shan State, Burma. *Journal of the Burma Research Society*, 49, 219–221, pls. 1–3.
- Kimura, S. (1934) Description of the fishes collected from the Yangtze-kiang, China, by the late Dr. K. Kishinouye and

- his party in 1927–1929. *Journal of the Shanghai Science Institute. Section 3, Systematic and morphological biology and pharmacognosy*, 1, 11–247, pls. 1–6.
- Kindle, E.M. (1895) The South American cat-fishes belonging to Cornell University. *Annals of the New York Academy of Sciences*, 7, for 1894, 249–256.
- Klausewitz, W. & Rössel, F. (1961) *Rhynchodoras xingui*, ein bemerkenswerter neuer Wels aus Brasilien (Pisces, Siluroidea, Doradidae). *Senckenbergiana Biologica*, 42, 45–48.
- Klein, J.T. (1775) *Neuer Schauplatz der Natur, nach den Richtigsten Beobachtungen und Versuchen, in Alphabetischer Ordnung, Vorgestellt durch eine Gesellschaft von Gelehrten*, Weidmann, Leipzig, xiv + 1044 p.
- Kleynhans, C.J. & James, N.P.E. (1995) Threatened fishes of the world: *Chiloglanis bifurcus* Jubb & le Roux, 1969 (Mochokidae). *Environmental Biology of Fishes*, 43, 120.
- Klunzinger, C.B. (1880) Die von Müller'sche Sammlung australischer Fische in Stuttgart. *Sitzungsberichte der Kaiserlichen Akademie der Wissenschaften, Mathematisch- Naturwissenschaftlichen Classe, Wien, Abt. 1., Botanik, Zoologie, Anatomie, Geologie und Paläontologie*, 80, 325–430, pls. 1–9.
- Knaack, J. (1961) Ein neuer Panzerwels aus Brasilien (*Corydoras guapore*) (Pisces, Teleostei, Callichthyidae). *Senckenbergiana Biologica*, 42, 135–138.
- Knaack, J. (1962) Zwei neue Panzerwelse, *Corydoras haraldschultzi* und *Corydoras sterbai* (Pisces, Teleostei, Callichthyidae). *Senckenbergiana Biologica*, 43, 129–135, pls. 11–12.
- Knaack, J. (1966a) Ein Zergpanzerwels — *Corydoras pygmaeus*. *Die Aquarien- und Terrarien- Zeitschrift*, 19, 168–169.
- Knaack, J. (1966b) Ein Zergpanzerwels aus Brasilien: *Corydoras pygmaeus*. Neubeschreibung (Pisces, Teleostei, Callichthyidae). *Aquarien-Terrar. Mschr. Ornith. Vivar*, 13, 364–365.
- Knaack, J. (1999a) A new species of bristlemouth catfish of the genus *Ancistrus* Kner, 1854 from the Mato Grosso, Brazil (Pisces, Siluriformes, Loricariidae). *Tropical Fish Hobbyist*, 47, 70–76.
- Knaack, J. (1999b) A new species of suckermouth catfish (*Hypostomus* Lacépède, 1803) from the Mato Grosso, Brazil (Pisces, Siluriformes, Loricariidae). *Tropical Fish Hobbyist*, 47, 102–108.
- Knaack, J. (1999c) Erstbeschreibung *Corydoras spectabilis* n. sp. *VDA-aktuell*, 1999, 74–79.
- Knaack, J. (1999d) New *Ancistrus* species from the Rio Cuiba System, Brazil (Pisces, Siluriformes, Loricariidae). *Tropical Fish Hobbyist*, 47, 150–155.
- Knaack, J. (1999e) Eine weitere neue Art der Gattung *Corydoras* Lacépède, 1803 aus dem Mato Grosso (Pisces, Siluriformes, Callichthyidae). *Aquaristik Aktuell*, 2000, 74–79. [Actually published in Dec. 1999.]
- Knaack, J. (2000a) Ein weiterer neuer Harnischwels aus dem Rio Guaporé: *Lasiancistrus guapore* n. sp. *Aquaristik Aktuell*, 2000, 56–61.
- Knaack, J. (2000b) Eine weitere neue Art der Gattung *Corydoras* La Cepède, 1803 aus dem Mato Grosso (Pisces, Siluriformes, Callichthyidae). *VDA-aktuell*, 2000, 45–56.
- Knaack, J. (2001) Ein weiterer neuer Panzerwels aus Bolivien: *Corydoras pantanalensis* n. sp. (Pisces, Siluriformes, Callichthyidae). *Aquaristik Aktuell*, 2001, 58–63.
- Knaack, J. (2002a) *Corydoras bilineatus* n. sp. (Pisces, Siluriformes, Callichthyidae). *Aquaristik Aktuell*, 10, 50–56.
- Knaack, J. (2002b) Ein neuer Prachthexenwels aus Paraguay: *Hemiloricaria aurata* n. sp. (Pisces, Siluriformes, Loricariidae). *Aquaristik aktuell*, 11, for 2003, 56–61.
- Knaack, J. (2002c) Ein weiterer neuer Panzerwels aus Bolivien: *Corydoras cruziensis* n. sp. (Pisces, Siluriformes, Callichthyidae). *VDA-aktuell*, 2002, 60–69.
- Knaack, J. (2002d) Ein weiterer neuer Panzerwels aus Bolivien: *Corydoras mamore* n. sp. (Pisces, Siluriformes, Callichthyidae). *VDA-aktuell*, 2003, 16–24.
- Knaack, J. (2004) Beschreibung von sechs neuen Arten der Gattung *Corydoras* La Cepède, 1803 (Teleostei: Siluriformes: Callichthyidae). *Zoologische Abhandlungen; Staatliches Museums für Tierkunde in Dresden*, 54, 55–105.
- Kner, R. (1853a) Die Panzerwelse des K.K. Hof-naturalien-Cabinetes zu Wien. I Abtheilung: Loricarinae. *Denkschriften der Mathematisch- Naturwissenschaftlichen Classe der Kaiserlichen Akademie der Wissenschaften in Wien*, 6, for 1854, 1–34, pls. 1–8. [separate issued before volume]
- Kner, R. (1853b) Über die Hypostomiden, oder die zweite Hauptgruppe der Panzerfische. *Sitzungsberichte der Kaiserlichen Akademie der Wissenschaften, Mathematisch- Naturwissenschaftlichen Classe, Wien*, 10, 278–282.
- Kner, R. (1853c) Ueber einige Sexual-Unterschiede bei der Gattung *Callichthys* und die Schwimmblase bei *Doras* C. Val. *Sitzungsberichte der Kaiserlichen Akademie der Wissenschaften, Mathematisch- Naturwissenschaftlichen Classe, Wien*, 11, 138–146, 1 pl.
- Kner, R. (1854) Die Hypostomiden. Zweite Hauptgruppe der Familie der Panzerfische. (Loricata vel Goniodontes). *Denkschriften der Mathematisch- Naturwissenschaftlichen Classe der Kaiserlichen Akademie der Wissenschaften in Wien*, 12, 1–10, 1 pl.

- kschriften der Mathematisch- Naturwissenschaftlichen Classe der Kaiserlichen Akademie der Wissenschaften in Wien*, 7, 251–286, pls. 1–5.
- Kner, R. (1855a) Ichthyologische Beiträge. *Sitzungsberichte der Kaiserlichen Akademie der Wissenschaften, Mathematisch- Naturwissenschaftlichen Classe, Wien*, 17, 92–162, pls. 1–6.
- Kner, R. (1855b) Über ein neues genus aus der familie der Welse, Siluroidei. *Sitzungsberichte der Kaiserlichen Akademie der Wissenschaften, Mathematisch- Naturwissenschaftlichen Classe, Wien*, 17, 313–316, pls. 1–2.
- Kner, R. (1857) Ichthyologische Beiträge. II Abtheilung. *Sitzungsberichte der Kaiserlichen Akademie der Wissenschaften, Mathematisch- Naturwissenschaftlichen Classe, Wien*, 26, 373–448, pls. 1–9.
- Kner, R. (1858) Kritische Bemerkungen über Castelnau's Siluroiden. *Archiv für Naturgeschichte*, 24, 344–350.
- Kner, R. (1859) Zur Familie der Characinen. III.— Folge der Ichthyologischen Beiträge. *Denkschriften der Mathematisch- Naturwissenschaftlichen Classe der Kaiserlichen Akademie der Wissenschaften in Wien*, 17, 137–182, pls. 1–9.
- Kner, R. (1863) Eine Uebersicht der ichthyologischen Ausbeute des Herrn Professors Dr. Mor. Wagner in Central-Amerika. *Sitzungsberichte der Königl. Bayerischen Akademie der Wissenschaften, München*, 2, 220–230.
- Kner, R. (1865) Fische. In: *Reise der Österreichischen Fregatte "Novara" um die Erde in den Jahren 1857–59, Unter den Befehlen des Commodore B. von Wüllerstorf-Urbain. Zoologischer Theil*, Wien, pp. 1–109, pls. 1–5.
- Kner, R. (1866) Specielles Verzeichniss der während der Reise der kaiserlichen Fregatte "Novara" gesammelten Fische. III., und Schlussabtheilung. *Sitzungsberichte der Kaiserlichen Akademie der Wissenschaften, Mathematisch- Naturwissenschaftlichen Classe, Wien, Abt. I, Botanik, Zoologie, Anatomie, Geologie und Paläontologie*, 53, 543–550.
- Kner, R. (1867) Fische [continued]. In: *Reise der Österreichischen Fregatte "Novara" um die Erde in den Jahren 1857–59, Unter den Befehlen des Commodore B. von Wüllerstorf-Urbain. Zoologischer Theil*, Wien, pp. 275–433, pls. 12–16.
- Kner, R. & Steindachner, F. (1864) Neue Gattungen und Arten von Fischen aus Central-Amerika; gesammelt von Prof. Moritz Wagner. *Abhandlungen der Mathematisch-physikalischen Classe der Königlich Bayerischen Akademie der Wissenschaften*, 10, 1–61, pls. 1–6.
- Kner, R. & Steindachner, F. (1867) Neue Fische aus dem Museum der Herren Joh. Ces. Godeffroy & Sohn in Hamburg. *Sitzungsberichte der Kaiserlichen Akademie der Wissenschaften, Mathematisch- Naturwissenschaftlichen Classe, Wien, Abth. I, Botanik, Zoologie, Anatomie, Geologie und Paläontologie*, 54, 356–395, pls. 1–5. [Separate issued with different pagination.]
- Kobayakawa, M. (1989) Systematic revision of the catfish genus *Silurus*, with description of a new species from Thailand and Burma. *Japanese Journal of Ichthyology*, 36, 155–186.
- Kobayakawa, M. & Okuyama, S. (1994) Fossils of *Silurus biwaensis* (Siluridae) from the Ueno Formation, ancient Lake Biwa, Japan. *Japanese Journal of Ichthyology*, 40, 500–503.
- Koch, W.R. (2002) Revisão taxonômica do gênero *Homodiaetus* (Teleostei, Siluriformes, Trichomycteridae). *Iheringia, Série Zoologia*, 92, 33–46.
- Koch, W.R. & Reis, R.E. (1996) *Tatia boemia*, a new species of authenipterid catfish (Teleostei: Siluriformes) from the rio Uruguay drainage, southern Brazil. *Ichthyological Exploration of Freshwaters*, 7, 85–90.
- Kochetov. (1998) [A new species of *Synodontis*]. *Aquarium*, 2, 44–45. [In Russian; not seen]
- Koken, E. (1884) Ueber Fish-Otolithen, insbesondere über diejenigen der nord-deutschen Oligocän-Ablagerungen. *Zeitschrift der Deutschen Geologischen Gesellschaft*, 36, 500–565, pls. 9–12.
- Koken, E. (1885) Otolithen. In: A.V. Koenen, Über eine palaeözane Fauna von Kopenhagen. *Abhandlungen der Königlichen Gesellschaft der Wissenschaften zu Göttingen. Physikalische Klasse*, 32, 113–116.
- Koken, E. (1888) Neue Untersuchungen am tertiären Fisch-Otolithen, I. *Zeitschrift der Deutschen Geologischen Gesellschaft*, 40, 274–305, pls. 17–19.
- Koken, E. (1891) Neue Untersuchungen am tertiären Fisch-Otolithen, II. *Zeitschrift der Deutschen Geologischen Gesellschaft*, 43, 77–170, pls. 1–10.
- Koller, O. (1926) Einige neue Fischformen von der Insel Hainan. *Anzeiger der Mathematisch- Naturwissenschaftlichen Classe der Kaiserlichen Akademie der Wissenschaften in Wien*, 63, 74–77.
- Koller, O. (1927) Fische von der Insel Hai-nan. *Annalen des Naturhistorischen Museums in Wien*, 41, 25–49, pl. 1.
- König, C.D.E. (1825) *Icones. Fossilium Sectiles. Centuria Prima*, Fol, London, 4 p., 19 pls.
- Kosygin, L. & Vishwanath, W. (2005) Validity and redescription of *Glyptothorax manipurensis* Menon and record of *Glyptothorax sinense* (Regan) from India. *Journal of the Bombay Natural History Society*, 102, 61–65.
- Kottelat, M. (1983) A new species of *Erethistes* Müller & Troschel from Thailand and Burma (Osteichthyes: Siluriformes: Sisoridae). *Hydrobiologia*, 107, 71–74.

- Kottelat, M. (1984a) Catalogue des types du Musée d'Historie Naturelle de Neuchâtel. I.—Pisces. *Bulletin de la Societe Neuchâteloise des Sciences Naturelles*, 107, 143–153.
- Kottelat, M. (1984b) A review of the species of Indochinese fresh-water fishes described by H. E. Sauvage. *Bulletin du Muséum National d'Histoire Naturelle (série 4, section A)*, 6, 791–822.
- Kottelat, M. (1985) Fresh-water fishes of Kampuchea. A provisory annotated check-list. *Hydrobiologia*, 121, 249–279.
- Kottelat, M. (1988) Authorship, dates of publication, status and types of Spix and Agassiz's Brazilian fishes. *Spixiana*, 11, 69–93.
- Kottelat, M. (1989a) Zoogeography of the fishes from Indochinese inland waters with an annotated check-list. *Bulletin Zoologisch Museum, Universiteit van Amsterdam*, 12, 1–56.
- Kottelat, M. (1989b) On the validity of *Phractocephalus* Agassiz, 1829, vs. *Pirarara* Agassiz, 1829 (Osteichthyes: Pimelodidae). *Spixiana*, 12, 321.
- Kottelat, M. (2000a) Diagnoses of a new genus and 64 new species of fishes from Laos (Teleostei: Cyprinidae, Balitoridae, Bagridae, Syngnathidae, Chaudhuriidae, and Tetraodontidae). *Journal of South Asian Natural History*, 5, 37–82.
- Kottelat, M. (2000b) Notes on taxonomy, nomenclature and distribution of some fishes of Laos. *Journal of South Asian Natural History*, 5, 83–90.
- Kottelat, M. (2001a) *Freshwater Fishes of Northern Vietnam. A Preliminary Check-list of the Fishes Known or Expected to Occur in Northern Vietnam with Comments on Systematics and Nomenclature*, The World Bank, Environment and Social Development Unit, East Asia and Pacific Region, Washington, D.C., iii + 123 + 18 p., 15 color pls.
- Kottelat, M. (2001b) *Fishes of Laos*, WHT Publications, Colombo, Sri Lanka, 198 p.
- Kottelat, M. & Lim, K.K.P. (1995) *Hemibagrus hoevenii*, a valid species of Sundaic catfish (Teleostei: Bagridae). *Malayan Nature Journal*, 49, 41–47.
- Kottelat, M., Lim, K.K.P. & Ng, P.K.L. (1994) *Bagrus hoevenii* Bleeker, 1846 (currently *Hemibagrus hoevenii*; Osteichthyes, Siluriformes): proposed designation of a neotype. *Bulletin of Zoological Nomenclature*, 51, 320–322.
- Kottelat, M. & Ng, H.H. (1999) *Belodontichthys truncatus*, a new species of silurid catfish from Indochina (Teleostei: Siluridae). *Ichthyological Exploration of Freshwaters*, 10, 387–391.
- Kottelat, M., Ng, H.H. & Ng, P.K.L. (1998) Notes on the identity of *Hemibagrus elongatus* (Günther, 1864) and other east Asian species allied to *H. guttatus* (La Cepède, 1803) (Teleostei, Bagridae). *The Raffles Bulletin of Zoology*, 46, 565–572.
- Kottelat, M. & Sutter, E. (1988) Catalogue des types de poissons du Musée d'histoire naturelle de Bâle (Naturhistorisches Museum Basel). *Verhandlungen der Naturforschenden Gesellschaft in Basil*, 98, 51–57.
- Kottelat, M., Whitten, A.J., Kartikasari, S.N. & Wirjoatmodjo, S. (1993) *Freshwater Fishes of Western Indonesia and Sulawesi*, Periplus Editions, Hong Kong, 259 p., 84 pl.
- Kowarsky, J. (1976) Clarification of the name and distribution of the plotsid catfish *Cnidoglanis macrocephalus*. *Copeia*, 1976, 593–594.
- Kulkarni, C. (1952) A new genus of schilbeid catfishes from the Deccan. *Records of the Indian Museum*, 49, 231–238.
- Kullander, S.O., Fang, F., Delling, B. & Åhlander, E. (1999) The fishes of the Kashmir Valley. In: Nyman, L. (Ed.), *River Jhelum, Kashmir Valley: Impacts of the Aquatic Environment*, SWEDMAR, pp. 99–167.
- La Cepède, B.G.E. (1803) *Histoire Naturelle des Poissons*, Vol. 5, Plassan, Paris, lxviii + 803 p. + index, 21 pls.
- La Monte, F. (1929) Two new fishes from Mt. Duida, Venezuela. *American Museum Novitates*, 373, 1–4.
- La Monte, F. (1933a) A new subgenus of *Plecostomus* from Brazil. *American Museum Novitates*, 591, 1–2.
- La Monte, F. (1933b) *Pimelodus valenciennis* Kr. the type of a new genus. *Copeia*, 1933, 226.
- La Monte, F. (1935) Fishes from Rio Jurua and Rio Purus, Brazilian Amazonas. *American Museum Novitates*, 784, 1–8.
- La Monte, F. (1939) *Tridentopsis tocantinsi*, a new pygidiid fish from Brazil. *American Museum Novitates*, 1024, 1–2.
- La Monte, F. (1941) A new *Corydoras* from Brazil. *Zoologica (New York)*, 26, 5–6.
- Lacerda, M.T.C. & Evers, H.-G. (1996) Parotocinclus-Arten aus Brasilien. *Die Aquarien- und Terrarien- Zeitschrift*, 49, 88–95.
- Ladiges, W. & Voelker, J. (1961) Untersuchungen über die Fischfauna in Gebirgsgewässern des Wasserscheidenhochlands in Angola. *Mitteilungen aus dem Hamburgischen Zoologischen Museum und Institut*, 59, 117–140, pls. 3–7.
- Ladiges, W., Wahlert, G. von & Mohr, E. (1958) Die Typen und Typoide der Fischsammlung des Hamburgischen Zoologischen Staatsinstituts und Zoologischen Museums. *Mitteilungen aus dem Zoologischen Museum in Berlin*, 56, 155–167.
- Lambert, J.G. (1958) Poissons Siluriformes et Cyprinodontiformes récoltés en Guinée Française, avec la description d'une nouvelle espèce de *Microsynodontis*. *Revue de Zoologie et de Botanique Africaines*, 57, 39–56.

- Landim, M.I. & Costa, W.J.E.M. (2002) *Listrura tetraradiata* (Siluriformes: Trichomycteridae): a new glanapterygine catfish from the southeastern Brazilian coastal plains. *Copeia*, 2002, 152–156.
- Langeani, F. (1990) Revisão do gênero *Neoplectostomus*, com a descrição de quatro espécies novas do sudeste brasileiro (Ostariophysi, Siluriformes, Loricariidae). *Comunicações do Museu de Ciências da PUCRS, Série Zoologia, Porto Alegre*, 3, 3–31.
- Langeani, F. & Araujo, R.B. (1994) O gênero *Rineloricaria* Bleeker, 1862 (Ostariophysi, Siluriformes) na bacia do Rio Paraná superior: *Rineloricaria pentamaculata* sp. n. e *Rineloricaria latirostris* (Boulenger, 1900). *Comunicações do Museu de Ciências da PUCRS, Série Zoologia, Porto Alegre*, 7, 151–166.
- Langeani, F., Oyakawa, O.T. & Montoya-Burgos, J.I. (2001) New species of *Harttia* (Loricariidae, Loricariinae) from the Rio São Francisco basin. *Copeia*, 2001, 136–142.
- Larrañaga, D.A. (1923) *Escritos de Don Dámaso Antonio Larrañaga. Los Publica el Instituto Histórico y Geográfico del Uruguay*, Edición Nacional, 512 p.
- Lasso, C.A. (1990) Los peces de la Gran Sabana, Alto Caroni, Venezuela. *Memoria, Sociedad de Ciencias Naturales La Salle*, 49/50, 209–285.
- Lasso, C.A. & Provenzano, F. (1998) *Chaetostoma vasquezi*, nueva especie de corroncho del Escudo de Guayana, Estado Bolívar, Venezuela (Siluroidei: Loricariidae): descripción y consideraciones biogeográficas. *Memoria, Sociedad de Ciencias Naturales La Salle*, 57, for 1997, 53–65.
- Lasso, C.A. & Provenzano, F. (2003) Dos nuevas especies de bagres del género *Trichomycterus* (Siluriformes: Trichomycteridae) de la Gran Sabana, Escudo de las Guayanás, Venezuela. *Revista de Biología Tropical*, 50, 1139–1149.
- Latrelle, P.A. (1825) *Familles Naturelles du Règne Animal Exposées Succinctement et dans un Ordre Analytique, avec l'Indication de leurs Genres*, 2^e ed., Paris, 570 p.
- Le Bail, P.-Y., Keith, P. & Planquette, P. (2000) *Atlas des Poissons d'Eau Douce de Guyane. Tome 2, Fascicule II, Siluriformes*, Publications scientifiques du MNHN, Service du Patrimoine Naturel, Paris, 307 p.
- Le Grande, W.H. (1981) Chromosomal evolution in North American catfishes (Siluriformes: Ictaluridae) with particular emphasis on the madtoms. *Copeia*, 1981, 33–52.
- Lee, C.L. & Kim, I.S. (1990) A taxonomic revision of the family Bagridae (Pisces: Siluriformes) from Korea. *Korean Journal of Ichthyology*, 2, 117–137.
- Leege, C.O. (1922) Der Rumpfpanzer der Panzerwelse und seine Skelettbeziehungen (*Plecostomus angipinnatus* n. sp., *Callichthys callichthys* L. und *Corydoras paleatus* Jen.). *Jenaische Zeitschrift für Naturwissenschaft*, 58, 145–270, pls. 9–10.
- Lehmann, A., P. & Reis, R.E. (2004) *Callichthys serralabium*: a new species of neotropical catfish from the upper Orinoco and Negro rivers (Siluriformes: Callichthyidae). *Copeia*, 2004, 336–343.
- Leidenfrost, G. (1925) Die fossilen Siluriden-Ungarns. *Mitteilungen aus dem Jahrbuch der Kgl. Ungarischen Geologischen Anstalt*, 24, 115–123, pls. 4–7.
- Leidy, J. (1873a) Notice of remains of fishes in the Bridger Tertiary Formation of Wyoming. *Proceedings of the Academy of Natural Sciences, Philadelphia*, 1873, 97–99.
- Leidy, J. (1873b) Contributions to the extinct vertebrate fauna of the western territories. *Annual Report, United States Geological and Geographic Survey of the Territories*, 1, for 1867, 1–358, 38 pl.
- Leidy, J. (1889) Notice and description of fossils in caves and crevices of the limestone rocks of Pennsylvania. *Annual Report, Geological Survey of Pennsylvania*, for 1887, 1–20.
- Lelek, A. (1987) *The Freshwater Fishes of Europe. Vol. 9: Threatened Fishes of Europe*, AULA-Verlag, Wiesbaden, 343 p.
- Leriche, M. (1900) Faune ichthyologique des Sables à unios et térédines des environs d'Épernay (Marne). *Annales de la Société Géologique du Nord*, 29, 173–200.
- Leriche, M. (1901) Contribution à l'étude des Siluridés fossiles. *Annales de la Société Géologique du Nord*, 30, 165–175.
- Lesson, R.P. (1826–1831) Poissons. In: Duperrey, L.I. (Ed.), *Voyage Autour du Monde, ..., sur la Corvette de La Majesté La Coquille, Pendant les Années 1822, 1823, 1824 et 1825...*, Vol. 5: *Zoologie, Tome 2, Part 1*, A. Bertrand, Paris, pp. 66–238, atlas: pls. 1–38. [Atlas to Vol. 5 apparently issued in 1826, text in 1831. See AMNH online catalog for details.]
- Lesueur, C.A. (1819) Notice de quelques poissons découverts dans les lacs du Haut-Canada, durant l'été de 1816. *Mémoires du Muséum d'Histoire Naturelle*, 5, 148–161, 2 pls.
- Lévêque, C., Paugy, D. & Teugels, G.G. (Eds.) (1992) *Faune des Poissons d'Eaux Douces et Saumâtres de l'Afrique de l'Ouest*. Vol. 2, Musée Royal de l'Afrique Central, Tervuren, and Editions de l'ORSTOM, Paris.
- Lévêque, C., Paugy, D., Teugels, G.G. & Romand, R. (1989) Inventaire taxonomique et distribution des poissons d'eau

- douce des bassins côtiers de Guinée et de Guinée Bissau. *Revue d'Hydrobiologie Tropicale*, 22, 107–127.
- Leviton, A.E., Gibbs, J., R. H., Heal, E. & Dawson, C.E. (1985) Standards in herpetology and ichthyology: Part I.—Standard symbolic codes for institutional resource collections in herpetology and ichthyology. *Copeia*, 1985, 802–832.
- Leybold, F. (1859) Descripción de una nueva especie de pez, descubierto por don Federico Leybold en el Río Seco de los baños de Colina. *Anales de la Universidad de Chile*, 16, 1083–1085.
- Li, G. & Wang, J. (1979) [The taxonomy and description of Early Tertiary fossil fishes from Sanshui Basin and the adjacent districts of Guangdong]. *Zhongshandaxue Xuebao, Zi Ran ke xue ban [=Acta Scientiarum Naturalium Universitatis Sunyatsenii]*, 1979, 41–50. [In Chinese, with title and very brief abstract in English.]
- Li, J., Chen, X. & Chan, B.P.L. (2005) A new species of *Pseudobagrus* (Teleostei: Siluriformes: Bagridae) from southern China. *Zootaxa*, 1067, 49–57.
- Li, S.-S. (1984a) [A study of infraspecies classification about *Glyptothorax fukiensis* (Rendahl) (comb. nov.)]. *Journal of Yunnan University*, 1984, 63–72. [In Chinese, with English summary]
- Li, S.-S. (1984b) [A study of the classification of the striped chest sisorids (*Glyptothorax* Blyth) in China]. *Journal of Yunnan University*, 1984, 75–89. [In Chinese, with English summary]
- Li, S.-S. (1986) Systematics, distribution and evolution of *Glyptothorax* (Siluriformes: Sisoridae). In: Uyeno, T., Arai, R., Taniuchi, T. & Matsuura, K. (Eds.), *Indo-Pacific Fish Biology: Proceedings of the Second International Conference on Indo-Pacific Fishes*, Ichthyological Society of Japan, pp. 521–528.
- Lichtenstein, M.H.C. (1819) Ueber einige neue Arten von Fishen aus der Gattung *Silurus*. *Zoologisches Magazin (Wiedemann)*, 1, 57–63.
- Lichtenstein, M.H.C. (1823) *Verzeichniss der Doubletten des Zoologischen Museums der Königl. Universität zu Berlin, nebst Beschreibung Vieler Bisher Unbekannter Arten von Säugethieren, Vögeln, Amphibien und Fischen*, In Commission bei T. Trautwein, Berlin, x + 118 p., pl. 1.
- Lilyestrom, C.G. (1984) Consideraciones sobre la taxonomía de las especies del género *Cochliodon* Heckel en Venezuela (Pisces, Loricariidae). *Revista UNELLEZ de Ciencia y Tecnología*, 2, 41–53.
- Lim, K.K.P. & Ng, H.H. (1999) *Clarias batu*, a new species of catfish (Teleostei: Clariidae) from Pulau Tioman, Peninsular Malaysia. *The Raffles Bulletin of Zoology, Supplement*, 6, 157–167.
- Lima, F.C.T. & Britto, M.R. (2001) New catfish of the genus *Aspidoras* (Siluriformes: Callichthyidae) from the upper Rio Paraguai system in Brazil. *Copeia*, 2001, 1010–1016.
- Lima, S.M.Q. & Costa, W.J.E.M. (2004) *Trichomycterus giganteus* (Siluriformes: Loricarioidea: Trichomycteridae): A new catfish from the Rio Guandu basin, southeastern Brazil. *Zootaxa*, 761, 1–6.
- Lin, S.-Y. (1934) Three new fresh-water fishes of Kwangtung Province. *Lingnan Science Journal*, 13, 225–230.
- Lin, Y.-H. (2003) [A new species of the genus *Glyptothorax* Blyth from Guangdong, China (Siluriformes, Sisoridae)]. *Acta Zootaxonomica Sinica*, 28, 159–162. [In Chinese, with English abstract]
- Linck, H.F. (1790) Versuch einer Eintheilung der Fische nach den Zähnen. *Magazin für das Neueste aus der Physik und Naturgeschichte, Gotha*, 6, 28–38.
- Linder, R.S. & Ng, H.H. (2002) Anmerkungen zu den Stachelwelsen der Gattung *Bagrichthys* (Teleostei, Bagridae). *BSSW Report*, 4, 4–9.
- Linnaeus, C. (1754) *Hans Maj:ts Adolf Frideriks vår Allernädigste Konungs Naturalie Samling Innehållande Sällsynta och Främmande Djur, som Bevaras på Kongl. Lust-slottet Ulriksdahl Beskrefne och Afritade Samt på Nädig Befallning Utgifne af Carl Linnaeus*, Holmiae, xxx + 96 + 7 p.
- Linnaeus, C. (1758) *Systema Naturae Sive Regna Tria Naturae, Secundum Classes, Ordines, Genera, Species, cum Characteribus, Differentiis, Synonymis, Locis. Tomus I. Editio decima, reformata*, Holmiae, ii + 824 p.
- Linnaeus, C. (1764) *Museum S:ae R:ae M:tis Adolphi Friderici Regis Svecorum, Gothorum, Vandalorumque &c. &c. &c. in quo Animalia Rariora Imprimis & Exotica: Aves, Amphibia, Piscis Describuntur. Tomi Secundi Prodromus*, Holmiae, 110 p.
- Linnaeus, C. (1766) *Systema Naturae Sive Regna Tria Naturae, Secundum Classes, Ordines, Genera, Species, cum Characteribus, Differentiis, Synonymis, Locis. 12th ed. Vol. 1, pt. 1*, Laurentii Salvii, Holmiae, 532 p.
- Littmann, M.W. (1998) *Revision of the South American Shovelnose Catfishes of the Genus Sorubim (Siluriformes: Pimelodidae), with Descriptions of Two New Species*. Unpublished M.Sc. thesis, Southern Illinois University, Carbondale.
- Littmann, M.W., Burr, B.M. & Buitrago-Suarez, U.A. (2001) A new cryptic species of *Sorubim* Cuvier (Siluriformes: Pimelodidae) from the upper and middle Amazon basin. *Proceedings of the Academy of Natural Sciences, Philadelphia*, 151, 87–93.
- Littmann, M.W., Burr, B.M. & Nass, P. (2000) *Sorubim cuspicaudus*, a new long-whiskered catfish from northwestern

- South America (Siluriformes: Pimelodidae). *Proceedings of the Biological Society of Washington*, 113, 900–917.
- Littmann, M.W., Burr, B.M., Schmidt, R.E. & Isern, E.R. (2001) *Sorubim elongatus*, a new species of catfish (Siluriformes: Pimelodidae) from tropical South America syntopic with *S. lima*. *Ichthyological Exploration of Freshwaters*, 12, 1–16.
- Liu, C.-X., Peng, Z.-G. & He, S.-P. (2005) Studies in species classification for genus *Cranoglanis* Peters with the method of morphometrics. *Acta Hydrobiologia Sinica*, 29, 507–512. [In Chinese, with English abstract]
- Liu, H.T. & Su, T.T. (1962) [Pliocene fishes from Yushe Basin, Shansi]. *Vertebrate Palasiatica (Beijing)*, 6, 1–47. [In Chinese, with English summary.]
- Lohberger, K. (1930) Fische aus dem Wamifluss. *Zoologischer Anzeiger*, 89, 88–94.
- Lönnberg, E. (1895) Notes on fishes collected in the Cameroons by Mr. Y. Sjöstedt. *Öfversigt af Kongl. Vetenskapsakademiens Forhandlingar*, 1895, 179–195.
- Lönnberg, E. (1924) Two new silurids from lower Congo. *Annals and Magazine of Natural History (Ser. 9)*, 14, 619–621.
- Lönnberg, E. & Rendahl, H. (1920) On some freshwater fishes from Lower Congo. *Annals and Magazine of Natural History (Ser. 9)*, 6, 167–176.
- Lönnberg, E. & Rendahl, H. (1922) Some new silurids from the Congo. *Annals and Magazine of Natural History (Ser. 9)*, 10, 122–127.
- Lönnberg, E. & Rendahl, H. (1930) Eine neue Art der Gattung *Corydoras*. *Arkiv för Zoologi*, 22 A, 1–6.
- López, H.L. & Miquelarena, A.M. (1991a) Los Hypostominae (Pisces: Loricariidae) de Argentina. *Fauna de Agua Dulce de la República Argentina*, 40, Pisces, 1–64.
- López, H.L. & Miquelarena, A.M. (1991b) Peces loricaridos de la Cuenca de Plata, Argentina. Parte I.— El genero *Cochliodon* Heckel, 1854 (Pisces: Siluriformes). *Gayana Zoologica*, 55, 3–11.
- Lopez-Fernandez, H. & Winemiller, K.O. (2000) A review of Venezuelan species of *Hypophthalmus* (Siluriformes: Pimelodidae). *Ichthyological Exploration of Freshwaters*, 11, 35–46.
- Loubens, G. & Panfili, J. (2000) Biologie de *Pseudoplatystoma fasciatum* et *P. tigrinum* (Teleostei: Pimelodidae) dans le bassin du Mamoré (Amazonie Bolivienne). *Ichthyological Exploration of Freshwaters*, 11, 13–34.
- Lowe-McConnell, R.H. (1964) The fishes of the Rupununi savanna district of British Guiana, South America. Part I.— Ecological groupings of fish species and effects of the seasonal cycle on the fish. *The Journal of the Linnean Society of London. Zoology*, 45, 103–144.
- Lucena, C.A.S. de & da Silva, J.F.P. (1991) Descrição de uma nova espécie do gênero *Rhamdella* Eigenmann & Eigenmann, 1888 (Siluriformes: Pimelodidae) para o médio rio Uruguai, sul do Brasil. *Comunicações do Museu de Ciências da PUCRS, Série Zoologia, Porto Alegre*, 4, 28–47.
- Lucena, C.A.S. de, Malabarba, L.R. & Reis, R.E. (1992) Resurrection of the Neotropical pimelodid catfish *Parapimelodus nigribarbis* (Boulenger), with a phylogenetic diagnosis of the genus *Parapimelodus* (Teleostei: Siluriformes). *Copeia*, 1992, 138–146.
- Lucena, Z.M.S. & Lucena, C.A.S. de. (1990) Sobre a localidade-tipo das espécies de peixes descritas por Steindachner (1907). *Comunicações do Museu de Ciências da PUCRS, Série Zoologia, Porto Alegre*, 3, 99–101.
- Lundberg, J.G. (1975) The fossil catfishes of North America. *Claude W. Hibbard Memorial, Volume 2. University of Michigan, Papers on Paleontology*, 11, i–iv, 1–51.
- Lundberg, J.G. (1982) The comparative anatomy of the toothless blindcat, *Troglagnathus pattersoni* Eigenmann, with a phylogenetic analysis of the ictalurid catfishes. *Miscellaneous Publications, Museum of Zoology, University of Michigan*, 163, 1–85.
- Lundberg, J.G. (1992) The phylogeny of ictalurid catfishes: A synthesis of recent work. In: Mayden, R.L. (Ed.), *Systematics, Historical Ecology, & North American Freshwater Fishes*, Stanford Univ. Press, Stanford, CA, pp. 392–420.
- Lundberg, J.G. (1997) Fishes of the La Venta fauna: additional taxa, biotic and paleoenvironmental implications, In: Kay, R.F., et al. (Eds.), *Vertebrate Paleontology in the Neotropics: The Miocene Fauna of La Venta, Colombia*, Smithsonian Institution Press, Washington, pp. 67–91.
- Lundberg, J.G. (1998) The temporal context for diversification of Neotropical fishes. In: Malabarba, L.R., Reis, R.E., Vari, R.P., Lucena, Z.M.S. & Lucena, C.A.S. (Eds.), *Phylogeny and Classification of Neotropical Fishes*, Edipucrs, Porto Alegre, pp. 49–68.
- Lundberg, J.G. (2005) *Brachyplatystoma promagdalena* n. sp., a fossil goliath catfish (Siluriformes: Pimelodidae) from the Miocene of Colombia, South America. *Neotropical Ichthyology*, 3, 597–605.
- Lundberg, J.G. & Aguilera, O. (2003) The late Miocene *Phractocephalus* catfish (Siluriformes: Pimelodidae) from Urumaco, Venezuela: additional specimens and reinterpretation as a distinct species. *Neotropical Ichthyology*, 1, 97–109.

- Lundberg, J.G. & Akama, A. (2005) *Brachyplatystoma capapretum*: a new species of goliath catfish from the Amazon basin, with a reclassification of allied catfishes (Siluriformes: Pimelodidae). *Copeia*, 2005, 492–516.
- Lundberg, J.G. & Baskin, J.N. (1969) The caudal skeleton of the catfishes, order Siluriformes. *American Museum Novitates*, 2398, 1–49.
- Lundberg, J.G., Bornbusch, A.H. & Mago-Leccia, F. (1991) *Gladioglanis conquistador* n. sp. from Ecuador with diagnoses of the subfamilies Rhamdiinae Bleeker and Pseudopimelodinae n. subf. (Siluriformes: Pimelodidae). *Copeia*, 1991, 190–209.
- Lundberg, J.G. & Case, G.R. (1970) A new catfish from the Eocene Green River Formation, Wyoming. *Journal of Paleontology*, 44, 451–457.
- Lundberg, J.G., Linares, O., Nass, P. & Antonio, M.E. (1988) *Phractocephalus hemiolopterus* (Pimelodidae: Siluriformes) from the Late Miocene Urumaco formation, Venezuela: A further case of evolutionary stasis and local extinction among South American fishes. *Journal of Vertebrate Paleontology*, 8, 131–138.
- Lundberg, J.G. & Littmann, M.W. (2003) Pimelodidae. In: Reis, R.E., Kullander, S.O. & Ferraris, C.J., Jr. (Eds.), *Check list of the Freshwater Fishes of South and Central America*, Edipucrs, Porto Alegre, Brazil, pp. 432–446.
- Lundberg, J.G., Mago-Leccia, F. & Nass, P. (1991) *Exallodontus aguanai*, a new genus and species of Pimelodidae (Pisces: Siluriformes) from deep river channels of South America, and delimitation of the subfamily Pimelodinae. *Proceedings of the Biological Society of Washington*, 104, 840–869.
- Lundberg, J.G. & McDade, L.A. (1986) On the South American catfish *Brachyrhamdia imitator* Myers (Siluriformes: Pimelodidae), with phylogenetic evidence for a large intrafamilial lineage. *Notulae Naturae (Philadelphia)*, 463, 1–24.
- Lundberg, J.G., Nass, P. & Mago-Leccia, F. (1989) *Pteroglanis manni* Eigenmann and Pearson, a juvenile of *Sorubimichthys planiceps* (Agassiz), with a review of the nominal species of *Sorubimichthys* (Pisces: Pimelodidae). *Copeia*, 1989,
- Lundberg, J.G. & Parisi, B.M. (2002) *Propimelodus*, new genus, and a description of *Pimelodus eigenmanni* van der Stigchel, 1946, a long recognized yet poorly-known South American catfish (Pimelodidae: Siluriformes). *Proceedings of the Academy of Natural Sciences, Philadelphia*, 152, 75–88.
- Lundberg, J.G. & Rapp Py-Daniel, L. (1994) *Bathycetopsis oliveirai*, gen. et sp. nov., a blind and depigmented catfish (Siluriformes: Cetopsidae) from the Brazilian Amazon. *Copeia*, 1994, 381–390.
- Lütken, C.F. (1874a) Ichthyographiske bidrag. I.— Nogle nye eller mindre fuldstændigt kjendte Pandsermaller, især fra det nordlige Sydamerica. *Videnskabelige Meddelelser fra den Naturhistorisk Forening i Kjøbenhavn*, 13/14, for 1873, 202–220, pl. 4.
- Lütken, C.F. (1874b) Ichthyographiske Bidrag. II.— Nye eller mindre vel kjendte Malleformer fra forskjellige Verdensdele. *Videnskabelige Meddelelser fra den Naturhistorisk Forening i Kjøbenhavn*, for 1874, 190–220.
- Lütken, C.F. (1874c) Siluridae novae Brasiliæ centralis a clarissimo J. Reinhardt in provincia Minas-geræs circa oppidulum Lagoa Santa, præcipue in flumine Rio das Velhas et affluentibus collectae, secundum characteres essentiales, breviter descriptæ. *Oversigt over Selskabets verksamhed, Kongelige Danske Videnskabernes selskab.*, Kjøbenhavn, 1873, 29–36.
- Lütken, C.F. (1875) Velhas-Flodens Fiske. Et Bidrag til Brasiliens Ichthyologi; efter Professor J. Reinhardts Indsamlinger og Optegnelser. *Det Kongelige Danske Videnskabernes Selskabs Skrifter, Raekke 5*, 12, 121–253, + 2 unnum., + i–xxi, pls. 1–5.
- Lütken, C.F. (1892) Om en med stegophiler og tricomycterer beslaegtet sydamerikansk mallefisk (*Acanthopoma annectens* Ltk. n. g. & sp.?). *Videnskabelige Meddelelser fra den Naturhistorisk Foreningi Kjøbenhavn*, for 1891, 53–60.
- Lydekker, R. (1886) Indian Tertiary and post-Tertiary vertebrata: Tertiary Fishes. *Memoirs of the Geological Society of India, Palaeontology Indica, Series 10*, 3, 241–258, pls. 35–37.
- Lynn, W.G. & Melland, A.M. (1939) A fossil catfish (*Felichthys stauroforus*) from the Maryland Miocene. *Journal of the Washington Academy of Sciences*, 29, 14–20.
- MacDonagh, E.J. (1931) Notas zoológicas de una excursión entre Patagones y San Blas. *Notas Preliminares del Museo de La Plata*, 1, 63–86.
- MacDonagh, E.J. (1937) Sobre el Manguruyú (gênero *Paulicea*, Silurideos). *Revista del Museo de La Plata (Nueva serie)*, 1, 3–30.
- MacDonagh, E.J. (1938) Contribución a la sistemática y etología de los peces fluviatiles Argentinos. *Revista del Museo de La Plata, Sección Zoología*, 1, 119–208, pls. 1–5.
- Mack. (1861) *Trav. Ussuri* (= *Puteschestvie po dolinge ryeki Usuri*), 1, 195–198. [Not seen]
- Macleay, W. (1881) A descriptive catalogue of Australian fishes.— Part IV. *Proceedings of the Linnean Society of New*

- South Wales*, 6, 202–387.
- Macleay, W. (1883) Contribution to a knowledge of the fishes of New Guinea.— No. 4. *Proceedings of the Linnean Society of New South Wales*, 8, 252–280.
- Magalhães, A.C. (1931) *Monographia Brazileira de Peixes Fluviaes*. Graphicars, São Paulo, 255 p.
- Mago-Leccia, F. (1984) *Entomocorus gameroi*, una nueva especie de bagre auqueniptero (Teleostei, Siluriformes) de Venezuela, incluyendo la descripción de su dimorfismo sexual secundario. *Acta Biologica Venezolana*, 11, 215–236.
- Mai, D.Y. (1978) [*Identification of the Fresh-water Fishes of North Viet Nam*], Scientific & Technology Publisher, Ha Noi, 340 p., 48 pl. [In Vietnamese. Selected parts translated into English in Kottelat (2001:74 et seq.).]
- Malabarba, L.R. & Mahler, J.K.F., Jr. (1998) Review of the genus *Microglanis* in the rio Uruguay and coastal drainages of southern Brazil (Ostariophysi: Pimelodidae). *Ichthyological Exploration of Freshwaters*, 9, 243–254.
- Maldonado-Ocampo, J.A., Ortega-Lara, A., Usma Oveido, J.S., Galvis Vergara, G., Villa-Navarro, F.A., Vásquez Gamboa, L., Prada-Pedreros, S. & Ardila-Rodríguez, C.A. (2005) *Peces de los Andes de Colombia: Guía de Campo*. Instituto de Investigación de Recursos Biológicos Alexander von Humboldt, Bogotá, Colombia, 346 pp.
- Manimekalan, A. & Arunachalam, M. (2002) Rediscovery of critically endangered air breathing cat fish *Clarias dayi* Hora (Pisces: Claridae) from Mudumalai Wildlife Sanctuary, Tamil Nadu. *Journal of the Bombay Natural History Society*, 99, 129–131.
- Manimekalan, A. & Das, H.S. (1998) *Glyptothorax davissinghi* (Pisces: Sisoridae) a new cat fish from Nilambur in the Nilgiri Biosphere Reserve, South India. *Journal of the Bombay Natural History Society*, 95, 87–91.
- Marceniuk, A.P. (2003) *Relações Filogenéticas e Revisão dos Gêneros da Família Ariidae (Ostariophysi, Siluriformes)*. Ph. D. Dissertation, Universidade de São Paulo, São Paulo.
- Marceniuk, A.P. (2005a) Chave para identificação das espécies de bagres marinhos (Siluriformes, Ariidae) da costa Brasileira. *Boletim do Instituto do Pesca, São Paulo*, 31, 89–101.
- Marceniuk, A.P. (2005b) Redescrição de *Genidens barbus* (Lacépède, 1803) e *Genidens machadoi* (Miranda-Ribeiro, 1918), bagres marinhos (Siluriformes, Ariidae) do Atlântico sul ocidental. *Papéis Avulsos de Zoologia*, 45, 111–125.
- Marceniuk, A.P. & Ferraris, C.J., Jr. (2003) Ariidae. In: Reis, R.E., Kullander, S.O. & Ferraris, C.J., Jr. (Eds.), *Check list of the Freshwater Fishes of South and Central America*, Edipucrs, Porto Alegre, Brazil, pp. 447–455.
- Marcgravius, G. (1648) *Historia Naturalis Brasiliæ, Auspicio et Beneficio Illustriss. I. Mauritii Com. Nassau Illius Provinciae et Maris Summi Praefecti Adornata in qua non Tantum Plantæ et Animalia, sed et Indigenarum Morbi, Ingenia et Mores Describuntur et Iconibus Supra Quingentas Illustrantur*, Lugduni Batavorum & Amstelodami, 300 p.
- Marck, W. von der. (1876a) Fossile fische von Sumatra. In: H.B. Geinitz & W. von der Marck, Zur Geologie von Sumatra. *Palaeontographica*, 22, 405–414, 2 pl. [Also: 1878, Jaarb. Mijnw. Ned. Oost-Indië, 7, 138–155]
- Marck, W. von der. (1876b) Fossile fische von Sumatra. *Mitteilungen aus dem Königlichen Mineralogisch-Geologischen und Prähistorischen Museum in Dresden*, 2, 7–16, 2 pls.
- Marck, W. von der & Schlüter, C. (1868) Neue Fische und Krebse aus der Kreide von Westphalen. *Palaeontographica*, 15, 269–305, pls. 41–44.
- Marini, T.L., Nichols, J.T. & La Monte, F.R. (1933) Six new eastern South American fishes examined in the American Museum of Natural History. *American Museum Novitates*, 618, 1–7.
- Markle, D.F. (1997) Audubon's hoax: Ohio River fishes described by Rafinesque. *Archives of Natural History*, 24, 439–447.
- Marschall, A. (1873) *Nomenclator Zoologicus. Continens Nomina Systematica Generum Animalium tam Viventium quam Fossilium, Secundum Ordinem Alphabeticum Disposita*, Vindobonae, iv + 482 p.
- Martín Salazar, F.J. (1964) Las especies del género *Farlowella* de Venezuela (Pisces — Nematognathi — Loricariidae) Con descripción de 5 especies y 1 sub-especies nuevas. *Memoria, Sociedad de Ciencias Naturales La Salle*, 24, 242–261.
- Martín Salazar, F.J., Isbrücker, I.J.H. & Nijssen, H. (1982) *Dentectus barbarmatus*, a new genus and species of mailed catfish from the Orinoco Basin of Venezuela (Pisces, Siluriformes, Loricariidae). *Beaufortia*, 32, 125–137.
- Matthes, H. (1959a) Poissons nouveaux du Ruanda. *Folia Scientifica Africæ Centralis*, 5, 62.
- Matthes, H. (1959b) Poissons nouveaux du Lac Tanganika. Descriptions préliminaires. *Folia Scientifica Africæ Centralis*, 5, 77–78.
- Matthes, H. (1962) Poissons nouveaux ou intéressants du lac Tanganyika et du Ruanda. *Annales du Musée Royal de l'Afrique Centrale, Series in 8°, Sciences Zoologiques*, 111, 27–88, pls. 1–4.
- Matthes, H. (1964) Les poissons du lac Tumba et de la région d'Ikela. *Étude systématique et écologique. Annales du Musée Royal de l'Afrique Centrale, Series in 8°, Sciences Zoologiques*, 126, 1–204, map, table, pls. 1–6.

- Matthes, H. (1965) Quelques poissons nouveaux du Congo. *Revue de Zoologie et de Botanique Africaines*, 71, 177–193.
- Mazzoni, R., Caramaschi, U. & Weber, C. (1994) Taxonomical revision of the species of *Hypostomus* Lacépède, 1803 (Siluriformes, Loricariidae) from the lower rio Paraiba do Sul, state of Rio de Janeiro, Brazil. *Revue suisse de Zoologie, Annales de la Société zoologique suisse et du Muséum d'Histoire naturelle de Genève*, 101, 3–18.
- Mbega, J.D. & Teugels, G.G. (1998) Révision systématique du genre *Anaspidoglanis* (Siluriformes: Claroteidae) de l'Afrique Centrale. *Cybium (3e série)*, 22, 223–236.
- M'Clelland, J. (1842) On the fresh-water fishes collected by William Griffith, Esq., F. L. S. Madras Medical Service, during his travels under the orders of the Supreme Government of India, from 1835 to 1842. *Calcutta Journal of Natural History, and Miscellany of the Arts and Sciences in India*, 2, 560–589, pls. 6, 15, 18, 20, 21.
- M'Clelland, J. (1844a) Description of a collection of fishes made at Chusan and Ningpo in China, by Dr. G. R. Playfair, Surgeon of the Phlegethon, war steamer, during the late military operations in that country. *Calcutta Journal of Natural History, and Miscellany of the Arts and Sciences in India*, 4, for 1843, 390–413, pls. 21–25.
- M'Clelland, J. (1844b) Apodal fishes of Bengal. *Calcutta Journal of Natural History, and Miscellany of the Arts and Sciences in India*, 5, 151–226, pls. 5–14.
- Meek, S.E. (1902) A contribution to the ichthyology of Mexico. *Field Columbian Museum, Zoological Series*, 3, 63–128, pls. 14–31.
- Meek, S.E. (1904) The fresh-water fishes of Mexico north of the isthmus of Tehuantepec. *Field Columbian Museum, Zoological Series*, 5, i–xiii + 1–252, pls. 1–17.
- Meek, S.E. (1905) Two new species of fishes from Brazil. *Proceedings of the Biological Society of Washington*, 18, 241–242.
- Meek, S.E. (1906) Description of three new species of fishes from Middle America. *Field Columbian Museum, Zoological Series*, 7, 91–95.
- Meek, S.E. (1907a) Synopsis of the fishes of the great lakes of Nicaragua. *Field Columbian Museum, Zoological Series*, 7, 97–132.
- Meek, S.E. (1907b) Notes on fresh-water fishes from Mexico and Central America. *Field Columbian Museum, Zoological Series*, 7, 133–157.
- Meek, S.E. (1909) New species of fishes from tropical America. *Field Columbian Museum, Zoological Series*, 7, 207–211.
- Meek, S.E. & Hildebrand, S.F. (1913) New species of fishes from Panama. *Publication. Zoological Series, Field Museum of Natural History*, 10, 77–91.
- Meek, S.E. & Hildebrand, S.F. (1916) The fishes of the fresh waters of Panama. *Publication. Zoological Series, Field Museum of Natural History*, 10, 217–374, pls. 6–32.
- Meek, S.E. & Hildebrand, S.F. (1923) The marine fishes of Panama.— Part I. *Publication. Zoological Series, Field Museum of Natural History*, 15, i–xi + 1–330, pls. 1–24.
- Mees, G.F. (1967) Freshwater fishes of Suriname: the genus *Heptapterus* (Pimelodidae). *Zoologische Mededelingen (Leiden)*, 42, 215–229.
- Mees, G.F. (1974) The Auchenipteridae and Pimelodidae of Suriname (Pisces, Nematognathi). *Zoologische Verhandelingen (Leiden)*, 132, 1–256, pls. 1–15.
- Mees, G.F. (1978a) On the identity of *Arius oncinus* R. H. Schomburgk (Pisces, Nematognathi, Auchenipteridae). *Zoologische Mededelingen (Leiden)*, 52, 267–276.
- Mees, G.F. (1978b) Two new species of Pimelodidae from northwestern South America (Pisces, Nematognathi). *Zoologische Mededelingen (Leiden)*, 53, 253–261, pls. 1–3.
- Mees, G.F. (1983) Naked catfishes from French Guiana (Pisces, Nematognathi). *Zoologische Mededelingen (Leiden)*, 57, 43–58.
- Mees, G.F. (1984) A note on the genus *Tocantinsia* (Pisces, Nematognathi, Auchenipteridae). *Amazoniana*, 9, 31–34.
- Mees, G.F. (1985) Further records of Auchenipteridae and Pimelodidae from Suriname (Pisces: Nematognathi). *Zoologische Mededelingen (Leiden)*, 59, 239–249.
- Mees, G.F. (1986) Records of Auchenipteridae and Pimelodidae from French Guiana (Pisces, Nematognathi). *Proceedings of the Koninklijke Nederlandse Akademie van Wetenschappen (Series C)*, 89, 311–325.
- Mees, G.F. (1987a) A new species of *Heptapterus* from Venezuela (Pisces, Nematognathi, Pimelodidae). *Proceedings of the Koninklijke Nederlandse Akademie van Wetenschappen (Series C)*, 90, 451–456.
- Mees, G.F. (1987b) The members of the subfamily Aspredininae, family Aspredinidae in Suriname (Pisces, Nematognathi). *Proceedings of the Koninklijke Nederlandse Akademie van Wetenschappen (Series C)*, 90, 173–192.
- Mees, G.F. (1988a) Notes on the genus *Tatia* (Pisces, Nematognathi, Auchenipteridae). *Proceedings of the Koninklijke*

- Nederlandse Akademie van Wetenschappen (Series C), 91, 405–414.
- Mees, G.F. (1988b) The genera of the subfamily Bunocephalinae (Pisces, Nematognathi, Aspredinidae). *Proceedings of the Koninklijke Nederlandse Akademie van Wetenschappen (Series C)*, 91, 85–102.
- Mees, G.F. (1989) Notes on the genus *Dysichthys*, subfamily Bunocephalidae, family Aspredinidae (Pisces, Nematognathi). *Proceedings of the Koninklijke Nederlandse Akademie van Wetenschappen (Series C)*, 92, 189–250.
- Mees, G.F. (1997) The type species of the genera *Bunocephalus* Kner, 1855, and *Bunocephalichthys* Bleeker, 1858 (Pisces, Aspredinidae, Bunocephalinae). *Proceedings of the Koninklijke Nederlandse Akademie van Wetenschappen (Series C)*, 99, 225–228.
- Mees, G.F. & Cala, P. (1989) Two new species of *Imparfinis* from northern South America (Pisces, Nematognathi, Pimelodidae). *Proceedings of the Koninklijke Nederlandse Akademie van Wetenschappen (Series C)*, 92, 379–394.
- Meinken, H. (1937) Beiträge zur Fischfauna des mittleren Paraná — III. *Blätter für Aquarien- und Terrarien-Kunde*, 48, 73–80.
- Meinken, H. (1957) Mitteilungen der Fischbestimmungsstelle des VDA. XXIV.— Über zwei der Liebhaberei bislang unbekannte *Corydoras*-Neuheiten (Callichthyidae — Ostariophysi). *Die Aquarien- und Terrarien-Zeitschrift*, 10, 4–7.
- Menon, A.G.K. (1950) On a remarkable blind siluroid fish of the family Clariidae from Kerala (India). *Records of the Indian Museum*, 48, 59–66, pl. 1.
- Menon, A.G.K. (1951) On certain features in the anatomy of *Horaglanis* Menon. *Journal of the Zoological Society of India*, 3, 249–253.
- Menon, A.G.K. (1955) Further observations on the fish fauna of the Manipur State. *Records of the Indian Museum*, 52, for 1954, 21–26.
- Menon, A.G.K. (1999) Check list, freshwater fishes of India. *Records of the Zoological Survey of India, Miscellaneous Publication, Occasional Paper*, 175, i–xxviii + 1–366.
- Menon, A.G.K. & Rama Rao, K.V. (1974) On the dates of the parts of Day's 'Fishes of India'. *Journal for the Society for the Bibliography of Natural History*, 7, 143.
- Menon, A.G.K. & Yazdani, G.M. (1968) Catalogue of type-specimens in the Zoological Survey of India, Part 2.— Fishes. *Records of the Zoological Survey of India*, 61, for 1963, 91–190.
- Menon, M.A.S. (1955) Notes on fishes of the genus *Glyptothorax* Blyth. *Records of the Indian Museum*, 52, for 1954, 27–54.
- Meuschen, F.C. (1778) *Museum Gronovianum. Sive, Index Rerum Naturalium, tam Mammalium, Amphibiorum, Pis- cium, Insectorum, . . .*, T. Haak, J. Meerburg, Lugundi Batavorum.
- Meyen, F.J.F. (1834–1835) *Reise um die Erde Ausgeführt auf dem Königlich Preussischen Seehandlungsschiffe Prinzess Louise, Commandirt von Captain W. Wendt, in den Jahren 1830, 1831 und 1832; Historischer Bericht*, Sander'sche Buchhandlung, Berlin.
- Miao, C.P. (1934) Notes on the fresh-water fishes of the southern part of Kiangsu, I.— Chinkiang. *Contributions from the Biological Laboratory of the Science Society of China, Zoological Series*, 10, 111–244.
- Miles, C. (1942) Descripción sistemática del "pez graso" del Lago de Tota (Boyacá). *Caldasia*, 5, 55–58.
- Miles, C. (1943a) *Estudio Económico y Ecológico de Los Peces de Agua Dulce del Valle de Cauca*, Departamento del Valle de Cauca, Cali, Colombia, 97 p.
- Miles, C. (1943b) On three recently described species and a new genus of pygidiid fishes from Colombia. *Revista de la Academia Colombiana de Ciencias Exactas, Fisicas y Naturales*, 5, 367–369.
- Miles, C. (1945) Some newly recorded fishes from the Magdalena River system. *Caldasia*, 3, 453–464.
- Miles, C. (1947) *Los Peces del Río Magdalena*, Ministerio de la Economía Nacional; sección de piscicultura, pesca y caza, Bogotá, 214 p., appendices.
- Miller, G.S., Jr. (1905) Notes on the generic names *Pteronotus* and *Dermonotus*. *Proceedings of the Biological Society of Washington*, 18, 223.
- Miller, R.R. (1984) *Rhamdia reddelli*, new species, the first blind pimelodid catfish from Middle America, with a key to the Mexican species. *Transactions of the San Diego Society of Natural History*, 20, 135–144.
- Miller, R.R., Minckley, W.L. & Norris, S.M. (2005) *Freshwater Fishes of México*, University of Chicago Press, Chicago, xxv + 490 p., 99 pl.
- Miller, R.R. & Smith, G.R. (1967) New fossil fishes from Plio-Pleistocene Lake Idaho. *Occasional Papers of the Museum of Zoology, University of Michigan*, 654, 1–24.
- Minding, J. (1832) *Lehrbuch der Naturgeschichte der Fische*, Berlin, xii + 132 p., 6 pls.
- Miquelarena, A.M. & Menni, R.C. (1999) *Rhamdella aymarae*, a new species from the Itiyuro River, northern Argentina

- (Siluriformes: Pimelodidae). *Ichthyological Exploration of Freshwaters*, 10, 201–210.
- Miranda Ribeiro, A. (1906) Vertebrados do Itatiaya (Peixes, Serpentes, Saurios, Aves e Mammiferos). Resultados de excursões do Sr. Carlos Moreira, Assistente de Secção de Zoologia do Museu Nacional. *Arquivos do Museu Nacional do Rio de Janeiro*, 13, for 1905, 165–190, pls. 1–3.
- Miranda Ribeiro, A. (1907a) Peixes do Iporanga — S. Paulo. Resultados de excursões do Sr. Ricardo Krone, membro correspondente do Museu Nacional do Rio de Janeiro. *Boletim Sociedade Nacional de Agricultura, Rio de Janeiro [Lavoura]*, 11, 185–190.
- Miranda Ribeiro, A. (1907b) Uma novidade ichthyologica.— *Typhlobagrus kronei*. *Kosmos, Rio de Janeiro*, 4, [1–3].
- Miranda Ribeiro, A. (1908a) On fishes from the Iporanga River. S. Paulo-Brasil. *Arkiv för Zoologi*, 4, for 1907, 1–5, pl. 1.
- Miranda Ribeiro, A. (1908b) Peixes da Ribeira. Resultados de excursão do Sr. Ricardo Krone, membro correspondente do Museu Nacional do Rio de Janeiro. *Kosmos, Rio de Janeiro*, 5, [1–5].
- Miranda Ribeiro, A. (1911) Fauna brasiliense. Peixes. Tomo IV (A) [Eleutherobranchios Aspirophoros]. *Arquivos do Museu Nacional do Rio de Janeiro*, 16, 1–504, pls. 22–54.
- Miranda Ribeiro, A. (1912) *Loricariidae, Callichthyidae, Doradidae e Trichomycteridae*. In: *Comissão de Linhas Telegraphicas Estrategicas de Matto-Grosso ao Amazonas, Matto-Grosso, Annexo no. 5 (Historia Natural: Zoologia)*. 1–31 p., 1 pl.
- Miranda Ribeiro, A. (1914) *Pimelodidae, Trachycorystidae, Cetopsidae, Bunocephalidae, Auchenipteridae, e Hypophthalmidae*. In: *Comissão de Linhas Telegraphicas Estrategicas de Matto-Grosso ao Amazonas, Matto-Grosso, Annexo no. 5 (Historia Natural: Zoologia)*. 1–13 p., pls. 1–2.
- Miranda Ribeiro, A. (1917) De scleracanthis.— Fluvio "Solinões" anno MCMVIII a cl. F. Machado da Silva duce brasiliense inventis et in Museu Urbis "Rio de Janeiro" servatis. *Revista da Sociedade Brasileira de Ciências*, 1, 49–52.
- Miranda Ribeiro, A. (1918a) *Hemipsilichthys*, Eignm. & Eignm., e generos alliedos. *Revista da Sociedade Brasileira de Ciências*, 2, 101–107, pls. 1–7.
- Miranda Ribeiro, A. (1918b) Nova chave para a determinação das espécies do gênero *Tachysurus*. *Revista da Sociedade Brasileira de Ciências*, 2, 108–111.
- Miranda Ribeiro, A. (1918c) *Ancistrus*. *Revista da Sociedade Brasileira de Ciências*, 2, 112–114.
- Miranda Ribeiro, A. (1918d) Tres generos e dezesete especies novas de peixes Brasilieros determinados nas colleções do Museu Paulista. *Revista do Museu Paulista*, 10, 631–646, 1 pl.
- Miranda Ribeiro, A. (1918e) Lista dos peixes Brasilieros do Museu Paulista, Primeira parte and Terceira parte. *Revista do Museu Paulista*, 10, 705–736, 759–783.
- Miranda Ribeiro, A. (1918f) Considerações sôbre o gênero *Brachyplatystoma* e *Platystomatichthys* de Bleeker. *Revista do Museu Paulista*, 10, 247–283.
- Miranda Ribeiro, A. (1920) Peixes (excl. Characinidae). *Comissão de Linhas Telegraphicas Estrategicas de Matto-Grosso ao Amazonas, Annexo no. 5 (Historia Natural: Zoologia)*, 58, 1–15, 17 unnum. pls.
- Miranda Ribeiro, A. (1924) Ainda "*Hemipsilichthys*" e generos alliedos. *Boletim do Museu Nacional do Rio de Janeiro, Nova Série, zoologia*, 1, 365–366.
- Miranda Ribeiro, A. (1937) Sobre uma colleção de vertebrados do nordeste brasileiro. Primeira parte: Peixes e batrachios. *O Campo, Rio de Janeiro*, 1, 54–56.
- Miranda Ribeiro, A. (1939) Alguns novos dados ictiológicos da nossa fauna. *Boletim Biológico, São Paulo (Nova Série)*, 4, 358–363.
- Miranda Ribeiro, P. (1939a) Sobre o gênero *Harttia*, Steind. (Peixes: Loricariidae). *Boletim Biológico, São Paulo (Nova Série)*, 4, 11–13, pl. 2.
- Miranda Ribeiro, P. (1939b) Um *Paraotocinclus* do Nordeste Brasileiro (Peixes — Larocaridae [sic, Loricariidae] — Hypoptopomatinae). *Boletim Biológico, São Paulo (Nova Série)*, 4, 364–365.
- Miranda Ribeiro, P. (1940) Alguns peixes do sul de Mato Grosso. *O Campo, Rio de Janeiro*, 60, 1.
- Miranda Ribeiro, P. (1942) Um novo "*Corydoras*" do Rio Javari, Amazonas, Brasil (Pisces, Callichthyidae). *Revista Brasileira de Biologia*, 2, 427–428.
- Miranda Ribeiro, P. (1943) Dois novos pigidídeos Brasileiros (Pisces — Pygidiidae). *Boletim do Museu Nacional do Rio de Janeiro, Nova Série, Zoologia*, 9, 1–3.
- Miranda Ribeiro, P. (1944a) Uma nova espécie para o gênero *Bunocephalus* Kner, 1855 (Pisces — Aspredinidae). *Boletim do Museu Nacional do Rio de Janeiro, Nova Série, Zoologia*, 13, 1–3.
- Miranda Ribeiro, P. (1944b) Um pigidídeo do Alto Amazonas (Pisces — Pygidiidae). *Boletim do Museu Nacional do Rio*

- de Janeiro, Nova Série, Zoologia*, 19, 1–3.
- Miranda Ribeiro, P. (1944c) Nova espécie para o gênero *Stegophilus* Reinhardt, 1858 (Pisces — Pygidiidae — Stegophilinae). *Boletim do Museu Nacional do Rio de Janeiro, Nova Série, Zoologia*, 20, 1–3.
- Miranda Ribeiro, P. (1946) Notas para o estudo dos Pygidiidae Brasileiros (Pisces — Pygidiidae — Stegophilinae).— I. *Boletim do Museu Nacional do Rio de Janeiro, Nova Série, Zoologia*, 58, 1–20, foldout table, pls. 1–7.
- Miranda Ribeiro, P. (1947) Notas para o estudo dos Pygidiidae Brasileiros (Pisces — Pygidiidae — Vandelliinae).— II. *Boletim do Museu Nacional do Rio de Janeiro, Nova Série, Zoologia*, 78, 1–8, pls. 1–2.
- Miranda Ribeiro, P. (1949a) Duas novas espécies de peixes na coleção ictiológica do Museu Nacional (Pisces, Callichthyidae et Pygidiidae). *Revista Brasileira de Biologia*, 9, 143–145.
- Miranda Ribeiro, P. (1949b) Notas para o estudo dos Pygidiidae Brasileiros (Pisces — Pygidiidae — Pygidiinae).— III. *Boletim do Museu Nacional do Rio de Janeiro, Nova Série, Zoologia*, 88, 1–3, 2 pls.
- Miranda Ribeiro, P. (1951a) Sobre *Oxyropsis* Eigenmann & Eigenmann, 1889 (Pisces — Nematognathi — Loricariidae). *Boletim do Museu Nacional do Rio de Janeiro, Nova Série, Zoologia*, 104, 1–3, pls. 1–4.
- Miranda Ribeiro, P. (1951b) Notas para o estudo dos Pygidiidae Brasileiros (Pisces — Pygidiidae — Stegophilinae).— IV. *Boletim do Museu Nacional do Rio de Janeiro, Nova Série, Zoologia*, 106, 1–16, table, pls. 1–3.
- Miranda Ribeiro, P. (1951c) Reedição de algumas publicações de Alípio de Miranda-Ribeiro. *Arquivos do Museu Nacional do Rio de Janeiro*, 42, xxxvii–lxxx.
- Miranda Ribeiro, P. (1953) Tipos das espécies e subespécies do Prof. Alípio de Miranda-Ribeiro depositados no Museu Nacional. *Arquivos do Museu Nacional do Rio de Janeiro*, 42, 389–417.
- Miranda Ribeiro, P. (1954) Catálogo dos Peixes do Museu Nacional. I.— Pygidiidae Eigenmann & Eigenmann, 1888. *Publicações Avulsas do Museu Nacional (Rio de Janeiro)*, 15, 1–17.
- Miranda Ribeiro, P. (1956) Notas para o estudo dos Pygidiidae Brasileiros (Pisces — Pygidiidae).— V. *Boletim do Museu Nacional do Rio de Janeiro, Nova Série, Zoologia*, 142, 1–6.
- Miranda Ribeiro, P. (1957) Notas para o estudo dos Pygidiidae Brasileiros (Pisces — Pygidiidae).— VI. *Publicações Avulsas do Museu Nacional (Rio de Janeiro)*, 13, 71–73.
- Miranda Ribeiro, P. (1959) Considerações sobre Callichthyidae Gill, 1872 (Nematognathi). *Boletim do Museu Nacional do Rio de Janeiro, Nova Série, Zoologia*, 206, 1–9.
- Miranda Ribeiro, P. (1959) Catálogo dos Peixes do Museu Nacional. III.— Callichthyidae Gill, 1872. *Publicações Avulsas do Museu Nacional (Rio de Janeiro)*, 27, 1–16.
- Miranda Ribeiro, P. (1962a) Apontamentos ictiológicos — I. *Boletim do Museu Nacional do Rio de Janeiro, Nova Série, Zoologia*, 240, 1–6.
- Miranda Ribeiro, P. (1962b) Sobre os gêneros *Heptapterus* Bleeker, 1858, e *Acentronichthys* Eigenmann & Eigenmann, 1889 (Pisces — Pimelodidae). *Boletim do Museu Nacional do Rio de Janeiro, Nova Série, Zoologia*, 236, 1–11.
- Miranda Ribeiro, P. (1964) *Rhamdella schultzi* sp. nov. (Pisces — Siluriformes — Pimelodidae). *Boletim do Museu Nacional do Rio de Janeiro, Nova Série, Zoologia*, 248, 1–4.
- Miranda Ribeiro, P. (1968) Apontamentos ictiológicos — IV. *Boletim do Museu Nacional do Rio de Janeiro, Nova Série, Zoologia*, 262, 1–7.
- Mirza, M.R. (1980) The systematics and zoogeography of the freshwater fishes of Pakistan and Azad Kashmir. *Proceeding of the first Pakistan Congress of Zoology*, 1–41.
- Mirza, M.R. (2003) Checklist of freshwater fishes of Pakistan. *Pakistan Journal of Zoology, Supplement Series*, 3, 1–30.
- Mirza, M.R. & Awan, M.I. (1973) Two new catfishes (Pisces, Siluriformes) from Pakistan. *Biologia (Lahore)*, 19, 145–159.
- Mirza, M.R. & Hameed, K. (1974) Sisorid fishes (Osteichthyes: Sisoridae) of Pakistan and Azad Kashmir. *Biologia (Lahore)*, 20, 83–89.
- Mirza, M.R. & Jan, M.A. (1989) *Batasio pakستانicus* new species, a new catfish (Pisces, Bagridae) from Pakistan. *Scientific Khyber*, 2, 283–286.
- Mirza, M.R. & Kashmiri, K.M. (1971) A note on the fishes of the genus *Glyptothorax* Blyth (Osteichthyes, Sisoridae) from West Pakistan with the description of a new subspecies. *Biologia (Lahore)*, 17, 87–93.
- Mirza, M.R. & Naik, I.U. (1969) Fishes of Zhob District with the description of a new species. *Pakistan Journal of Science*, 21, 121–125.
- Mirza, M.R., Nawaz, H. & Javed, M.N. (1992) A note on the fishes of genus *Aorichthys* Wu with the description of a new subspecies from Pakistan. *Pakistan Journal of Zoology*, 24, 211–213.
- Mirza, M.R. & Nijssen, H. (1978) *Glyptothorax stocki*, a new sisorid catfish from Pakistan & Azad Kashmir (Siluriformes, Sisoridae). *Bulletin Zoölogisch Museum, Universiteit van Amsterdam*, 6, 79–85.

- Mirza, M.R., Parveen, A. & Javed, M.N. (1999) *Gagata pakistanica*, a new catfish from Pakistan (Pisces: Sisoridae). *Punjab University, Journal of Zoology*, 14, 1–4.
- Misra, K.S. (1976) *Teleostomi: Cypriniformes; Siluri. The Fauna of India and the Adjacent Countries (Pisces, 2nd Edition)*, Vol. 3, Zoological Survey of India, Calcutta, xxi + 367 p., 5 pl.
- Mitchell, T.L. (1838) *Three Expeditions into the Interior of Eastern Australia, with Descriptions of Recently Explored Regions of Australia Felix, and of the Present Colony of New South Wales*, Vol. 1, T. & W. Boone, London, 351 p. + 51 pls.
- Mitchill, S.L. (1815) The fishes of New York described and arranged. *Transactions of the Literary and Philosophical Society of New-York*, 1, 355–492, pls. 1–6.
- Mitchill, S.L. (1817) [Report on the ichthyology of the Wallkill, from specimens of fishes presented to the society (Lyceum of Natural History)...by Dr. B. Akerly...]. *American Monthly Magazine and Critical Review*, 1, 289–290.
- Mo, T. (1991) *Anatomy, Relationships and Systematics of the Bagridae (Teleostei: Siluroidei) with a Hypothesis of Siluroid Phylogeny (Theses Zoologicae, 17)*, Koeltz, Koenigstein, 216 p., 63 figs.
- Mo, T.-P. & Chu, X.-L. (1986) [A revision of the sisorid catfish genus *Glyptothorax* from China]. *Zoological Research*, 7, 339–350. [In Chinese]
- Mohsin, A.K.M. & Ambak, M.A. (1983) *Freshwater Fishes of Peninsular Malaysia*, Penerbit Universiti Pertanian Malaysia, 284 p.
- Mojica, J.I., Castellanos-C., C. & Polanco-F, A. (2000) Redescubrimiento de *Dupouyichthys sapito* (Pisces: Siluriformes: Aspredinidae) en la cuenca del Río Magdalena (Colombia). *Caldasia*, 22, 353–355.
- Molina, G.I. (1782) *Saggio Sulla Storia Naturale del Chile, del Signor Abate Giovanni Ignazio Molina*, Bologna, v + 306 p. + errata + map.
- Montoya-Burgos, J.-I. (2003) Historical biogeography of the catfish genus *Hypostomus* (Siluriformes: Loricariidae), with implications on the diversification of Neotropical ichthyofauna. *Molecular Ecology*, 12, 1855–1867.
- Montoya-Burgos, J.-I., Muller, S., Weber, C. & Pawlowski, J. (1997) Phylogenetic relationships between Hypostominae and Ancistrinae (Siluroidei: Loricariidae): first results from mitochondrial 12S and 16S rRNA gene sequences. *Revue suisse de Zoologie, Annales de la Société zoologique suisse et du Muséum d'Histoire naturelle de Genève*, 104, 185–198.
- Montoya-Burgos, J.-I., Muller, S., Weber, C. & Pawlowski, J. (1998) Phylogenetic relationships of the Loricariidae (Siluriformes) based on mitochondrial rRNA gene sequences. In: Malabarba, L.R., Reis, R.E., Vari, R.P., Lucena, Z.M.S. & Lucena, C.A.S. (Eds.), *Phylogeny and Classification of Neotropical Fishes*, Edipucrs, Porto Alegre, pp. 363–374.
- Montoya-Burgos, J.-I., Weber, C. & Le Bail, P.-Y. (2002) Phylogenetic relationships within *Hypostomus* (Siluriformes: Loricariidae) and related genera based on mitochondrial D-loop sequences. *Revue suisse de Zoologie, Annales de la Société zoologique suisse et du Muséum d'Histoire naturelle de Genève*, 109, 369–382.
- Mori, T. (1933) Second addition to the fish fauna of Tsing-tau, China, with descriptions of three new species. *Japanese Journal of Zoology*, 5, 165–169.
- Mori, T. (1936) [Descriptions of one new genus and three new species of Siluroidea from Chosen]. *Dobutsugaku Zasshi*, 48, 671–675, pl. 24. [In Japanese and English].
- Mori, T. (1952) Check list of the fishes of Korea. *Memoirs of the Hyogo University of Agriculture*, 1, 1–228.
- Moyer, G.R., Burr, B.M. & Krajewski, C. (2004) Phylogenetic relationships of thorny catfishes (Siluriformes: Doradidae) inferred from molecular and morphological data. *Zoological Journal of the Linnean Society*, 140, 551–575.
- Mukerji, D.D. (1932) On a collection of fish from lower Burma. *Records of the Indian Museum*, 34, 281–286.
- Müller, J. (1840) Vergleichende Anatomie der Myxinoiden, der Cyclostomen mit durchhohrten Gaumen. IV — Über das Gefäss-system. *Abhandlungen der Deutschen Akademie der Wissenschaften zu Berlin*, 1839, 175–304, pls. 1–5. [Abstracts apparently published in Berlin Akad. Wissenschaften, Berlin, 1839: 184–186 (see Dean, 1917), Archiv für Naturgeschichte (see Jordan, 1919: 196), and Archiv für Anatomie, Physiologie und wissenschaftliche Medizin (see Eschmeyer, 1998). Availability of names from abstracts, and priority of publication, not established.]
- Müller, J. (1842a) Über die Schwimmblase der Fische, mit Bezug auf einige neue Fishgattungen. *Königlichen Preussischen Akademie des Wissenschaften zu Berlin*, 1842, 202–210.
- Müller, J. (1842b) Beobachtungen über die Schwimmblase der Fische, mit Bezug auf einige neue Fischgattungen. *Archiv für Anatomie, Physiologie und wissenschaftliche Medicin (Müller)*, Jahr, 1842, 307–329.
- Müller, J. (1843) Beiträge zur Kenntniss der natürlichen Familien der Fische. *Archiv für Naturgeschichte*, 9, 292–330.
- Müller, J. (1865) *Reisen in den Vereinigten Staaten, Canada und Mexico*, 3 vols [1864–65], F.A. Brockhaus, Leipzig.
- Müller, J. & Troschel, F.H. (1848) Fische. In: *Reisen in Britisch-Guiana in den Jahren 1840–44. Im Auftrag Sr. Mäjestat*

- des Königs von Preussen ausgeführt von Richard Schomburgk. [Versuch einer Fauna und Flora von Britisch-Guiana.]* Vol. 3, Berlin, pp. 618–644.
- Müller, J. & Troschel, F.H. (1849) *Horae Ichthyologicae: Beschreibung und Abbildung neuer Fische, Drittes Heft*, Verlag von Veit & Co, Berlin, 28 p., 5 pls.
- Muller, S. (1989) Description de deux nouvelles espèces paraguayennes du genre *Ancistrus* Kner, 1854 (Pisces, Siluriformes, Loricariidae). *Revue suisse de Zoologie, Annales de la Société zoologique suisse et du Muséum d'Histoire naturelle de Genève*, 96, 885–904.
- Muller, S. (1990) Étude méristique et morphométrique d'*Ancistrus piriformis* Muller et *Ancistrus pirareta* Muller. *Revue suisse de Zoologie, Annales de la Société zoologique suisse et du Muséum d'Histoire naturelle de Genève*, 97, 153–168.
- Muller, S. & Isbrücker, I.J.H. (1993) *Lithoxus boujardi* (Siluriformes, Loricariidae), une espèce nouvelle du Bassin de l'Approuague, Guyane Française. *Cybium (3e série)*, 17, 71–76.
- Muller, S., Rapp Py-Daniel, L.H. & Zuanon, J. (1994) *Ancistrus ranunculus*, a new species of loricariid fish (Siluriformes: Loricariidae) from the Xingú and Tocantins rivers, Brazil. *Ichthyological Exploration of Freshwaters*, 5, 289–296.
- Muller, S. & Weber, C. (1992) Les dents des sous-familles Hypostominae et Ancistrinae (Pisces, Siluriformes, Loricariidae) et leur valeur taxonomique. *Revue suisse de Zoologie, Annales de la Société zoologique suisse et du Muséum d'Histoire naturelle de Genève*, 99, 747–754.
- Muriel-Cunha, J. & Pinna, M., de (2005) New data on cistern catfish, *Phreatobius cisternarum*, from subterranean waters at the mouth of the Amazon River (Siluriformes, Incertae sedis). *Papéis Avulsos de Zoologia*, 45, 327–339.
- Murray, A. (1855) On electrical fishes; with a description of a new species of *Malapterurus* from Old Calabar, received from the Rev. Hope M. Waddell, missionary there. *Proceedings of the Royal Physical Society of Edinburgh*, 1, 20–21, 1 pl.
- Murray, A.M. & Budney, L.A. (2003) A new species of catfish (Claroteidae, *Chrysichthys*) from an Eocene crater lake in East Africa. *Canadian Journal of Earth Science*, 40, 983–993.
- Musikasinthorn, P., Utsugi, K. & Watanabe, K. (1998) Rediscovery of the pangasid catfish *Helicophagus typus* in Borneo. *Natural History Bulletin of the Siam Society*, 46, 197–201.
- Musyl, M.K. & Keenan, C.P. (1996) Evidence for cryptic speciation in Australian freshwater eel-tailed catfish, *Tandanus tandanus* (Teleostei: Plotosidae). *Copeia*, 1996, 526–534.
- Myers, G.S. (1925a) Description of a new catfish from Abyssinia. *Copeia*, no. 139, 12–13.
- Myers, G.S. (1925b) *Tridentopsis pearsoni* a new pygidiid catfish from Bolivia. *Copeia*, no. 148, 83–86.
- Myers, G.S. (1926) Descriptions of a new characin fish and a new pygidiid catfish from the Amazon Basin. *Copeia*, no. 156, 150–152.
- Myers, G.S. (1927) Descriptions of new South American fresh-water fishes collected by Dr. Carl Ternetz. *Bulletin of the Museum of Comparative Zoology*, 68, 107–135.
- Myers, G.S. (1928) New fresh-water fishes from Peru, Venezuela, and Brazil. *Annals and Magazine of Natural History (Ser. 10)*, 2, 83–90.
- Myers, G.S. (1931) On the fishes described by Koller from Hainan in 1926 and 1927. *Lingnan Science Journal*, 10, 255–262.
- Myers, G.S. (1932) Notes on Colombian fresh-water fishes, with description of a new *Astroblepus*. *Copeia*, 1932, 137–138.
- Myers, G.S. (1933) New importations: Leopard *Corydoras*. *Aquarium, Philadelphia*, 2, 188–189.
- Myers, G.S. (1938) Notes on *Ansorgia*, *Clarisilurus*, *Wallago*, and *Ceratoglanis*, four genera of African and Indo-Malayan catfishes. *Copeia*, 1938, 98.
- Myers, G.S. (1941) A new name for *Taenionema*, a genus of Amazonian siluroid fishes. *Stanford Ichthyological Bulletin*, 2, 88.
- Myers, G.S. (1942) Studies on South American fresh-water fishes.— I. *Stanford Ichthyological Bulletin*, 2, 89–114.
- Myers, G.S. (1944) Two extraordinary new blind nematognath fishes from the Rio Negro, representing a new subfamily of Pygidiidae, with a rearrangement of the genera of the family, and illustrations of some previously described genera and species from Venezuela and Brazil. *Proceedings of the California Academy of Sciences, Series 4*, 23, 591–602, pls. 52–56.
- Myers, G.S. (1948) Note on two generic names of Indo-Malayan silurid fishes, *Wallago* and *Wallagonia*. *Proceedings of the California Zoological Club*, 1, 19–20.
- Myers, G.S. (1953) A note on the habits and classification of *Corydoras hastatus*. *The Aquarium Journal*, 24, 268–270.

- Myers, G.S. (1960) The genera and ecological geography of the South American banjo catfishes, family Aspredinidae. *Stanford Ichthyological Bulletin*, 7, 132–139.
- Myers, G.S. & Weitzman, S.H. (1954) Another new *Corydoras* from Brazil. *The Aquarium Journal*, 25, 93–94.
- Myers, G.S. & Weitzman, S.H. (1956) Two new Brazilian fresh water fishes. *Stanford Ichthyological Bulletin*, 7, 1–4.
- Myers, G.S. & Weitzman, S.H. (1960) Two new fishes collected by General Thomas D. White in eastern Colombia. *Stanford Ichthyological Bulletin*, 7, 98–109.
- Myers, G.S. & Weitzman, S.H. (1966) Two remarkable new trichomycterid catfishes from the Amazon basin in Brazil and Colombia. *Journal of Zoology (London)*, 149, 277–287.
- Nani, A. & Fuster, M.L. (1947) *Hypophthalmus oremaculatus* una nueva especie del orden "Nematognathi" (Pisces, Hypophthal.). *Comunicaciones del Museo Argentino de Ciencias Naturales "Bernardino Rivadavia"*, Serie Ciencias Zoológicas, 2, 1–9.
- Naseka, A.M. & Bogutskaya, N.G. (2004) Contribution to taxonomy and nomenclature of freshwater fishes of the Amur drainage area and the Far East (Pisces, Osteichthyes). *Zoosystematica Rossica*, 12, 279–290.
- Nass, P. (1991) *Anatomía Comparada del Bagre Cunagaro Brachyplatystoma juruense (Boulenger, 1898)*, Incluyendo un Análisis Filogenético de la Familia Pimelodidae, Unpublished Ph.D. dissertation, Universidad Central de Venezuela, Caracas.
- Nath, P. & Dey, S.C. (1989) Two new fish species of the genus *Amblyceps* Blyth from Arunachal Pradesh, India. *Journal of the Assam Science Society*, 32, 1–6.
- Neave, S.A. (1940) *Nomenclator Zoologicus*, Vol. 3, The Zoological Society of London, London.
- Nelson, E.W. (1876) A partial catalogue of the fishes of Illinois. *Bulletin of the Illinois Museum of Natural History*, 1, 33–52.
- Nelson, J.S. (2006) *Fishes of the World*, Fourth Edition, John Wiley and Sons, New York, 601 p.
- Newton, E.T. (1889) A contribution to the history of Eocene siluroid fishes. *Proceedings of the Zoological Society of London*, 1889, 201–207, 1 pl.
- Ng, H.H. (1996) *Akysis heterurus*, a new species of catfish (Teleostei: Akysidae) from eastern Sumatra, with notes on *Akysis hendricksoni* Alfred. *The Raffles Bulletin of Zoology*, 44, 3–10.
- Ng, H.H. (1999a) *Pterocryptis inusitata*, a new species of silurid catfish from Laos (Teleostei: Siluriformes). *Ichthyological Exploration of Freshwaters*, 10, 371–374.
- Ng, H.H. (1999b) *Laides longibarbis*, a valid species of schilbeid catfish from Indochina (Teleostei: Siluriformes). *Ichthyological Exploration of Freshwaters*, 10, 381–385.
- Ng, H.H. (1999c) A review of the southeast Asian catfish genus *Ceratoglanis* (Siluriformes: Siluridae), with the description of a new species from Thailand. *Proceedings of the California Academy of Sciences, Series 4*, 51, 385–395.
- Ng, H.H. (1999d) Two new species of catfishes of the genus *Clarias* from Borneo (Teleostei: Clariidae). *The Raffles Bulletin of Zoology*, 47, 17–32.
- Ng, H.H. (1999e) The *Akysis* of Myanmar: a review (Teleostei: Akysidae). *The Raffles Bulletin of Zoology*, 47, 541–548.
- Ng, H.H. (1999f) *Bagrichthys obscurus*, a new species of bagrid catfish from Indochina (Teleostei: Bagridae). *Revista de Biología Tropical*, 47, 545–552.
- Ng, H.H. (2000) *Bagrichthys vaillantii* (Popa, 1906), a valid species of bagrid catfish from eastern Borneo (Teleostei: Siluriformes). *Zoologische Mededelingen (Leiden)*, 73, 327–332.
- Ng, H.H. (2001a) *Kryptopterus dissitus*, a new silurid catfish from Indochina (Teleostei, Siluridae). *Folia Zoologica*, 50, 197–200.
- Ng, H.H. (2001b) *Amblyceps macropterus*, a new species of amblycipitid catfish (Osteichthyes: Amblycipitidae) from Pakistan. *Ichthyological Exploration of Freshwaters*, 12, 201–204.
- Ng, H.H. (2001c) *Clarias microstomus*, a new species of clariid catfish from eastern Borneo (Teleostei: Siluriformes). *Zoological Studies*, 40, 158–162.
- Ng, H.H. (2002a) *Bagrichthys majusculus*, a new catfish from Indochina (Teleostei, Bagridae). *Folia Zoologica*, 51, 49–54.
- Ng, H.H. (2002b) *Ompok binotatus* and *Ompok pluriradiatus*, two new species of silurid catfish from western Borneo (Teleostei: Siluriformes). *Ichthyological Exploration of Freshwaters*, 13, 25–32.
- Ng, H.H. (2002c) Descriptions of two new species of *Kryptopterus* from Thailand and Borneo in the *K. limpok* species group (Teleostei: Siluridae). *Ichthyological Exploration of Freshwaters*, 13, 69–74.
- Ng, H.H. (2002d) The identity of *Mystus nigriceps* (Valenciennes in Cuvier & Valenciennes, 1840), with the description of a new bagrid catfish (Teleostei: Siluriformes) from Southeast Asia. *The Raffles Bulletin of Zoology*, 50, 161–168.
- Ng, H.H. (2003a) *Mystus impluviatus*: a new species of bagrid catfish (Teleostei: Bagridae) from eastern Borneo. *Copeia*,

- 2003, 373–378.
- Ng, H.H. (2003b) A review of the *Ompok hypophthalmus* group of silurid catfishes with the description of a new species from South-east Asia. *Journal of Fish Biology*, 62, 1296–1311.
- Ng, H.H. (2003c) A revision of the south Asian sisorid catfish genus *Sisor* (Teleostei: Siluriformes). *Journal of Natural History*, 37, 2871–2883.
- Ng, H.H. (2003d) *Kryptopterus paraschilbeides*, a new species of catfish (Teleostei: Siluridae) from mainland Southeast Asia. *The Natural History Journal of the Chulalongkorn University*, 3, 1–8.
- Ng, H.H. (2003e) *Arius verrucosus*, a new species of freshwater ariid catfish (Teleostei: Ariidae) from the Mekong River. *Occasional Papers of the Museum of Zoology, University of Michigan*, 734, 1–14.
- Ng, H.H. (2003f) *Ompok pinnatus*, a new species of silurid catfish (Teleostei: Siluriformes: Siluridae) from mainland Southeast Asia. *Proceedings of the Biological Society of Washington*, 116, 47–51.
- Ng, H.H. (2003g) *Clarias nigricans*, a new species of clariid catfish (Teleostei: Siluriformes) from eastern Borneo. *The Raffles Bulletin of Zoology*, 51, 393–398.
- Ng, H.H. (2003h) *Clarias insolitus*, a new species of clariid catfish (Teleostei: Siluriformes) from southern Borneo. *Zootaxa*, 284, 1–8.
- Ng, H.H. (2003i) *Kryptopterus geminus*, a new species of silurid catfish (Teleostei: Siluridae) from mainland Southeast Asia. *Zootaxa*, 305, 1–11.
- Ng, H.H. (2003j) Phylogeny and systematics of Bagridae. In: Arratia, G., Kapoor, B.G., Chardon, M. & Diogo, R. (Eds.), *Catfishes, Vol. 1*, Science Publishers, Enfield, NH, USA, pp. 439–464.
- Ng, H.H. (2004a) *Wallago micropogon*: a new species of silurid catfish (Teleostei: Siluridae) from mainland Southeast Asia. *Copeia*, 2004, 92–97.
- Ng, H.H. (2004b) New estuarine species of *Mystus* (Teleostei: Bagridae) from the Malay Peninsula. *Copeia*, 2004, 883–887.
- Ng, H.H. (2004c) *Batasio elongatus*, a new species of bagrid catfish from southwest Myanmar (Siluriformes: Bagridae). *Ichthyological Exploration of Freshwaters*, 15, 67–70.
- Ng, H.H. (2004d) *Clarias sulcatus*, a new walking catfish (Teleostei: Clariidae) from Pulau Redang. *Ichthyological Exploration of Freshwaters*, 15, 289–294.
- Ng, H.H. (2004e) *Oreoglanis macronemus*, a new species of glyptosternine catfish (Teleostei: Siluriformes: Sisoridae) from northern Laos. *The Raffles Bulletin of Zoology*, 52, 209–213.
- Ng, H.H. (2004f) *Kryptopterus platypogon*, a new silurid catfish (Teleostei: Siluridae) from Borneo. *Zootaxa*, 398, 1–8.
- Ng, H.H. (2004g) Two new glyptosternine catfishes (Teleostei: Sisoridae) from Vietnam and China. *Zootaxa*, 428, 1–12.
- Ng, H.H. (2004h) The *Microsynodontis* (Teleostei: Siluriformes: Mochokidae) of the lower Guinea region, west central Africa, with the description of eight new species. *Zootaxa*, 531, 1–52.
- Ng, H.H. (2004i) *Rita macracanthus*, a new riverine catfish (Teleostei: Bagridae) from South Asia. *Zootaxa*, 568, 1–12.
- Ng, H.H. (2005a) *Conta pectinata*, a new erethistid catfish (Teleostei: Eretistidae) from northeast India. *Ichthyological Exploration of Freshwaters*, 16, 23–28.
- Ng, H.H. (2005b) *Pseudolaguvia foveolata*, a new catfish (Teleostei: Eretistidae) from northeast India. *Ichthyological Exploration of Freshwaters*, 16, 173–178.
- Ng, H.H. (2005c) *Gogangra laevis*, a new species of riverine catfish from Bangladesh (Teleostei: Sisoridae). *Ichthyological Exploration of Freshwaters*, 16, 279–286.
- Ng, H.H. (2005d) *Amblyceps carinatum*, a new species of hillstream catfish from Myanmar (Teleostei: Amblycipitidae). *The Raffles Bulletin of Zoology*, 53, 243–249.
- Ng, H.H. (2005e) *Glyptothorax botius* (Hamilton, 1822), a valid species of catfish (Teleostei: Sisoridae) from northeast India. *Zootaxa*, 930, 1–19.
- Ng, H.H. (2005f) *Erethistoides sicula*, a new catfish (Teleostei: Eretistidae) from India. *Zootaxa*, 1021, 1–12.
- Ng, H.H. (2005g) Two new species of *Pseudolaguvia* (Teleostei: Eretistidae) from Bangladesh. *Zootaxa*, 1044, 35–47.
- Ng, H.H. & Chan, B.P.-L. (2005) Revalidation and redescription of *Pterocryptis anomala* (Herre, 1933), a catfish (Teleostei: Siluridae) from southern China. *Zootaxa*, 1060, 51–64.
- Ng, H.H. & Dodson, J.J. (1999) Morphological and genetic descriptions of a new species of catfish, *Hemibagrus chrysops*, from Sarawak, east Malaysia, with an assessment of phylogenetic relationships (Teleostei: Bagridae). *The Raffles Bulletin of Zoology*, 47, 45–57.
- Ng, H.H. & Edds, D.R. (2004) *Batasio macronotus*, a new species of bagrid catfish from Nepal (Teleostei: Bagridae). *Ichthyological Exploration of Freshwaters*, 15, 295–300.
- Ng, H.H. & Edds, D.R. (2005a) Two new species of *Erethistoides* (Teleostei: Eretistidae) from Nepal. *Ichthyological*

- Exploration of Freshwaters*, 16, 239–248.
- Ng, H.H. & Edds, D.R. (2005b) Two new species of *Pseudecheneis*, rheophilic catfishes (Teleostei: Sisoridae) from Nepal. *Zootaxa*, 1047, 1–19.
- Ng, H.H. & Ferraris, C.J., Jr. (2000) A review of the genus *Hemibagrus* in southern Asia, with descriptions of two new species. *Proceedings of the California Academy of Sciences, Series 4*, 52, 125–142.
- Ng, H.H. & Freyhof, J. (2001a) A review of the catfish genus *Pterocryptis* (Siluridae) in Vietnam, with the description of two new species. *Journal of Fish Biology*, 59, 624–644.
- Ng, H.H. & Freyhof, J. (2001b) *Oreoglanis insulatus*, a new species of glyptosternine catfish (Siluriformes: Sisoridae) from central Vietnam. *Journal of Fish Biology*, 59, 1164–1169.
- Ng, H.H. & Freyhof, J. (2003) *Akysis clavulus*, a new species of catfish (Teleostei: Akysidae) from central Vietnam. *Ichthyological Exploration of Freshwaters*, 14, 311–316.
- Ng, H.H. & Freyhof, J. (2005) A new species of *Pseudomystus* (Teleostei: Bagridae) from Central Vietnam. *Copeia*, 2005, 745–750.
- Ng, H.H. & Hadiaty, R.K. (2005) Two new bagrid catfishes (Teleostei: Bagridae) from the Alas River drainage, northern Sumatra. *Ichthyological Exploration of Freshwaters*, 16, 83–92.
- Ng, H.H. & Kottelat, M. (1996) *Akysis fuscus*, a new species of catfish (Teleostei: Akysidae) from the Kapuas basin, Borneo. *Ichthyological Exploration of Freshwaters*, 7, 19–26.
- Ng, H.H. & Kottelat, M. (1997) *Silurichthys citatus*, a new species of silurid catfish from western Borneo (Teleostei: Siluriformes). *Ichthyological Exploration of Freshwaters*, 7, 203–208.
- Ng, H.H. & Kottelat, M. (1998a) *Hyalobagrus*, a new genus of miniature bagrid catfish from southeast Asia (Teleostei: Siluriformes). *Ichthyological Exploration of Freshwaters*, 9, 335–346.
- Ng, H.H. & Kottelat, M. (1998b) *Pterocryptis buccata*, a new species of catfish from western Thailand (Teleostei: Siluridae) with epigean and hypogean populations. *Ichthyological Research*, 45, 393–399.
- Ng, H.H. & Kottelat, M. (1998c) The catfish genus *Akysis* Bleeker (Teleostei: Akysidae) in Indochina, with descriptions of six new species. *Journal of Natural History*, 32, 1057–1097.
- Ng, H.H. & Kottelat, M. (1999) *Oreoglanis hypsiurus*, a new species of glyptosternine catfish (Teleostei: Sisoridae). *Ichthyological Exploration of Freshwaters*, 10, 375–380.
- Ng, H.H. & Kottelat, M. (2000a) A review of the genus *Amblyceps* (Osteichthyes: Amblycipitidae) in Indochina, with descriptions of five new species. *Ichthyological Exploration of Freshwaters*, 11, 335–348.
- Ng, H.H. & Kottelat, M. (2000b) Description of three new species of catfishes (Teleostei: Akysidae and Sisoridae) from Laos and Vietnam. *Journal of South Asian Natural History*, 5, 7–15.
- Ng, H.H. & Kottelat, M. (2000c) *Helicophagus leptorhynchus*, a new species of molluscivorous catfish from Indochina (Teleostei: Pangasiidae). *The Raffles Bulletin of Zoology*, 48, 55–58.
- Ng, H.H. & Kottelat, M. (2000d) *Cranoglanis henrici* (Vaillant, 1893) a valid species of cranoglanidid catfish from Indochina (Teleostei: Cranoglanididae). *Zoosystema*, 22, 847–852.
- Ng, H.H. & Kottelat, M. (2001) A review of the genus *Batasio* (Teleostei: Bagridae) in Indochina, with the description of *B. tigrinus* sp. n. from Thailand. *Revue suisse de Zoologie, Annales de la Société zoologique suisse et du Muséum d'Histoire naturelle de Genève*, 108, 495–511.
- Ng, H.H. & Kottelat, M. (2003) *Parakysis notialis*, a new species of akysid catfish from Borneo (Siluriformes: Akysidae). *Ichthyological Research*, 50, 48–51.
- Ng, H.H. & Kottelat, M. (2004) *Akysis vespa*, a new species of catfish (Siluriformes: Akysidae) from the Ataran River drainage (Myanmar). *Ichthyological Exploration of Freshwaters*, 25, 193–200.
- Ng, H.H. & Kottelat, M. (2005) *Caelatoglanis zontatus*, a new genus and species of the Erythistidae (Teleostei: Siluriformes) from Myanmar, with comments on the nomenclature of *Laguvia* and *Hara* species. *Ichthyological Exploration of Freshwaters*, 16, 13–22.
- Ng, H.H. & Lim, K.K.P. (1995) A revision of the southeast Asian catfish genus *Parakysis* (Teleostei: Akysidae), with descriptions of two new species. *Ichthyological Exploration of Freshwaters*, 6, 255–266.
- Ng, H.H. & Lim, K.K.P. (2000) *Hemileiocassis panjang*, a new genus and new species of bagrid catfish from Java (Teleostei, Siluriformes). *Beaufortia*, 50, 191–195.
- Ng, H.H. & Lim, K.K.P. (2005) The identity of *Pseudomystus moeschii* (Boulenger, 1890), with the description of two new species of bagrid catfishes from Southeast Asia (Teleostei: Bagridae). *Zootaxa*, 851, 1–18.
- Ng, H.H., Martin-Smith, K.M. & Ng, P.K.L. (2000) *Hemibagrus furcatus*, a new species of bagrid catfish (Teleostei: Siluriformes) from Sabah, east Malaysia. *The Raffles Bulletin of Zoology*, 48, 65–69.
- Ng, H.H. & Ng, P.K.L. (1998) A revision of the South-east Asian catfish genus *Silurichthys*. *Journal of Fish Biology*, 52,

- Ng, H.H. & Ng, P.K.L. (2001) A revision of the akysid catfish genus *Acrochordonichthys* Bleeker. *Journal of Fish Biology*, 58, 386–418.
- Ng, H.H. & Rachmatika, I. (1999) The catfishes (Teleostei: Siluriformes) of Bentuang Karimun National Park, West Kalimantan, Indonesia. *The Raffles Bulletin of Zoology*, 47, 167–183.
- Ng, H.H. & Rachmatika, I. (2005) *Glyptothorax exodon*, a new species of rheophilic catfish from Borneo (Teleostei: Sisoridae). *The Raffles Bulletin of Zoology*, 53, 251–255.
- Ng, H.H. & Rainboth, W.J. (1999) The bagrid catfish genus *Hemibagrus* (Teleostei: Siluriformes) in central Indochina with a new species from the Mekong River. *The Raffles Bulletin of Zoology*, 47, 555–576.
- Ng, H.H. & Rainboth, W.J. (2001) A review of the sisorid catfish genus *Oreoglanis* (Siluriformes: Sisoridae) with descriptions of four new species. *Occasional Papers of the Museum of Zoology, University of Michigan*, 732, 1–34.
- Ng, H.H. & Rainboth, W.J. (2005) Four new species of *Akysis* (Teleostei: Siluriformes: Akysidae) from mainland Southeast Asia, with comments on *A. similis*. *The Raffles Bulletin of Zoology*, Supplement 13, 33–42.
- Ng, H.H. & Sabaj, M.H. (2005) *Akysis hardmani* (Siluriformes: Akysidae), a new species of catfish from Thailand. *Ichthyological Exploration of Freshwaters*, 16, 215–222.
- Ng, H.H. & Siebert, D.J. (1998) A revision of the akysid catfish genus *Breitensteinia* Steindachner with descriptions of two new species. *Journal of Fish Biology*, 53, 645–657.
- Ng, H.H. & Siebert, D.J. (2002) Rediscovery and redescription of *Ompok weberi* (Hardenberg, 1936), a poorly known species of silurid catfish (Teleostei: Siluriformes) from Borneo. *The Raffles Bulletin of Zoology*, 50, 169–173.
- Ng, H.H. & Siebert, D.J. (2004) A new species of the catfish genus *Akysis* (Siluriformes: Akysidae) from southern Borneo. *Zootaxa*, 733, 1–8.
- Ng, H.H. & Siebert, D.J. (2005) *Pseudomystus stenogrammus*, a new species of bagrid catfish from Borneo (Teleostei, Bagridae). *Zootaxa*, 813, 1–7.
- Ng, H.H. & Sparks, J.S. (2002) *Plotosus fisadoha*, a new species of marine catfish (Teleostei: Siluriformes: Plotosidae) from Madagascar. *Proceedings of the Biological Society of Washington*, 115, 80–85.
- Ng, H.H. & Sparks, J.S. (2003) The ariid catfishes (Teleostei: Siluriformes: Ariidae) of Madagascar, with the description of two new species. *Occasional Papers of the Museum of Zoology, University of Michigan*, 735, 1–21.
- Ng, H.H. & Sparks, J.S. (2005) Revision of the endemic Malagasy catfish family Anchariidae (Teleostei: Siluriformes), with descriptions of a new genus and three new species. *Ichthyological Exploration of Freshwaters*, 16, 303–323.
- Ng, H.H. & Tan, H.H. (1999) The fishes of the Endau drainage, Peninsular Malaysia with descriptions of two new species of catfishes (Teleostei: Akysidae, Bagridae). *Zoological Studies*, 38, 350–366.
- Ng, H.H. & Tan, H.H. (2000) A new species of *Encheloclarias* (Siluriformes: Clariidae) from Sumatra. *Journal of Fish Biology*, 57, 536–539.
- Ng, H.H. & Tan, H.H. (2002) Redescription of *Acrochordonichthys ischnosoma* Bleeker, 1858, a poorly known species of akysid catfish (Teleostei: Siluriformes) from Sumatra and Java. *The Raffles Bulletin of Zoology*, 50, 449–452.
- Ng, H.H. & Tan, H.H. (2004) *Ompok platyrhynchus*, a new silurid catfish (Teleostei: Siluridae) from Borneo. *Zootaxa*, 580, 1–11.
- Ng, H.H., Wirjoatmodjo, S. & Hadiaty, R.K. (2001a) *Mystus punctifer*, a new species of bagrid catfish (Teleostei: Siluriformes) from northern Sumatra. *The Raffles Bulletin of Zoology*, 49, 355–358.
- Ng, H.H., Wirjoatmodjo, S. & Hadiaty, R.K. (2001b) *Hemibagrus caveatus*, a new species of bagrid catfish (Teleostei: Siluriformes) from northern Sumatra. *The Raffles Bulletin of Zoology*, 49, 359–361.
- Ng, H.H., Wirjoatmodjo, S. & Hadiaty, R.K. (2004) *Kryptopterus piperatus*, a new species of silurid catfish (Teleostei: Siluridae) from northern Sumatra. *Ichthyological Exploration of Freshwaters*, 15, 91–95.
- Ng, P.K.L. & Lim, K.K.P. (1993) The Southeast Asian catfish genus *Encheloclarias* (Teleostei: Clariidae), with descriptions of four new species. *Ichthyological Exploration of Freshwaters*, 4, 21–37.
- Ng, P.K.L. & Ng, H.H. (1995) *Hemibagrus gracilis*, a new species of large riverine catfish (Teleostei: Bagridae) from Peninsular Malaysia. *The Raffles Bulletin of Zoology*, 43, 133–142.
- Nguyen, H.D. & Nguyen, V.H. (2001) [Two newly found fish species of *Pareuchiloglanis* genus (Sisoridae, Siluriformes) in Vietnam]. *Tap chí Sinh Học [Journal of Biology]*, 23, 66–71. [In Vietnamese, with English title and summary; English title only roughly agrees with Vietnamese title]
- Nichols, J.T. (1919a) "Cascudos" brasileiros do genero *Plecostomus* do Museu Paulista (Brasilian catfishes of the genus *Plecostomus* from the Museu Paulista). *Revista do Museu Paulista*, 11, 409–426.
- Nichols, J.T. (1919b) Um novo genero de cascudos da familia Loricariidae. *Revista do Museu Paulista*, 11, 533–535, 539–540.

- Nichols, J.T. (1925) Some Chinese fresh-water fishes. X.— Subgenera of bagrin catfishes. XI.— Certain apparently undescribed carps from Fukien. XII.— A small goby from the central Yangtze. XIII.— A new minnow referred to *Leucogobio*. XIV.— Two apparently undescribed fishes from Yunnan. *American Museum Novitates*, 185, 1–7.
- Nichols, J.T. (1926a) Some Chinese fresh-water fishes. XV.— Two apparently undescribed catfishes from Fukien. XVI.— Concerning gudgeons related to *Pseudogobio*, and two new species of it. XVII.— Two new rhodeins. *American Museum Novitates*, 214, 1–7.
- Nichols, J.T. (1926b) Some Chinese fresh-water fishes. XVIII.— New species in recent and earlier Fukien collections. *American Museum Novitates*, 224, 1–7.
- Nichols, J.T. (1930) Some Chinese fresh-water fishes. XXVI.— Two new species of *Pseudogobio*. XXVII.—A new catfish from northeastern Kiangsi. *American Museum Novitates*, 440, 1–5.
- Nichols, J.T. (1931) Some Chinese fresh-water fishes. XXVIII.— A collection from Chungan Hsien, northwestern Fukien. *American Museum Novitates*, 449, 1–3.
- Nichols, J.T. (1940) Results of the Archbold expeditions. No. 30.— New catfishes from northern New Guinea. *American Museum Novitates*, 1093, 1–3.
- Nichols, J.T. (1941) Four new fishes from western China. *American Museum Novitates*, 1107, 1–3.
- Nichols, J.T. (1943) *The Fresh-water Fishes of China, Natural History of Central Asia, Vol. 9*, American Museum of Natural History, New York, 322 p., 10 pl.
- Nichols, J.T. (1956) A new pygidiin catfish from Argentina. *American Museum Novitates*, 1760, 1–3.
- Nichols, J.T. & Griscom, L. (1917) Fresh-water fishes of the Congo basin obtained by the American Museum Congo expedition, 1909–1915. *Bulletin of the American Museum of Natural History*, 37, 653–756, pls. 64–83.
- Nichols, J.T. & La Monte, F.R. (1933a) A new catfish, *Amphilus pictus*, and a discussion of a small Liberian collection of fishes. *American Museum Novitates*, 626, 1–3.
- Nichols, J.T. & La Monte, F.R. (1933b) New fishes from the Kasai district of the Belgian Congo. *American Museum Novitates*, 656, 1–6.
- Nichols, J.T. & La Monte, F.R. (1934) More new fishes from the Kasai district of the Belgian Congo. *American Museum Novitates*, 723, 1–6.
- Nichols, J.T. & La Monte, F.R. (1953) Two new African catfishes from eastern French Equatorial Africa. *American Museum Novitates*, 1648, 1–4.
- Nichols, J.T. & Pope, C.H. (1927) The fishes of Hainan. *Bulletin of the American Museum of Natural History*, 54, 321–394, pl. 26.
- Nico, L.G. & Pinna, M.C.C., de. (1996) Confirmation of *Glanapteryx anguilla* (Siluriformes, Trichomycteridae) in the Orinoco River basin, with notes on the distribution and habits of the Glanapteryginae. *Ichthyological Exploration of Freshwaters*, 7, 27–32.
- Nielsen, J.G. (1974) *Fish Types in the Zoological Museum of Copenhagen*, Copenhagen, 115 p.
- Nijssen, H. (1970) Revision of the Surinam catfishes of the genus *Corydoras* La Cepède, 1803 (Pisces, Siluriformes, Callichthyidae). *Beaufortia*, 18, 1–75.
- Nijssen, H. (1971) Two new species and one new subspecies of the South American catfish genus *Corydoras* (Pisces, Siluriformes, Callichthyidae). *Beaufortia*, 19, 89–98.
- Nijssen, H. (1972) Records of the catfish genus *Corydoras* from Brazil and French Guiana with descriptions of eight new species (Pisces, Siluriformes, Callichthyidae). *Netherlands Journal of Zoology*, 21, 412–433, pls. 1–3.
- Nijssen, H. & Isbrücker, I.J.H. (1967) Notes on the Guiana species of *Corydoras* La Cepède, 1803, with descriptions of seven new species and designation of a neotype for *Corydoras punctatus* (Bloch, 1794) –(Pisces, Cypriniformes, Callichthyidae). *Zoologische Mededelingen (Leiden)*, 42, 21–50, pls. 1–5.
- Nijssen, H. & Isbrücker, I.J.H. (1970) The South American catfish genus *Brochis* Cope, 1872 (Pisces, Siluriformes, Callichthyidae). *Beaufortia*, 18, 151–168.
- Nijssen, H. & Isbrücker, I.J.H. (1971) Two new species of the catfish genus *Corydoras* from Brazil and Peru (Pisces, Siluriformes, Callichthyidae). *Beaufortia*, 18, 183–189.
- Nijssen, H. & Isbrücker, I.J.H. (1975) *Cataphractus punctatus* Bloch, 1794 (Pisces, Siluriformes, Callichthyidae): request for invalidation of neotype and validation of a rediscovered syntype as lectotype. *Bulletin of Zoological Nomenclature*, 32, 63–64, pl. 1.
- Nijssen, H. & Isbrücker, I.J.H. (1976a) The South American plated catfish genus *Aspidoras* R. von Ihering, 1907, with descriptions of nine new species from Brazil (Pisces, Siluriformes, Callichthyidae). *Bijdragen tot de Dierkunde*, 46, 107–131.
- Nijssen, H. & Isbrücker, I.J.H. (1976b) *Corydoras ornatus*, a new species of callichthyid catfish from the Rio Tapajós

- drainage, Brazil (Pisces, Siluriformes, Callichthyidae). *Bulletin Zoölogisch Museum, Universiteit van Amsterdam*, 5, 125–129.
- Nijssen, H. & Isbrücker, I.J.H. (1976c) A new callichthyid catfish, *Corydoras gracilis*, from Brazil. *Tropical Fish Hobbyist*, 25, 90–98.
- Nijssen, H. & Isbrücker, I.J.H. (1979) Chronological enumeration of nominal species and subspecies of *Corydoras* (Pisces, Siluriformes, Callichthyidae). *Bulletin Zoölogisch Museum, Universiteit van Amsterdam*, 6, 129–135.
- Nijssen, H. & Isbrücker, I.J.H. (1980a) On the identity of *Corydoras nattereri* Steindachner 1877 with the description of a new species *Corydoras prionotos* (Pisces, Siluriformes, Callichthyidae). *Beaufortia*, 30, 1–9.
- Nijssen, H. & Isbrücker, I.J.H. (1980b) A review of the genus *Corydoras* La Cepède, 1803 (Pisces, Siluriformes, Callichthyidae). *Bijdragen tot de Dierkunde*, 50, 190–220.
- Nijssen, H. & Isbrücker, I.J.H. (1980c) *Aspidoras virgulatus* n. sp., a plated catfish from Espírito Santo, Brazil (Pisces, Siluriformes, Callichthyidae). *Bulletin Zoölogisch Museum, Universiteit van Amsterdam*, 7, 133–139.
- Nijssen, H. & Isbrücker, I.J.H. (1980d) Three new *Corydoras* species from French Guiana and Brazil (Pisces, Siluriformes, Callichthyidae). *Netherlands Journal of Zoology*, 30, 494–503.
- Nijssen, H. & Isbrücker, I.J.H. (1982) *Corydoras boehlkei*, a new catfish from the Rio Caura system in Venezuela (Pisces, Siluriformes, Callichthyidae). *Proceedings of the Academy of Natural Sciences, Philadelphia*, 134, 139–142.
- Nijssen, H. & Isbrücker, I.J.H. (1983a) *Brochis britskii*, a new species of plated catfish from the upper Rio Paraguai system, Brazil (Pisces, Siluriformes, Callichthyidae). *Bulletin Zoölogisch Museum, Universiteit van Amsterdam*, 9, 177–186.
- Nijssen, H. & Isbrücker, I.J.H. (1983b) Review of the genus *Corydoras* from Colombia, with descriptions of two new species (Pisces, Siluriformes, Callichthyidae). *Beaufortia*, 33, 53–71.
- Nijssen, H. & Isbrücker, I.J.H. (1983c) Sept espèces nouvelles de poissons-chats cuirassés du genre *Corydoras* La Cepède, 1803, de Guyane française, de Bolivie, d'Argentine, du Surinam et du Brésil (Pisces, Siluriformes, Callichthyidae). *Revue Française d'Aquariologie et Herpetologie*, 10, 73–84.
- Nijssen, H. & Isbrücker, I.J.H. (1985) *Lasiancistrus scolymus*, a new species of mailed catfish from Rio Aripuanã, Est. Mato Grosso do Sul, Brazil (Pisces, Siluriformes, Loricariidae). *Bijdragen tot de Dierkunde*, 55, 242–248.
- Nijssen, H. & Isbrücker, I.J.H. (1986a) Cinq espèces nouvelles de poissons-chats cuirassés du genre *Corydoras* Lacepede, 1803, du Pérou et de l'Equateur (Pisces, Siluriformes, Callichthyidae). *Revue Française d'Aquariologie et Herpetologie*, 12, for 1985, 65–76.
- Nijssen, H. & Isbrücker, I.J.H. (1986b) Review of the genus *Corydoras* from Peru and Ecuador (Pisces, Siluriformes, Callichthyidae). *Studies on Neotropical Fauna and Environment*, 21, 1–68.
- Nijssen, H. & Isbrücker, I.J.H. (1987) *Spectracanthicus murinus*, nouveaux genre et espèce de poisson-chat cuirassé du Rio Tapajós, Est. Pará, Brésil, avec des remarques sur d'autres genres de Loricariidés (Pisces, Siluriformes, Loricariidae). *Revue Française d'Aquariologie et Herpetologie*, 13, for 1986, 93–98.
- Nijssen, H. & Isbrücker, I.J.H. (1988) Trois nouvelles espèces du genre *Aristoloricaria* de Colombie et du Pérou, avec illustration du dimorphisme sexuel secondaire des lèvres de *A. condei* (Pisces, Siluriformes, Loricariidae). *Revue Française d'Aquariologie et Herpetologie*, 15, 33–38.
- Nijssen, H. & Isbrücker, I.J.H. (1990) *Lithoxus stocki*, a species new to science of ancistrin loricariid catfish from the Maroni River drainage, with a comparison of the primary type specimens of the six species of *Lithoxus* (syn. *Paralithoxus*) (Pisces, Siluriformes, Loricariidae). *Bijdragen tot de Dierkunde*, 60, 327–333.
- Nikolsky, G. & Soin, S.G. (1948) [On catfishes (family Siluridae) in the Amur basin]. *Doklady Akademii nauk SSSR / Comptes Rendus de l'Académie des Sciences de l'URSS*, Moscow, 59, 1357–1360. [In Russian]
- Nolf, D. (1981) Revision des types d'otolithes de poissons fossiles décrits par R. Schubert. *Verhandlungen der Geologischen Bundesanstalt*, 1981, 133–183.
- Norman, J.R. (1922) Four new fishes from Tanganyika Territory. *Annals and Magazine of Natural History (Ser. 9)*, 9, 686–688.
- Norman, J.R. (1923a) Three new fishes from Yunnan, collected by Prof. J. W. Gregory, F.R.S. *Annals and Magazine of Natural History (Ser. 9)*, 11, 561–563.
- Norman, J.R. (1923b) Two new siluroid fishes from the Ivory Coast, collected by Mr. Willoughby P. Lowe. *Annals and Magazine of Natural History (Ser. 9)*, 11, 582–583.
- Norman, J.R. (1923c) A new cyprinoid fish from Tanganyika Territory, and two new fishes from Angola. *Annals and Magazine of Natural History (Ser. 9)*, 12, 694–696.
- Norman, J.R. (1925a) A new siluroid fish of the genus *Clarias* from southwestern Uganda. *Occasional Papers of the*

- Boston Society of Natural History, 5, 189–190, pl. 11.
- Norman, J.R. (1925b) Two new fishes from Tonkin, with notes on the siluroid genera *Glyptosternum*, *Exostoma*, etc. *Annals and Magazine of Natural History (Ser. 9)*, 15, 570–575.
- Norman, J.R. (1926a) A new catfish of the genus *Cetopsis* from the Rio das Velhas, Brazil. *Annals and Magazine of Natural History (Ser. 9)*, 17, 116.
- Norman, J.R. (1926b) Descriptions of nine new freshwater fishes from French Guiana and Brazil. *Annals and Magazine of Natural History (Ser. 9)*, 18, 91–97.
- Norman, J.R. (1926c) A new blind catfish from Trinidad, with a list of the blind cave-fishes. *Annals and Magazine of Natural History (Ser. 9)*, 18, 324–331.
- Norman, J.R. (1928a) Two new fishes from Lake Victoria. *Annals and Magazine of Natural History (Ser. 10)*, 2, 104–106.
- Norman, J.R. (1928b) A new siluroid fish of the genus *Chrysichthys* from Nigeria. *Annals and Magazine of Natural History (Ser. 10)*, 2, 385.
- Norman, J.R. (1932) A collection of fishes from Sierra Leone. *Annals and Magazine of Natural History (Ser. 10)*, 10, 180–185.
- Norman, J.R. (1935a) A collection of fishes from the Ashanti Forest, Gold Coast. *Annals and Magazine of Natural History (Ser. 10)*, 15, 215–221.
- Norman, J.R. (1935b) Description of a new loricariid catfish from Ecuador. *Annals and Magazine of Natural History (Ser. 10)*, 15, 627–629.
- Norris, S.M. (2002) A revision of the African electric catfishes, family Malapteruridae (Teleostei, Siluriformes), with erection of a new genus and descriptions of fourteen new species, and an annotated bibliography. *Musée Royal de l'Afrique Centrale, Annales, Sciences Zoologiques*, 289, 1–155.
- Ogilby, J.D. (1898a) New genera and species of fishes. *Proceedings of the Linnean Society of New South Wales*, 23, 32–41.
- Ogilby, J.D. (1898b) New genera and species of fishes [continued from p. 41]. *Proceedings of the Linnean Society of New South Wales*, 23, 280–299.
- Ogilby, J.D. (1899) Contribution to Australian ichthyology. *Proceedings of the Linnean Society of New South Wales*, 24, 154–186.
- Ogilby, J.D. (1908) New or little known fishes in the Queensland Museum. *Annals of the Queensland Museum*, 9, 1–41.
- Ogilby, J.D. (1910) On new or insufficiently described fishes. *Proceedings of the Royal Society of Queensland*, 23, 1–55.
- Okada, Y. & Kubota, S.S. (1957) Description of a new fresh-water catfish, *Coreobagrus ichikawai*, with an emendation of the genus *Coreobagrus* Mori. *Japanese Journal of Ichthyology*, 5, 143–145.
- Oken, L. (1817) Cuviers und Okens Zoologien neben einalder gestellt. *Isis (Oken)*, 8, col.1145–1184 [incl.1779–1782, sic. 1179–1182].
- Olazarri, J., Mones, A., Ximénez, A. & Philippi, M.E. (1970) Lista de los ejemplares-tipo depositados en el Museo Nacional de Historia Natural de Montevideo, Uruguay. *Comunicaciones Zoológicas del Museo de Historia Natural de Montevideo*, 10, 1–12.
- Oliveira, J.C. de. (1997) Redescrição de *Hemipsilichthys garbei* Ihering, 1911, com designação de lectótipo e comentários sobre a sua distribuição e da de *Hemipsilichthys gobio* (Lütken, 1874) (Teleostei: Siluriformes: Loricariidae). *Papeis Avulsos de Zoologia, São Paulo*, 40, 113–126.
- Oliveira, J.C. de, & Britski, H.A. (2000) Redescrição de *Taunayia bifasciata* (Eigenmann & Norris, 1900), comb. nova, um bagre enigmático do Estado de São Paulo (Siluriformes, Pimelodidae, Heptapterinae). *Papeis Avulsos de Zoologia, São Paulo*, 41, 119–133.
- Oliveira, J.C. de, & de Moraes, D.F., Jr. (1997) Presença de *Hoplosternum* Gill, 1858 (Teleostei, Siluriformes, Callichthyidae) nas bacias dos rios São Francisco, Paraíba do Sul e Alto Paraná: primeiro registro e comentários. *Boletim do Museu Nacional do Rio de Janeiro, Nova Série, zoologia*, 383, 1–8.
- Oliveira, J.C. de, & de Moraes, D.F., Jr. (1997) Dados adicionais à descrição de *Steindachneridion parahybae* (Steindachner, 1876) (Teleostei, Siluroidei, Pimelodidae). *Boletim do Museu Nacional do Rio de Janeiro, Nova Série, zoologia*, 384, 1–11.
- Oliveira, J.C. de, & Oyakawa, O.T. (1999) Two new species of *Hemipsilichthys* (Teleostei: Loricariidae) from Serra do Espinhaço, Minas Gerais, Brazil. *Ichthyological Exploration of Freshwaters*, 10, 73–80.
- Oliviera, J.C. de, Vari, R.P. & Ferraris, C.J., Jr. (2001) A new species of “whale catfish” (Siluriformes, Cetopsidae) from the western portions of the Amazon basin. *Proceedings of the Biological Society of Washington*, 114, 574–578.
- Orcés V., G. (1960) Peces ecuatorianos de la familia Callichthyidae, con la descripción de una especie nueva. *Ciencia y*

- Naturaleza (Quito)*, 3, 2–6, 1 pl.
- Orcés V., G. (1961) Hallazgo de peces de los géneros *Xiliphius* y *Hoplomyzon* en el sistema del Amazonas. Descripción de una nueva especie. *Ciencia y Naturaleza (Quito)*, 4, 3–6.
- Orcés V., G. (1962) Dos nuevos peces del género *Xyliphius*. *Ciencia y Naturaleza (Quito)*, 5, 50–54, 1 pl.
- Orcés V., G. (1977) Contribuciones al conocimiento de los peces del Ecuador. I.— Especies de la subfamilia Sorubiminae. *Politécnica, Revista de Información Técnico-científica*, Quito, 3, for 1976–1977, 76–91.
- Orton, J. (1871) *Contributions to the Natural History of the Valley of Quito*. McCalla & Stavely, Philadelphia.
- Oshima, M. (1919) Contributions to the study of the fresh water fishes of the island of Formosa. *Annals of the Carnegie Museum*, 12, 169–328, pls. 48–53.
- Oshima, M. (1926) Notes on a collection of fishes from Hainan, obtained by Prof. S. F. Light. *Annotationes Zoologicae Japonenses*, 11, 1–25.
- Osório, B. (1909) Peixes d'água doce da Guiné Portuguesa. *Memorias do Museu Bocage, Lisboa*, 1, 95–107, pl. 1.
- Ovchynnyk, M.M. (1971) Unrecorded and new species of fishes from fresh waters of Ecuador. *Zoologischer Anzeiger*, 187, 82–122.
- Owen, R. (1853) *Descriptive Catalogue of the Osteological Series Contained in the Museum of the Royal College of Surgeons of England, Vol. 1.—Pisces, Reptilia, Aves, Marsupialia*, London, 350 p.
- Oyakawa, O.T. (1993) Cinco espécies novas de *Harttia* Steindachner, 1876 da região sudeste do Brasil, de comentários sobre o gênero (Teleostei, Siluriformes, Loricariidae). *Comunicações do Museu de Ciências da PUCRS, Série Zoológia, Porto Alegre*, 6, 3–27.
- Oyakawa, O.T., Akama, A. & Zanata, A.M. (2005) Review of the genus *Hypostomus* Lacépède, 1803 from rio Ribeira de Iguaçu basin, with description of a new species (Pisces, Siluriformes, Loricariidae). *Zootaxa*, 921, 1–27.
- Paepke, H.-J. (1999) *Bloch's Fish Collection in the Museum für Naturkunde der Humboldt Universität zu Berlin: an Illustrated Catalog and Historical Account [Theses Zoologicae, 32]*, A. R. G. Gantner Verlag, KG, Ruggell, Liechtenstein, 216 p., 32 pl.
- Page, L.M., Armbruster, J.W. & Sabaj, M.H. (1996) Redescription of *Glyptoperichthys scrophus*, a loricariid catfish from Peru. *Ichthyological Exploration of Freshwaters*, 7, 185–191.
- Page, L.M. & Burr, B.M. (1991) *A field guide to freshwater fishes: North America north of Mexico. The Peterson Field Guide Series Vol. 42*, Houghton Mifflin Co., Boston, xii + 432 p.
- Pallas, P.S. (1787) Pisium novae species descriptae. *Nova Acta Academiae Scientiarum Imperialis Petropolitanae*, 1, for 1783, 347–360, pls. 9–11. [Cover of journal lists date of publication as 1787; Agassiz (1854), lists date as 1783, Dean (1916) says 1788, AMNH library says vol. 1 published in 1783]
- Pappenheim, P. (1911) Zoologische Ergebnisse der Expedition des Herrn G. Tessmann nach Süd-Kamerun und Spanisch-Guinea.—Fische. *Mitteilungen aus dem Zoologischen Museum in Berlin*, 5, 505–528.
- Pappenheim, P. & Boulenger, G.A. (1914) Fische. In: Schubotz, H. (Ed.), *Wissenschaftliche Ergebnisse der Deutschen Zentral-Afrika Expedition, 1907–1908*. Vol. 5 (Zoologie III), Klinkhardt & Biermann, Leipzig, pp. 225–260, pls. 1–10. [In 8 vols. 1912–1914.]
- Parameswaran, S., Selvaraj, C. & Radhakrishnam, S. (1967) A review of the Indian freshwater fishes of the genus *Ompok* Lacépède. *Journal of the Zoological Society of India*, 19, 89–98.
- Parisi, B.M. (2003) *Contribution à L'Etude de la Taxonomie du Groupe de Poissons-chats Néotropicaux Calophysus-Pimelodus (Pimelodidae, Siluriformes) et Hypothèses Phylogénétiques*, Doctoral dissertation, Muséum National d'Histoire Naturelle, Paris.
- Patterson, C. (1993) Osteichthyes: Teleostei. In: Benton, M.J. (Ed.), *The Fossil Record*, Vol. 2, Chapman & Hall, London.
- Paugy, D. (1987) Description de deux nouvelles espèces de *Synodontis* du bassin du Konkouré (Guinée), *S. dekimpei* et *S. levequei* (Pisces, Mochokidae). *Cybium (3e série)*, 11, 357–364.
- Paugy, D. & Bénech, V. (1989) Les poissons d'eau douce des bassins côtiers du Togo (Afrique de l'Ouest). *Revue d'Hydrobiologie Tropicale*, 22, 295–316.
- Paugy, D. & Roberts, T.R. (1992) Mochokidae. In: Lévêque, C., Paugy, D. & Teugels, G.G. (Eds.), *Faune des Poissons d'Eaux Douces et Saumâtres de l'Afrique de l'Ouest Vol. 2*, Musée Royal de l'Afrique Centrale, Tervuren, and Editions de l'ORSTOM, Paris, pp. 500–563.
- Paxton, J.R., Hoese, D.F., Allen, G.R. & Hanley, J.E. (1989) *Zoological Catalogue of Australia. Volume 7.—Pisces. Petromyzontidae to Carangidae*, Australian Government Publishing Service, Canberra, xii + 665 p.
- Pearson, N.E. (1924) The fishes of the eastern slope of the Andes. I.—The fishes of the Rio Beni basin, Bolivia, collected by the Mulford expedition. *Indiana University Studies*, 11, 1–83, pls. 1–12.

- Pearson, N.E. (1937) The fishes of the Atlantic and Pacific slopes near Cajamarca, Peru. *Proceedings of the California Academy of Sciences, Series 4*, 23, 87–98, pls. 12–13.
- Pellegrin, J. (1900) Poissons nouveaux ou rares du Congo français. *Bulletin du Muséum d'Histoire Naturelle, Paris*, 6, 177–182.
- Pellegrin, J. (1901) Poissons nouveaux ou rares du Congo français. *Bulletin du Muséum d'Histoire Naturelle, Paris*, 7, 328–332.
- Pellegrin, J. (1906a) Collections recueillies par M. E. Haug dans l'Ogôoué (Poissons). *Bulletin du Muséum d'Histoire Naturelle, Paris*, 12, 467–471.
- Pellegrin, J. (1906b) Poissons nouveaux du Soudan. *Bulletin du Muséum d'Histoire Naturelle, Paris*, 12, 472–474.
- Pellegrin, J. (1907a) Siluridé nouveau du Fouta-Djalon. *Bulletin du Muséum National d'Histoire Naturelle*, 13, 23–25.
- Pellegrin, J. (1907b) Poissons du sud Cameroun, recueillis par la mission Cottes. *Bulletin du Muséum National d'Histoire Naturelle*, 13, 319–322.
- Pellegrin, J. (1908) Description de deux poissons nouveaux de l'Amérique du Sud, de la famille des Loricariidés. *Bulletin de la Société Zoologique de France*, 33, 125–127.
- Pellegrin, J. (1909a) Collections recueillies par M. E. Haug dans l'Ogôoué (Poissons). *Bulletin du Muséum National d'Histoire Naturelle*, 15, 66–68.
- Pellegrin, J. (1909b) Poissons de la Komadougou et du lac Tchad récoltés par la mission Tilho-Gaillard. *Bulletin du Muséum National d'Histoire Naturelle*, 15, 240–245.
- Pellegrin, J. (1909c) Mission géodésique de l'Équateur. Collections recueillies par M. le Dr. Rivet. Description de deux poissons nouveaux de la famille des Loricariidae. *Bulletin du Muséum National d'Histoire Naturelle*, 15, 517–519.
- Pellegrin, J. (1909d) Note complémentaire sur une seconde collections de poissons recueillie par M. E. Haug, à Ngomo (Ogôoué). *Bulletin de la Société Philomathique (Ser. 10)*, 1, 45–53.
- Pellegrin, J. (1909e) Les poissons du genre *Vandellia* C. V. *Bulletin de la Société Philomathique (Ser. 10)*, 1, 197–204.
- Pellegrin, J. (1911) Poissons de l'Equateur recueillis par M. le Dr. Rivet. In: *Mission du Service Géographique de l'Armée pour la Mesure d'un Arc de Méridien Équatorial en Amérique du Sud sous le Contrôle Scientifique de l'Académie des Sciences, 1899–1906, Tome 9 (Zoologie), Fasc. 2 (Reptiles, Poissons, Batraciens)*, Ministère de l'Instruction Publique, Paris, pp. 1–15, pl. 1.
- Pellegrin, J. (1912) Description d'un poisson nouveau de l'Orénoque appartenant au genre *Xenocara*. *Bulletin de la Société Zoologique de France*, 37, 271–272.
- Pellegrin, J. (1913a) Poissons nouveaux de Guinée française recueillis par M. Pobéguin. *Bulletin de la Société Zoologique de France*, 38, 236–241.
- Pellegrin, J. (1913b) Poissons nouveaux de l'Ogôoué recueillis par Mr. Ernest Haug. *Bulletin de la Société Zoologique de France*, 38, 272–275.
- Pellegrin, J. (1914) Poissons nouveaux du haut Zambèze recueillis par M. V. Ellenberger. *Bulletin de la Société Zoologique de France*, 39, 24–28.
- Pellegrin, J. (1919a) Poissons nouveaux du Mozambique. *Bulletin de la Société Zoologique de France*, 44, 397–401.
- Pellegrin, J. (1919b) Poissons du Gribingui recueillis par M. Baudon. Description de sept espèces nouvelles. *Bulletin de la Société Zoologique de France*, 44, 201–214.
- Pellegrin, J. (1920) Poissons des lagunes de la Côte d'Ivoire. Description de deux espèces nouvelles. *Bulletin de la Société Zoologique de France*, 45, 115–121.
- Pellegrin, J. (1922a) Poissons de l'Oubanghi-Chari recueillis par M. Baudon. Description d'un genre, de cinq espèces et d'une variété. *Bulletin de la Société Zoologique de France*, 47, 64–76.
- Pellegrin, J. (1922b) Poissons du Gribingui recueillis par M. Baudon. Description d'un Mormyridé et d'un Characiniidé nouveaux. *Bulletin de la Société Zoologique de France*, 47, 220–223.
- Pellegrin, J. (1922c) Poissons nouveaux de l'Afrique orientale. *Bulletin du Muséum National d'Histoire Naturelle*, 28, 349–351.
- Pellegrin, J. (1922d) Poissons nouveaux ou rares du musée du Congo. *Revue de Zoologie Africaine*, 10, 272–280.
- Pellegrin, J. (1923a) Présentation d'un crâne de *Clarias* géant du Niger. *Bulletin du Muséum National d'Histoire Naturelle*, 29, 211–213.
- Pellegrin, J. (1923b) *Les Poissons des Eaux Douces de l'Afrique Occidentale (du Sénégal au Niger)*. Publications du Comité d'Etudes Historiques et Scientifiques, Emile Larose, Paris, 373 p.
- Pellegrin, J. (1924a) Poissons du Niger recueillis par M. Jean Thomas. Description de deux espèces nouvelles. *Bulletin du Muséum National d'Histoire Naturelle*, 30, 457–463.
- Pellegrin, J. (1924b) Description d'un poisson nouveau du Gabon appartenant au genre *Synodontis*. *Bulletin de la Société*

- Zoologique de France*, 49, 320–322.
- Pellegrin, J. (1924c) Description d'un Siluridé nouveau récolté au Congo Belge. *Revue de Zoologie Africaine*, 12, 487–489.
- Pellegrin, J. (1925) Poissons du Nord du Gabon et de la Sangha recueillis par M. Baudon. Description de deux espèces et d'une variété nouvelles. *Bulletin de la Société Zoologique de France*, 50, 97–106.
- Pellegrin, J. (1926) Description de Siluridés, d'un Cyprinodontidé et d'un Tétrodontidé récoltés au Congo Belge par le Dr. Schouteden. *Revue de Zoologie Africaine*, 14, 201–208.
- Pellegrin, J. (1927) Description d'un Siluridé nouveau du Cameroun appartenant au genre *Synodontis*. *Bulletin de la Société Zoologique de France*, 52, 365–366.
- Pellegrin, J. (1928a) Sur une collection de poissons du Cameroun recueillie par M. Chamaulte. *Annales. Société des Sciences Naturelles de la Charente-Inférieure (nouv. sér)*, 1, 1–11.
- Pellegrin, J. (1928b) Poisson du Kasai (Congo Belge). Description d'un genre nouveau et de quatre espèces nouvelles. *Bulletin de la Société Zoologique de France*, 53, 103–113.
- Pellegrin, J. (1928c) Description d'un poisson nouveau du Gabon appartenant au genre *Amphilius*. *Bulletin de la Société Zoologique de France*, 53, 453–454.
- Pellegrin, J. (1929a) Mission Saharienne Augiéras-Draper, 1927–1928. Poissons. *Bulletin du Muséum National d'Histoire Naturelle (2^e Série)*, 1, 134–139.
- Pellegrin, J. (1929b) Siluridés, Cyprinodontidés, Acanthoptérygiens du Cameroun recueillis par M. Th. Monod; description de cinq espèces et deux variétés nouvelles. *Bulletin de la Société Zoologique de France*, 54, 358–369.
- Pellegrin, J. (1929c) Siluridé et Cyprinodontidé nouveaux du Gabon recueillis par M. A. Bandon. *Bulletin de la Société Zoologique de France*, 54, 640–643.
- Pellegrin, J. (1930) Poissons de l'Ogôoué, du Kouilou, de l'Alima et de la Sangha recueillis par M. A. Baudon; description de cinq espèces et cinq variétés nouvelles. *Bulletin de la Société Zoologique de France*, 55, 196–210.
- Pellegrin, J. (1931a) Poissons du Kouilou et de la Nyanga recueillis par M. A. Baudon. *Bulletin de la Société Zoologique de France*, 56, 205–211.
- Pellegrin, J. (1931b) Description d'un poisson nouveau de l'Equateur appartenant à la famille des Loricariidés. *Revue suisse de Zoologie, Annales de la Société zoologique suisse et du Muséum d'Histoire naturelle de Genève*, 38, 113–115.
- Pellegrin, J. (1932) Description d'un *Chrysichthys* géant du Congo. *Bulletin du Muséum National d'Histoire Naturelle (2^e Série)*, 4, 165–168.
- Pellegrin, J. (1933a) Voyage de Ch. Alluaud et P. A. Chappuis en Afrique occidentale Française (Dec. 1930–Mars 1931). IV.—Poissons. *Archiv für Hydrobiologie*, 26, 101–120.
- Pellegrin, J. (1933b) Poissons de la région du Kivu adressés par M. Guy Babault. *Bulletin de la Société Zoologique de France*, 58, 169–175.
- Pellegrin, J. (1935) Poissons de Guinée Française recueillis par M. Waterlot; description d'une espèce et de deux variétés nouvelles. *Bulletin de la Société Zoologique de France*, 60, 462–466.
- Pellegrin, J. (1936a) Contribution à l'Ictyologie de l'Angola. *Arquivos do Museu Bocage, Lisboa*, 7, 45–62.
- Pellegrin, J. (1936b) Poissons nouveaux du haut-Laos et de l'Annam. *Bulletin de la Société Zoologique de France*, 61, 243–248.
- Pellegrin, J. (1938a) Poissons de l'Afrique équatoriale Française de Jean Thomas. *Bulletin de la Société Zoologique de France*, 63, 369–378.
- Pellegrin, J. (1938b) Descrizione d'un Siluride nuovo del Giuba, appartenente al genere *Clarotes*. *Bollettino di Pesca, Piscicoltura e Idrobiologia*, 14, 218–222.
- Pellegrin, J. & Chevey, P. (1937) Poissons d'Indochine recueillis par MM. J. Delacour et Lowe; description d'une espèce nouvelle. *Bulletin de la Société Zoologique de France*, 62, 313–318.
- Pellegrin, J. & Fang, P.W. (1940) Poissons de Chine de M. Ho, description de deux espèces nouvelles. *Bulletin de la Société Zoologique de France*, 64, 338–343.
- Peng, Z.G., He, S.P. & Zhang, Y.G. (2002) Mitochondrial cytochrome b sequence variation and phylogeny of the East Asian bagrid catfishes. *Progress in Natural Science*, 12, 421–425.
- Peng, Z.G., He, S.P. & Zhang, Y.G. (2004) Phylogenetic relationships of glyptosternoid fishes (Siluriformes: Sisoridae) inferred from mitochondrial cytochrome b gene sequences. *Molecular Phylogenetics and Evolution*, 31, 979–987.
- Peng, Z.G., Zhang, Y.G., He, S.P. & Chen, Y.Y. (2005) Phylogeny of Chinese catfishes inferred from mitochondrial cytochrome b sequences. *Acta Genetica Sinica*, 32, 145–154.
- Perdices, A., Bermingham, E., Montilla, A. & Doadrio, I. (2002) Evolutionary history of the genus *Rhamdia* (Teleostei:

- Pimelodidae) in Central America. *Molecular Phylogenetics and Evolution*, 25, 172–189.
- Pereira, E.H.L. (2005) Resurrection of *Pareiorhaphis* Miranda Ribeiro, 1918 (Teleostei: Siluriformes: Loricariidae), and description of a new species from the rio Iguaçu basin, Brazil. *Neotropical Ichthyology*, 3, 271–276.
- Pereira, E.H.L., Oliveira, J.C. & Oyakawa, O.T. (2000) *Hemipsilichthys papillatus*, a new species of loricariid catfish (Teleostei: Siluriformes) from Minas Gerais, Brazil. *Ichthyological Exploration of Freshwaters*, 11, 377–383.
- Pereira, E.H.L. & Oyakawa, O.T. (2003) *Isbrueckerichthys epakmos*, a new species of loricariid catfish from the rio Ribeira de Iguape basin, Brazil (Teleostei, Siluriformes). *Neotropical Ichthyology*, 1, 3–9.
- Pereira, E.H.L. & Reis, R.E. (1992) *Hemipsilichthys vestigipinnis* sp. n. (Teleostei, Siluriformes) a new loricariid catfish from the rio Uruguay basin, southern Brazil. *Revue Française d'Aquariologie et Herpetologie*, 18, for 1991, 111–116.
- Pereira, E.H.L. & Reis, R.E. (2002) Revision of the loricariid genera *Hemipsilichthys* and *Isbrueckerichthys* (Teleostei: Siluriformes), with descriptions of five new species of *Hemipsilichthys*. *Ichthyological Exploration of Freshwaters*, 13, 97–146.
- Pereira, E.H.L., Reis, R.E., Souza, P.F.M. & Lazzarotto, H. (2003) A new species of the loricariid catfish genus *Hemipsilichthys* from southern Rio de Janeiro coastal rivers, southeastern Brazil (Teleostei: Siluriformes). *Zootaxa*, 285, 1–10.
- Pereira, S.M. (1984) Siluriformes (Osteichthyes, Teleostei) del Terciario tardio de los alrededores de Paraná (Entre Ríos). *Jorn. Argent. Paleontol. Vertebr., Res*, 1, 2.
- Pérez, A. & Provenzano R, F. (1996) *Cordylancistrus perijae*, a new species of armored catfish (Siluroidei: Loricariidae) from the Maracaibo basin, Venezuela. *Studies on Neotropical Fauna and Environment*, 31, 27–34.
- Pérez, A. & Viloria, A. (1994) *Ancistrus galani* n. sp. (Siluriformes: Loricariidae), with comments on biospeleological explorations in western Venezuela. *Mémoires de Biospéologie*, 21, 103–107.
- Perugia, A. (1891) Appunti sopra alcuni pesci sud-americani conservati nel Museo Civico di Storia Naturale di Genova. *Annali del Museo Civico de Storia Naturale di Genova (Ser. 2a)*, 10, 605–657.
- Perugia, A. (1892) Intorno ad alcuni pesci raccolti al Congo dal Capitano Giacomo Bove. *Annali del Museo Civico de Storia Naturale di Genova (Ser. 2a)*, 10, 967–977.
- Perugia, A. (1894) Viaggio di Lamberto Loria nella Papuasia orientale. XIII.— Pesci d'acqua dolce. *Annali del Museo Civico de Storia Naturale di Genova (Ser. 2a)*, 14, 546–553.
- Perugia, A. (1897) Di alcuni pesci raccolti in Bolivia dal Prof. Luigi Balzan. *Annali del Museo Civico de Storia Naturale di Genova (Ser. 2a)*, 18, 16–27.
- Peters, W. (1852) Diagnosen von neuen Flussfischen aus Mossambique. *Monatsberichte der Königlichen Preussische Akademie des Wissenschaften zu Berlin*, 1852, 275–276, 681–685.
- Peters, W. (1861) Über zwei neue Gattungen von Fischen aus dem Ganges. *Monatsberichte der Königlichen Preussische Akademie des Wissenschaften zu Berlin*, 1861, 712–713.
- Peters, W. (1868a) Über eine neue Nagergattung, *Chiropodomys penicillatus*, so wie über einige neue oder weniger bekannte Amphibien und Fische. *Monatsberichte der Königlichen Preussische Akademie des Wissenschaften zu Berlin*, 1868, 448–460, pl. 2.
- Peters, W. (1868b) Ueber eine von dem Baron Carl von der Decken entdeckte neue Gattung von Welsen, *Chiloglanis deckenii*, und einige andere Süßwasserfische aus Ostafrika. *Monatsberichte der Königlichen Preussische Akademie des Wissenschaften zu Berlin*, 1868, 598–602, pl. 2.
- Peters, W. (1868c) *Flussfische. Naturwissenschaftliche Reise nach Mossambique auf befehl Seiner Mäjestat des Königs Friedrich Wilhelm IV, in den Jahren 1842 bis 1848 Ausgeführt. Zoologie, Vol. IV*, G. Reimer, Berlin, xii + 116 p., 20 pls.
- Peters, W. (1877) Über die von Dr. C. Sachs in Venezuela gesammelten Fische. *Monatsberichte der Königlichen Preussische Akademie des Wissenschaften zu Berlin*, 1877, 469–473.
- Peters, W. (1881a) Über vier neue Fische. *Sitzungsberichte der Gesellschaft Naturforschender Freunde zu Berlin*, 1881, 17–19.
- Peters, W. (1881b) Über eine Sammlung von Fischen, welche Hr. Dr. Gerlach in Hongkong gesandt hat. *Monatsberichte der Königlichen Preussische Akademie des Wissenschaften zu Berlin*, 1880, 1029–1037, 1 pl.
- Peters, W. (1882) Über drei neue Arten von *Mormyrus* aus Ost- und Westafrika, und eine Art von *Clarias* aus Westafrika. *Sitzungsberichte der Gesellschaft Naturforschender Freunde zu Berlin*, 1882, 72–74.
- Pethiyagoda, R. & Bahir, M.M. (1998) *Heteropneustes microps*, a junior synonym of *H. fossilis* (Osteichthyes: Heteropneustidae). *Journal of South Asian Natural History*, 3, 113–114.
- Pethiyagoda, R. & Kottelat, M. (1994) Three new species of fishes of the genera *Osteochilichthys* (Cyprinidae), *Travan-*

- coria* (Balitoridae) and *Horabagrus* (Bagridae) from the Chalakudy River, Kerala, India. *Journal of South Asian Natural History*, 1, 97–116.
- Pethon, P. (1969) List of type specimens of fishes, amphibians and reptiles in the Zoological Museum, University of Oslo. *Rhizocrinus, Occasional Papers of the Zoological Museum, University of Oslo*, 1, 1–17.
- Peyer, B. (1928) Ergebnisse der Forschungsreisen Prof. E. Stromers in den Wüsten Ägyptens. V.—Tertiäre Wirbeltiere. 2. Die Welse des ägyptischen Alttertiärs nebst einer kritischen Übersicht über alle fossilen Welse. *Abhandlungen der Bayerischen Akademie der Wissenschaften, Mathematisch-Naturwissenschaftliche Abteilung*, 32, 1–62, pls. 1–6.
- Pfaff, J.R. (1933) Report on the fishes collected by Mr. Harry Madsen during Professor O. Olufsen's Expedition to French Sudan in the years 1927–28. *Videnskabelige Meddelelser fra Dansk Naturhistorisk Forening i København*, 94, 273–315, pl. 6.
- Pfeffer, G.J. (1889) Übersicht der von Herrn Dr. Franz Stuhlmann in Ägypten, auf Sanzibar und dem gegenüberliegenden Festlande gesammelten Reptilien, Amphibien, Fische, Mollusken und Krebse. *Jahrbuch der Hamburgischen Wissenschaftlichen Anstalten*, 6, 1–36.
- Pfeffer, G.J. (1896) Die Fische Ost-Afrikas. In: Möbius, K. (Ed.), *Die Thierwelt Ost-Afrikas und der Nachbargebiete, Vol. 1, Lief. V*. D. Reimer, Berlin, pp. xviii + 72 p.
- Pfeiffer, W. & Eisenberg, J.F. (1965) Die lauterzeugung der dornwelse (Doradidae) und der fiederbartwelse (Mochokidae). *Zeitschrift für Morphologie und Ökologie der Tiere*, 54, 669–679.
- Philippi, R.A. (1866) Bemerkungen über die chilischen Flussfische. *Monatsberichte der Königlichen Preussische Akademie des Wissenschaften zu Berlin*, 1866, 708–717.
- Pietschmann, V. (1913a) Eine neue *Glyptosternum*-Art aus dem Tigris. *Anzeiger der Mathematisch- Naturwissenschaftlichen Classe der Kaiserlichen Akademie der Wissenschaften in Wien*, 1913, 93–95.
- Pietschmann, V. (1913b) Fische des Wiesbadener Museums. *Jahrbücher des Nassauischen Vereins für Naturkunde, Wiesbaden*, 66, 170–201, pls. 1–2.
- Pietschmann, V. (1932) Ein neuer Wels aus dem Nil. *Zoologischer Anzeiger*, 100, 92–94.
- Pietschmann, V. (1939) Matériali zoologici dell' Eritrea raccolti da G. Müller durante la spedizione dell' istituto sieroterapico milanese e conservati al museo di Trieste. Parte VI.—Pisces. *Atti del Museo Civico di Storia Naturale di Trieste*, 14, 179–186.
- Pinna, M.C.C. de. (1988) A new genus of trichomycterid catfish (Siluroidei, Glanapteryginae), with comments on its phylogenetic relationships. *Revue suisse de Zoologie, Annales de la Société zoologique suisse et du Muséum d'Histoire naturelle de Genève*, 95, 113–128.
- Pinna, M.C.C. de. (1989a) A new sarcoglanidine catfish, phylogeny of its subfamily, and an appraisal of the phyletic status of the Trichomycterinae (Teleostei, Trichomycteridae). *American Museum Novitates*, 2950, 1–39.
- Pinna, M.C.C. de. (1989b) Redescription of *Glanapteryx anguilla*, with notes on the phylogeny of Glanapteryginae (Siluriformes, Trichomycteridae). *Proceedings of the Academy of Natural Sciences, Philadelphia*, 141, 361–374.
- Pinna, M.C.C. de. (1992a) *Trichomycterus castroi*, a new species of trichomycterid catfish from the Rio Iguaçu of South-eastern Brazil (Teleostei: Siluriformes). *Ichthyological Exploration of Freshwaters*, 3, 89–95.
- Pinna, M.C.C. de. (1992b) A new subfamily of Trichomycteridae (Teleostei, Siluriformes), lower loricarioid relationships and a discussion on the impact of additional taxa for phylogenetic analysis. *Zoological Journal of the Linnean Society*, 106, 175–229.
- Pinna, M.C.C. de. (1993) *Higher-level Phylogeny of Siluriformes (Teleostei: Ostariophysi), with a New Classification of the Order*; Unpublished Ph.D. dissertation, City University of New York, New York.
- Pinna, M.C.C. de. (1996) A phylogenetic analysis of the Asian catfish families Sisoridae, Akysidae, and Amblycipitidae, with a hypothesis on the relationships of the neotropical Aspredinidae (Teleostei, Ostariophysi). *Fieldiana, Zoology (New Series)*, 84, i–iv + 1–83.
- Pinna, M.C.C. de. (1998a) Phylogenetic relationships of Neotropical Siluriformes (Teleostei: Ostariophysi): Historical overview and synthesis of hypotheses. In: Malabarba, L.R., Reis, R.E., Vari, R.P., Lucena, Z.M. & Lucena, C.A.S. (Eds.), *Phylogeny and Classification of Neotropical Fishes*, Edipucrs, Porto Alegre, pp. 279–330.
- Pinna, M.C.C. de. (1998b) A new species of the catfish genus *Glanapteryx* (Siluriformes: Trichomycteridae). *Proceedings of the Biological Society of Washington*, 111, 35–42.
- Pinna, M.C.C. de. (2003) Nematogenyidae. In: Reis, R.E., Kullander, S.O. & Ferraris, C.J., Jr. (Eds.), *Check list of the Freshwater Fishes of South and Central America*, Edipucrs, Porto Alegre, Brazil, pp. 268–269.
- Pinna, M.C.C. de & Britski, H.A. (1991) *Megalocentor*, a new genus of parasitic catfish from the Amazon basin: the sister group of *Apomatoceros* (Trichomycteridae: Stegophilinae). *Ichthyological Exploration of Freshwaters*, 2, 113–128.

- Pinna, M.C.C. de & Keith, P. (2003) A new species of the catfish genus *Ituglanis* from French Guyana (Osteichthyes; Siluriformes: Trichomycteridae). *Proceedings of the Biological Society of Washington*, 116, 873–882.
- Pinna, M.C.C. de & Ng, H.H. (2004) The second ural centrum in Siluriformes and its implication for the monophyly of superfamily Sisoroidea (Teleostei, Ostariophysii). *American Museum Novitates*, 3437, 1–23.
- Pinna, M.C.C. de & Starnes, W.C. (1990) A new genus and species of Sarcoglanidinae from the Río Mamoré, Amazon Basin, with comments on subfamilial phylogeny (Teleostei, Trichomycteridae). *Journal of Zoology (London)*, 222, 75–88.
- Pinna, M.C.C., de & Vari, R.P. (1995) Monophyly and phylogenetic diagnosis of the family Cetopsidae, with synonymization of the Helogenidae (Teleostei: Siluriformes). *Smithsonian Contributions to Zoology*, 571, i–iii + 1–26.
- Pinna, M.C.C. de & Winemiller, K.O. (2000) A new species of *Ammoglanis* (Siluriformes: Trichomycteridae) from Venezuela. *Ichthyological Exploration of Freshwaters*, 11, 255–264.
- Pinna, M.C.C. de & Wosiacki, W. (2002) A new interstitial catfish of the genus *Listrura* from southern Brazil (Siluriformes: Trichomycteridae: Glanapteryginae). *Proceedings of the Biological Society of Washington*, 114, 720–726.
- Pinna, M.C.C. de & Wosiacki, W. (2003) Trichomycteridae. In: Reis, R.E., Kullander, S.O. & Ferraris, C.J., Jr. (Eds.), *Check list of the Freshwater Fishes of South and Central America*, Edipucrs, Porto Alegre, Brazil, pp. 270–290.
- Pinto, S.Y. & Marzulo, D. (1975) Estudos morfológicos. VI.— Sobre *Lophiosilurus alexandri* Steindachner, 1876. (Actinopterygii, Cypriniformes, Pimelodidae). *Boletim do Museu de História Natural da Universidade Federal de Minas Gerais, Zoologia*, 21, 1–9, 4 pl.
- Piorski, N.M. (1999) Diferenciação morfométrica entre as espécies *Platydoras costatus* (Linnaeus, 1766) e *P. armatus* (Cuvier & Valenciennes, 1840) (Pisces; Siluriformes; Doradidae). *Comunicações do Museu de Ciências da PUCRS, Série Zoologia, Porto Alegre*, 12, 19–30.
- Playfair, R.L. (1867) On the fishes of Cachar. *Proceedings of the Zoological Society of London*, 1867, 14–17, pl. 3.
- Playfair, R.L. & Günther, A. (1867) *The Fishes of Zanzibar, with a List of the Fishes of the Whole East Coast of Africa*, Van Voorst, London, xix + 153 p., 21 pl.
- Poll, M. (1933) Contribution à la faune ichthyologique du Katanga. *Annales du Musée du Congo Belge, Zoologie (Ser. I)*, 3, 101–152.
- Poll, M. (1938) Poissons du Katanga (bassin du Congo) récoltés par le professeur Paul Brien. *Revue de Zoologie et de Botanique Africaines*, 30, 389–423.
- Poll, M. (1941a) Étude systématique et morphologique d'une collection de poissons de l'Uele (Congo Belge), comprenant trois espèces nouvelles. *Bulletin du Musée Royal d'Histoire Naturelle de Belgique*, 17, 1–18.
- Poll, M. (1941b) Poissons nouveaux de la Côte d'Ivoire. *Revue de Zoologie et de Botanique Africaines*, 34, 133–143.
- Poll, M. (1942a) Les poissons du Lac Tumba, Congo belge. *Bulletin du Musée Royal d'Histoire Naturelle de Belgique*, 18, 1–25.
- Poll, M. (1942b) Description d'un genre nouveau de Bagridae du lac Tanganyika. *Revue de Zoologie et de Botanique Africaines*, 35, 318–322.
- Poll, M. (1942c) Description d'un genre nouveau de Clariidae originaire du Congo belge. *Revue de Zoologie et de Botanique Africaines*, 36, 94–100.
- Poll, M. (1943a) Description du *Tanganikallabes mortiauxi*, gen. nov., sp. n., de la famille des Clariidae. *Revue de Zoologie et de Botanique Africaines*, 37, 126–133.
- Poll, M. (1943b) Descriptions de poissons nouveaux du Lac Tanganyika, appartenant aux familles des Clariidae et Cichlidae. *Revue de Zoologie et de Botanique Africaines*, 37, 305–318.
- Poll, M. (1944a) Descriptions de poissons nouveaux recueillis dans la région d'Albertville (Congo belge) par le Dr. G. Pojer. *Bulletin du Musée Royal d'Histoire Naturelle de Belgique*, 20, 1–12.
- Poll, M. (1944b) Description préliminaire d'un Clariidae nouveau observé à Léopoldville par M. L. Tihon. *Revue de Zoologie et de Botanique Africaines*, 38, 79–82, 1 pl.
- Poll, M. (1946) Révision de la faune ichthyologique du Lac Tanganyika. *Annales du Musée du Congo Belge, Zoologie (Ser. I)*, 4, 146–364, pls. 1–3 + map.
- Poll, M. (1948) Poissons recueillis au Katanga par H. J. Bredo. *Bulletin du Musée Royal d'Histoire Naturelle de Belgique*, 24, 1–24.
- Poll, M. (1949) Résultats scientifiques des croisières du Navire-École Belge "Mercator" IV.— Poissons. *Mémoires, Institut Royal des Sciences Naturelles de Belgique, série 2*, 33, 173–269.
- Poll, M. (1952) Poissons de rivières de la région des lacs Tanganyika et Kivu recueillis par G. Marlier. *Revue de Zoologie et de Botanique Africaines*, 46, 221–236.
- Poll, M. (1953) Poissons non Cichlidae. *Exploration Hydrobiologique du Lac Tanganyika (1946–1947)*, Vol. 3 (part 5a),

- Institut Royal des Sciences Naturelles de Belgique, Bruxelles, pp. 251 p., 11 pls.
- Poll, M. (1954) Poissons de forêt des environs de Yangambi (Stanleyville) recueillis par A. Hulot. *Annales du Musée royal du Congo Belge*, (Série in 8°), *Sciences Zoologiques*, 1, 56–68.
- Poll, M. (1957) Redescription du *Gymnallabes tihoni* Poll, 1944, Clariidae microphthalmes du Stanley-Pool. *Revue de Zoologie et de Botanique Africaines*, 55, 237–248.
- Poll, M. (1959) Recherches sur la faune ichthyologique de la région du Stanley Pool. *Annales du Musée royal du Congo Belge*, (Série in 8°), *Sciences Zoologiques*, 71, 75–174, pls. 12–26.
- Poll, M. (1966) Genre et espèce nouveaux de Bagridae du fleuve Congo en région de Léopoldville. *Revue de Zoologie et de Botanique Africaines*, 74, 425–428.
- Poll, M. (1967) Contribution à la faune ichthyologique de l'Angola. *Publicações Culturais, Companhia de Diamantes de Angola*, 75, 1–381, pls. 1–20.
- Poll, M. (1971) Révision des *Synodontis* Africains (Famille Mochocidae). *Annales du Musée Royal de l'Afrique Centrale, Series in 8°, Sciences Zoologiques*, 191, 1–497.
- Poll, M. (1974) Un nouveau *Synodontis* (Pisces Mochocidae) de la rivière Lukenie (République du Zaïre). *Revue de Zoologie Africaine*, 88, 441–444.
- Poll, M. (1976) Poissons. *Exploration Parc National Upemba Mission G. F. de Witte*, 73, 1–127, 43 pls.
- Poll, M. (1977) Les genres nouveaux *Platyallabes* et *Platyclarias* comparés au genre *Gymnallabes* Gthr. Synopsis nouveau des genres de Clariidae. *Bulletin de la Classe des Sciences. Académie Royale de Belgique* (Ser. 5), 63, 122–149.
- Poll, M. & Gosse, J.-P. (1963) Contribution à l'étude systématique de la faune ichthyologique du Congo Central. *Annales du Musée Royal de l'Afrique Centrale, Series in 8°, Sciences Zoologiques*, 116, 43–111, pls. 1–4.
- Poll, M. & Gosse, J.-P. (1969) Révision des Malapteruridae (Pisces, Siluriformes) et description d'une deuxième espèce de silure électrique: *Malapterurus microstoma* sp. n. *Bulletin, Institut royal des Sciences naturelles de Belgique*, 45, 1–12, pls. 1–3.
- Poll, M. & Gosse, J.-P. (1994) Genera des poissons d'eau douce de l'Afrique. *Mémoire de la Classe des Sciences, Académie royale de Belgique*, 9, 1–324.
- Poll, M. & Lambert, J. (1958) Un Cyprinodontide et un Clariide nouveaux de la grande forêt congolaise. *Revue de Zoologie et de Botanique Africaines*, 58, 328–339.
- Poll, M., Lanza, B. & Romoli Sassi, A. (1972) Genre nouveau extraordinaire de Bagridae du fleuve Juba: *Pardiglanis tarabinii* gen. n. sp. n. (Pisces Siluriformes). *Monitore Zoologico Italiano*, N. S., suppl. 4, 327–345.
- Poll, M. & Roberts, T.R. (1968) Description d'une espèce nouvelle de *Synodontis* du Bassin du Congo. *Revue de Zoologie et de Botanique Africaines*, 77, 296–302.
- Poll, M. & Roman, B. (1967) Poissons nouveaux de la Haute Comoé. *Revue de Zoologie et de Botanique Africaines*, 75, 179–187.
- Poll, M. & Stewart, D.J. (1975) Un Mochocidae et un Kneriidae nouveaux de la rivière Luongo (Zambia), affluent du bassin du Congo (Pisces). *Revue de Zoologie et de Botanique Africaines*, 89, 151–158.
- Popta, C.M.L. (1900) A new species of *Arius*. *Notes from the Leyden Museum*, 22, 71–74.
- Popta, C.M.L. (1904) Descriptions préliminaires des nouvelles espèces de poissons recueillies au Bornéo central par M. le Dr. A. W. Nieuwenhuis en 1898 et en 1900. *Notes from the Leyden Museum*, 24, for 1902–04, 179–202.
- Popta, C.M.L. (1906) Résultats ichthyologiques des voyages scientifiques de Monsieur le Professeur Dr. A. W. Nieuwenhuis dans le centre e Bornéo (1898 et 1900). *Notes from the Leyden Museum*, 27, 1–304, 10 pls.
- Popta, C.M.L. (1911) Ueber Fische von Wladiwostok und von Blagoweschtensk a. Amur, gesammelt von Herrn Dr. P. v. Wittenburg. *Jahresheshefte des Vereins für Vaterländische Naturkunde in Württemberg*, 75, 333–353.
- Popta, C.M.L. (1913) *Auchenoglanis büttikoferi* n. sp. from West Africa. *Notes from the Leyden Museum*, 35, 237–240, pl. 10.
- Popta, C.M.L. (1919) Description of *Clarias nigeriae* n. sp. from the Wari, mouth of the Niger, West Africa. *Zoologische Mededelingen (Leiden)*, 5, 4.
- Posada, A. (1909) Los peces. In: *Estudios científicos del doctor Andres Posada con algunos otros escritos suyos sobre diversos temas*, Medellin, Colombia, pp. 285–322.
- Pouyaud, L., Gustiano, R. & Teugels, G.G. (2004) Contribution to the phylogeny of the Pangasiidae based on mitochondrial 12S rRNA. *Indonesian Journal of Agricultural Science*, 5, 45–62.
- Pouyaud, L. & Teugels, G.G. (2000) Description of a new pangasiid catfish from east Kalimantan, Indonesia (Siluriformes: Pangasiidae). *Ichthyological Exploration of Freshwaters*, 11, 193–200.
- Pouyaud, L., Teugels, G.G., Gustiano, R. & Legendre, M. (2000) Contributions to the phylogeny of pangasiid catfishes

- based on allozymes and mitochondrial DNA. *Journal of Fish Biology*, 56, 1509–1538.
- Pouyaud, L., Teugels, G.G. & Legendre, M. (1999) Description of a new pangasiid catfish from South-east Asia (Siluriformes). *Cybium (3e série)*, 23, 247–258.
- Prashad, B. & Mukerji, D.D. (1929) The fish of the Indawgyi Lake and the streams of the Myitkyina District (Upper Burma). *Records of the Indian Museum*, 31, 161–223, pls. 7–10.
- Priem, F. (1904) Sur les poissons du Bartonien et les siluridés et acipenséridés de l'Éocène du bassin de Paris. *Bulletin de la Société Géologique de France, 4e serie*, 4, 42–47.
- Priem, F. (1906) Sur les Otolithes des Poissons éocènes du Bassin parisien. *Bulletin de la Société Géologique de France, 4e serie*, 6, 265–280.
- Priem, F. (1908) *Étude des Poissons Fossiles du Bassin Parisien*. Publications des Annales de Paléontologie, Paris 144 p., 74 figs., 5 pls.
- Priem, F. (1914) Sur des poissons fossiles et en particulier des Siluridés du Tertiaire supérieur et des couches récentes d'Afrique. *Mémoires de la Société géologique de France*, 21, 1–13, pls. 1–5.
- Provenzano R., F. (1995) *Leporacanthicus galaxias* Isbrücker y Nijssen (1989) (Pisces: Siluriformes: Loricariidae), nueva cita de un bagre loricarido para Venezuela. *Acta Biologica Venezuelica*, 15, 97–98.
- Provenzano R., F. (1997) *Gelanoglanis stroudi* new record of a catfish (Siluroidei: Auchenipteridae) for the Venezuelan continental ichthyofauna. *Acta Biologica Venezuelica*, 17, 79–81.
- Provenzano R., F., Lasso, C. & Ponte, V. (1995) *Neblinichthys roraima*, a new species of armored catfish (Siluroidei: Loricariidae) from río Kukenan, Venezuela, with considerations about the biogeography of the Guyana Shield. *Ichthyological Exploration of Freshwaters*, 6, 243–254.
- Provenzano R., F., Machado-Allison, A., Chernoff, B., Willink, P. & Petry, P. (2005) *Harttia merevari* a new species of catfish (Siluriformes: Loricariidae) from Venezuela. *Neotropical Ichthyology*, 3, 519–524.
- Provenzano R., F., Schaefer, S.A., Baskin, J.N. & Royero-Leon, R. (2003) New, possibly extinct lithogenine loricariid (Siluriformes, Loricariidae) from northern Venezuela. *Copeia*, 2003, 562–575.
- Puyo, J. (1936) Contribution à l'étude ichthyologique de la Guyane française. *Pêches et pêcheries. Bulletin de la Société d'Histoire Naturelle de Toulouse*, 70, 5–258.
- Quevedo, R. & Reis, R. (2002) *Pogonopoma obscurum*: a new species of loricariid catfish (Siluriformes: Loricariidae) from southern Brazil, with comments on the genus *Pogonopoma*. *Copeia*, 2002, 402–410.
- Quoy, J.R.C. & Gaimard, J.P. (1824–1825) Description des Poissons, chapter IX. In: Freycinet, L., de (Ed.), *Voyage Autour du Monde... Exécuté sur les Corvettes de L. M. L'Uranie et La Physicienne, Pendant les Années 1817, 1818, 1819 et 1820*, Chez Pillet aîné, Paris, pp. 192–401, pls. 43–65. [Pp.192–328 issued in 1824; 329–401 in 1825]
- Rafinesque, C.S. (1815) *Analyse de la Nature, ou Tableau de l'Univers et des Corps Organisés*, Palerme, 224 p.
- Rafinesque, C.S. (1818a) Discoveries in natural history, made during a journey through the western region of the United States. *American Monthly Magazine and Critical Review*, 3, 354–356.
- Rafinesque, C.S. (1818b) Further account of discoveries in natural history, in the western states, made during a journey through the western region of the United States. *American Monthly Magazine and Critical Review*, 4, 39–42.
- Rafinesque, C.S. (1819) Prodrome de 70 nouveaux genres d'animaux découverts dans l'intérieur des États-Unis d'Amérique, durant l'année 1818. *Journal de Physique, de Chimie et d'Histoire Naturelle*, 88, 417–429.
- Rafinesque, C.S. (1820a) *Ichthyologia Ohiensis, or Natural History of the Fishes Inhabiting the River Ohio and its Tributary Streams, Preceded by a Physical Description of the Ohio and its Branches*, Privately printed, Lexington, Kentucky, 90 p.
- Rafinesque, C.S. (1820b) Description of the Silures or catfishes of the River Ohio. *Quarterly Journal of Science, Literature and the Arts*, 9, 48–52.
- Rafinesque, C.S. (1820c) Ichthyologia Ohiensis [Part 7]. *Western Review and Miscellaneous Magazine*, 2, 355–363.
- Rafinesque, C.S. (1832) Extracts from a second series of zoological letters written to Baron Cuvier of Paris, by Prof. Rafinesque in 1831. *Atlantic Journal and Friend of Knowledge*, 1, 19–22.
- Rainboth, W.J. (1996) *Fishes of the Cambodian Mekong. FAO Species Identification Field Guide for Fishery Purposes*, FAO, Rome, 265 p.
- Ramakrishniah, M. (1988) A new bagrid fish of the genus *Mystus* (Scopoli) from Krishna River system. *Matsya*, 12/13, 139–143.
- Ramsay, E.P. & Ogilby, J.D. (1886) A contribution to the knowledge of the fish-fauna of New Guinea. *Proceedings of the Linnean Society of New South Wales (Ser. 2)*, 1, 8–20.
- Ranzani, C. (1841) [De nonnullis novis speciebus Piscium. Opusculum tertium]. *Nuovi Annali delle Scienze Naturali (Bologna)*, 3, 60–66.

- Ranzani, C. (1842) De nonnullis novis speciebus piscium. Opusculum tertium. *Novi Commentarii Academiae Scientiarum Instituti Bononiensis*, 5, 307–338, pls. 23–28.
- Rao, V.R. (1956) The skull of an Eocene siluroid fish from Western Kutch, India. *Journal of the Paleontological Society of India*, 1, 181–185, pl. 28.
- Rapp Py-Daniel, L.H. (1981) *Furcodontichthys novaesi* n. gen., n. sp. (Osteichthyes, Siluriformes; Loricariidae) na bacia Amazônia, Brasil. *Boletim do Museu Paraense Emílio Goeldi, Nova Serie, Zoologia*, 105, 1–17.
- Rapp Py-Daniel, L.H. (1985) *Dekeyseria amazonica*, novo gênero e nova espécie na região amazônica, Brasil, e *Dekeyseria scaphirhyncha* (Kner, 1854) nova combinação (Loricariidae: Siluriformes). *Amazoniana*, 9, 177–191.
- Rapp Py-Daniel, L.H. (1988) *Hypostomus hoplonites* sp. n. da bacia amazônica, Brasil (Pisces, Siluroidea, Loricariidae). *Iheringia, Série Zoologia*, 68, 13–23.
- Rapp Py-Daniel, L.H. (1989) Redescription of *Parancistrus aurantiacus* (Castelnau, 1855) and preliminary establishment of two new genera: *Baryancistrus* and *Oligancistrus* (Siluriformes, Loricariidae). *Cybium (3e série)*, 13, 235–246.
- Rapp Py-Daniel, L.H. (1991) *Chaetostoma jegui*, a new mailed catfish from Rio Uraricoera, Brazil (Osteichthyes: Loricariidae). *Ichthyological Exploration of Freshwaters*, 2, 239–246.
- Rapp Py-Daniel, L.H. & Oliveira, E.C. (2001) Seven new species of *Harttia* from the Amazonian-Guyana region (Siluriformes: Loricariidae). *Ichthyological Exploration of Freshwaters*, 12, 79–96.
- Rapp Py-Daniel, L.H. & Zuanon, J. (2005) Description of a new species of *Parancistrus* (Siluriformes: Loricariidae) from the rio Xingu, Brazil. *Neotropical Ichthyology*, 3, 571–577.
- Rashida, M., Mirza, M.R. & Saleem, M. (1996) A contribution to the systematics and biology of *Glyptothorax kashmiriensis* Hora (Pisces: Sisoridae) from Pakistan and Azad Kashmir. *Biologia (Lahore)*, 42, 59–60.
- Regan, C.T. (1903a) Description of a new fish of the genus *Chaetostomus* from Venezuela. *Annals and Magazine of Natural History (Ser. 7)*, 11, 599.
- Regan, C.T. (1903b) Descriptions of new South-American fishes in the collection of the British Museum. *Annals and Magazine of Natural History (Ser. 7)*, 12, 621–630.
- Regan, C.T. (1904a) On a collection of fishes made by Mr. John Graham at Yunnan Fu. *Annals and Magazine of Natural History (Ser. 7)*, 13, 190–194.
- Regan, C.T. (1904b) A monograph of the fishes of the family Loricariidae. *Transactions of the Zoological Society of London*, 17, 191–350, pls. 9–21.
- Regan, C.T. (1905a) A synopsis of the species of the silurid genera *Parexostoma*, *Chimarrhichthys*, and *Exostoma*. *Annals and Magazine of Natural History (Ser. 7)*, 15, 182–185.
- Regan, C.T. (1905b) Description of a new loricariid fish of the genus *Xenocara* from Venezuela. *Novitates Zoologicae (Tring)*, 12, 242.
- Regan, C.T. (1905c) Description de six poissons nouveaux faisant partie de la collection du Musée d'Histoire Naturelle de Genève. *Revue suisse de Zoologie, Annales de la Société zoologique suisse et du Muséum d'Histoire naturelle de Genève*, 13, 389–393, pls. 5–6.
- Regan, C.T. (1906–1908) Pisces. In: Godman, F.D. & Salvin, O. (Eds.), *Biologia Central-Americana, Part 193*, London, pp. 1–203, 25 pls. [signatures dated to month and year]
- Regan, C.T. (1906a) Notes on some loricariid fishes, with descriptions of two new species. *Annals and Magazine of Natural History (Ser. 7)*, 17, 94–98.
- Regan, C.T. (1906b) Descriptions of five new freshwater fishes from Sarawak, Borneo, collected by Dr. D. Hose. *Annals and Magazine of Natural History (Ser. 7)*, 18, 66–68.
- Regan, C.T. (1906c) On the fresh-water fishes of the island of Trinidad, based on the collection, notes, and sketches, made by Mr. Lechmere Guppy, Junr. *Proceedings of the Zoological Society of London*, 1906, pt 2, 378–393, pls. 21–25.
- Regan, C.T. (1907a) Descriptions of three new fishes from Yunnan, collected by Mr. J. Graham. *Annals and Magazine of Natural History (Ser. 7)*, 19, 63–64.
- Regan, C.T. (1907b) Descriptions of six new freshwater fishes from Mexico and Central America. *Annals and Magazine of Natural History (Ser. 7)*, 19, 258–260.
- Regan, C.T. (1907c) Fishes. Pp. 157–158, in: Reports on a collection of Batrachia, reptiles and fish from Nepal and the western Himalayas. *Records of the Indian Museum*, 1, 149–158, pl. 6.
- Regan, C.T. (1908a) Descriptions of three new freshwater fishes from China. *Annals and Magazine of Natural History (Ser. 8)*, 1, 109–111, pl. 4.
- Regan, C.T. (1908b) Descriptions of new freshwater fishes from China and Japan. *Annals and Magazine of Natural His-*

- tory (Ser. 8), 1, 149–153.
- Regan, C.T. (1908c) Descriptions of four new freshwater fishes from British New Guinea. *Annals and Magazine of Natural History* (Ser. 8), 1, 153–156.
- Regan, C.T. (1908d) Description of a new loricariid fish of the genus *Plecostomus* from Argentina. *Annals and Magazine of Natural History* (Ser. 8), 2, 358.
- Regan, C.T. (1908e) Description of new fishes from Lake Candidius, Formosa, collected by Dr. A. Moltrecht. *Annals and Magazine of Natural History* (Ser. 8), 2, 358–360.
- Regan, C.T. (1908f) Descriptions of new loricariid fishes from South America. *Proceedings of the Zoological Society of London*, 1907, 795–800, pls. 47–49.
- Regan, C.T. (1908g) The Duke of Bedford's Zoological Exploration in eastern Asia.— VIII. A collection of freshwater fishes from Corea. *Proceedings of the Zoological Society of London*, 1908, 59–63, pls. 2–3.
- Regan, C.T. (1909) Descriptions of three new freshwater fishes from South America, presented to the British Museum by Herr J. Paul Arnold. *Annals and Magazine of Natural History* (Ser. 8), 3, 234–235.
- Regan, C.T. (1911) The classification of the teleostean fishes of the order Ostariophysi. 2. Siluroidea. *Annals and Magazine of Natural History* (Ser. 8), 8, 553–577.
- Regan, C.T. (1912a) A revision of the South-American siluroid fishes of the genus *Corydoras*, with a list of the specimens in the British Museum (Natural History). *Annals and Magazine of Natural History* (Ser. 8), 10, 209–220.
- Regan, C.T. (1912b) Descriptions of new fishes of the family Loricariidae in the British Museum Collection. *Proceedings of the Zoological Society of London*, 1912, 666–670, pls. 75–77.
- Regan, C.T. (1913a) Descriptions of two new fishes from Paranagua, Brazil, presented to the British Museum by Herr A. Rachow. *Annals and Magazine of Natural History* (Ser. 8), 11, 231–232.
- Regan, C.T. (1913b) A synopsis of the siluroid fishes of the genus *Liocassis*, with descriptions of new species. *Annals and Magazine of Natural History* (Ser. 8), 11, 547–554.
- Regan, C.T. (1913c) Fishes from the River Ucayali, Peru, collected by W. Mounsey. *Annals and Magazine of Natural History* (Ser. 8), 12, 281–283.
- Regan, C.T. (1913d) The fishes of the San Juan River, Colombia. *Annals and Magazine of Natural History* (Ser. 8), 12, 462–473.
- Regan, C.T. (1913e) Description of a new loricariid fish of the genus *Plecostomus* from Rio Janeiro. *Annals and Magazine of Natural History* (Ser. 8), 12, 555.
- Regan, C.T. (1916) A new loricariid fish of the genus *Cyclopium* from Ecuador. *Annals and Magazine of Natural History* (Ser. 8), 18, 80.
- Regan, C.T. (1920a) Three new fishes from the Tanganyika Territory. *Annals and Magazine of Natural History* (Ser. 9), 6, 104–105.
- Regan, C.T. (1920b) Pisces. *Zoological Record*, 55, for 1918, 1–19.
- Regan, C.T. (1923) Note on the siluroid fishes of the genera *Glyptosternum* and *Exostoma*. *Annals and Magazine of Natural History* (Ser. 9), 11, 608–610.
- Reichel, M. (1927) Étude anatomique du *Phreatobius cisternarum* Goeldi, silure aveugle du Brésil. *Revue suisse de Zoologie, Annales de la Société zoologique suisse et du Muséum d'Histoire naturelle de Genève*, 34, 285–403, pl. 2–6.
- Reinhardt, J.T. (1859) *Stegophilus insidiosus*, en ny Mallefish fra Brasilien of dens Levemaade. *Videnskabelige Meddelelser fra den Naturhistorisk Forening i Kjøbenhavn, Aaret*, for 1858, 79–97, pl. 2.
- Reis, R.E. (1983) *Rineloricaria longicauda* e *Rineloricaria quadrensis*, duas novas espécies de Loricariinae do sul do Brasil (Pisces, Siluriformes, Loricariidae). *Iheringia, Série Zoologia*, 62, 61–80.
- Reis, R.E. (1987) *Ancistrus cryptophthalmus* sp. n., a blind mailed catfish from the Tocantins River basin, Brazil (Pisces, Siluriformes, Loricariidae). *Revue Française d'Aquariologie et Herpetologie*, 14, 81–84.
- Reis, R.E. (1992) Wir Fingen den blinden Antennenwels, *Ancistrus cryptophthalmus*. In: Stawikowski, R. (Ed.), *Harnischwelse, Die Aquarien- und Terrarien Zeitschrift, Sonderheft*, Eugen Ulmer, Stuttgart, pp. 54–56.
- Reis, R.E. (1997) Revision of the neotropical catfish genus *Hoplosternum* (Ostariophysi: Siluriformes: Callichthyidae), with the description of two new genera and three new species. *Ichthyological Exploration of Freshwaters*, 7, 299–326.
- Reis, R.E. (1998a) Anatomy and phylogenetic analysis of the Neotropical callichthyid catfishes (Ostariophysi, Siluriformes). *Zoological Journal of the Linnean Society*, 124, 105–168.
- Reis, R.E. (1998b) Systematics, biogeography, and the fossil record of the Callichthyidae: A review of the available data. In: Malabarba, L.R., Reis, R.E., Vari, R.P., Lucena, Z.M. & Lucena, C.A.S. (Eds.), *Phylogeny and Classification of*

- Neotropical Fishes*, Edipucrs, Porto Alegre, pp. 351–362.
- Reis, R.E. (2003) Callichthyidae. In: Reis, R.E., Kullander, S.O. & Ferraris, C.J., Jr. (Eds.), *Check list of the Freshwater Fishes of South and Central America*, Edipucrs, Porto Alegre, Brazil, pp. 291–309.
- Reis, R.E. (2004) *Otocinclus cocama*, a new uniquely colored loricariid catfish from Peru (Teleostei: Siluriformes), with comments on the impact of taxonomic revisions to the discovery of new taxa. *Neotropical Ichthyology*, 2, 109–115.
- Reis, R.E. & Cardoso, A.R. (2001) Two new species of *Rineloricaria* from southern Santa Catarina and northeastern Rio Grande do Sul, Brazil (Teleostei: Loricariidae). *Ichthyological Exploration of Freshwaters*, 12, 319–332.
- Reis, R.E. & Kaefer, C.C. (2005) Two new species of the Neotropical catfish genus *Lepthoplosternum* (Ostariophysi: Siluriformes: Callichthyidae). *Copeia*, 2005, 724–731.
- Reis, R.E., Kullander, S.O. & Ferraris, C.J., Jr. (Eds.) (2003) *Check List of the Freshwater Fishes of South and Central America*, Edipucrs, Porto Alegre, Brazil, 729 p.
- Reis, R.E., Le Bail, P.-Y. & Mol, J.H.A. (2005) New arrangement in the synonymy of *Megalechis* Reis, 1997 (Siluriformes: Callichthyidae). *Copeia*, 2005, 678–682.
- Reis, R.E. & Pereira, E.H.L. (1999) *Hemipsilichthys nudulus*, a new, uniquely-plated species of loricariid catfish from the rio Araranguá basin, Brazil (Teleostei: Siluriformes). *Ichthyological Exploration of Freshwaters*, 10, 45–51.
- Reis, R.E. & Pereira, E.H.L. (2000) Three new species of the loricariid catfish genus *Loricariichthys* (Teleostei: Siluriformes) from southern South America. *Copeia*, 2000, 1029–1047.
- Reis, R.E. & Schaefer, S.A. (1992) *Eurycheilus pantherinus* (Siluroidei: Loricariidae), a new genus and species of Hypoptopomatinae from southern Brazil. *Copeia*, 1992, 215–223.
- Reis, R.E. & Schaefer, S.A. (1993) *Eurycheilichthys* nom. nov., a substitute name for *Eurycheilus* Reis and Schaefer, 1992 (Siluroidei: Loricariidae). *Copeia*, 1993, 894.
- Reis, R.E. & Schaefer, S.A. (1998) New cascudinhos from southern Brazil: Systematics, endemism, and relationships (Siluriformes, Loricariidae, Hypoptopomatinae). *American Museum Novitates*, 3254, 1–25.
- Reis, R.E., Weber, C. & Malabarba, L.R. (1990) Review of the genus *Hypostomus* Lacep  de, 1803 from southern Brazil, with descriptions of three new species (Pisces: Siluriformes: Loricariidae). *Revue suisse de Zoologie, Annales de la Soci  t   zoologique suisse et du Mus  um d'Histoire naturelle de Gen  ve*, 97, 729–766.
- Reizer, C., Mattei, X. & De Vos, L. (1980) Contribution    l'  tude de la faune ichtyologique du bassin du fleuve S  n  gal; V.—Schilbeidae. *Bulletin de l'Institut fondamental d'Afrique noire. S  rie A, Sciences naturelles*, 42, 181–200.
- Rema Devi, K. & Emilyamma, K.G. (1997) On the specific identity of *Ompok bimaculatus* (Siluriformes, Siluridae). *Journal of the Bombay Natural History Society*, 94, 421–422, 1 pl.
- Rema Devi, K. & Raghunathan, M.B. (1999) *Heteropneustes longipectoralis* (Siluriformes: Heteropneustidae) a new species from the Anamalai Hills, in the western Ghats. *Records of the Zoological Survey of India*, 97, 109–115, pls. 1–2.
- Rendahl, H. (1922) A contribution to the ichthyology of north-west Australia. *Nyt Magazin for Naturvidenskaberne, Kristiania*, 60, 163–197.
- Rendahl, H. (1925) Eine neue Art der Gattung *Glyptosternum* aus China. *Zoologischer Anzeiger*, 64, 307–308.
- Rendahl, H. (1928) Beitr  ge zur Kenntnis der Chinesischen s  sswasserfische. I.—Systematischer teil. *Arkiv f  r Zoologi*, 20 A, 1–194.
- Rendahl, H. (1932) Die Fischfauna der chinesischen Provinz Szetschwan. *Arkiv f  r Zoologi*, 24 A, 1–134.
- Rendahl, H. (1937) Einige Fische aus Ecuador und Bolivia. *Arkiv f  r Zoologi*, 29 A, 1–11.
- Rendahl, H. (1941) Fische aus dem pazifischen Abflussgebiet Kolumbiens. *Arkiv f  r Zoologi*, 33 A, 1–15.
- Rendahl, H. & Vestergren, G. (1941) Eine neue Art der Gattung *Glyptosternon* s. str. aus dem nord  stlichen Birma. *Zoologischer Anzeiger*, 133, 213–214.
- Retzer, M.E. (2005) Description of a new species of *Acestridium* (Siluriformes: Loricariidae) from Colombia. *Zootaxa*, 972, 1–6.
- Retzer, M.E., Nico, L.G. & Provenzano R, F. (1999) Two new species of *Acestridium* (Siluriformes: Loricariidae) from southern Venezuela, with observations on camouflage and color change. *Ichthyological Exploration of Freshwaters*, 10, 313–326.
- Retzer, M.E. & Page, L.M. (1997) Systematics of the stick catfishes, *Farlowella* Eigenmann & Eigenmann (Pisces, Loricariidae). *Proceedings of the Academy of Natural Sciences, Philadelphia*, 147, 33–88.
- Ribeiro, A.C., Carvalho, M. & Melo, A.L.A. (2005) Description and relationships of *Otothyropsis marapoama*, a new genus and species of hypoptopomatine catfish (Siluriformes: Loricariidae) from rio Tiet   basin, southeastern Brazil. *Neotropical Ichthyology*, 3, 489–498.
- Ribeiro, A.C., Melo, A.L.A. & Pereira, E.H.L. (2002) A new species of *Parotocinclus* (Siluriformes: Loricariidae) from

- the rio São Francisco basin, southeastern Brazil. *Ichthyological Exploration of Freshwaters*, 13, 217–224.
- Ricardo-Bertram, C.K. (1943) The fishes of the Bangweulu region. *The Journal of the Linnean Society of London. Zoology*, 41, 183–217.
- Richardson, J. (1836) The Fish. In: *Fauna Boreali-Americana; or the Zoology of the Northern Parts of British America: Containing Descriptions of the Objects of Natural History Collected on the Late Northern Land Expeditions, Under the Command of Sir John Franklin, R.N., Part 3*, John Murray, Albemarle-Street; R. Bentley, London, pp. i–xv + 1–327, pls. 74–97.
- Richardson, J. (1844–1848) Ichthyology of the voyage of H. M. S. Erebus & Terror. In: Richardson, J. & Gray, J. E. (Eds.), *The Zoology of the Voyage of H. M. S. Erebus & Terror, Under the Command of Captain Sir J. C. Ross ... During ... 1839–43*, Vol. 2, no. 2, London, pp. i–viii + 1–139, pls. 1–60. [Issued over several years, as follows: 1844: 1–16; 1845: 17–52; 1846: 53–74; 1848: i–viii + 75–139.]
- Richardson, J. (1845) Ichthyology.— Part 3. In: Hinds, R.B. (Ed.), *The Zoology of the Voyage of H. M. S. Sulphur, Under the Command of Captain Sir Edward Belcher, R. N., C. B., F. R. G. S., etc., During the Years 1836–42, No. 10*, Smith, Elder & Co, London, pp. 99–150, pls. 55–64.
- Richardson, J. (1846) Report on the ichthyology of the seas of China and Japan. *Report of the British Association for the Advancement of Science, 15th meeting*, 1845, 187–320.
- Richter. (1928). *Abhandlungen und Berichte der Pommerschen Naturforschenden Gesellschaft*, 9 [not seen; citation incomplete]
- Ringuelet, R.A. (1965) Diferenciacion geografica de "otuno", *Diplomystes viedmensis* MacDonagh, 1931 (Pisces Siluriformes). *Physis (Buenos Aires)*, 25, 89–92.
- Ringuelet, R.A. (1982) Una nueva subespecie del bagre patagonico *Diplomystes viedmensis* Mac Donagh, 1931 en el Rio Senguer (Chubut, Argentina). *Limnobiós*, 2, 349–351.
- Ringuelet, R.A., Arámburu, R.H. & Alonso de Arámburu, A. (1967) *Los Peces Argentinos de Agua Dulce*. Comision de Investigacion Cientifica, La Plata, 602 p.
- Risch, L. (1981) The systematic status of *Gephyroglanis longipinnis* Boulenger, 1899, *Chrysichthys magnus* Pellegrin, 1922, and *Gephyroglanis gigas* Pellegrin, 1922 (Pisces, Bagridae). *Revue de Zoologie Africaine*, 95, 508–524.
- Risch, L. (1983) Note on the synonyms of *Eutropius mandibularis* Günther, 1867, and *Eutropius multitaeniatus* Pellegrin, 1913 (Pisces, Schilbeidae). *Revue de Zoologie Africaine*, 97, 268–287.
- Risch, L. (1984) Preliminary data of a systematic revision of the African species of the family Schilbeidae (Pisces: Siluriformes). *Revue de Zoologie Africaine*, 98, 424–433.
- Risch, L. (1985a) Redécouverte de l'espèce *Gephyroglanis ogooensis* Pellegrin, 1900 (Pisces, Bagridae) das l'Ogôoué. *Revue de Zoologie Africaine*, 99, 97–104.
- Risch, L. (1985b) Description of two new species in the genus *Chrysichthys* Bleeker, 1858 (Pisces, Bagridae). *Revue de Zoologie Africaine*, 99, 185–193.
- Risch, L. (1986) Bagridae. In: Daget, J., Gosse, J.-P. & Thys van den Audenaerde, D.F.E. (Eds.), *Check-list of the Freshwater Fishes of Africa*, Vol. 2, ISBN Bruxelles; MRAC Tervuren; ORSTOM, Paris, pp. 2–35.
- Risch, L. (1987) Description of four new bagrid catfishes from Africa (Siluriformes: Bagridae). *Cybium (3e série)*, 11, 20–38.
- Risch, L. (1988) Description d'une espèce nouvelle de *Chrysichthys* (Pisces, Bagridae), provenant de la rivière Konkouré (République de Guinée). *Cybium (3e série)*, 12, 3–7.
- Risch, L. (1992a) Description de *Chrysichthys dageti* sp. n. (Teleostei, Bagridae) du bassin du Kouilou (République du Congo). *Cybium (3e série)*, 16, 151–157.
- Risch, L. (1992b) Bagridae. In: Lévêque, C., Paugy, D. & Teugels, G.G. (Eds.), *Faune des Poissons d'Eaux Douces et Saumâtres de l'Afrique de l'Ouest*, Vol. 2, Musée Royal de l'Afrique Central, Tervuren, and Editions de l'ORSTOM, Paris, pp. 394–431.
- Risch, L. & Thys van den Audenaerde, D.F.E. (1981) Note on the systematical status of *Gephyroglanis velifer* Thys, 1965 (Pisces, Bagridae). *Revue de Zoologie Africaine*, 95, 245–251.
- Risch, L. & Thys van den Audenaerde, D.F.E. (1985a) Nouvelle description, distribution zoogéographique et affinités de *Chrysichthys johnelsi* Daget, 1959 (Pisces, Bagridae). *Cybium (3e série)*, 9, 243–254.
- Risch, L. & Thys van den Audenaerde, D.F.E. (1985b) Note sur la présence de *Chrysichthys longidorsalis* (Pisces, Bagridae) et de *Sanagia velifera* (Pisces, Cyprinidae) dans le Nyong (Cameroun). *Revue de Zoologie Africaine*, 99, 87–96.
- Risso, E.N.P. de & Morra, M.I. (1964) *Parapterodoras paranensis*: nuevo género, nueva especie de Doradidae (Pisces — Nematognathi). *Notas del Museo de Ciencias Naturales del Chaco, Resistencia*, 1, 1–5, pl. 1.

- Risso, F.J.J. & de Risso, E.N.P. (1962) *Epapterus chaquensis*, nueva especie de Auchenipteridae (Pisces, Nematognathi). *Notas Biológicas de la Facultad de Ciencias Exactas, Físicas y Naturales, Corrientes, Zoología*, 3, 1–8.
- Risso, F.J.J. & de Risso, E.N.P. (1964a) Hallazgo de una nueva especie de *Xyliphius* en el Paraná (Pisces — Aspredinidae). *Notas del Museo de Ciencias Naturales del Chaco, Resistencia*, 1, 11–16.
- Risso, F.J.J. & de Risso, E.N.P. (1964b) Los Siluriformes conocidos como "Manduré" y otras especies afines (Pisces: Auchenipteridae, Hypophthalmidae, y Pimelodidae). *Notas del Museo de Ciencias Naturales del Chaco, Resistencia*, 1, 1–31.
- Rivière, E. (1886) Exposition de la mission Brazza au Muséum. *Revue Scientifique*, 3^e ser., 38, 13–23.
- Roberts, T.R. (1967) *Rheoglanis dendrophorus* and *Zaireichthys zonatus*, bagrid catfishes from the lower rapids of the Congo River. *Ichthyologia, The Aquarium Journal*, 39, 119–131.
- Roberts, T.R. (1978) An ichthyological survey of the Fly River in Papua New Guinea with descriptions of new species. *Smithsonian Contributions to Zoology*, 281, 1–72.
- Roberts, T.R. (1982a) A revision of the south and southeastern Asian angler-catfishes (Chacidae). *Copeia*, 1982, 895–901.
- Roberts, T.R. (1982b) Systematics and geographical distribution of the Asian silurid catfish genus *Wallago*, with a key to the species. *Copeia*, 1982, 890–894.
- Roberts, T.R. (1983) Revision of the south and southeast Asian sisorid catfish genus *Bagarius*, with description of a new species from the Mekong. *Copeia*, 1983, 435–445.
- Roberts, T.R. (1989a) The freshwater fishes of western Borneo (Kalimantan Barat, Indonesia). *Memoirs of the California Academy of Sciences*, 14, i–xii + 1–210.
- Roberts, T.R. (1989b) Systematic revision and description of new species of suckermouth catfishes (*Chiloglanis*, Mochokidae) from Cameroun. *Proceedings of the California Academy of Sciences, Series 4*, 46, 151–178.
- Roberts, T.R. (1992) Revision of the striped catfishes of Thailand misidentified as *Mystus vittatus*, with descriptions of two new species (Pisces: Bagridae). *Ichthyological Exploration of Freshwaters*, 3, 77–88.
- Roberts, T.R. (1993) The freshwater fishes of Java, as observed by Kuhl and van Hasselt in 1820–23. *Zoologische Verhandelingen (Leiden)*, 285, 1–94.
- Roberts, T.R. (1994) Systematic revision of Asian bagrid catfishes of the genus *Mystus* sensu stricto, with a new species from Thailand and Cambodia. *Ichthyological Exploration of Freshwaters*, 5, 241–256.
- Roberts, T.R. (1998) *Pseudecheneis sympelvicus*, a new species of rheophilic sisorid catfish from Laos (Mekong basin). *The Raffles Bulletin of Zoology*, 46, 289–292.
- Roberts, T.R. (1999) *Pangasius bedado*, a new species of molluscivorous catfish from Sumatra (Pisces, Siluriformes, Pangasiidae). *Natural History Bulletin of the Siam Society*, 47, 109–115.
- Roberts, T.R. (2000) A review of the African electric catfish family Malapteruridae, with descriptions of new species. *Occasional Papers in Ichthyology*, 1, 1–15.
- Roberts, T.R. (2001) *Ayarnangra estuarius*, a new genus and species of sisorid catfish from the Ayeyarwaddy basin, Myanmar. *Natural History Bulletin of the Siam Society*, 49, 81–87.
- Roberts, T.R. (2003) Systematics and osteology of Leptoglaninae, a new subfamily of the African catfish family Amphiliidae, with descriptions of three new genera and six new species. *Proceedings of the California Academy of Sciences, Series 4*, 54, 81–132.
- Roberts, T.R. & Ferraris, C.J., Jr. (1998) Review of South Asian sisorid catfish genera *Gagata* and *Nangra*, with descriptions of a new genus and five new species. *Proceedings of the California Academy of Sciences, Series 4*, 50, 315–345.
- Roberts, T.R. & Jumnongthai, J. (1999) Miocene fishes from Lake Phetchabun in north central Thailand, with descriptions of new taxa of Cyprinidae, Pangasiidae, and Chandidae. *Natural History Bulletin of the Siam Society*, 47, 153–189.
- Roberts, T.R. & Stewart, D.J. (1976) An ecological and systematic survey of fishes in the rapids of the lower Zaire or Congo River. *Bulletin of the Museum of Comparative Zoology*, 147, 239–317, pls. 1–14.
- Roberts, T.R. & Vidhayanon, C. (1991) Systematic revision of the Asian catfish family Pangasiidae, with biological observations and descriptions of three new species. *Proceedings of the Academy of Natural Sciences, Philadelphia*, 143, 97–144.
- Rochebrune, A.T., de. (1885) Vertebratorum novorum vel minus cognitorum orae Africæ occidentalis incolarum. Diagnoses. *Bulletin de la Société Philomathique (Ser. 7)*, 9, 86–99.
- Rodiles-Hernández, R., Hendrickson, D.A., Lundberg, J.G. & Humphries, J.M. (2005) *Lacantunia enigmatica* (Teleostei: Siluriformes) a new and phylogenetically puzzling freshwater fish from Mesoamerica. *Zootaxa*, 1000, 1–24.

- Rodríguez, M.S. & Miquelarena, A.M. (2005) A new species of *Rineloricaria* (Siluriformes: Loricariidae) from the Paraná and Uruguay river basins, Misiones, Argentina. *Zootaxa*, 945, 1–15.
- Roedel, H. (1930) Fischotolithen aus Paläozängeschieben. *Zeitschrift für Geschiebeforschung*, 6, 49–77, 1 pl.
- Röhl, E. (1942) *Fauna Descriptiva de Venezuela*. Tipografia Americana, Caracas, 432 p.
- Roman, B. (1966) Les poissons des Haut-Bassins de la Volta. *Annales du Musée Royal de l'Afrique Centrale, Series in 8°, Sciences Zoologiques*, 150, 1–191, pls. 1–7.
- Roman, B. (1970) Nuevas especies de peces de Río Muni (Guinea Ecuatorial). *Publicaciones del Instituto de Biología Aplicada, Barcelona*, 49, 5–23.
- Roman, B. (1975) Une nouvelle espèce de *Synodontis* (poissons, Mochocides) dans la Bougouriba, affluent de la Volta Noire: *Synodontis voltae. Notes Docum. Volta*, 8, 45–52.
- Román-Valencia, C. (1998) Redescripcion de *Branchioica phaneronomema* Miles, 1943 (Pisces, Trichomycteridae) de la cuene del rio Magdalena, Colombia. *Revista de la Academia Colombiana de Ciencias Exactas, Fisicas y Naturales*, 22, 299–303.
- Román-Valencia, C., Lehmann A, P. & Muñoz, A. (1999) Presencia del genero *Callichthys* (Siluriformes: Callichthyidae) en Colombia y descripcion de una nueva especie para el alto Rio Cauca. *Dahlia — Revista de la Asociacion Colombiana de Ictiologos*, 3, 53–62.
- Römer, W. (1990) *Ancistrus*, deine Kinder! Nach Beobachtungen von Ewald Kuß. *Die Aquarien- und Terrarienzeitschrift*, 43, 146–147.
- Rössel, F. (1961) *Corydoras caudimaculatus* ein neuer panzerwels aus Brasilien. (Pisces, Teleostei, Callichthyidae). *Senckenbergiana Biologica*, 42, 49–50.
- Rössel, F. (1962a) *Centromochlus schultzi*, ein neues Wels aus Brasilien (Pisces, Teleostei, Auchenipteridae). *Senckenbergiana Biologica*, 43, 27–30.
- Rössel, F. (1962b) *Corydoras cervinus*, ein neuer panzerwels aus Brasilien (Pisces, Teleostei, Callichthyidae). *Senckenbergiana Biologica*, 43, 31–33.
- Rössel, F. (1962c) *Corydoras axelrodi*, ein neuer Panzerwels aus Columbien (Pisces, Teleostei, Callichthyidae). *Senckenbergiana Biologica*, 43, 335–337.
- Rössel, F. (1963) Neue und seltene *Corydoras*-Arten aus Brasilien (Pisces, Teleostei, Callichthyidae). *Senckenbergiana Biologica*, 44, 359–363.
- Rössel, F. (1964) Welse (Siluroidea) gesammelt von der deutschen Indien-Expedition 1955/58. *Mitteilungen aus dem Hamburgischen Zoologischen Museum und Institut*, 61, 145–158.
- Royer, R. (1987) *Morfología de la aleta dorsal en los bagres (Teleostei: Siluriformes), con especial referencia a las familias americanas*. Escuela de Biología, Undergraduate thesis, Universidad Central de Venezuela, Facultad de Ciencias, Caracas.
- Royer, R. (1992) *Tatia musaica*, una neuva especie de bagre auqueniptero (Siluriformes — Auchenipteridae) de la cuena del Rio Orinoco, Territorio Federal Amazonas, Venezuela. *Acta Cientifica Venezolana*, 43, 300–306.
- Royer, R. (1999) *Studies on the systematics and phylogeny of the catfish family Auchenipteridae (Teleostei: Siluriformes)*, Doctoral dissertation, University of Bristol, Bristol, U. K.
- Royer, R. & Hureau, J.-C. (1996) The type specimens of auchenipterid catfishes (Siluriformes: Auchenipteridae) in the Muséum National d'Histoire Naturelle, Paris. *Cybium (3e série)*, 20, 369–377.
- Ruiz, V.H. & Berra, T.M. (1994) Fishes of the High Biobio River of south-central Chile with notes on diet and speculations on the origin of the ichthyofauna. *Ichthyological Exploration of Freshwaters*, 5, 5–18.
- Rüppell, W.P.E.S. (1829) *Beschreibung und Abbildung Mehrerer Neuer Fische im Nil Entdeckt*, Brönnner, Frankfurt-am-Main, 12 p., 3 pls. [Issued in 3 memoirs: in 1829, 1832, and 1835].
- Rüppell, W.P.E.S. (1832) *Fortsetzung der Beschreibung und Abbildung Mehrerer Neuer Fische, im Nil Entdeckt*, Brönnner, Frankfurt-am-Main, 14 p., 3 pls. [Issued in 3 parts, in 1829, 1832, and 1835].
- Rüppell, W.P.E.S. (1835–1838) *Neue Wirbelthiere zu der Fauna von Abyssinien Gehörig. Fische des Rothen Meeres*, Frankfurt-am-Main, 148 p., 33 pls. [1835:1–28, pls. 1–7; 1836:29–52, pls. 8–14; 1837:53–80, pls. 15–21; 1838:81–148, pls. 22–33.]
- Russell, A. (1794) *Natural History of Aleppo, Containing a Description of the City, and the Principal Natural Productions in its Neighbourhood. Together with an Account of the Climate, Inhabitants, and Diseases, Particularly of the Plague; Second Edition, Revised, Enlarged, and Illustrated by P. Russell*, Vol. 2, G.G. and J. Robinson, London, vii + 430 + xxxiv p. + 26 p. index, 16 pl.
- Russell, P. (1803) *Descriptions and Figures of Two Hundred Fishes Collected at Vizagapatam on the Coast of Coromandel*, East India Company, London, vii + 162 p., 197 pl. [In two volumes].

- Rutter, C.M. (1896) Notes on fresh water fishes of the Pacific slope of North America. II.— The fishes of Rio Yaqui, Sonora, with the description of a new genus of Siluridae. *Proceedings of the California Academy of Sciences, Series 2*, 6, 255–262.
- Sabaj, M.H. (2002) *Taxonomy of the Neotropical Thorny Catfishes (Siluriformes: Doradidae) and Revision of Genus Leptodoras*, Unpublished Ph.D. dissertation, University of Illinois at Urbana-Champaign.
- Sabaj, M.H. (2005) Taxonomic assessment of *Leptodoras* (Siluriformes: Doradidae) with descriptions of three new species. *Neotropical Ichthyology*, 3, 637–678.
- Sabaj, M.H. & Ferraris, C.J., Jr. (2003) Doradidae. In: Reis, R.E., Kullander, S.O. & Ferraris, C.J., Jr. (Eds.), *Check list of the Freshwater Fishes of South and Central America*, Edipucrs, Porto Alegre, Brazil, pp. 456–469.
- Sabino, J. & Trajano, E. (1997) A new species of blind armoured catfish, genus *Ancistrus*, from caves of Bodoquena region, Mato Grosso do Sul, southwestern Brazil (Siluriformes, Loricariidae, Ancistrinae). *Revue Française d'Aquariologie et Herpetologie*, 24, 73–78.
- Sagua, O. (1987) On a new species of electric catfish from Kainji, Nigeria, with some observations on its biology. *Journal of Fish Biology*, 30, 75–89.
- Sahni, A. & Mishra, V.P. (1975) Lower Tertiary vertebrates from western India. *Monographs of the Palaeontological Society of India*, 3, 1–48.
- Sanders, M. (1934) Die fossilen Fische der Alttertiären Süßwasserablagerungen aus Mittel-Sumatra. *Verhandelingen van het Geologisch Mijnbouwkundig Genootschap voor Nederland en Koloniën. Geologische serie*, 11, 1–144, 9 pls. [Also as published Dissertation, University of Amsterdam.]
- Sands, D. (1990) A new species of callichthyid catfish, *Corydoras araguaiaensis* (Siluriformes, Callichthyidae) from the Rio Araguaia, Brazil. *D. Sands, Catfishes of the World, Suppl.*, Self published, pp. [1–2].
- Sands, D.D. (1995) Four new *Corydoras* (Callichthyidae) species from upper Negro River tributaries and a range extension, together with a discussion of *C. bicolor* Nijssen & Isbrücker. *Freshwater and Marine Aquarium*, 18, 8–12, 14, 16, 18.
- Sands, D.D. & Black, B.K. (1985) Two new species of *Brachyrhamdia*, Myers, 1927, from Brazil and Peru, together with a redefinition of the genus. In: Sands, D. (Ed.), *Catfishes of the World, Vol. 3, Suppl (first set)*, Self published, pp. 58(1)–58(8).
- Santos, R.S. (1973) *Steindachneridion iheringi* (Woodward), um siluriforme da bacia do Paraíba, Estado de São Paulo. *Anais da Academia Brasiliera de Ciências*, 43, 667.
- Sarmento-Soares, L.M. & Buckup, P.A. (2005) A new *Glanidium* from the Rio São Francisco basin, Brazil (Siluriformes: Auchenipteridae: Centromochlinae). *Copeia*, 2005, 846–853.
- Sarmento-Soares, L.M., Martins-Pinheiro, R.F., Aranda, A.T. & Chamon, C.C. (2005) *Trichomycterus pradensis*, a new catfish from southern Bahia coastal rivers, northeastern Brazil (Siluriformes: Trichomycteridae). *Ichthyological Exploration of Freshwaters*, 16, 289–302.
- Sauvage, H.E. (1874) Notices ichthyologiques. *Revue et Magasin de Zoologie (Ser. 3)*, 2, 332–340.
- Sauvage, H.E. (1878) Note sur quelques poissons d'espèces nouvelles provenant des eaux douces de l'Indo-Chine. *Bulletin de la Société Philomathique (Ser. 7)*, 2, 233–242.
- Sauvage, H.E. (1879) Notice sur la faune ichthyologique de l'Ogôoué. *Bulletin de la Société Philomathique (Ser. 7)*, 3, 90–103.
- Sauvage, H.E. (1880a) Description de quelques poissons de la collection du Muséum d'histoire naturelle. *Bulletin de la Société Philomathique (Ser. 7)*, 4, 220–228.
- Sauvage, H.E. (1880b) Notice sur quelques poissons de l'île Campbell et de l'Indo-Chine. *Bulletin de la Société Philomathique (Ser. 7)*, 4, 228–233.
- Sauvage, H.E. (1880c) Étude sur la faune ichthyologique de l'Ogôoué. *Nouvelles Archives du Muséum d'Histoire Naturelle, Paris (Ser. 2)*, 3, 5–56, pls. 1–3.
- Sauvage, H.E. (1881) Recherches sur la faune ichthyologique de l'Asie et description d'espèces nouvelles de l'Indo-Chine. *Nouvelles Archives du Muséum d'Histoire Naturelle, Paris (Ser. 2)*, 4, 123–194, pls 5–8.
- Sauvage, H.E. (1882) Catalogue des poissons recueillis par M. E. Chantre pendant son voyage en Syrie, Haute-Mésopotamie, Kurdistan et Caucase. *Bulletin de la Société Philomathique (Ser. 7)*, 6, 163–168.
- Sauvage, H.E. (1883a) Sur une collection de poissons recueillie dans le lac Biwako (Japon) par M. F. Steenackers. *Bulletin de la Société Philomathique (Ser. 7)*, 7, 144–150.
- Sauvage, H.E. (1883b) Sur une collection de poissons recueillie dans Mé-Nam (Siam) par M. Harmand. *Bulletin de la Société Philomathique (Ser. 7)*, 7, 150–155.
- Sauvage, H.E. (1883c) Descriptions de quelques poissons de la collection du Muséum d'histoire naturelle. *Bulletin de la*

- Société Philomathique (Ser. 7), 7, 156–161.
- Sauvage, H.E. (1884a) Sur un siluroïde de la Réunion. *Bulletin de la Société Philomathique* (Ser. 7), 8, 147.
- Sauvage, H.E. (1884b) Notice sur la Faune ichthyologique d' l'ouest de l'Asie et plus particulièrement sur les poissons recueillis par M. Chantre pendant son voyage dans cette région. *Nouvelles Archives du Muséum d'Histoire Naturelle, Paris* (Ser. 2), 7, 1–41, pls. 1–3.
- Sauvage, H.E. & Dabry de Thiersant, P. (1874) Notes sur les poissons des eaux douces de la Chine. *Annales des sciences naturelles. Zoologie et biologie animale* (Sér. 6), 1, 1–18.
- Saville-Kent, W. (1889) *Preliminary Report on the Food-fishes of Queensland*, Government Printer, Brisbane, 16 p., 16 pls.
- Saville-Kent, W. (1893) *The Great Barrier Reef of Australia; its Products and Potentialities*, W.H. Allen, London, xiii + 387 p., 48 pl.; 16 color plates.
- Sawyer, F.C. (1953) The dates of issue of J. E. Gray's "Illustrations of Indian Zoology" (London, 1830–1835). *Journal of the Society for the Bibliography of Natural History*, 3, 48–55.
- Schaefer, S.A. (1986) *Historical Biology of the Loricariid Catfishes: Phylogenetics and Functional Morphology*, Ph. D. Thesis, University of Chicago.
- Schaefer, S.A. (1987) Osteology of *Hypostomus plecostomus* (Linnaeus), with a phylogenetic analysis of the loricariid subfamilies (Pisces: Siluriformes). *Contributions in Science, Los Angeles County Museum*, 394, 1–31.
- Schaefer, S.A. (1988) A new species of the loricariid genus *Parotocinclus* from southern Venezuela (Pisces: Siluroidei). *Copeia*, 1988, 182–188.
- Schaefer, S.A. (1990) Anatomy and relationships of the scoloplacid catfishes. *Proceedings of the Academy of Natural Sciences, Philadelphia*, 142, 167–210.
- Schaefer, S.A. (1991) Phylogenetic analysis of the loricariid subfamily Hypoptopomatinae (Pisces: Siluroidei: Loricariidae), with comments on generic diagnoses and geographic distribution. *Zoological Journal of the Linnean Society*, 102, 1–41.
- Schaefer, S.A. (1996a) *Nannoptopoma*, a new genus of loricariid catfishes (Siluriformes: Loricariidae) from the Amazon and Orinoco River Basins. *Copeia*, 1996, 913–926.
- Schaefer, S.A. (1996b) Type designations for some Steindachner loricariid material (Siluriformes: Loricariidae) in the Natural History Museum, Vienna. *Copeia*, 1996, 1031–1035.
- Schaefer, S.A. (1997) The neotropical cascudinhos: Systematics and biogeography of the *Otocinclus* catfishes (Siluriformes: Loricariidae). *Proceedings of the Academy of Natural Sciences, Philadelphia*, 148, 1–120.
- Schaefer, S.A. (1998) Conflict and resolution: Impact of new taxa on phylogenetic studies of the neotropical cascudinhos (Siluroidei: Loricariidae). In: Malabarba, L.R., Reis, R.E., Vari, R.P., Lucena, Z.M. & Lucena, C.A.S. (Eds.), *Phylogeny and Classification of Neotropical Fishes*, Edipucrs, Porto Alegre, pp. 375–400.
- Schaefer, S.A. (2003a) Scoloplacidae. In: Reis, R.E., Kullander, S.O. & Ferraris, C.J., Jr. (Eds.), *Check list of the Freshwater Fishes of South and Central America*, Edipucrs, Porto Alegre, Brazil, pp. 310–311.
- Schaefer, S.A. (2003b) Astroblepidae. In: Reis, R.E., Kullander, S.O. & Ferraris, C.J., Jr. (Eds.), *Check list of the Freshwater Fishes of South and Central America*, Edipucrs, Porto Alegre, Brazil, pp. 312–317.
- Schaefer, S.A. (2003c) Hypoptopomatinae. In: Reis, R.E., Kullander, S.O. & Ferraris, C.J., Jr. (Eds.), *Check list of the Freshwater Fishes of South and Central America*, Edipucrs, Porto Alegre, Brazil, pp. 321–329.
- Schaefer, S.A. (2003d) Relationships of *Lithogenes villosus* Eigenmann, 1909 (Siluriformes, Loricariidae): Evidence from high-resolution computed microtomography. *American Museum Novitates*, 3402, 1–55.
- Schaefer, S.A. & Provenzano, R.F. (1998) *Niobichthys ferrarisi*, a new genus and species of armored catfish from southern Venezuela (Siluriformes: Loricariidae). *Ichthyological Exploration of Freshwaters*, 8, 221–230.
- Schaefer, S.A. & Provenzano, R.F. (1993) The Guyana Shield *Parotocinclus*: Systematics, biogeography, and description of a new Venezuelan species (Siluroidei: Loricariidae). *Ichthyological Exploration of Freshwaters*, 4, 39–56.
- Schaefer, S.A., Provenzano, R.F., Pinna, M., de & Baskin, J.N. (2005) Noteworthy Venezuelan glanapterygine catfishes (Siluriformes, Trichomycteridae), with discussion of their biogeography and psammophily. *American Museum Novitates*, 3496, 1–27.
- Schaefer, S.A. & Stewart, D.J. (1993) Systematics of the *Panaque dentex* species group (Siluriformes: Loricariidae), wood-eating armored catfishes from tropical South America. *Ichthyological Exploration of Freshwaters*, 4, 309–342.
- Schaefer, S.A., Weitzman, S.H. & Britski, H.A. (1989) Review of the Neotropical catfish genus *Scoloplax* (Pisces: Loricarioidea: Scoloplacidae) with comments on reductive characters in phylogenetic analysis. *Proceedings of the Academy of Natural Sciences, Philadelphia*, 141, 181–211.

- Schilthuis, L. (1891) On a collection of fishes from the Congo; with description of some new species. *Tijdschrift der Nederlandse Dierkundige Vereeniging*, Ser. 2, 3, 83–92, pl. 6.
- Schindler, I. (1996) Ergänzende Beschreibung von *Lithoxus bovallii* (Regan, 1906) aus Guyana (Siluriformes, Loricariidae). *Zeitschrift für Fischkunde*, 3, 223–233.
- Schindler, O. (1959) *Loricariichthys melini* nov. spec. *Arkiv för Zoologi*, 12, 387–389.
- Schmidt, P. (1928) On three rare cat-fishes of the Magdalena River (South America, Columbia [sic]). *Comptes Rendus de l'Académie des Sciences de l'URSS*, [1928], 9–13.
- Schmidt, R.E. (1985) New distribution records and complementary description of *Haemomaster venezuelae* (Siluriformes: Trichomycteridae), a rare and poorly known fish from northern South America. *Studies on Neotropical Fauna and Environment*, 20, 93–96.
- Schmidt, R.E. (1987) Redescription of *Vandellia beccarii* (Siluriformes: Trichomycteridae) from Guyana. *Copeia*, 1987, 234–237.
- Schmidt, R.E. (1993) Relationships and notes on the biology of *Paracanthopoma parva* (Pisces: Trichomycteridae). *Ichthyological Exploration of Freshwaters*, 4, 185–191.
- Schmidt, R.E. & Ferraris, C.J., Jr. (1985) A new species of *Parotocinclus* (Pisces: Loricariidae) from Guyana. *Proceedings of the Biological Society of Washington*, 98, 341–346.
- Schomburgk, R.H. (1841) The Natural History of Fishes of Guiana.— Part I. In: Jardine, W. (Ed.), *The Naturalists' Library*, Vol. 3, W.H. Lizars, Edinburgh, 263 p., pls. 1–30.
- Schomburgk, R.H. (1848) *Reisen in Britisch-Guiana in den Jahren 1840–1844. Im Auftrag Sr. Mäjestat des Königs von Preussen Ausgeführt von Richard Schomburgk. Versuch einer Fauna und Flora von Britisch-Guiana*, Nach Vorlagen von Johannes Muller, Ehrenberg, Erichson, Klotzsch, Troschel, Cabanis und Andern, Leipzig.
- Schubart, O. (1964a) Duas novas espécies de peixe da família Pimelodidae do Rio Mogi Guaçu (Pisces, Nematognathi). *Boletim do Museu Nacional do Rio de Janeiro, Nova Série, zoologia*, 244, 1–22.
- Schubart, O. (1964b) Sobre algumas Loricariidae da bacia do Rio Mogi Guaçu. *Boletim do Museu Nacional do Rio de Janeiro, Nova Série, zoologia*, 251, 1–19.
- Schubart, O. & Gomes, A.L. (1959) Descrição de *Cetopsorhamdia iheringi* sp. n. (Pisces, Nematognathi, Pimelodidae, Luciopimelodinae). *Revista Brasileira de Biologia*, 19, 1–7.
- Schubert, R. (1908) Die Fischotolithen des Pausramer Mergels.— 2. *Zeitschrift des Mährischen Landesmuseums*, 8, 102–120.
- Schubert, R. (1916) Obereocäne Otolithen von Barton Cliff bei Christchurch (Hampshire). *Jahrbuch der Kaiserlich Königlichen Geologischen Reichs-Anstalt*, 65, 277–288.
- Schultz, L.P. (1942) The fresh-water fishes of Liberia. *Proceedings of the United States National Museum*, 92, 301–348, pls. 35–36.
- Schultz, L.P. (1944a) A new loricariid catfish from the Rio Truando, Colombia. *Copeia*, 1944, 155–156.
- Schultz, L.P. (1944b) A new genus and species of pimelodid catfish from Colombia. *Journal of the Washington Academy of Sciences*, 34, 93–95.
- Schultz, L.P. (1944c) The catfishes of Venezuela, with descriptions of thirty-eight new forms. *Proceedings of the United States National Museum*, 94, 173–338, pls. 1–14.
- Schultz, L.P. (1944d) Two new species of fishes (Gymnotidae, Loricariidae) from Caripito, Venezuela. *Zoologica (New York)*, 29, 39–44, pl. 1.
- Schultz, L.P. (1945) *Pygidium mondolfi*, a new catfish from Venezuela. *Journal of the Washington Academy of Sciences*, 35, 29–31.
- Scopoli, G.A. (1777) *Introductio ad Historiam Naturalem, Sistens Genera Lapidum, Plantarum et Animalium Hactenus Detecta, Characteribus Essentialibus Donata, in Tribus Divisa, Subinde ad Leges Naturae*, Prague, x + 506 p.
- Seba, A. (1734–1765) *Locupletissimi Rerum Naturalium Thesauri Accurata Descriptio, et Iconibus Artificiosissimis Expressio, per Universam Physices Historiam. Opus, cui, in hoc Rerum Genere, Nullum par Exstitit. Ex Toto Terrarum Orbe Collegit, Digessit, Descripsit, et de Pingendum Curavit Albertus Seba*, 4 vols, 449 pls, J. Wetstenium & J. Gul. Smith et Janssonio-Waesbergios [etc], Amstelaedami. [Fish in vol. 3, published in 1758.]
- Seegers, L. (1989) *Leptoglanis wamiensis* spec. nov. (Bagridae, Siluriformes) aus dem Wami- Einzug in Tanzania. *Die Aquarien- und Terrarien- Zeitschrift*, 42, 284–287.
- Seegers, L. (1996) The fishes of the Lake Rukwa drainage. *Musée Royal de l'Afrique Centrale, Annales, Sciences Zoologiques*, 278, 1–407.
- Seegers, L. (2003) *Atopochilus vogti*: Ein Afrika-Saugwels aus dem Wami-River, Tanzania. *Aquarium (Bornheim)*, 405, 8–14.

- Sen, N. & Biswas, B.K. (1994) On a new species of *Nangra* Day (Pisces: Siluriformes: Sisoridae) from Assam, north east India with a note on comparative studies of other known species. *Records of the Zoological Survey of India*, 94, 441–446.
- Serpa, A.T. (1967) Pesces del Oriente Peruano. Algunas especies de Loricariidae, con referencia especial de la “Caramacha” *Pterygoplichthys multiradiatus* (Hancock). *Ecología y Utilidad. Biota, Lima*, 50, 201–259.
- Shaw, G. (1804) *General Zoology or Systematic Natural History. Vol. 5.—Pisces, in Two Parts*, G. Kearsley, London. [(pt 1): i–v + 1–250, pls. 93–132, 43+, 65+, 6+, 74+ and (pt 2): i–vi + 251–463, pls. 132–182, 158+.]
- Shaw, G.E. & Shebbeare, E.O. (1936) Description of a new species of fish from northern Bengal. *Journal of the Bombay Natural History Society*, 39, 188–189, 1 pl.
- Shaw, T.-H. (1930) Notes on some fishes from Ka-Shing and Shing-Tsong, Chekiang Province. *Bulletin of the Fan Memorial Institute of Biology, Peiping*, 1, 109–121.
- Shepherd, C.E. (1916) Fossil otoliths. *Knowledge*, 39 (new series, 13), 177–184, 203–205, figs. 154–157.
- Sherborn, C.D. & Griffin, F.J. (1934) On the dates of publication of the natural history portions of Alcide d'Orbigny's "Voyage Amerique meridionale". *Annals and Magazine of Natural History (Ser. 10)*, 13, 130–134.
- Shibatta, O.A. (1998) *Sistemática e Evolução da Família Pseudopimelodidae (Ostariophysi, Siluriformes), com a Revisão Taxonómica de Gênero Pseudopimelodus*, Ph.D. dissertation, Univ. Fed. São Carlos, São Paulo, Brazil.
- Shibatta, O.A. (2003a) Pseudopimelodidae. In: Reis, R.E., Kullander, S.O. & Ferraris, C.J., Jr. (Eds.), *Check list of the Freshwater Fishes of South and Central America*, Edipucrs, Porto Alegre, Brazil, pp. 401–405.
- Shibatta, O.A. (2003b) Phylogeny and classification of “Pimelodidae”. In: Arratia, G., Kapoor, B.G., Chardon, M. & Diogo, R. (Eds.), *Catfishes*, Science Publishers, Enfield, NH, USA, pp. 385–400.
- Shibatta, O.A. & Benine, R.C. (2005) A new species of *Microglanis* Eigenmann, 1912 (Siluriformes: Pseudopimelodidae) from upper rio Paraná basin, Brazil. *Neotropical Ichthyology*, 3, 579–585.
- Shibatta, O.A. & Pavanelli, C.S. (2005) Description of a new *Batrochoglanis* species (Siluriformes, Pseudopimelodidae) from the rio Paraguai basin, State of Mato Grosso, Brazil. *Zootaxa*, 1092, 21–30.
- Shimabukuro-Dias, C.K., Oliveira, C., Reis, R.E. & Foresti, F. (2004) Molecular phylogeny of the armored catfish family Callichthyidae (Ostariophysi, Siluriformes). *Molecular Phylogenetics and Evolution*, 32, 152–163.
- Shrestha, J. (1980) *Fishes of Nepal*. Curriculum Development Centre, Tribhuvan University, Kathmandu.
- Sielfeld-K., W.H. (1979) Nuevo registro de *Netuma barbus* (La Cepède, 1803) para águas chilenas (Pisces: Siluriformes). *Anales del Instituto de la Patagónia*, 10, 189–192.
- Silas, E.G. (1951a) On a collection of fish from the Anamalai and Nelliampathy Hill ranges, Western Ghats, with notes on their zoogeographic significance. *Journal of the Bombay Natural History Society*, 49, 679–681.
- Silas, E.G. (1951b) Notes on fishes of the genus *Glyptothorax* Blyth from Peninsular India, with description of a new species. *Journal of the Bombay Natural History Society*, 50, 367–370.
- Silfvergrip, A.M.C. (1992) Zungaro, a senior synonym of *Paulicea* (Teleostei: Pimelodidae). *Ichthyological Exploration of Freshwaters*, 3, 305–310.
- Silfvergrip, A.M.C. (1996) *A Systematic Revision of the Neotropical Catfish Genus Rhamdia (Teleostei, Pimelodidae)*, Stockholm University, Stockholm, 156 p., 8 pl.
- Silfvergrip, A.M.C. & Paepke, H.-J. (1997) Kritischer Katalog der Typen der Fischsammlung des Zoologischen Museums Berlin. Teil 7: Pimelodidae (Siluriformes). [Critical Catalogue of the Types of the Fish Collection of the Zoological Museum of Berlin. Part 7: Pimelodidae (Siluriformes)]. *Mitteilungen aus dem Zoologischen Museum in Berlin*, 73, 165–173.
- Skelton, P.H. (1981) The description and osteology of a new species of *Gephyroglanis* (Siluriformes, Bagridae) from the Olifants River, South West Cape, South Africa. *Annals of the Cape Provincial Museums. Natural History*, 13, 217–249.
- Skelton, P.H. (1984) A systematic revision of species of the catfish genus *Amphilinus* (Siluroidei, Amphiliidae) from east and southern Africa. *Annals of the Cape Provincial Museums. Natural History*, 16, 41–71.
- Skelton, P.H. (1986) Two new *Amphilinus* (Pisces, Siluroidei, Amphiliidae) from the Zaïre River system, Africa. *Revue de Zoologie Africaine*, 99, 263–291.
- Skelton, P.H. (1989) Descriptions of two new species of West African amphiliid catfishes (Siluroidei: Amphiliidae). *J. L. B. Smith Institute of Ichthyology, Special Publication*, 48, 1–13.
- Skelton, P.H. (1992) Amphiliidae. In: Lévéque, C., Paugy, D. & Teugels, G.G. (Eds.), *Faune des Poissons d'Eaux Douces et Saumâtres de l'Afrique de l'Ouest*, Vol. 2, Musée Royal de l'Afrique Centrale, Tervuren, and Editions de l'ORS-TOM, Paris, pp. 451–467.
- Skelton, P.H. (1993) *A Complete Guide to the Freshwater Fishes of Southern Africa*, Southern Book Publishers, Ltd.,

- Halfway House, South Africa, xiii + 388 p.
- Skelton, P.H. (2001) *A Complete Guide to the Freshwater Fishes of Southern Africa*, Second Edition, Struik Publishers, Cape Town, xiv + 395 p.
- Skelton, P.H., Bruton, M.N. & al. (1985) The fishes of the Okavango drainage system in Angola, South West Africa and Botswana: Taxonomy and distribution. *Ichthyological Bulletin of the J. L. B. Smith Institute of Ichthyology*, 50, 1–21.
- Skelton, P.H., Risch, L. & De Vos, L. (1984) On the generic identity of the *Gephyroglanis* catfishes from southern Africa (Pisces, Siluroidei, Bagridae). *Revue de Zoologie Africaine*, 98, 337–372.
- Skelton, P.H. & Teugels, G.G. (1986) Amphiliidae. In: Daget, J., Gosse, J.-P. & Thys van den Audenaerde, D.F.E. (Eds.), *Check-list of the Freshwater Fishes of Africa*, Vol. 2, ISBN, Bruxelles; MRAC, Tervuren; and ORSTOM, Paris, pp. 54–65.
- Skelton, P.H. & Teugels, G.G. (1991) A review of the clariid catfishes (Siluroidei, Clariidae) occurring in southern Africa. *Revue d'Hydrobiologie Tropicale*, 24, 241–260.
- Skelton, P.H. & Teugels, G.G. (1992) Neotype description for the African catfish *Clarias gariepinus* (Burchell, 1822) (Pisces: Siluroidei: Clariidae). *Ichthyological Bulletin of the J. L. B. Smith Institute of Ichthyology*, 56, 1–7.
- Skelton, P.H. & White, P.N. (1990) Two new species of *Synodontis* (Pisces: Siluroidei: Mochokidae) from southern Africa. *Ichthyological Exploration of Freshwaters*, 1, 277–287.
- Smith, A. (1838–1849) Pisces. In: *Illustrations of the Zoology of South Africa; Consisting Chiefly of Figures and Descriptions of the Objects of Natural History Collected During an Expedition into the Interior of South Africa in 1834, 1835, and 1836, Fitted Out by the Cape of Good Hope Association for Exploring Central Africa*, Vol. 4, Smith, Elder, London, pp. 1–77, 31 pl. In 5 vols.
- Smith, C.L. (1962) Some Pliocene fishes from Kansas, Oklahoma, and Nebraska. *Copeia*, 1962, 505–520.
- Smith, G.R., Morgan, N. & Gustafson, E. (2000) Fishes of the Mio-Pliocene Ringold Formation, Washington: Pliocene capture of the Snake River by the Columbia River. *University of Michigan Papers on Paleontology*, 32, 1–47.
- Smith, H.M. (1931) Descriptions of new genera and species of Siamese fishes. *Proceedings of the United States National Museum*, 79, 1–48, pl. 1.
- Smith, H.M. (1933) Contributions to the ichthyology of Siam [II–VI]. *Journal of the Siam Society, Natural History Supplement*, 9, 53–87, pls. 1–3.
- Smith, H.M. (1934) Contributions to the ichthyology of Siam: IX–XIX. *Journal of the Siam Society, Natural History Supplement*, 9, 287–325, pls. 10–14.
- Smith, H.M. (1939) A new genus of clariid catfishes. *Copeia*, 1939, 236.
- Smith, H.M. (1941) The proper name for the oriental catfish usually called *Plotosus anguillaris* (Bloch). *Proceedings of the Biological Society of Washington*, 54, 15–16.
- Smith, H.M. (1945) The fresh-water fishes of Siam, or Thailand. *United States National Museum, Bulletin*, 188, i–xi + 1–622, pls. 1–9.
- Smith, H.M. & Seale, A. (1906) Notes on a collection of fishes from the island of Mindanao, Philippine Archipelago, with descriptions of new genera and species. *Proceedings of the Biological Society of Washington*, 19, 73–82.
- Smith, J.A. (1875) Notice of new fishes from West Africa. I.—*Ophiocephalus obscurus*, Günther. II.—*Synodontis robbianus* nov. spec. mihi. *Proceedings of the Royal Physical Society of Edinburgh*, 8, 89–95.
- Smith, M.L. (1987) Osteology and systematics of the fossil catfishes (genus *Ictalurus*) of central Mexico. *Journal of Paleontology*, 61, 380–387.
- Soares-Porto, L.M. (1994) *Auchenipterichthys dantei*, a new species of catfish from the Amazon basin (Siluriformes: Auchenipteridae). *Ichthyological Exploration of Freshwaters*, 5, 281–287.
- Soares-Porto, L.M. (1995) A new species of *Tatia* from the Amazon basin (Siluriformes: Auchenipteridae). *Ichthyological Exploration of Freshwaters*, 6, 201–206.
- Soares-Porto, L.M. (1998) Monophyly and interrelationships of the Centromochlinae (Siluriformes: Auchenipteridae). In: Malabarba, L.R., Reis, R.E., Vari, R.P., Lucena, Z.M. & Lucena, C.A.S. (Eds.), *Phylogeny and Classification of Neotropical Fishes*, Edipucrs, Porto Alegre, pp. 331–350.
- Soares-Porto, L.M. (2000) A new species of *Centromochlus* (Siluriformes: Auchenipteridae) from the rio Negro drainage, Amazon basin, Brazil, with comments on its relationships. *Ichthyological Exploration of Freshwaters*, 11, 279–287.
- Soares-Porto, L.M., Walsh, S.J., Nico, L.G. & Netto, J.M. (1999) A new species of *Gelanoglanis* from the Orinoco and Amazon river basins, with comments on miniaturization within the genus (Siluriformes: Auchenipteridae: Centromochlinae). *Ichthyological Exploration of Freshwaters*, 10, 63–72.

- Son, Y.-M., Kim, I.-S. & Choo, I.-Y. (1987) A new species of torrent catfish, *Liobagrus obesus* from Korea. *Korean Journal of Limnology*, 20, 21–29.
- Sonnini, C.S. ([1799]) *Voyage dans la Haute et Basse Egypte, fait par Ordre de l'Ancien Gouvernement*, F. Buisson, Paris.
- Soriano-Señorans, J. (1950) Nota preliminar sobre *Loricaria* (*Loricaria*) *devicenzi* [sic] n. sp. de la Cuenca del Río Uruguay. *Revista de la Facultad de Humanidades y Ciencias*, Montevideo, 4, 265–266.
- Sousa, L.M., de & Rapp Py-Daniel, L.H. (2005) Description of two new species of *Physopyxis* Cope, 1871 and redescription of *P. lyra* (Siluriformes: Doradidae). *Neotropical Ichthyology*, 3, 625–636.
- Sowerby, A. de C. (1921) On a new silurid fish from the Yalu River, South Manchuria. *Proceedings of the United States National Museum*, 60, 1–2.
- Speirs, J.M. (1952) Nomenclature of the channel catfish and the burbot of North America. *Copeia*, 1952, 99–103.
- Spix, J.B., von & Agassiz, L. (1829–1831) *Selecta Genera et Species Piscium quos in Itinere per Brasiliam Annos MDC-CXXVII–MDCCCXX Jussu et Auspiciis Maximiliani Josephi I.... Colleget et Pingendo Curavit Dr J. B. de Spix, Typis C. Wolf, Monachii*. [Issued in two parts: part 1: i–xvi + i–ii + 1–82, pls. 1–48, in 1829; part 2: 83–138, pls. 49–101, in 1831]
- Srivastava, G.J. (1968) *Fishes of Eastern Uttar Pradesh [India]*, Vishwavidyalaya Prakashan, Varanasi, xxii + 163 p.
- Srivastava, M.P. (1989) A new hill-stream sisorid fish of the genus *Hara* (Blyth) 1860 from Kosi Belt, North Bihar, India. *Journal of Freshwater Biology*, 1, 121–125.
- Stansch, K. (1914) *Die Exotischen Zierfische in Wort und Bild. Part 8*. G. Wenzel & Sohn, Braunschweig, 349 p.
- Starks, E.C. (1906) On a collection of fishes made by P. O. Simons in Ecuador and Peru. *Proceedings of the United States National Museum*, 30, 761–800, pls. 65–66.
- Starks, E.C. (1913) *The Fishes of the Stanford Expedition to Brazil*, Stanford University Publication, University Series, 77 p., 15 pl.
- Stauffer, J. (1869) [Fishes]. In: Mombert, J.I. (Ed.), *An Authentic History of Lancaster County, in the State of Pennsylvania*, J.E. Barr & Co, Lancaster [PA], pp. 576–579.
- Steindachner, F. (1864a) Ichthyologische Notizen. *Anzeiger der Mathematisch- Naturwissenschaftlichen Classe der Kaiserlichen Akademie der Wissenschaften in Wien*, 1, 37–38.
- Steindachner, F. (1864b) Ichthyologische Notizen. *Sitzungsberichte der Kaiserlichen Akademie der Wissenschaften, Mathematisch- Naturwissenschaftlichen Classe, Wien, Abt. 1, Botanik, Zoologie, Anatomie, Geologie und Paläontologie*, 49, 200–214, pls. 1–2.
- Steindachner, F. (1866) Ichthyologische Mittheilungen, IX. *Verhandlungen der Kaiserlich Zoologisch-Botanischen Gesellschaft in Wien*, 16, 761–796, pls. 13–18.
- Steindachner, F. (1867a) Über einige Fische aus dem Fitzroy-Flusse bei Rockhampton in Ost-Australien. *Anzeiger der Mathematisch- Naturwissenschaftlichen Classe der Kaiserlichen Akademie der Wissenschaften in Wien*, 4, 7.
- Steindachner, F. (1867b) Ichthyologische Notizen, vierte Folge. *Anzeiger der Mathematisch- Naturwissenschaftlichen Classe der Kaiserlichen Akademie der Wissenschaften in Wien*, 4, 63–64.
- Steindachner, F. (1867c) Ichthyologische Notizen (5. Folge). *Anzeiger der Mathematisch- Naturwissenschaftlichen Classe der Kaiserlichen Akademie der Wissenschaften in Wien*, 4, 119–120.
- Steindachner, F. (1867d) Über einige Fische aus dem Fitzroy-Flusse bei Rockhampton in Ost-Australien. *Sitzungsberichte der Kaiserlichen Akademie der Wissenschaften, Mathematisch- Naturwissenschaftlichen Classe, Wien, Abt. 1, Botanik, Zoologie, Anatomie, Geologie und Paläontologie*, 55, 9–16, pl. 1.
- Steindachner, F. (1867e) Ichthyologische Notizen, IV. *Sitzungsberichte der Kaiserlichen Akademie der Wissenschaften, Mathematisch- Naturwissenschaftlichen Classe, Wien, Abt. 1, Botanik, Zoologie, Anatomie, Geologie und Paläontologie*, 55, 517–534, pls. 1–6.
- Steindachner, F. (1867f) Ichthyologische Notizen, V. *Sitzungsberichte der Kaiserlichen Akademie der Wissenschaften, Mathematisch- Naturwissenschaftlichen Classe, Wien, Abt. 1, Botanik, Zoologie, Anatomie, Geologie und Paläontologie*, 55, 701–717, pls. 1–3.
- Steindachner, F. (1867g) Characters of new fishes. *Annals and Magazine of Natural History (Ser. 3)*, 19, 441–442.
- Steindachner, F. (1870) Zur Fischfauna des Senegal, Zweite Abtheilung. *Sitzungsberichte der Kaiserlichen Akademie der Wissenschaften, Mathematisch- Naturwissenschaftlichen Classe, Wien, Abt. 1, Botanik, Zoologie, Anatomie, Geologie und Paläontologie*, 60, 945–995, pls. 1–8.
- Steindachner, F. (1875) Über einige neue brasilienische Siluroiden aus der Gruppe der Doradinen. *Sitzungsberichte der Kaiserlichen Akademie der Wissenschaften, Mathematisch- Naturwissenschaftlichen Classe, Wien, Abt. 1, Botanik, Zoologie, Anatomie, Geologie und Paläontologie*, 71, 138–151, pls. 1–4.

- Steindachner, F. (1876) Ichthyologische Beiträge, IV. *Sitzungsberichte der Kaiserlichen Akademie der Wissenschaften, Mathematisch- Naturwissenschaftlichen Classe, Wien, Abt. I, Botanik, Zoologie, Anatomie, Geologie und Paläontologie*, 72, 551–616, pls. 1–13.
- Steindachner, F. (1877a) Ichthyologische Beiträge, V. *Sitzungsberichte der Kaiserlichen Akademie der Wissenschaften, Mathematisch- Naturwissenschaftlichen Classe, Wien, Abt. I, Botanik, Zoologie, Anatomie, Geologie und Paläontologie*, 74, 49–240, pls. 1–15.
- Steindachner, F. (1877b) Die Süßwasserfische des südöstlichen Brasilien, III. *Sitzungsberichte der Kaiserlichen Akademie der Wissenschaften, Mathematisch- Naturwissenschaftlichen Classe, Wien, Abt. I, Botanik, Zoologie, Anatomie, Geologie und Paläontologie*, 74, 559–694, pls. 1–13.
- Steindachner, F. (1877c) Die Süßwasserfische des südöstlichen Brasilien, IV. *Sitzungsberichte der Kaiserlichen Akademie der Wissenschaften, Mathematisch- Naturwissenschaftlichen Classe, Wien, Abt. I, Botanik, Zoologie, Anatomie, Geologie und Paläontologie*, 76, 217–230, pls. 1–2.
- Steindachner, F. (1878a) Zur Fischfauna des Magdalenen-Stromes. *Anzeiger der Mathematisch-Naturwissenschaftlichen Classe der Kaiserlichen Akademie der Wissenschaften in Wien*, 15, 88–91.
- Steindachner, F. (1878b) Ichthyologische Beiträge, VI. *Sitzungsberichte der Kaiserlichen Akademie der Wissenschaften, Mathematisch- Naturwissenschaftlichen Classe, Wien, Abt. I, Botanik, Zoologie, Anatomie, Geologie und Paläontologie*, 77, 379–392, pls. 1–3.
- Steindachner, F. (1878c) Ichthyologische Beiträge, VII. *Sitzungsberichte der Kaiserlichen Akademie der Wissenschaften, Mathematisch- Naturwissenschaftlichen Classe, Wien, Abt. I, Botanik, Zoologie, Anatomie, Geologie und Paläontologie*, 78, for 1879 [separate apparently released in 1878], 377–400.
- Steindachner, F. (1879a) Über einige neue und seltene Fischarten aus den zoologischen Museum zu Wien, Stuttgart und Warschau. *Anzeiger der Kaiserlichen Akademie der Wissenschaften, Mathematisch- Naturwissenschaftlichen Classe*, 16, 29–34.
- Steindachner, F. (1879b) Über die Fauna des Orinoco bei Ciudad Bolivar, des Mamoni-Flusses bei Chepo im Isthmus von Panama und einiger Flüsse Peru's; Beiträge zur Kenntniss der Süßwasserfische Südamerikas. *Anzeiger der Kaiserlichen Akademie der Wissenschaften, Mathematisch- Naturwissenschaftlichen Classe*, 16, 149–152.
- Steindachner, F. (1879c) Ichthyologische Beiträge, VIII. *Anzeiger der Kaiserlichen Akademie der Wissenschaften, Mathematisch- Naturwissenschaftlichen Classe*, 16, 194–195.
- Steindachner, F. (1879d) Zur Fischfauna des Magdalenen-Stromes. *Denkschriften der Mathematisch- Naturwissenschaftlichen Classe der Kaiserlichen Akademie der Wissenschaften in Wien*, 39, 19–78, pls. 1–15.
- Steindachner, F. (1879e) Über einige neue und seltene Fisch-Arten aus den k. k. zoologischen Museum zu Wien, Stuttgart, und Warschau. *Denkschriften der Mathematisch- Naturwissenschaftlichen Classe der Kaiserlichen Akademie der Wissenschaften in Wien*, 41, 1–52, pls. 1–9.
- Steindachner, F. (1879f) Beiträge zur Kenntniss der Flussfische Südamerika's. *Denkschriften der Mathematisch- Naturwissenschaftlichen Classe der Kaiserlichen Akademie der Wissenschaften in Wien*, 41, 151–172, pls. 1–4.
- Steindachner, F. (1879g) Ichthyologische Beiträge, VIII. *Sitzungsberichte der Kaiserlichen Akademie der Wissenschaften, Mathematisch- Naturwissenschaftlichen Classe, Wien, Abt. I, Botanik, Zoologie, Anatomie, Geologie und Paläontologie*, 80, 119–191, pls. 1–3.
- Steindachner, F. (1880a) Beiträge zur Kenntniss der Flussfische Südamerikas, II, und Ichthyologische Beiträge, IX. *Anzeiger der Kaiserlichen Akademie der Wissenschaften, Mathematisch- Naturwissenschaftlichen Classe*, 17, 157–159.
- Steindachner, F. (1880b) Zur Fisch-fauna des Cauca und der Flüsse bei Guayaquil. *Denkschriften der Mathematisch- Naturwissenschaftlichen Classe der Kaiserlichen Akademie der Wissenschaften in Wien*, 42, 55–104, pls. 1–9.
- Steindachner, F. (1880c) Ichthyologische Beiträge, IX. *Sitzungsberichte der Kaiserlichen Akademie der Wissenschaften, Mathematisch- Naturwissenschaftlichen Classe, Wien, Abt. I, Botanik, Zoologie, Anatomie, Geologie und Paläontologie*, 82, 238–266, pls. 1–6.
- Steindachner, F. (1881a) Ichthyologische Beiträge, X. *Anzeiger der Kaiserlichen Akademie der Wissenschaften, Mathematisch- Naturwissenschaftlichen Classe*, 18, 45–46.
- Steindachner, F. (1881b) Beiträge zur Kenntniss der Flussfische Südamerika's, III, und Ichthyologische Beiträge, XI. *Anzeiger der Kaiserlichen Akademie der Wissenschaften, Mathematisch- Naturwissenschaftlichen Classe*, 18, 97–100.
- Steindachner, F. (1881c) Beiträge zur Kenntniss der Flussfische Südamerika's, II. *Denkschriften der Mathematisch- Naturwissenschaftlichen Classe der Kaiserlichen Akademie der Wissenschaften in Wien*, 43, 103–146, pls. 1–7.
- Steindachner, F. (1881d) Beiträge zur Kenntniss der Flussfische Südamerika's, III. *Denkschriften der Mathematisch-*

- Naturwissenschaftlichen Classe der Kaiserlichen Akademie der Wissenschaften in Wien*, 44, for 1882, 1–18, pls. 1–5.
- Steindachner, F. (1881e) Ichthyologische Beiträge, X. *Sitzungsberichte der Kaiserlichen Akademie der Wissenschaften, Mathematisch- Naturwissenschaftlichen Classe, Wien, Abt. I, Botanik, Zoologie, Anatomie, Geologie und Paläontologie*, 83, 179–219, pls. 1–8.
- Steindachner, F. (1881f) Ichthyologische Beiträge, XI. *Sitzungsberichte der Kaiserlichen Akademie der Wissenschaften, Mathematisch- Naturwissenschaftlichen Classe, Wien, Abt. I, Botanik, Zoologie, Anatomie, Geologie und Paläontologie*, 83, 393–408, pl. 1.
- Steindachner, F. (1882a) Ichthyologische Beiträge, XII. *Anzeiger der Kaiserlichen Akademie der Wissenschaften, Mathematisch- Naturwissenschaftlichen Classe*, 19, 142–143.
- Steindachner, F. (1882b) Beiträge zur Kenntniss der Flussfische Südamerika's, IV. *Anzeiger der Kaiserlichen Akademie der Wissenschaften, Mathematisch- Naturwissenschaftlichen Classe*, 19, 175–180.
- Steindachner, F. (1882c) Beiträge zur Kenntniss der Flussfische Südamerikas, IV. *Denkschriften der Mathematisch- Naturwissenschaftlichen Classe der Kaiserlichen Akademie der Wissenschaften in Wien*, 46, for 1883, 1–44, pls. 1–7.
- Steindachner, F. (1882d) Ichthyologische Beiträge, XII. *Sitzungsberichte der Kaiserlichen Akademie der Wissenschaften, Mathematisch- Naturwissenschaftlichen Classe, Wien, Abt. I, Botanik, Zoologie, Anatomie, Geologie und Paläontologie*, 86, 61–82, pls. 1–5.
- Steindachner, F. (1883) Ichthyologische Beiträge, XIII. *Anzeiger der Kaiserlichen Akademie der Wissenschaften, Mathematisch- Naturwissenschaftlichen Classe*, 20, 194–197.
- Steindachner, F. (1884) Ichthyologische Beiträge, XIII. *Sitzungsberichte der Kaiserlichen Akademie der Wissenschaften, Mathematisch- Naturwissenschaftlichen Classe, Wien, Abt. I, Botanik, Zoologie, Anatomie, Geologie und Paläontologie*, 88, 1065–1114, pls. 1–8.
- Steindachner, F. (1894) Die Fische Liberia's. *Notes from the Leyden Museum*, 16, 1–96, pls. 1–4.
- Steindachner, F. (1900) Erstattungen eines vorläufigen Berichtes über einige von Ihrer königlichen Hoheit Frau Prinzessin Therese von Bayern während einer Reise nach Südamerika 1898 gesammelte neue Fischarten. *Anzeiger der Kaiserlichen Akademie der Wissenschaften, Mathematisch- Naturwissenschaftlichen Classe*, 37, 206–208.
- Steindachner, F. (1901) Kükenthal's Ergebnisse einer zoologischen Forschungreise in den Molukken und Borneo. Fische. *Abhandlungen Senckenbergische Naturforschende Gesellschaft*, 25, 409–464, pls. 17–18.
- Steindachner, F. (1902) Herpetologische und ichthyologische Ergebnisse einer Reise nach Südamerika, mit einer Einleitung con Therese Prinzessin von Bayern. *Denkschriften der Mathematisch- Naturwissenschaftlichen Classe der Kaiserlichen Akademie der Wissenschaften in Wien*, 72, 89–148, pls. 1–6.
- Steindachner, F. (1906) Ueber zwei neue *Corydoras*-Arten aus dem Parnahyba und Parahimflusse im Staate Piauh. *Anzeiger der Kaiserlichen Akademie der Wissenschaften, Mathematisch- Naturwissenschaftlichen Classe*, 1906, 478–480.
- Steindachner, F. (1907a) Über eine neue *Psilichthys*-Art, *Ps. cameroni* aus dem Flusse Cubatão im Staate S. Catharina, Brasilien. *Anzeiger der Kaiserlichen Akademie der Wissenschaften, Mathematisch- Naturwissenschaftlichen Classe*, 44, 82–85.
- Steindachner, F. (1907b) Über zwei neue Arten von Süßwasserfischen aus dem Stromgebiete des Parnahyba. *Anzeiger der Kaiserlichen Akademie der Wissenschaften, Mathematisch- Naturwissenschaftlichen Classe*, 44, 152–155.
- Steindachner, F. (1907c) Ueber eine neue *Arges*-Art aus den Hohen Anden von Cayendelet. *Anzeiger der Kaiserlichen Akademie der Wissenschaften, Mathematisch- Naturwissenschaftlichen Classe*, 44, 228–229.
- Steindachner, F. (1907d) Über eine neue *Coridoras* [sic, *Corydoras*]-Art aus dem Rio Preto, einem sekundären Nebenflusse des Rio San Francisco, und eine *Xenocara*-Art aus dem Parnahyba bei Victoria und Sa. Filomena. *Anzeiger der Kaiserlichen Akademie der Wissenschaften, Mathematisch- Naturwissenschaftlichen Classe*, 44, 290–293.
- Steindachner, F. (1907e) Ueber einige Fischarten aus dem Flusse Cubatão im Staate Santa Catharina bei Theresopolis (Brasilien). *Sitzungsberichte der Kaiserlichen Akademie der Wissenschaften, Mathematisch- Naturwissenschaftlichen Classe, Wien, Abt. I, Botanik, Zoologie, Anatomie, Geologie und Paläontologie*, 116, 475–492, pls. 1–2.
- Steindachner, F. (1908a) Über eine im Rio Juraguá bei Joinville im Staate S. Catharina (Brasilien) vorkommende noch unbeschriebene *Pseudochalceus*-Art, *Ch. affinis*, sowie über eine neue Characinegattung und -art, *Joinvillea rosae*, von gleichem Fundorte. *Anzeiger der Kaiserlichen Akademie der Wissenschaften, Mathematisch- Naturwissenschaftlichen Classe*, 45, 28–31.
- Steindachner, F. (1908b) Über drei neue Characinen und drei Siluroiden aus dem Stromgebiete des Amazonas innerhalb Brasilien. *Anzeiger der Kaiserlichen Akademie der Wissenschaften, Mathematisch- Naturwissenschaftlichen Classe*,

- 45, 61–69.
- Steindachner, F. (1908c) Über drei neue Arten von Süßwasserfischen aus dem Amazonasgebiet und aus dem See Candiðus auf der Insel Formosa, ferner über die vorgerückte Altersform von *Loricaria acuta* C. V. *Anzeiger der Kaiserlichen Akademie der Wissenschaften, Mathematisch- Naturwissenschaftlichen Classe*, 45, 82–87.
- Steindachner, F. (1908d) Ueber eine noch unbekannte Art der Gattung *Bergiella* Eig. aus dem La Plata. *Anzeiger der Kaiserlichen Akademie der Wissenschaften, Mathematisch- Naturwissenschaftlichen Classe*, 45, 110–113.
- Steindachner, F. (1908e) Über eine während der brasilianischen Expedition entdeckte *Brachyplatystoma*-Art aus dem Rio Parnahyba und über eine dicht gefleckte und gestrichelte Varietät von *Giton fasciatus* aus den Gewässern von Santos (Staate Sao Paulo). *Anzeiger der Kaiserlichen Akademie der Wissenschaften, Mathematisch- Naturwissenschaftlichen Classe*, 45, 126–130.
- Steindachner, F. (1908f) Über zwei neue Siluroiden und zwei *Curimatus*-Arten, sowie über eine Varietät von *Ancistrus vittalus* [sic, *vittatus*] aus dem Amazonasgebiete innerhalb Brasiliens. *Anzeiger der Kaiserlichen Akademie der Wissenschaften, Mathematisch- Naturwissenschaftlichen Classe*, 45, 163–168.
- Steindachner, F. (1909a) Eine neue *Brachyplatystoma* und eine neue *Loricaria*-Art. *Anzeiger der Kaiserlichen Akademie der Wissenschaften, Mathematisch- Naturwissenschaftlichen Classe*, 46, 195–197.
- Steindachner, F. (1909b) Über eine *Ageneiosus* (*Pseudageneiosus*)-Art im Rio Parnahyba und Rio Puty bei Therezina. *Anzeiger der Kaiserlichen Akademie der Wissenschaften, Mathematisch- Naturwissenschaftlichen Classe*, 46, 341–342.
- Steindachner, F. (1910a) Über eine neue *Loricaria*-Art aus dem Flußgebiete des Jaraguá und der Ribeira im Staate S. Paulo und Sa. Catharina, über eine mit *Ancistrus aculeatus* (Perugia) = *Ancistrus gigas* (Blgr.) Reg. sehr nahe verwandte *Ancistrus*-Art aus dem Rio S. Francisco bei Barra, über eine neue *Corydoras*-Art aus dem Jaraguá und über die äußeren Geschlechtunterschiede von *Corydoras kronei*, Ribeira [sic]. *Anzeiger der Kaiserlichen Akademie der Wissenschaften, Mathematisch- Naturwissenschaftlichen Classe*, 47, 57–62.
- Steindachner, F. (1910b) Über eine noch unbeschriebene *Oxyloricaria* (= *Sturisoma*)-Art aus dem Rio Meta im Venezuela und über die relativen Längenmasse bei *O. rostrata* (Sp.). *Anzeiger der Kaiserlichen Akademie der Wissenschaften, Mathematisch- Naturwissenschaftlichen Classe*, 47, 410–411.
- Steindachner, F. (1910c) Über einige *Ageneiosus*- und *Farlowella*-Arten, etc. *Annalen des K. K. Naturhistorischen Hofmuseums, Wien*, 24, 399–408.
- Steindachner, F. (1910d) Die Fische des Itapocu und seiner Zuflüsse im Staate S^a Catharina (Brasilien). *Annalen des K. K. Naturhistorischen Hofmuseums, Wien*, 24, 419–433, pl. 5.
- Steindachner, F. (1911a) Über vier neue Siluroiden und Characinen aus dem Amazonasgebiete und von Ceará aus der Sammlung des Museums Göldi in Pará. *Anzeiger der Kaiserlichen Akademie der Wissenschaften, Mathematisch- Naturwissenschaftlichen Classe*, 48, 324–331.
- Steindachner, F. (1911b) Über einige neue und seltene südamerikanische Süßwasserfische. *Anzeiger der Kaiserlichen Akademie der Wissenschaften, Mathematisch- Naturwissenschaftlichen Classe*, 48, 369–376.
- Steindachner, F. (1911c) Beiträge zur Kenntniss der Fischfauna des Tanganyikasees und des Kongogebietes. *Anzeiger der Kaiserlichen Akademie der Wissenschaften, Mathematisch- Naturwissenschaftlichen Classe*, 48, 528–530.
- Steindachner, F. (1911d) Über einige neue und seltene afrikanische Süßwasserfische. *Anzeiger der Kaiserlichen Akademie der Wissenschaften, Mathematisch- Naturwissenschaftlichen Classe*, 48, 531–535.
- Steindachner, F. (1911e) Beiträge zur Kenntniss der Fischfauna des Tanganyikasees und des Kongogebietes. *Sitzungsberichte der Kaiserlichen Akademie der Wissenschaften, Mathematisch- Naturwissenschaftlichen Classe, Wien, Abt. I, Botanik, Zoologie, Anatomie, Geologie und Paläontologie*, 120, 1171–1186, pls. 1–3.
- Steindachner, F. (1912) Zur Fischfauna des Dscha, eines sekundären Nebenflusses des Congo im Bezirke Molundu des südlichen Kamerun. *Anzeiger der Kaiserlichen Akademie der Wissenschaften, Mathematisch- Naturwissenschaftlichen Classe*, 49, 443–449.
- Steindachner, F. (1913) Zur Fischfauna des Dscha, eines sekundären Nebenflusses des Kongo, im Bezirke Molundu, Kamerun. *Denkschriften der Mathematisch- Naturwissenschaftlichen Classe der Kaiserlichen Akademie der Wissenschaften in Wien*, 89, 1–64, pls. 1–9.
- Steindachner, F. (1914) Bericht über die ichthyologischen Aufsammlungen der Brüder Adolf und Albin Horn während einer im Sommer 1913 ausgeführten Reise nach Deutsch-Ostafrika. *Anzeiger der Kaiserlichen Akademie der Wissenschaften, Mathematisch- Naturwissenschaftlichen Classe*, 51, 536–538.
- Steindachner, F. (1915a) Vorläufigen bericht über einige neue Süßwasserfische aus Südamerika. *Anzeiger der Kaiserlichen Akademie der Wissenschaften, Mathematisch- Naturwissenschaftlichen Classe*, 52, 199–202.
- Steindachner, F. (1915b) Beiträge zur Kenntnis der Flussfische Südamerikas, V. *Anzeiger der Kaiserlichen Akademie der*

- Wissenschaften, Mathematisch- Naturwissenschaftlichen Classe*, 52, 217–219.
- Steindachner, F. (1915c) Ichthyologische Beiträge, XVIII. *Anzeiger der Kaiserlichen Akademie der Wissenschaften, Mathematisch- Naturwissenschaftlichen Classe*, 52, 346–349.
- Steindachner, F. (1915d) Bericht über die ichthyologischen Aufsammlungen der Brüder Adolf und Albin Horn während einer im Sommer 1913 ausgeführten Reise nach Deutsch-Ostafrika. *Denkschriften der Mathematisch- Naturwissenschaftlichen Classe der Kaiserlichen Akademie der Wissenschaften in Wien*, 92, 59–86, pls. 1–5.
- Steindachner, F. (1915e) Beiträge zur Kenntniss der Flussfische Südamerikas, V. *Denkschriften der Mathematisch- Naturwissenschaftlichen Classe der Kaiserlichen Akademie der Wissenschaften in Wien*, 93, for 1917, 15–106, pls. 1–13. [Issued first as a separate in 1915, with dual pagination.]
- Steindachner, F. & Döderlein, L. (1887) Beiträge zur Kenntniss der Fische Japan's, IV. *Denkschriften der Mathematisch- Naturwissenschaftlichen Classe der Kaiserlichen Akademie der Wissenschaften in Wien*, 53, 257–296, pls. 1–4.
- Steinitz, H. (1961) Notes on some glyptosternoid fishes (Sisoridae, Siluroidea). *Mitteilungen aus dem Hamburgischen Zoologischen Museum und Institut*, 59, 105–115.
- Stewart, D.J. (1985a) A new species of *Cetopsorhamdia* (Pisces: Pimelodidae) from the Río Napo basin of eastern Ecuador. *Copeia*, 1985, 339–344.
- Stewart, D.J. (1985b) A review of the South American catfish tribe Hoplomyzontini (Pisces, Aspredinidae), with descriptions of new species from Ecuador. *Fieldiana, Zoology (New Series)*, 25, i–iii + 1–19.
- Stewart, D.J. (1986a) Revision of *Pimelodina* and description of a new genus and species from the Peruvian Amazon (Pisces: Pimelodidae). *Copeia*, 1986, 653–672.
- Stewart, D.J. (1986b) A new pimelodid catfish from the deep-river channel of the Río Napo, eastern Ecuador (Pisces: Pimelodidae). *Proceedings of the Academy of Natural Sciences, Philadelphia*, 138, 46–52.
- Stewart, D.J. & Pavlik, M.J. (1985) Revision of *Cheirocerus* (Pisces: Pimelodidae) from tropical freshwaters of South America. *Copeia*, 1985, 356–367.
- Steykal, G.C. (1980) The grammar of family-group names as exemplified by those of fishes. *Proceedings of the Biological Society of Washington*, 93, 168–177.
- Stinton, F.C. (1962) Teleostean otoliths from the upper Tertiary strata of Sarawak, Brunei, and North Borneo. *British Borneo Geological Survey Department, Annual Report for 1962*, 75–92, figs. 1–18.
- Stinton, F.C. (1966) Fish otoliths from the London Clay. In: Casier, E. (Ed.), *Faune Ichtyologique du London Clay*, British Museum (Natural History), London, pp. 404–478, tables 1–4.
- Stinton, F.C. (1977) Fish otoliths from the English Eocene. *Palaeontographical Society Monographs*, Part 2, for 1976, 57–26, pls. 4–8.
- Stromer, E. (1904) Nematognathi aus de Fajûm und dem Natronthale in Ägypten. *Neues Jahrbuch für Mineralogie, Geologie und Paläontologie*, 1, 1–7, pl. 1.
- Subhash Babu, K.K. & Nayar, C.K.G. (2004) A new species of the blind fish *Horaglanis Menon* (Siluroidea: Clariidae) from Parappukara (Trichur District) and a new report of *Horaglanis krishnai* Menon from Ettumanur (Kottayam district) Kerala. *Journal of the Bombay Natural History Society*, 101, 296–298.
- Sudarto, Teugel, G.G. & Pouyaud, L. (2003) Description of two new *Clarias* species from Borneo (Siluriformes, Clariidae). *Cybium (3e série)*, 27 153–161.
- Sudarto, Teugels, G.G. & Pouyaud, L. (2004) Description of a new clariid catfish, *Clarias pseudonieuhofii* from west Borneo (Siluriformes, Clariidae). *Zoological Studies*, 43, 8–19.
- Sufi, S.M.K. (1963) Checklist of the fishes of the genus *Glyptothorax* Blyth of W. Pakistan with record of *Glyptothorax platypogonoides* [sic] (Bleeker) from Peshwar district (U. P.). *Biologia (Lahore)*, 9, 23–27.
- Suttkus, R.D. & Taylor, W.R. (1965) *Noturus munitus*, a new species of madtom, family Ictaluridae, from southern United States. *Proceedings of the Biological Society of Washington*, 78, 169–178.
- Svensson, G.S.O. (1933) Fresh water fishes from the Gambia River (British West Africa). *Results of the Swedish Expedition 1931. Kungl. Svenska Vetenskapsakademiens Handlingar*, 12, 1–102, pls. 1–8.
- Svetovidova, A.A. (1978) [List of holotypes, syntypes and paratypes kept in the Department of Ichthyology of the Moscow State University Museum]. *Sbornik Trudov Zoologicheskogo Muzeua Mgu*, 16, 256–263. [In Russian.]
- Swain, J. (1882) A review of Swainson's genera of fishes. *Proceedings of the Academy of Natural Sciences, Philadelphia*, 1882, 272–284.
- Swain, J. & Kalb, G.B. (1883) A review of the genus *Noturus* with a description of one new species. *Proceedings of the United States National Museum*, 5, 638–644.
- Swainson, W. (1838) *The Natural History and Classification of Fishes, Amphibians, & Reptiles, or Monocardian Animals*, vol. 1, Longman, Orme, Brown, Green and Longmans, and John Taylor, London, vi + 368 p.

- Swainson, W. (1839) *The Natural History and Classification of Fishes, Amphibians, & Reptiles, or Monocardian Animals*, vol. 2, Vol. 2, Longman, Orme, Brown, Green and Longmans, and John Taylor, London, vi + 448 p.
- Sydenham, D.H.J. (1978) Redescriptions of the type specimens of six clariid species (Pisces) from western Africa. *Zoological Journal of the Linnean Society*, 64, 347–371.
- Sydenham, D.H.J. (1980) New species of *Clarias* from West Africa (Pisces, Clariidae). *Revue de Zoologie Africaine*, 94, 659–677.
- Sydenham, D.H.J. & Olawoye, O.F. (1981) A new species of *Clarias* from Nigeria (Pisces, Clariidae). *Revue de Zoologie Africaine*, 95, 234–244.
- Sykes, W.H. (1839) On the fishes of the Deccan. *Proceedings of the Zoological Society of London*, 1838, 157–165.
- Sykes, W.H. (1840) An account of the fishes of Dukhun. *Annals and Magazine of Natural History (n. s.)*, 4, 54–62.
- Sykes, W.H. (1841) On the fishes of the Dukhun. *Transactions of the Zoological Society of London*, 2, 349–378, pls. 60–67.
- Sytchevskaya, E.K. (1986) [Palaeogene freshwater fish fauna of the USSR and Mongolia]. *Sovmestnaya Sovetsko-Mongolskaya Paleontologicheskaya Ekspeditsiya Trudy*, 29, 1–157. [In Russian.]
- Sytchevskaya, E.K. (1989) [Neogene freshwater fish fauna of Mongolia]. *The joint Soviet-Mongolian Paleontological Expedition; Transactions*, 39, 8–142 [In Russian] [not seen]
- Takako, A.K., Oliveira, C. & Oyakawa, O.T. (2005) Revision of the genus *Pseudotocinclus* (Loricariidae: Hypoptopomatinae), with the description of two new species. *Neotropical Ichthyology*, 3, 499–508.
- Talwar, P.K. (1976) A contribution to the systematics of *Arius tenuispinis* Day, 1877 (Pisces: Ariidae). *Records of the Zoological Survey of India*, 69, 291–294.
- Talwar, P.K. & Jhingran, A.G. (1991) *Inland Fishes of India and Adjacent Countries*, Oxford & IBH Publishing Co, New Delhi, Bombay, Calcutta, xvii + [36] + 1158 p., 1 map.
- Tamang, P. (1993) *A Preliminary Annotated List of Fish Expected to Occur in Bhutanese River Systems*. National Environmental Commission, Thimphu, Bhutan, 17 p.
- Tan, H.H. & Ng, H.H. (2000) The catfishes (Teleostei: Siluriformes) of central Sumatra. *Journal of Natural History*, 34, 267–303.
- Tan, T.H.T. & Ng, P.K.L. (1996) Catfishes of the *Ompok leiacanthus* (Bleeker, 1853) species group (Teleostei: Siluridae) from southeast Asia, with description of a new species. *The Raffles Bulletin of Zoology*, 44, 531–542.
- Taphorn, D., Royero, R., Machado-Allison, A. & Mago-Leccia, F. (1997) Lista actualizada de los peces de agua dulce de Venezuela. In: La Marca, E. (Ed.), *Vertebrados Actuales y Fósiles de Venezuela*, Museo de Ciencia y Tecnología de Mérida, Mérida, Venezuela, pp. 55–100.
- Taphorn, D.C. & Lilyestrom, C.G. (1983) Un nuevo pez del género *Xiliphius* (Aspredinidae) de Venezuela. *Revista UNELLEZ de Ciencia y Tecnología*, 1, 43–44, unnumbered plate.
- Taphorn, D.C. & Lilyestrom, C.G. (1984a) *Rhinodoras thomersoni*: un bagre sierra nuevo en Venezuela (Pisces, Doradidae). *Revista UNELLEZ de Ciencia y Tecnología*, 2, 87–92.
- Taphorn, D.C. & Lilyestrom, C.G. (1984b) *Lamontichthys maracaibero* y *L. llanero* dos especies nuevas para Venezuela (Pisces, Loricariidae). *Revista UNELLEZ de Ciencia y Tecnología*, 2, 93–100.
- Taphorn, D.C. & Marrero, C. (1990) *Hoplomyzon sexpapilostoma*, a new species of Venezuelan catfish (Pisces: Aspredinidae), with comments on the Hoplomyzontini. *Fieldiana, Zoology (New Series)*, 61, 1–9.
- Taverne, L. & Aloulou-Triki, A. (1974) Étude anatomique, myologique et ostéologique du genre *Synodontis* Cuvier (Pisces: Siluriformes, Mochocidae). *Annales du Musée Royal de l'Afrique Centrale, Series in 8°, Sciences Zoologiques*, 210, pl. 1–2.
- Taverne, L. & Vos, L. de (1998) *Clariallabes mutsindziensis* (Teleostei: Clariidae), nouveau silure du bassin de la Malagarasi (bassin du lac Tanganyika), Burundi. *Ichthyological Exploration of Freshwaters*, 8, 211–220.
- Taylor, W.R. (1969) A revision of the catfish genus *Noturus* Rafinesque with an analysis of higher groups in the Ictaluridae. *Bulletin of the U. S. National Museum*, 282, i–vi + 1–315, pls. 1–21.
- Taylor, W.R. (1986a) Ariidae. In: Daget, J., Gosse, J.-P. & Thys van den Audenaerde, D.F.E. (Eds.), *Check-list of the Freshwater Fishes of Africa*, Vol. 2, ISBN Bruxelles, MRAC Tervuren, ORSTOM, Paris, pp. 153–159.
- Taylor, W.R. (1986b) Ariidae. In: Smith, M.M. & Heemstra, P.C. (Eds.), *Smiths' Sea Fishes*, Macmillan South Africa, Johannesburg, pp. 211–213.
- Taylor, W.R. (1990) Ariidae. In: Quéro, J.-C., Hureau, J.-C., Karrer, C., Post, A. & Saldanha, L. (Eds.), *Check-list of the Fishes of the Eastern Tropical Atlantic*, UNESCO, Paris, pp. 230–234.
- Taylor, W.R. & Gomon, J.R. (1986) Plotosidae. In: Daget, J., Gosse, J.-P. & Thys van den Audenaerde, D.F.E. (Eds.), *Check-list of the Freshwater Fishes of Africa*, Vol. 2, ISBN Bruxelles; MRAC Tervuren; ORSTOM, Paris, pp. 160–

- Taylor, W.R. & Menezes, N.A. (1978) Family Ariidae. In: Fischer, W. (Ed.), *FAO Species Identification Sheets Fishery Purposes. Western Central Atlantic (Fishing Area 31)*, FAO, Rome.
- Tchang, T.-L. (1934) Notes on a new catfish from Kaifeng. *Bulletin of the Fan Memorial Institute of Biology, Peiping (Zoological series)*, 5, 41–43.
- Tchang, T.-L. (1935a) A new catfish from Yunnan. *Bulletin of the Fan Memorial Institute of Biology, Peiping (Zoological series)*, 6, 95–97.
- Tchang, T.-L. (1935b) Two new catfishes from south China. *Bulletin of the Fan Memorial Institute of Biology, Peiping (Zoological series)*, 6, 174–177.
- Tchang, T.-L. (1936) Study on some Chinese catfishes. *Bulletin of the Fan Memorial Institute of Biology, Peiping (Zoological series)*, 7, 35.
- Tchang, T.-L. (1960) *The Siluriform Fishes of China*, People Education Press. [In Chinese, with English summary]
- Tchang, T.-L. & Shih, H.-J. (1934) Notes on the fishes of the valley of lower Kialingkiang. *Lingnan Science Journal*, 13, 431–435.
- Tchernavin, V.V. (1944) A revision of some Trichomycterinae based on material preserved in the British Museum (Natural History). *Proceedings of the Zoological Society of London*, 114, 234–275.
- Temminck, C.J. & Schlegel, H. (1843–1850) Pisces. In: *Fauna Japonica, sive Descriptio Animalium quae in Itinere per Japoniam Suscepit Annis 1823–30 Collegit, Notis Observationibus et Adumbrationibus Illustravit P. F. de Siebold*, [Issued in parts: Parts 1–4: 1–72 (1843); parts 5–6, 73–112 (1844); parts 7–9, 113–172 and pls. 1–143 and pl. A (1845); parts 10–14, 173–269 (1846); part 15, 270–324 (1850)]
- Terofal, F. (1983) Die Fischausbeute der Brasilien-Expedition 1817–1820 con J. B. Spix und C. F. Ph. V. Martius. *Spixiana*, 9, 313–317.
- Teugels, G.G. (1980) Notes on the status and the synonyms of *Clarias pachynema* Boulenger, 1903 (Pisces, Clariidae). *Revue de Zoologie Africaine*, 94, 678–692.
- Teugels, G.G. (1981) Notes on the status and the synonyms of *Clarias laeviceps* Gill, 1862 (Pisces, Clariidae). *Revue de Zoologie Africaine*, 95, 359–370.
- Teugels, G.G. (1982a) A systematic outline of the African species of the genus *Clarias* (Pisces; Clariidae), with an annotated bibliography. *Musée Royal de l'Afrique Centrale, Annales, Sciences Zoologiques*, 236, 1–249.
- Teugels, G.G. (1982b) On the rehabilitation of *Clarias gabonensis* Günther, 1867, with a redescription and notes on its systematical status (Pisces, Clariidae). *Revue de Zoologie Africaine*, 96, 45–60.
- Teugels, G.G. (1982c) Preliminary data of a systematic outline of the African species of the genus *Clarias* (Pisces, Clariidae). *Revue de Zoologie Africaine*, 96, 731–748.
- Teugels, G.G. (1983) Notes on the status of *Clarias ngamensis* Castelnau, 1861, *C. mellandi* Boulenger, 1905, *C. prentissgrayi* (Fowler, 1930) and *C. lamottei* Daget and Planquette, 1967 (Pisces, Clariidae) with the rehabilitation of *Dinotopterooides* Fowler, 1930, as a subgenus of *Clarias*. *Cybium (3e série)*, 7 (1), 15–28.
- Teugels, G.G. (1986a) A systematic revision of the African species of the genus *Clarias* (Pisces; Clariidae). *Musée Royal de l'Afrique Centrale, Annales, Sciences Zoologiques*, 247, 1–199.
- Teugels, G.G. (1986b) Clariidae. In: Daget, J., Gosse, J.-P. & Thys van den Audenaerde, D.F.E. (Eds.), *Check-list of the Freshwater Fishes of Africa, Vol. 2*, ISBN Bruxelles; MRAC Tervuren; ORSTOM, Paris, pp. 66–101.
- Teugels, G.G. (1992a) Clariidae. In: Lévêque, C., Paugy, D. & Teugels, G.G. (Eds.), *Faune des Poissons d'Eaux Douces et Saumâtres de l'Afrique de l'Ouest, Vol. 2*, Musée Royal de l'Afrique Central, Tervuren, and Editions de l'ORSTOM, Paris, pp. 468–495.
- Teugels, G.G. (1992b) Malapteruridae. In: Lévêque, C., Paugy, D. & Teugels, G.G. (Eds.), *Faune des Poissons d'Eaux Douces et Saumâtres de l'Afrique de l'Ouest, Vol. 2*, Musée Royal de l'Afrique Central, Tervuren, and Editions de l'ORSTOM, Paris, pp. 496–499.
- Teugels, G.G. (1996) Taxonomy, phylogeny and biogeography of catfishes (Ostariophysi, Siluroidei): an overview. *Aquatic Living Resources*, 9, 9–34.
- Teugels, G.G. (2003a) State of the art of Recent siluriform systematics. In: Arratia, G., Kapoor, B.G., Chardon, M. & Diogo, R. (Eds.), *Catfishes*, Science Publishers, Enfield, NH, USA, pp. 317–352.
- Teugels, G.G. (2003b) *Clarias lamottei* (Siluriformes: Clariidae), a natural intergeneric hybrid from West Africa. *Cybium (3e série)*, 27, 11–15.
- Teugels, G.G. & Adriens, D. (2003) Taxonomy and phylogeny of Clariidae: an overview. In: Arratia, G., Kapoor, B.G., Chardon, M. & Diogo, R. (Eds.), *Catfishes*, Science Publishers, Enfield, NH, USA, pp. 465–487.
- Teugels, G.G., Denayer, B. & Legendre, M. (1990) A systematic revision of the African catfish genus *Heterobranchus*

- Geoffroy-Saint-Hilaire, 1809 (Pisces: Clariidae). *Zoological Journal of the Linnean Society*, 98, 237–257.
- Teugels, G.G., Diego, R.C., Pouyaud, L. & Legendre, M. (1999) Redescription of *Clarias macrocephalus* (Siluriformes: Clariidae) from South-East Asia. *Cybium (3e série)*, 23, 285–295.
- Teugels, G.G., Risch, L., De Vos, L. & Thys van den Audenaerde, D.F.E. (1991) Generic review of the African bagrid genera *Auchenoglanis* and *Parauchenoglanis*, with description of a new genus. *Journal of Natural History*, 25, 499–517.
- Teugels, G.G. & Roberts, T.R. (1987) *Silurus anguillaris* Linnaeus, 1758: designation as type species of *Clarias* Scopoli, 1777 and rediscovery of holotype (Pisces: Clariidae). *Zoological Journal of the Linnean Society*, 90, 95–98.
- Teugels, G.G., Skelton, P.H. & Lévéque, C. (1987) A new species of *Amphilius* (Pisces, Amphiliidae) from the Konkoure Basin, Guinea, West Africa. *Cybium (3e série)*, 11, 93–101.
- Teugels, G.G., Sudarto & Pouyaud, L. (2001) Description of a new *Clarias* species from Southeast Asia based on morphological and genetical evidence (Siluriformes, Clariidae). *Cybium (3e série)*, 25, 81–92.
- Teugels, G.G. & Thys van den Audenaerde, D.F.E. (1981) On the synonymy of the West African species *Clarias ebriensis* and *Clarias dahomeyensis* (Pisces, Clariidae). *Revue de Zoologie Africaine*, 95, 11–28.
- Thenius, E. (1952) Welsreste aus dem Unter-Pliozän des Wiener Beckens. *Neues Jahrbuch für Geologie und Paläontologie. Monatshefte*, 1952, 80–94.
- Thomas, M.R. & Burr, B.M. (2004) *Noturus gladiator*, a new species of madtom (Siluriformes: Ictaluridae) from coastal streams of Tennessee and Mississippi. *Ichthyological Exploration of Freshwaters*, 15, 351–368.
- Thominot, A. (1886) Sur quelques poissons nouveaux appartenant à la collection du Muséum d'Histoire Naturelle. *Bulletin de la Société Philomathique (Ser. 7)*, 10, 161–168.
- Thompson, Z. (1842) *History of Vermont, Natural, Civil, and Statistical, in Three Parts, with a New Map of the State, and 200 Engravings*, Published for the author, Burlington.
- Thunberg, C.P. (1787–1821) *Museum Naturalium Academiae Upsaliensis ... Praesidae. C. P. Thunberg, etc., 33 parts, Upsaliae*. [New species of fishes are in part 1, published in 1787.]
- Thunberg, C.P. (1791) Tvånné utländska fiskar. *Kongliga Svenska Vetenskaps Akademiens Handlingar, Stockholm*, 12, 190–192, pl. 6.
- Thunberg, C.P. (1792) Tvånné Japanske fiskar beskrifne. *Kongliga Svenska Vetenskaps Akademiens Handlingar, Stockholm*, 13, 29–32, pl. 1.
- Thys van den Audenaerde, D.F.E. (1964) Revision of the genus *Eutropiellus* Nichols and Lamonte (Pisces, Schilbeidae) with a description of a new species from Lower Nigeria *Eutropiellus vandeweyeri* sp. nov. *Revue de Zoologie et de Botanique Africaines*, 69, 214–230.
- Thys van den Audenaerde, D.F.E. (1965a) Description d'une nouvelle espèce de *Gephyroglanis* (Pisces, Bagridae) de la rivière Sanaga (Cameroun). *Revue de Zoologie et de Botanique Africaines*, 71, 264–273.
- Thys van den Audenaerde, D.F.E. (1965b) Description de *Parauchenoglanis boutchangai* sp. n. (Pisces, Bagridae) du bassin de la rivière Ngounié (Gabon). *Revue de Zoologie et de Botanique Africaines*, 72, 346–352.
- Thys van den Audenaerde, D.F.E. & De Vos, L.D.G. (1982) Description of *Eutropius djeremi* spec. nov. from the Cameroons (Pisces, Schilbeidae). *Revue de Zoologie Africaine*, 96, 179–184.
- Tilak, R. (1967) Studies of the osteocranum and the Weberian apparatus of the fishes of the genus *Batasio* Blyth, 1860 (Pisces: Siluroidei) with remarks on the systematic position of the genus. *Anatomischer Anzeiger*, 121, 415–434.
- Tilak, R. (1969) Descriptions of two new sisorids and a hybrid carp from Pauri Garhwal (Kamaon Hills) Uttar Pradesh. *Journal of the Inland Fishery Society of India*, 1, 37–48.
- Tilak, R. (1970a) Siluroid fishes of India, Pakistan, Burma, and Ceylon: Family Plotosidae. *Annals of Zoology (Warsaw)*, 27, 261–273.
- Tilak, R. (1970b) A new sisorid catfish of the genus *Gagata* Bleeker from India. *Zoologische Mededelingen (Leiden)*, 44, 207–215.
- Tilak, R. (1978) Redescription of *Hara hara* (Hamilton) and *Hara horai* Misra with a key to the species of *Hara* Blyth (Pisces: Sisoridae). *Bulletin of the Zoological Survey of India*, 1, 295–301.
- Tilak, R. (1982) Description of a new schilbeid fish, *Proeutropiichthys taakree burmanicus* from Burma. (Schilbeidae: Siluriformes). *Uttar Pradesh Journal of Zoology*, 1, for 1981, 34–39.
- Tilak, R. (1987) Studies on the fish fauna of Uttar Pradesh Terai. I.—On the extension of range of distribution of *Conta conta* (Hamilton) and *Chandramara chandramara* (Hamilton) (Sisoridae: Bagridae: Siluriformes). *Matsya*, 12/13, 84–92.
- Tilak, R. & Husain, A. (1975) A new sisorid catfish, *Laguvia ribeiroi kapuri* (Sisoridae: Siluriformes) from Uttar Pradesh. *Journal of the Inland Fishery Society of India*, 6, 1–5.

- Tilak, R. & Husain, A. (1976) Description of a new species of the genus *Glyptothorax* Blyth from River Yamuna, India (Pisces, Siluriformes: Sisoridae). *Annals of Zoology (Warsaw)*, 33, 229–234.
- Tilak, R. & Husain, A. (1978) Redescription of *Glyptothorax saisii* (Jenkins) (Sisoridae: Siluriformes) with remarks on its discontinuous distribution. *Annals of Zoology (Agra)*, 14, 33–40.
- Tilak, R. & Talwar, P.K. (1976) A taxonomic reassessment of *Hara horai* Misra (Pisces: Sisoridae) with a designation of neotype. *Newsletter of the Zoological Survey of India*, 2, 245–247.
- Tomoda, Y. (1961) Two new catfishes of the genus *Parasilurus* found in Lake Biwa-ko. *Memoirs of the College of Science, Kyoto University (Ser. B)*, 28, 347–354.
- Tortonese, E. (1942) Descrizione di una nuova specie ecuadoriana del genere *Pygidium* (Teleostei: Nematognathi). *Bullettino dei Musei di Zoologia ed Anatomia Comparata della Università di Torino (Ser. 4)*, 49, 1–3, pl. 1.
- Toula, F. (1889) Untersuchungen im Centralen Balkan. *Denkschriften der Kaiserlichen Akademie der Wissenschaften, Mathematisch-Naturwissenschaftliche Classe, Wien*, 55, 1–108, pls. 1–9, + map.
- Traill, T.S. (1832) Description of a *Silurus*, known in Demerara by the name of Gilbacke, more properly Geelbuik. *Memoirs of the Wernerian Natural History Society, Edinburgh*, 6, 377–380, pl. 6.
- Trajano, E. & Britski, H.A. (1992) *Pimelodella kronei* (Ribeiro, 1907) e seu sinônimo *Caecorhamdella brasiliensis* Borodin, 1927: Morfologia externa, taxonomia e evolução (Teleostomi, Siluriformes). *Boletim de Zoologia, Universidade de São Paulo*, 12, 53–89.
- Trajano, E. & Pinna, M.C.C., de. (1996) A new cave species of *Trichomycterus* from eastern Brazil (Siluriformes, Trichomycteridae). *Revue Française d'Aquariologie et Herpetologie*, 23, 85–90.
- Trajano, E., Reis, R.E. & Bichuette, M.E. (2004) *Pimelodella spelaea*: a new cave catfish from central Brazil, with data on ecology and evolutionary considerations (Siluriformes: Heptapteridae). *Copeia*, 2004, 315–325.
- Trewavas, E. (1936) Dr. Karl Jordan's expedition to South-West Africa and Angola: The fresh-water fishes. *Novitates Zoologicae (Tring)*, 40, 63–74, pls. 1–2.
- Trewavas, E. (1943) New schilbeid fishes from the Gold Coast, with a synopsis of the African genera. *Proceedings of the Zoological Society of London (Series B)*, 113, 164–171.
- Trewavas, E. (1962) Fishes of the crater lakes of the northwestern Cameroons. *Bonner Zoologische Beiträge*, 13, 146–192.
- Trewavas, E. (1964) A new species of *Irvineia*, an African genus of schilbeid fishes. *Annali del Museo Civico de Storia Naturale di Genova 'Giacomo Doria'*, 74, 388–396.
- Trewavas, E. (1974) The freshwater fishes of rivers Mungo and Meme and Lakes Kotto, Mboandong and Soden, west Cameroon. *Bulletin of the British Museum (Natural History), Zoology*, 26, 331–419, pls. 1–5.
- Triques, M.L. & Vono, V. (2004) Three new species of *Trichomycterus* (Teleostei: Siluriformes: Trichomycteridae) from the Rio Jequitinhonha basin, Minas Gerais, Brazil. *Ichthyological Exploration of Freshwaters*, 15, 161–172.
- Tschudi, J.J.von. (1846) Ichthyologie. In: *Untersuchungen über die Fauna Peruana*, Scheitlin & Zollikofer, St. Gallen, pp. ii–xxx + 1–35, pls. 1–6. [In 12 parts, 1844–46]
- Tuckey, J.K. (1818) A general notice of the animals taken by Mr. John Cranch, during the expedition to explore the source of the River Zaire. Appendix 4. In: *Narrative of an Expedition to Explore the River Zaire, Usually Called the Congo, in South Africa, in 1816*, John Murray, London, pp. 407–419.
- Turner, B.J., Difffoot, N. & Rasch, E.M. (1992) The callichthyid catfish *Corydoras aeneus* is an unresolved diploid-tetraploid sibling species complex. *Ichthyological Exploration of Freshwaters*, 3, 17–23.
- Turner, W. (1867) On a remarkable mode of gestation in an undescribed species of *Arius* (*A. boakeii*). *Journal of Anatomy and Physiology*, 1, 78–82.
- Ünlü, E. & Bozkurt, R. (1996) Notes on the catfish, *Silurus triostegus* (Siluridae) from the Euphrates River in Turkey. *Cybium (3e série)*, 20, 315–317.
- Unmack, P.J. (2001) Corrected identifications and lectotype designation for *Porochilus argenteus* (Zietz) (Osteichthyes: Siluriformes: Plotosidae). *Records of the South Australian Museum*, 34, 57–59.
- Vaillant, L.L. (1880a) Note sur le genre *Otocinclus* et description d'une espèce nouvelle. *Bulletin de la Société Philomathique (Ser. 7)*, 4, 145–148.
- Vaillant, L.L. (1880b) Synopsis des espèces de Siluridae recueillies par M. le Dr. Jobert, à Caldéron (Haute-Amazone). *Bulletin de la Société Philomathique (Ser. 7)*, 4, 150–159.
- Vaillant, L.L. (1891) Note sur un nouveau genre de Siluroïdes (*Diastatomycter*) de Bornéo. *Bulletin de la Société Philomathique (Ser. 8)*, 3, 181–182.
- Vaillant, L.L. (1892a) Sur quelques poissons rapportés du haut-Tonkin, par M. Pavie. *Bulletin de la Société Philomathique (Ser. 8)*, 4, 125–127.

- Vaillant, L.L. (1892b) [Poissons de l'Oubanghi rapportés par M. Dybowski]. *Bulletin de la Société Philomathique* (Ser. 8), 4, 2.
- Vaillant, L.L. (1893a) Notes ichthyologiques. *Bulletin de la Société Philomathique* (Ser. 8), 5, 13–17.
- Vaillant, L.L. (1893b) Sur les poissons provenant du voyage de M. Bonvalot et du Prince Henri d'Orléans. *Bulletin de la Société Philomathique* (Ser. 8), 5, 197–204.
- Vaillant, L.L. (1893c) Sur une collection de poissons recueillie par M. Chaper, à Bornéo. *Bulletin de la Société Zoologique de France*, 18, 55–62.
- Vaillant, L.L. (1894a) Note sur les poissons de la famille des Siluridées appartenant à la Faune Madécasse et description d'une espèce nouvelle. *Bulletin de la Société Philomathique* (Ser. 8), 6, 75–80.
- Vaillant, L.L. (1894b) Contribution à l'étude de la faune ichthyologique de Bornéo. *Nouvelles Archives du Muséum d'Histoire Naturelle, Paris* (Ser. 3), 5, 23–114, pls. 1–2.
- Vaillant, L.L. (1895a) Sur la dentition des *Synodontis* et diagnose d'espèces nouvelles. *Comptes Rendus de la Société Philomathique de Paris*, 16, 47–48.
- Vaillant, L.L. (1895b) Essai monographique sur les silures du genre *Synodontis*. *Nouvelles Archives du Muséum d'Histoire Naturelle, Paris* (Ser. 3), 7, 223–284, pls. 9–14.
- Vaillant, L.L. (1896a) Note sur l'œuvre ichthyologique de C. A. Lesueur. *Bulletin de la Société Philomathique* (Ser. 8), 8, 15–32, pls. 1–35.
- Vaillant, L.L. (1896b) Essai monographique sur les silures du genre *Synodontis*, suite et fin. *Nouvelles Archives du Muséum d'Histoire Naturelle, Paris* (Ser. 3), 8, 87–178.
- Vaillant, L.L. (1897) Siluroïde nouveau de l'Afrique orientale (*Chimarrhoglanis leroyi*). *Bulletin du Muséum d'Histoire Naturelle, Paris*, 3, 81–84.
- Vaillant, L.L. (1899) Note préliminaire sur les collections ichthyologiques recueillies par M. Geay en 1897 et 1898 dans la Guyane française et le Contesté franco-brésilien. *Bulletin du Muséum d'Histoire Naturelle, Paris*, 5, 154–156.
- Vaillant, L.L. (1900) Contribution à l'étude de la faune ichthyologique de la Guyane française et du contesté Franco-Brésilien. *Nouvelles Archives du Muséum d'Histoire Naturelle, Paris* (Ser. 4), 4, 126–136.
- Vaillant, L.L. (1902) Résultats zoologiques de l'expédition scientifique Néerlandaise au Bornéo central. Poissons. *Notes from the Leyden Museum*, 24, 1–166, pls. 1–2.
- Vaillant, L.L. (1904) Poissons Recueillis par M.A. Pavie en Indo-Chine. In: Pavie, A. (Ed.), *Mission Pavie Indo-Chine, 1879–1895; Vol. 3, Recherches sur l'Histoire Naturelle de l'Indo-Chine Orientale*, E. Leroux, Paris, pp. 459–470.
- Valenciennes, A. (1832a) Nouvelles observations sur le capitán de Bogota, *Eremophilus mutisii*. In: *Voyage de Humboldt et Bonpland, Deuxième Partie: Observations de Zoologie et d'Anatomie Comparée*, Paris, pp. 341–348.
- Valenciennes, A. (1832b) Poissons. In: *Voyage aux Indes-Orientales, Pendant les Années 1825–1829*, Charles Bélanger, Paris, pp. 339–399, poissons pls. 1–4.
- Valenciennes, A. (1835–1847) Poissons; Catalogue des principales espèces de poissons, rapportées de l'Amérique méridionale. In: d'Orbigny, A. (Ed.), *Voyage dans L'Amérique Méridionale (le Brésil, la République Orientale de l'Uruguay, la République Argentine, la Patagonie, la République du Chili, la République de Bolivia, la République du Pérou), Exécuté Pendant les Années 1826, 1827, 1828, 1829, 1830, 1832 et 1833. Vol. 5 (pt. 2)*, Bertrand et Levraut, Paris. [16 plates in poissons series issued between 1835 and 1842, plates 1–3 issued in 1835; 4–7 issued in 1836; text issued in 1847.]
- Van der Hoeven, J. (1849) *Handboek der Dierkunde; Tweede Verbeterde Uitgave; met Bijvoegsels en Aanmerkingen door Leuckart*, Edition 2, Amsterdam.
- Van der Horst, C.J. (1931) Some South African siluroid fishes. *Annals of the Transvaal Museum*, 14, 246–250.
- Van der Stigchel, J.W.R. (1946) *The South American Nematognathi of the Museums at Leiden and Amsterdam*, E.J. Brill, Leiden, 204 p., 3 tabs. [Also published in 1947, with the same title and pagination, in *Zoologische Mededelingen* (Leiden), 27:1–204.]
- Van der Stigchel, J.W.R. (1964) A new species of pimelodid catfish from eastern Brazil, *Pimelodella boschmai* nov. spec. *Zoologische Mededelingen* (Leiden), 39, 327–330.
- Van Neer, W. (1994) Cenozoic fish fossils from the Albertine Rift Valley in Uganda. In: Senut, B. & Pickford, M. (Eds.), *Geology and Palaeobiology of the Albertine Rift Valley, Uganda-Zaire; vol. 2, Palaeobiology-paléobiologie*, Occasional Publication, International Center for Training and Exchanges in the Geosciences, Orléans, France, pp. 89–128.
- Vanni, S. (1991) Cataloghi del Museo Zoologico "La Specola" dell'Università di Firenze. VIII.—Osteichthyes: Tipi. *Atti della Società Toscana di Scienze Naturali, Pisa (Serie B)*, 96, 219–229.
- Vari, R.P. & Ferraris, C.J., Jr. (1998) The neotropical catfish genus *Epapterus* Cope (Siluriformes: Auchenipteridae): a

- reappraisal. *Proceedings of the Biological Society of Washington*, 111, 992–1007.
- Vari, R.P. & Ferraris, C.J., Jr. (2003) Cetopsidae. In: Reis, R.E., Kullander, S.O. & Ferraris, C.J., Jr. (Eds.), *Check list of the Freshwater Fishes of South and Central America*, Edipucrs, Porto Alegre, Brazil, pp. 257–260.
- Vari, R.P., Ferraris, C.J., Jr. & Keith, P. (2003) A new *Pseudocetopsis* species (Siluriformes: Cetopsidae) from Suriname and French Guiana. *Proceedings of the Biological Society of Washington*, 116, 692–698.
- Vari, R.P., Ferraris, C.J., Jr. & Pinna, M.C.C., de. (2005) The Neotropical whale catfishes (Siluriformes: Cetopsidae: Cetopsinae), a revisionary study. *Neotropical Ichthyology*, 3, 127–238.
- Vari, R.P., Jewett, S.L., Taphorn, D.C. & Gilbert, C.R. (1984) A new catfish of the genus *Epapterus* (Siluriformes: Auchenipteridae) from the Orinoco River basin. *Proceedings of the Biological Society of Washington*, 97, 462–472.
- Vari, R.P. & Ortega, H. (1986) The catfishes of the Neotropical family Helogenidae (Ostariophysi: Siluroidei). *Smithsonian Contributions to Zoology*, 442, i–iii + 1–20.
- Vélez-Espino, L.A. (2004) Taxonomic revision, ecology, and endangerment categorization of the Andean catfish *Astroblepus ubidai* (Teleostei: Astroblepidae). *Reviews in Fish Biology and Fisheries*, 13 (for 2003), 367–378.
- Vidal, E.S. & de Lucena, C.A.S. (1999) *Pimelodus atrobrunneus*, um nova espécie de pimelodídeo do rio Uruguai superior (Teleostei, Siluriformes, Pimelodidae). *Biociências, Porto Alegre*, 7, 121–134.
- Vidthayanon, C. & Ng, H.H. (2003) *Acrochordonichthys gyrinus*, a new species of akysid catfish (Teleostei: Siluriformes) from Thailand. *Zootaxa*, 183, 1–7.
- Vidthayanon, C. & Roongthongbaisuree, S. (1993) [Taxonomy of Thai Riverine Catfishes Family Schilbeidae and Pangasiidae], *National Inland Fisheries Institute. Department of Fisheries, Bangkok, Technical Paper*, 150, 1–57. [In Thai with English summary]
- Villa, J. (1977) A new species of pimelodid catfish of the genus *Rhamdia* from Nicaragua, Central America. *Brenesia*, 133–142.
- Vinciguerra, D. (1881) Appunti ittiologici sulle collezioni del Museo Civico di Genova. IV.— Prima contribuzione alla Fauna Ittiologica dell' isola di Borneo. Siluroidei raccolti durante il viaggio dei Signori Mse G. Doria e Dr. O. Becari. *Annali del Museo Civico de Storia Naturale di Genova*, 16, 161–182.
- Vinciguerra, D. (1890) Viaggio di Leonardo Fea in Birmania e regioni vicine. XXIV.— Pesci. *Annali del Museo Civico de Storia Naturale di Genova (Ser. 2a)*, 9, 129–362, pls. 7–11.
- Vinciguerra, D. (1893) Di alcuni pesci raccolti nel paese dei Somali dall' Ing.r. L. Brichetti-Robecchi. *Annali del Museo Civico de Storia Naturale di Genova (Ser. 2a)*, 13, 448–455.
- Vinciguerra, D. (1895) Esplorazione del Giuba e dei suoi affluenti compiuta dal Cap. V. Bottego durante gli anni 1892–93 sotto gli auspicii della Società geografica Italiana. III.— Pesci. *Annali del Museo Civico de Storia Naturale di Genova (Ser. 2a)*, 15, 21–60, pl. 5.
- Vinciguerra, D. (1897) Pesci raccolti dal Cap. V. Bottego durante la sua seconda spedizione nelle regioni dei Somali e dei Galla. *Annali del Museo Civico de Storia Naturale di Genova (Ser. 2a)*, 17, 343–364.
- Vinciguerra, D. (1898) I pesci dell'ultima spedizione del Cap. Bottego. *Annali del Museo Civico de Storia Naturale di Genova (Ser. 2a)*, 19, 240–261.
- Vinciguerra, D. (1928) Pesci raccolti dal March.^{se} Saverio Patrizi nel bacino del Congo. *Annali del Museo Civico de Storia Naturale di Genova 'Giacomo Doria'*, 53, for 1928–29, 5–29, pl. 1.
- Vishwanath, W. & Darshan, A. (2005) A new catfish species of the genus *Sisor* Hamilton (Teleostei: Siluriformes) from Manipur, India. *Zoos'Print Journal*, 20, 1952–1954.
- Vishwanath, W. & Kosygin, L. (1999) A new sisomid catfish of the genus *Myersglanis* Hora & Silas, 1951, from Manipur, India. *Journal of the Bombay Natural History Society*, 96, 291–296, pl. 1.
- Vishwanath, W. & Kosygin, L. (2000) On a new species of the genus *Hara* Blyth from Manipur, India. *Indian Journal of Fisheries*, 47, 143–147.
- Vishwanath, W. & Linthoingambi, I. (2005) A new sisomid catfish of the genus *Glyptothorax* Blyth from Manipur, India. *Journal of the Bombay Natural History Society*, 102, 201–203.
- Voigt, E. (1926) Über ein bemerkenswertes Vorkommen neuer Fischolithen in einem Senongeschiebe von Cöthen in Anhalt. *Zeitschrift für Geschiebeforschung*, 2, 172–187, pl. 2.
- Volz, W. (1903a) Neue Fische aus Sumatra. *Zoologischer Anzeiger*, 26, 553–559.
- Volz, W. (1903b) Fische von Sumatra. *Zoologische Jahrbücher. Abteilung für Systematik, Geographie und Biologie der Tiere (Jena)*, 19, 347–419, pls. 25–26.
- Volz, W. (1904) Fische von Sumatra, gesammelt von Herrn G. Schneider. *Revue suisse de Zoologie, Annales de la Société zoologique suisse et du Muséum d'Histoire naturelle de Genève*, 12, 451–493.
- Walbaum, J.J. (1792) *Petri Artedi Sueci Genera Piscium. In Quibus Systema Totum Ichthyologiae Proponitur cum Clas-*

- sibus, Ordinibus, Generum Characteribus, Specierum Differentiis, Observationibus Plurimis. Redactis Speciebus 242 ad Genera 52. Ichthyologiae, pars iii, Grypeswaldiae, impensis A. F. Röse, 723 p., 3 pl.*
- Walschaerts, L. (1987) Catalogue des types de poissons récents de l'Institut Royal des Sciences naturelles de Belgique. *Documents de Travail*, 40, 1–67.
- Walsh, S.J. (1990) *A Systematic Revision of the Neotropical Catfish Family Ageneiosidae (Teleostei: Ostariophysi: Siluriformes)*, Doctoral dissertation, University of Florida, Gainesville.
- Walsh, S.J., Chapman, L.J., Rosenberger, A.E. & Chapman, C.A. (2000) Redescription of *Amphilius jacksonii* (Siluriformes: Amphiliidae) with habitat and life-history notes. *Ichthyological Exploration of Freshwaters*, 11, 163–174.
- Walsh, S.J. & Gilbert, C.R. (1995) New species of troglobitic catfish of the genus *Prietella* (Siluriformes: Ictaluridae) from northeastern México. *Copeia*, 1995, 850–861.
- Wandolleck, B. (1916) *Arges stübeli* n. sp. *Zoologischer Anzeiger*, 47, 158–162.
- Wang, D., Zhao, Y.-H. & Zhang, C.-G. (2005) Revision of giant sea-catfish *Arius thalassinus* in China and a new Chinese record of the species *A. bilineatus* (Siluriformes: Ariidae). *Acta Zoologica Sinica*, 51, 423–430.
- Wang, D., Zhao, Y.-H. & Zhang, C.-G. (2005) Revision of *Arius arius* (formerly *Arius sinensis*) in China and its sexual differences (Siluriformes: Ariidae). *Acta Zoologica Sinica*, 51, 431–439. [In Chinese, with English abstract]
- Wang, J., Li, G. & Wang, J. (1981) [The Early Tertiary fossil fishes from Sanshui and its adjacent basin, Guangdong]. *Zhongguo Gushengwu Zhi Xin Bing Zhong [= Palaeontologia Sinica], Series C*, 22, 1–90. [In Chinese.]
- Watanabe, K. (1995) *Pseudobagrus pratti* (Günther, 1892), a senior synonym of *P. emarginatus* (Regan, 1913) (Siluriformes: Bagridae). *Japanese Journal of Ichthyology*, 42, 321–324.
- Watanabe, K. (1998) Meristic variation in the endangered bagrid catfish, *Pseudobagrus ichikawai*. *Ichthyological Research*, 45, 99–104.
- Watanabe, K. & Ito, S. (1999) Population size and distribution of *Pseudobagrus ichikawai*, an endangered bagrid catfish in the Kawaura River. *Japanese Journal of Ichthyology*, 46, 15–30.
- Watanabe, K. & Maeda, H. (1995) Redescription of two ambiguous Japanese bagrids, *Pseudobagrus aurantiacus* (Temminck and Schlegel) and *P. tokiensis* Döderlein. *Japanese Journal of Ichthyology*, 41, 409–420.
- Watanabe, K., Mori, S., Nagoshi, M., Jeon, S.-R. & Shimizu, Y. (1992) Morphological differences between two bagrid catfishes, *Coreobagrus ichikawai* and *C. brevicorpus*. *Japanese Journal of Ichthyology*, 39, 157–162.
- Watanabe, K. & Okuyama, S. (1994) [Fossil remains of a bagrid catfish from the Lower Pliocene Ueno Formation of the Kobiwako Group at Ohyamada-mura, Mie Prefecture, Japan]. *Bulletin of the Mizunami Fossil Museum*, 21, 57–61, pl. 5. [In Japanese, with English abstract.]
- Watanabe, K. & Uyeno, T. (1999) Fossil bagrid catfishes from Japan and their zoogeography, with description of a new species, *Pseudobagrus ikiensis*. *Ichthyological Research*, 46, 397–412.
- Watanabe, K., Zhang, C.-G. & Zhao, Y.-H. (2002) Redescription of the East Asian bagrid catfish *Pseudobagrus kyphus* Mai, 1978, with a new record from China. *Ichthyological Research*, 49, 384–388.
- Weber, A., Allegretti, G. & Sbordoni, V. (2003) *Rhamdia laluchensis*, a new species of troglobitic catfish (Siluriformes: Pimelodidae) from Chiapas, Mexico. *Ichthyological Exploration of Freshwaters*, 14, 273–280.
- Weber, A. & Wilkens, H. (1998) *Rhamdia macuspanensis*: a new species of troglobitic pimelodid catfish (Siluriformes; Pimelodidae) from a cave in Tabasco, Mexico. *Copeia*, 1998, 998–1004.
- Weber, C. (1985) *Hypostomus dlouhyi* nouvelle espèce de poisson-chat cuirassé du Paraguay (Pisces, Siluriformes, Loricariidae). *Revue suisse de Zoologie, Annales de la Société zoologique suisse et du Muséum d'Histoire naturelle de Genève*, 92, 955–968.
- Weber, C. (1986) Révision de *Hypostomus boulengeri* (Eigenmann & Kennedy), et deux espèces nouvelles de poissons-chats du Paraguay (Pisces, Siluriformes, Loricariidae). *Revue suisse de Zoologie, Annales de la Société zoologique suisse et du Muséum d'Histoire naturelle de Genève*, 93, 979–1007.
- Weber, C. (1987) *Hypostomus microstomus* sp. nov. et autres poissons-chats cuirassés du Rio Parana (Pisces, Siluriformes, Loricariidae). *Archives des Sciences (Geneva)*, 40, 273–284.
- Weber, C. (1991) Nouveaux taxa dans *Pterygoplichthys* sensu lato (Pisces, Siluriformes, Loricariidae). *Revue suisse de Zoologie, Annales de la Société zoologique suisse et du Muséum d'Histoire naturelle de Genève*, 98, 637–643.
- Weber, C. (1992) Révision du genre *Pterygoplichthys* sensu lato (Pisces, Siluriformes, Loricariidae). *Revue Française d'Aquariologie et Herpetologie*, 19, 1–36.
- Weber, C. (2003) Hypostominae. In: Reis, R.E., Kullander, S.O. & Ferraris, C.J., Jr. (Eds.), *Check list of the Freshwater Fishes of South and Central America*, Edipucrs, Porto Alegre, Brazil, pp. 351–372.
- Weber, C. & Montoya-Burgos, J.-I. (2002) *Hypostomus fonchii* n. sp. (Siluriformes: Loricariidae) from Peru, a key species suggesting the synonymy of *Cochliodon* with *Hypostomus*. *Revue suisse de Zoologie, Annales de la Société*

- zoologique suisse et du Muséum d'Histoire naturelle de Genève*, 109, 355–368.
- Weber, C., Muller, S. & Mahnert, V. (1992) Harnischwelse Paraguays. In: Stawikowski, R. (Ed.), *Harnischwelse*. Die Aquarien- und Terrarien- Zeitschrift Sonderheft. Verlag Eugen Ulmer, Stuttgart, pp. 10–13.
- Weber, M. (1897) Beiträge zur Kenntniss der Fauna von Süd-Afrika. I.— Zur Kenntniss der Süßwasser-Fauna von Süd-Afrika. *Zoologische Jahrbücher. Abteilung für Systematik, Geographie und Biologie der Tiere, Suppl. (Jena)*, 10, 135–199.
- Weber, M. (1907) Süßwasserfische von Neu-Guinea ein Beitrag zur Frage nach dem früheren Zusammenhang von Neu-Guinea und Australien. In: *Nova Guinea. Résultats de l'expédition scientifique Néerlandaise à la Nouvelle-Guinée Vol. 5 (Zool.) pt 2*, E. J. Brill, Leiden, pp. 201–267, pls. 11–13.
- Weber, M. (1910) Neue Fische aus Niederländisch Süd-Neu-Guinea. *Notes from the Leyden Museum*, 32, 225–240, pl. 3.
- Weber, M. (1913) Süßwasserfische aus Niederländisch Süd- und Nord-Neu-Guinea. In: *Nova Guinea. Résultats de l'expédition scientifique Néerlandaise à la Nouvelle-Guinée*, 9, *Zoologie, livr. 4*, E. J. Brill, Leiden, pp. 513–613, pls. 12–14.
- Weber, M. & Beaufort, L.F. de. (1912) Der Fische. In: Maass, A. (Ed.), *Durch Zentral-Sumatra, Vol. 2*, W. Süsserott, Berlin, pp. 522–541, pls. 11–12.
- Weber, M. & Beaufort, L.F. de. (1913) *The Fishes of the Indo-Australian Archipelago. II. Malacopterygii, Myctophidea, Ostariophysi: I Siluroidea*, E. J. Brill, Leiden, xx + 404 p.
- Weber, S.I.F. (1999) *Étude Comparative de Deux Genres de Pimelodidae (Actionopterygii, Siluriformes) du Paraguay: Phenacorhamdia Dahl et Imparfinis Eigenmann & Norris*, Graduate thesis, Museum d'histoire naturelle de la Ville de Genève, Genève.
- Weiler, W. (1935) Ergebnisse der Forschungsreisen Prof. E. Stromers in den Wüsten Ägyptens. II.— Wirbeltierreste der Baharje-Stufe (unterstes Cenoman). 16. Neue Untersuchungen an den Fischresten. *Abhandlungen der Bayerischen Akademie der Wissenschaften, Mathematisch-Naturwissenschaftliche Abteilung, (n. f.)*, 32, 1–57, 3 pls.
- Weiler, W. (1956) Über eine neue Gattung der Welse (Fam. Siluridae) aus dem Pliozän von Wellershäusen. *Paläontologische Zeitschrift*, 30, 180–189.
- Weiler, W. (1968) Otolithi Piscium (Neubearbeitung). In: Westphal, F. (Ed.), *Fossilium Catalogus, Vol. 1.— Animalia; Pars 117*, Dr. W. Junk, N.V., Gravenhage.
- Weitzman, S.H. (1956) A description, supplementary notes and a figure of *Corydoras cochui* Myers and Weitzman, a Brazilian catfish. *Stanford Ichthyological Bulletin*, 7, 14–18.
- Weitzman, S.H. (1960a) Figures and description of four South American catfishes of the genus *Corydoras*, including two new species. *Stanford Ichthyological Bulletin*, 7, 140–154.
- Weitzman, S.H. (1960b) Figures and description of a South American catfish, *Corydoras reticulatus* Fraser-Brunner. *Stanford Ichthyological Bulletin*, 7, 155–161.
- Weitzman, S.H. (1961) A new catfish, *Corydoras concolor* (Callichthyidae) from Venezuela. *Proceedings of the Biological Society of Washington*, 74, 105–110.
- Weitzman, S.H. (1963) A new catfish, *Corydoras pastazensis* (Callichthyidae) from Ecuador. *Proceedings of the Biological Society of Washington*, 76, 59–64.
- Weitzman, S.H. (1964) One new species and two redescriptions of catfishes of the South American callichthyid genus *Corydoras*. *Proceedings of the United States National Museum*, 116, 115–126.
- Weitzman, S.H. & Nijssen, H. (1970) Four new species and one new subspecies of the catfish genus *Corydoras* from Ecuador, Colombia and Brazil (Pisces, Siluriformes, Callichthyidae). *Beaufortia*, 18, 119–133.
- Weitzman, S.H. & Vari, R.P. (1988) Miniaturization in South American freshwater fishes; an overview and discussion. *Proceedings of the Biological Society of Washington*, 101, 444–465.
- Werneke, D.C., Armbruster, J.W., Lujan, N.K. & Taphorn, D.C. (2005) *Hemiancistrus guahiborum*, a new suckermouth armored catfish from Southern Venezuela (Siluriformes: Loricariidae). *Neotropical Ichthyology*, 3, 543–548.
- Werneke, D.C., Sabaj, M.H., Lujan, N.K. & Armbruster, J.W. (2005) *Baryancistrus demantoides* and *Hemiancistrus subviridis*, two new uniquely colored species of loricariids from Venezuela (Siluriformes: Loricariidae). *Neotropical Ichthyology*, 3, 533–542.
- Werner, F. (1906) Ergebnisse der mit Subvention aus der Erbschaft Treitl unternommenen zoologischen Forschungsreise Dr. Franz Werner's in den ägyptischen Sudan und nach Nord-Uganda. V.— Beiträge zur kenntnis der Fischfauna des Nils. *Anzeiger der Kaiserlichen Akademie der Wissenschaften, Mathematisch- Naturwissenschaftlichen Classe*, 43, 325–327, pls. 1–4.
- Weyenbergh, H. (1877) Algunos nuevos pescados del Museo Nacional, y algunas noticias ictiológicas. *Actas de la Academia Nacional de Ciencias Exactas, Buenos Aires*, 3, 1–21, pls. 1–4.

- Wheeler, A. (1985) The Linnaean fish collection in the Linnean Society of London. *Zoological Journal of the Linnean Society*, 84, 1–76.
- Wheeler, A. & Baddokwaya, A. (1981) The generic nomenclature of the marine catfishes usually referred to the genus *Arius* (Osteichthyes — Siluriformes). *Journal of Natural History*, 15, 769–773.
- White, E.I. (1926) Eocene fishes from Nigeria. *Bulletin, Geological Survey of Nigeria*, 10, 1–82, pls. 1–18.
- White, E.I. (1934) Fossil fishes of Sokoto Province. *Bulletin, Geological Survey of Nigeria*, 14, 1–78, 15 figs., 10 pls.
- White, E.I. (1937) The name of a fossil cat-fish. *The Geological Magazine*, 74, 144.
- White, E.I. & Moy-Thomas, J.A. (1940) Notes on the nomenclature of fossil fishes. Part I.— Homonyms A–C. *Annals and Magazine of Natural History (Ser. 11)*, 5, 502–507.
- White, E.I. & Moy-Thomas, J.A. (1941) Notes on the nomenclature of fossil fishes. Part III.— Homonyms M–Z. *Annals and Magazine of Natural History (Ser. 11)*, 7, 395–400.
- Whitehead, P.J.P. (1958) A new species of *Chiloglanis* (Pisces, Mochocidae) in Kenya. *Annals and Magazine of Natural History (Ser. 13)*, 1, 197–208.
- Whitehead, P.J.P. (1962a) Two new river fishes from eastern Kenya. *Annali del Museo Civico de Storia Naturale di Genova 'Giacomo Doria'*, 73, 98–108.
- Whitehead, P.J.P. (1962b) A new species of *Synodontis* (Pisces: Mochocidae) and notes on a mormyrid fish from the eastern rivers of Kenya. *Revue de Zoologie et de Botanique Africaines*, 65, 97–120.
- Whitehead, P.J.P. (1970) The Reeves Collection of Chinese fish drawings. *Bulletin of the British Museum (Natural History), Historical Series*, 3, for 1969, 191–233, pls. 1–29.
- Whitley, G.P. (1928) Studies in ichthyology, no. 2. *Records of the Australian Museum*, 16, 211–239, pls. 16–18.
- Whitley, G.P. (1933) Studies in ichthyology, no. 7. *Records of the Australian Museum*, 19, 60–112, pls. 11–15.
- Whitley, G.P. (1935) Studies in ichthyology, no. 9. *Records of the Australian Museum*, 19, 215–250, pl. 18.
- Whitley, G.P. (1938) Descriptions of some New Guinea fishes. *Records of the Australian Museum*, 20, 223–233.
- Whitley, G.P. (1939) Ichthyological genotypes: Desmarest's designations, 1874. *Australian Zoologist*, 9, 222–226.
- Whitley, G.P. (1940a) The Nomenclator Zoologicus and some new fish names. *Australian Naturalist*, 10, 241–243.
- Whitley, G.P. (1940b) Illustrations of some Australian fishes. *Australian Zoologist*, 9, 397–428, pls. 30–31.
- Whitley, G.P. (1941a) The catfish and its kittens. *Australian Museum Magazine*, 7, 306–313.
- Whitley, G.P. (1941b) Ichthyological notes and illustrations. *Australian Zoologist*, 10, 1–50, pls. 1–2.
- Whitley, G.P. (1944) New sharks and fishes from Western Australia. *Australian Zoologist*, 10, 252–273.
- Whitley, G.P. (1947) New sharks and fishes from Western Australia.— Part 3. *Australian Zoologist*, 11, 129–150, pl. 11.
- Whitley, G.P. (1956a) List of the native freshwater fishes of Australia. *Proceedings of the Royal Zoological Society of New South Wales*, 1954–55, 39–47.
- Whitley, G.P. (1956b) A new catfish from New Guinea. *Proceedings of the Royal Zoological Society of New South Wales*, 1954–55, 68.
- Wilkens, H. (1993) A new species of *Rhamdia* (Pisces: Pimelodidae) from a cave in the Sierra de Zongolica (Veracruz, Mexico). *Mitteilungen aus dem Hamburgischen Zoologischen Museum und Institut*, 90, 375–378.
- Willoughby, N.G. (1994) The taxonomy of the genus *Synodontis* (Pisces: Siluroidea) in Lake Kainji, Nigeria. *The African Journal of Tropical Hydrobiology and Fisheries*, 5, 25–30.
- Woodward, A.S. (1887) On some remains of siluroid fishes from British Eocene formations. *The Geological Magazine, new series, decade 3*, 4, 303–307.
- Woodward, A.S. (1889a) *Catalog of the Fossil Fishes in the British Museum Natural History*, Vol. 1, Trustees, London, xlvi + 474 p., 17 pl.
- Woodward, A.S. (1889b) Note on *Bucklandium diluvii* König, a siluroid fish from the London Clay of Sheppey. *Proceedings of the Zoological Society of London*, 1889, 208–210, pl. 22.
- Woodward, A.S. (1899) Considerações sobre alguns peixes Terciários dos schistos de Taubaté, Estado de S. Paulo, Brasil. *Revista do Museu Paulista*, 3, for 1898, 63–70, pls. 2–4.
- Woodward, A.S. (1901) *Catalogue of the Fossil Fishes in the British Museum. Part IV.— Containing the Actinopterygian Teleostomi of the Suborders Isospondyli (in part), Ostariophysi, Apodes, Percesoces, Hemibranchii, Acanthopterygii, and Anacanthini*, Trustees, London, xxxviii + 636 p., 19 pls., 22 tables.
- Worthington, E.B. (1933a) The fishes (other than Cichlidae) of Lake Bangweulu and adjoining regions, including descriptions of three new species. *Annals and Magazine of Natural History (Ser. 10)*, 12, 34–52.
- Worthington, E.B. (1933b) The fishes of Lake Nyasa (other than Cichlidae). *Proceedings of the Zoological Society of London*, 1933, 285–316.
- Worthington, E.B. & Ricardo, C.K. (1937) The fish of Lake Tanganyika (other than Cichlidae). *Proceedings of the Zoo-*

- logical Society of London*, 1936, 1061–1112.
- Wosiacki, W.B. (2004) New species of the catfish genus *Trichomycterus* (Siluriformes: Trichomycteridae) from the headwaters of the rio São Francisco basin, Brazil. *Zootaxa*, 592, 1–12.
- Wosiacki, W.B. (2005) A new species of *Trichomycterus* (Siluriformes: Trichomycteridae) from south Brazil and redescription of *T. iheringi* (Eigenmann). *Zootaxa*, 1040, 49–64.
- Wosiacki, W.B. & Garavello, J.C. (2004) Five new species of *Trichomycterus* from the rio Iguaçu (rio Paraná Basin), southern Brazil (Siluriformes: Trichomycteridae). *Ichthyological Exploration of Freshwaters*, 15, 1–16.
- Wosiacki, W.B. & Oyakawa, O.T. (2005) Two new species of catfish genus *Trichomycterus* (Siluriformes — Trichomycteridae) from the rio Ribeira de Iguaçu Basin, Southeastern Brazil. *Neotropical Ichthyology*, 3, 465–472.
- Wu, H.-W. (1930a) Description de poissons nouveaux de Chine. *Bulletin du Muséum National d'Histoire Naturelle (2e Série)*, 2, 255–259.
- Wu, H.-W. (1930b) Notes on some fishes collected by the Biological Laboratory Science Society of China. *Contributions from the Biological Laboratory of the Science Society of China, Zoological Series*, 6, 45–57.
- Wu, H.-W. (1930c) On some fishes collected from the upper Yangtse Valley. *Sinensis*, 1, 65–86.
- Wu, H.-W. (1939) On the fishes of Li-Kiang. *Sinensis*, 10, 92–142, pls. 1–3.
- Wu, X.-W., He, M.J. & Chu, S.-L. (1981) [On the fishes of Sisoridae from the region of Xizang]. *Oceanologia et Limnologia Sinica*, 12, 74–79. [In Chinese.]
- Wu, Y.-F. & Chen, Y. (1979) [Notes on fishes from Golog and Yushu region of Qinghai Province, China]. *Acta Zootaxonomica Sinica*, 4, 287–296. [In Chinese.]
- Wulff, J.C. (1765) *Ichthyologia, cum Amphibiis Regni Borsussici. Methodo Linnaeana Disposita*. Regiomonti, xii + 60 p.
- Xie, Z., Zhang, E. & He, S.-P. (2001) Study on species validation for *Glytothorax sinense* [sic] (Regan) and *G. fukiensis* [sic] (Rendahl) with the method of morphometrics. *Journal of Huazhong Agricultural University*, 20, 169–172. [In Chinese, with English abstract]
- Yerger, R.W. & Relyea, K. (1968) The flat-headed bullheads (Pisces: Ictaluridae) of the southeastern United States, and a new species of *Ictalurus* from the Gulf Coast. *Copeia*, 1968, 361–384.
- Young, T.S. & Laerm, J. (1993) A Late Pleistocene vertebrate assemblage from the St. Marks River, Wakulla County, Florida. *Brimleyana*, 18, 15–57.
- Zakaria-Ismail, M. (1992) Notes on the catfish *Lrides sinensis* (Pisces: Pangasiidae) from Peninsular Malaysia. *Malayan Nature Journal*, 46, 111–114.
- Zarske, A. (1998) *Phenacorhamdia nigrolineata* spec. nov., ein neuer Antennenwels aus dem Einzugsgebiet des Rio Ucayali in Peru (Teleostei: Siluriformes: Pimelodidae). *Zoologische Abhandlungen; Staatliches Museums für Tierkunde in Dresden*, 50, 27–31.
- Zarske, A. (2003) Wiederbeschreibung von *Rhamdia marthae* (Sands & Black, 1985) (Teleostei, Siluriformes, Pimelodidae). Redescription of *Rhamdia marthae* (Sands & Black, 1985) (Teleostei, Siluriformes, Pimelodidae). *Zoologische Abhandlungen; Staatliches Museums für Tierkunde in Dresden*, 53, 47–55.
- Zhang, Y. & Wang, D. (1996) Studies on the taxonomical position of *Pseudobagrus brevicaudatus* (Bagridae, Siluriformes). *Acta Hydrobiologica Sinica*, 20, 379–382. [In Chinese.]
- Zhao, Y., Lan, J. & Zhang, C. (2004) A new species of amblycipitid catfish, *Xiurenbagrus gigas* (Teleostei: Siluriformes), from Guangxi, China. *Ichthyological Research*, 51, 228–232.
- Zheng, C.-Y. (1979) [On a new catfish of the genus *Leiocassis* from Guangdong Province, China]. *Acta Zootaxonomica Sinica*, 4, 182–184. [In Chinese.]
- Zheng, P.-S. (Ed.) (1981) [*Freshwater Fishes of Guangxi Province*], Guangxi People's Publishers, 257 p. [In Chinese.]
- Zhou, W. & Chu, X.-I. (1992) [A new species of *Pseudecheneis* with comments on osteological differentiations at species level (Siluriformes: Sisoridae)]. *Acta Zootaxonomica Sinica*, 17, 110–115. [In Chinese, with English abstract.]
- Zhou, W. & Zhou, Y.-W. (2005) Phylogeny of the genus *Pseudecheneis* (Sisoridae) with an explanation of its distribution pattern. *Zoological Studies*, 44, 417–433.
- Zietz, A.H.C. (1896) Description of an additional new species of fish from the Finke and Barcoo rivers. In: Spencer, B. (Ed.), *Report on the Work of the Horn Scientific Expedition to Central Australia; Part 2.—Zoology*, Dulau and Co., London, and Melville, Mullen and Slade, Melbourne, Australia, pp. 410–411, pl. 16.
- Zolezzi, G. (1939) Descrizione di tre nuovi pesci del Giuba raccolti dalla Missione Ittiologica in A.O.I. *Bollettino di Pesca, Piscicoltura e Idrobiologia*, 15, 168–173.
- Zuanon, J.A.S., Rapp Py-Daniel, L.H. & Jégu, M. (1993) Two new species of *Aguarunichthys* from the Amazon basin (Siluroidei: Pimelodidae). *Ichthyological Exploration of Freshwaters*, 4, 251–260.

INDEX

Names in bold indicate valid taxa; page numbers in italics indicate primary entry for the name.

- 5-tentaculatus, *Heterobranchus*, 197
6-tentaculatus, *Heterobranchus*, 197,
200
7-radiatus, *Silurus*, 122
9-radiatus, *Silurus*, 201
11-radiatus, *Silurus*, 335
12-radiatus, *Silurus*, 177
16-radiatus, *Silurus*, 44
38-radiatus, *Silurus*, 70
A
aaldereni, *Hemipimelodus*, 32
abbreviatus, *Bagrus*, 95
abbreviatus, *Clarias*, 148
abbreviatus, *Plotosus*, 351
aboinensis, *Clarias (Clarioides)*, 144
Abron, 342
absconditus, *Pimelodus*, 336
abuelo, *Cheirocerus*, 332
abuelo, *Sovichthys*, 332
aburrense, *Chaetostoma*, 227
aburrensis, *Hypostomus*, 227
Acanthicini, 217
Acanthicus, 217, 218
acanthicus, *Rinelepis*, 218
Acanthicus adonis, 218
Acanthicus canensis, 266
Acanthicus hystrix, 218
Acanthobunocephalus, 57
Acanthobunocephalus nicoi, 57
acanthochiroides, *Batrochoglanis*,
352
acanthochiroides, *Pseudopimelodus*,
352
acanthochirus, *Pseudopimelodus*, 353
Acanthocleithron, 303
Acanthocleithron chapini, 303
Acanthodemus, 278
Acanthodoras, 167
Acanthodoras cataphractus, 167
Acanthodoras depressus, 167
Acanthodoras spinosissimus, 168
acanthomias, *Synodontis*, 311
Acanthonotus, 356
Acanthonotus Hardwickii, 356
acanthoperca, *Synodontis*, 441
Acanthopoma, 403
Acanthopoma annectens, 403
Acanthopoma bondi, 413
accipenser, *Loricaria*, 270
Acentronichthys, 180
Acentronichthys leptos, 180
Acestra, 217, 237
Acestra acus, 237
Acestra amazonum, 237
Acestra gladiolus, 237
Acestra gladius, 239
Acestra Knerii, 238
Acestra oxyrrhyncha, 239
Acestra oxyrryncha, 239
acestrichthys, *Farlowella*, 238
Acestridiinae, 217
Acestridium, 217, 218
Acestridium colombiense, 218
Acestridium colombiensis, 218
Acestridium dichromum, 218
Acestridium discus, 218
Acestridium martini, 218
Acestrini, 217
acicularis, *Sperata*, 106
Acipenser plecostomus, 258, 259
acienserinus, *Hemiodontichthys*, 247
acienserinus, *Leptodoras*, 172
acienserinus, *Oxydoras*, 172
acrensis, *Corydoras*, 112
acrocephalus, *Arius*, 47
Acrochordonichthyini, 12
Acrochordonichthys, 12
Acrochordonichthys Büttikoferi, 13
Acrochordonichthys chamaeleon, 12
Acrochordonichthys falcifer, 12
Acrochordonichthys guttatus, 12
Acrochordonichthys gyrinus, 12
Acrochordonichthys ischnosoma, 12
Acrochordonichthys mahakamensis,
12
Acrochordonichthys melanogaster, 13
Acrochordonichthys obscurus, 13
Acrochordonichthys pachyderma, 13
Acrochordonichthys platycephalus, 13
Acrochordonichthys rugosus, 13
Acrochordonichthys septentrionalis,
13
Acrochordonichthys strigosus, 13
Acrochordonichthys varius, 13
Acrochordonichthys zonatus, 13
aculeatus, *Chaetostomus*, 272
aculeatus, *Leioccasis*, 92
aculeatus, *Plecostomus*, 224
aculeatus, *Pterygoplichthys*, 272
acus, *Acestra*, 237
acus, *Farlowella*, 237
acus venezuelensis, *Farlowella*, 239
acuta, *Loricaria*, 269
acuticeps, *Auchenoglanis*, 154
acutirostris, *Oxyropsis*, 276
acutirostris, *Arius*, 31
acutirostris, *Chrysichthys*, 158
acutirostris, *Neotropius*, 359
acutirostris, *Oxyropsis*, 276
acutirostris, *Pseudeutropius*, 359
acutivelis, *Arius*, 155, 158
acutus, *Arius*, 51
acutus, *Corydoras*, 112
acutus, *Loricariichthys*, 269
Adansonii, *Bagrus*, 365
Adelopeltis, 107
Adelopeltis angusticeps, 107
Adelopeltis laticeps, 107
adiposalis, *Pseudobagrus*, 101
adolfoi, *Corydoras*, 112
adonis, *Acanthicus*, 218
adspersus, *Callichthys*, 127
aegyptiaca, *Eopeyeria*, 428
aegyptiacus, *Ariopsis*, 428
Aelurichthys, 37
Aelurichthys isthmensis, 38
Ælurichthys longispinis, 38
Ælurichthys nuchalis, 38
Aelurichthys panamensis, 37, 38
Aelurichthys pinnimaculatus, 38
Aelurichthys scutatus, 38
aeneum, *Hoplosoma*, 112
aeneus, *Ariodes*, 49
aeneus, *Corydoras*, 112
aeneus, *Pimelodus*, 215
aequalicuspis, *Rineloricaria*, 293
aequibarbis, *Arius*, 55
aequilabilis, *Pangasius*, 327
aequinoctiale, *Chaetostoma*, 227
aequinoctialis, *Chaetostomus*, 227
aequus, *Otolithus (Arius)*, 435
affine, *Platystoma*, 330
affinis, *Ailia*, 356
affinis, *Amblydoras*, 168
affinis, *Auchenipterus*
(*Pseudauchenipterus*), 76
affinis, *Bagrus*, 107
affinis, *Batasio*, 85
affinis, *Callichthys*, 111
affinis, *Doras*, 168
affinis, *Hassar*, 171
affinis, *Hypostomus*, 251
affinis, *Macrotocinclus*, 271
affinis, *Malapterurus*, 301
affinis, *Otocinclus*, 271
affinis, *Oxydoras*, 171
affinis, *Pimelodus*, 208
affinis, *Plecostomus*, 251
affinis, *Pseudauchenipterus*, 76
affinis, *Silurus (Callichrus)*, 374
afghana, *Silurus*, 376
africana, *Arius*, 31
africana, *Arius falcarius*, 31
africanus, *Otolithus (Arius)*, 435

- afrofischeri*, *Synodontis*, 311
Afro-fischeri, *Synodontis*, 311
Agamyxis, 168
Agamyxis albomaculatus, 168
Agamyxis pectinifrons, 168
Agassizi, *Pseudopimelodus*, 353
agassizii, *Cathorops*, 39
agassizii, *Corydoras*, 112
Agassizii, *Pinirampus*, 339
Agassizii, *Rhinelepis*, 289
agassizii, *Tachisurus*, 39
agastor, *Paraloricaria*, 278
agboyiensis, *Clarias*, 139
agboyiensis, *Clarias (Clarioides)*, 139
Agenciosus polystictus, 70
Ageneiosi, 68
Ageneiosus, 68
Ageneiosus (Pseudageneiosus) therezinae, 69
Ageneiosus armatus, 68, 69
Ageneiosus atronasus, 68
Ageneiosus barranquerensis, 69
Ageneiosus brevifilis, 68, 69
Ageneiosus brevis, 69
Ageneiosus caucanus, 70
Ageneiosus childreni, 366
Ageneiosus dentatus, 70
Ageneiosus freiei, 70
Ageneiosus gabardini, 69
Ageneiosus guianensis, 70
Ageneiosus inermis, 69
Ageneiosus madeirensis, 69
Ageneiosus magoi, 69
Ageneiosus marmoratus, 69
Ageneiosus marquesi, 70
Ageneiosus melanopogon, 68
Ageneiosus militaris, 68, 69
Ageneiosus mino, 38
Ageneiosus ogilviei, 69
Ageneiosus pardalis, 70
Ageneiosus paraguensis, 70
Ageneiosus piperatus, 70
Ageneiosus polystictus, 70
Ageneiosus quadrifilis, 78
Ageneiosus rondoni, 69
Ageneiosus sebae, 69
Ageneiosus ucayalensis, 70
Ageneiosus uruguayanus, 70
Ageneiosus valenciennesi, 69
Ageneiosus virgo, 70
Ageneiosus vittatus, 70
Ageneius polystictus, 70
Ageniosus (Silonia) diaphina, 86
Ageniosus (Silonia) lurida, 366
Ageniosus axillaris, 69
Ageniosus militaris, 69
Ageniosus porphyreus, 70
ageneiosus, *Batrachocephalus*, 38
Agenius, 68
Aglyptosternon, 387
Agmus, 58
Agmus lyriiformis, 60
agna, *Hypostomus*, 251
agnā, *Plecostomus*, 251
agricolus, *Bagrus*, 107
aguaboensis, *Ancistrus*, 219
aguadulce, *Cathorops*, 39
aguadulce, *Galeichthys*, 39
aguanaí, *Exallodontus*, 333
aguuarague, *Trichomycterus*, 441
Aguarunichthys, 329
Aguarunichthys inpai, 329
Aguarunichthys tocantinsensis, 329
Aguarunichthys torosus, 329
aguilerae, *Silvaichthys*, 76
agustini, *Farlowella*, 240
ahli, *Auchenoglanis*, 162
ahli, *Parauchenoglanis*, 162
Ailia, 356
Ailia affinis, 356
Ailia coila, 356
Ailia occidentalis, 360
Ailia punctata, 356
Ailia somalensis, 360
Ailichthyoidei, 356
Ailiichthys, 356
Ailiichthys punctata, 356
Ailurichthys, 37
ailurus, *Pimelodus*, 205
akamai, *Micromyzon*, 61
akhtari, *Glyptosternon*, 387
akhtari, *Glyptosternum*, 387
akiri, *Anaspidoglanis*, 154
akiri, *Auchenoglanis*, 154
aky, *Epauctionotus*, 236
Akyses, 12
Akysidae, 12
Akysis, 12
Akysis alfredi, 16
Akysis armatus, 98
Akysis baramensis, 16
Akysis brachybarbatus, 13
Akysis clavulus, 14
Akysis clinatus, 14
Akysis ephippifer, 14
Akysis filifer, 16
Akysis fuliginatus, 14
Akysis fuscus, 16
Akysis hendricksoni, 14
Akysis heterurus, 14
Akysis inermis, 16
Akysis kurzii, 18
Akysis leucorhynchus, 16
Akysis longifilis, 442
Akysis macronema, 16, 17
Akysis maculipinnis, 14
Akysis major, 391
Akysis meridionalis, 17
Akysis microps, 14
Akysis nitidus, 17
Akysis pictus, 14
Akysis prashadi, 14
Akysis pseudobagarius, 16, 17
Akysis recavus, 14
Akysis similis, 17
Akysis sinensis, 17
Akysis sinesis, 17
Akysis variegatus, 15
Akysis variegatus variegatus, 14
Akysis varius, 15
Akysis vespa, 15
alaknandi, *Glyptothorax*, 388
alaknandi, *Glyptothorax brevipinnis*, 388
alasensis, *Mystus*, 94
alatus, *Arius*, 53
alatus, *Hypostomus*, 251
albater, *Aspidoras*, 108
albater, *Noturus*, 211, 441
alberti, *Synodontis*, 311
albescens, *Glanidium*, 75
albicans, *Arius*, 337
albicans, *Bagrus*, 54
albicans, *Pimelodus*, 337
albicollaris, *Leiocassis*, 104
albicrux, *Trachelyopterus*, 79
albicrux, *Trachycorystes*, 79
albidus, *Arius*, 337
albidus, *Callichthys*, 128
albidus, *Pimelodus*, 40, 204
albifasciatus, *Bunocephalus*, 62
albilabris, *Bagrus*, 95
albilabris, *Paraplotosus*, 349
albilabris, *Plotosus*, 349
albinotatus, *Trichomycterus*, 414
albocinctus, *Hemiancistrus*, 223
albofasciatus, *Pimelodus*, 337
alboleata, *Synodontis*, 311
alboleatus, *Corydoras*, 112
alboleatus, *Mystus*, 94
alboleatus, *Synodontis*, 311
albomaculatus, *Agamyxis*, 168
albomaculatus, *Doras*, 168
albomaculatus, *Panaqolus*, 277
albomaculatus, *Panaque*, 277
albomarginatus, *Cephalosilurus*, 353
albomarginatus, *Leiocassis*, 101
albomarginatus, *Pseudobagrus*, 101
albomarginatus, *Pseudopimelodus*, 353
albopunctatus, *Clarias*, 139
albopunctatus, *Hypostomus*, 251
albopunctatus, *Plecostomus*, 251
aleppensis, *Macrones*, 97
aleuropsis, *Bunocephalus*, 58
alexandri, *Lophiosilurus*, 353
alfaroi, *Rhamdia*, 199

- alfredi*, *Akysis*, 16
alfredi, *Pseudobagarius*, 16
alga, *Chaetostomus*, 221
alikunhii, *Horaglanis*, 152
alipionis, *Hemipsilichthys*, 262
alipionis, *Isbrueckerichthys*, 262
Allabenchelys, 137
Allabenchelys attensi, 137
Allabenchelys brevior, 137
Allabenchelys dhonti, 142
Allabenchelys engelseni, 142
Allabenchelys laticeps, 137
Allabenchelys longicauda, 137
Allabenchelys manyangae, 137
Allabenchelys pietschmanni, 138
allenii, *Apomatoceros*, 403
alluaudi, *Clarias*, 139
aloikae, *Bunocephalus amaurus*, 58
alta, *Tymanopleura*, 69
altae, *Centromochlus*, 73
altamazonicum, *Leptoplosternum*, 128
alternatum, *Pygidium*, 414
alternatus, *Trichomycterus*, 414
alternifasciatum, *Chaetostoma*, 227
alternus, *Ochmacanthus*, 409
alterum, *Pygidium*, 415
alterus, *Trichomycterus*, 415
altifrons, *Pangasius*, 325
altipinnis, *Auchenoglanis*, 162
altipinnis, *Eutropius*, 365
altipinnis, *Hemiloricaria*, 245
altipinnis, *Liposarcus*, 291
altipinnis, *Loricaria*, 245
altipinnis, *Parauchenoglanis*, 162
altipinnis, *Pimelodella*, 189
altipinnis, *Pimelodus*, 189
altissimus, *Pimelodus*, 337
altocorpus, *Farlowella*, 444
altus, *Silurus*, 381
atula, *Doumea*, 24
aluensis, *Chrysichthys*, 155
alvarezi, *Gymnallabes*, 150
amandajanea, *Corydoras*, 112
amapaensis, *Corydoras*, 113
Amaralia, 57
Amaralia hypsiura, 57
Amarginops, 153
Amarginops hildae, 153
Amarginops platus, 153
amatitlanensis, *Rhamdia*, 198
amaurus, *Bunocephalus* 58
amazona, *Farlowella*, 237
amazonae, *Helogenes*, 134
amazonensis, *Parotocinclus*, 281
amazonica, *Dekeyseria*, 118
amazonica, *Loricaria*, 270
amazonicus, *Bunocephalus*, 59
amazonicus, *Dysichthys*, 59
amazonicus, *Ituglanis*, 406
amazonicus, *Trichomycterus*, 406
amazonum, *Acesta*, 237
amazonum, *Loricaria apeltogaster*, 288
amazonum, *Rhinodoras*, 170
amazonus, *Pseudohemiodon*, 288
ambiacus, *Corydoras*, 113
amblops, *Felichthys*, 79
amblops, *Trachelyopterus*, 79
Amblycepinae, 17
Amblyceps, 17, 98
Amblyceps apangi, 17
Amblyceps arnunachalensis, 17
Amblyceps caecutiens, 17, 18
Amblyceps cæcutiens, 18
Amblyceps carinatum, 18
Amblyceps deyi, 17
Amblyceps foratum, 18
Amblyceps horae, 99
Amblyceps inermis, 17
Amblyceps laticeps, 18
Amblyceps macropterus, 18
Amblyceps mangois, 18
Amblyceps marginatoides, 19
Amblyceps marginatus, 19
Amblyceps mucronatum, 18
Amblyceps murraystuarti, 18
Amblyceps murray-stuarti, 18
Amblyceps platycephalus, 18
Amblyceps serratum, 18
Amblyceps tenuispinis, 18
Amblyceps variegatum, 19
Amblycipitidae, 17
Amblydoras, 168, 179
Amblydoras affinis, 168
Amblydoras bolivarensis, 168
Amblydoras gonzalezi, 168
Amblydoras monitor, 169, 169
Amblydoras nauticus, 169
Amblydoras truncatus, 168
amblyurum, *Steindachneridion*, 343
amblyurus, *Steindachneria*, 343
amboinensis, *Cryptopterus*, 369
ambrosetti, *Liposarcus*, 291
ambrosetti, *Pterygoplichthys*, 291
amblyiacus, *Auchenipterus*, 71
Ameiurina, 203
Ameiurus, 203, 204
Ameiurus brunneus, 204
Ameiurus cancellatus, 211
Ameiurus catus, 204
Ameiurus hazenensis, 204
Ameiurus lavetti, 204
Ameiurus leidyi, 204
Ameiurus macgrewi, 204
Ameiurus maconnellii, 211
Ameiurus melas, 205
Ameiurus natalis, 205
Ameiurus nebulosus, 205
Ameiurus pectinatus, 206
Ameiurus platycephalus, 206
Ameiurus primaevus, 207
Ameiurus reticulatus, 206
Ameiurus sawrockensis, 206
Ameiurus serracanthus, 206
Ameiurus vespertinus, 206
amekiensis, *Otolithus (Arius)*, 435
amemiyae, *Aoria*, 91
amemiyai, *Aoria*, 91
americanus, *Cataphractus*, 167
Amissidens, 32
Amissidens hainesi, 32
Amiurus, 204
Amiurus australis, 208
Amiurus bolli, 205
Amiurus brachyacanthus, 205
Amiurus brunneus, 204
Amiurus cragini, 205
Amiurus decorus, 210
Amiurus dugèsii, 208
Amiurus erubennus, 205
Amiurus lophius, 204
Amiurus meeki, 209
Amiurus meridionalis, 208
Amiurus mexicanus, 209
Amiurus mispiliensis, 206
Amiurus natalis analis, 205
Amiurus niveiventris, 204
Amiurus obesus, 205
Amiurus prosthistius, 205
Ammoglanis, 403
Ammoglanis diaphanus, 403
Ammoglanis pulex, 403
ammophilus, *Aphanotorulus*, 225
amphibelus, *Corydoras*, 113
Amphiliidae, 20
Amphilus, 20, 21
Amphilus atesuensis, 21
Amphilus baudoni, 21
Amphilus baudoni uniformis, 21
Amphilus baudoni var. *uniformis.*, 21
Amphilus brevidorsalis, 23
Amphilus brevis, 21
Amphilus cryptobullatus, 21
Amphilus grammatorhus, 22, 23
Amphilus grammatorhus
brevipinna, 22
Amphilus grammatorhus
inaequalis, 23
Amphilus grammatorhus
marmoratus, 23
Amphilus grammatorhus var.
brevipinna, 23
Amphilus grammatorhus var.
inaequalis, 23
Amphilus grandis, 23
Amphilus hargeri, 23

- Amphilios jacksonii*, 21
Amphilios kakrimensis, 21
Amphilios kivuensis, 21
Amphilios krefftii, 23
Amphilios lamani, 22
Amphilios lampei, 22
Amphilios laticaudatus, 22
Amphilios lentiginosus, 22
Amphilios longirostris, 22
Amphilios maesii, 22
Amphilios natalensis, 22
Amphilios nigricaudatus, 21
Amphilios nigricaudatus multipunctata, 21
Amphilios nigricaudatus var. *multipunctata*, 21
Amphilios notatus, 27
Amphilios opistophthalmus, 22
Amphilios oxyrhinus, 23
Amphilios pictus, 21
Amphilios platychir, 22
Amphilios platychir cubangoensis, 23
Amphilios platychir var. *cubangoensis*, 23
Amphilios pulcher, 23
Amphilios pulcher ephippiata, 23
Amphilios rheophilus, 23
Amphilios transvaaliensis, 23
Amphilios uranoscopus, 23
Amphilios zairensis, 23
amphioxoa, *Cetopsis*, 131
amphiloxus, *Hemicetopsis*, 131
amplexicauda, *Clarias*, 148
Anacanthus, 301
Anadoras, 169, 179
Anadoras grypus, 169
Anadoras regani, 169
Anadoras weddelli, 169
anale, *Chaetostoma*, 227
analis, *Amiurus natalis*, 205
analis, *Hypocolpter*, 227
analis, *Leiocassis (Dermocassis)*, 101
analis, *Pseudobagrus*, 101
analis, *Trachycorystes*, 80
anamalaiensis, *Glyptothorax*, 388
ananas, *Physopyxis*, 176
Anaspidoglanis, 153
Anaspidoglanis akiri, 154
Anaspidoglanis boutchangai, 154
Anaspidoglanis macrostomus, 154
Anchariidae, 29
Ancharius, 29
Ancharius brevibarbis, 29
Ancharius fuscus, 29
Ancharius griseus, 29
Ancistomus, 283
Ancistri, 216
ancistroides, *Hypostomus*, 251
ancistroides, *Plecostomus*, 251
Ancistrus, 216, 219
Ancistrus (Hemiancistrus) yaravi, 273
Ancistrus (Pseudancistrus) coquenani, 287
Ancistrus aguaboensis, 219
Ancistrus annectens, 242
Ancistrus barrae, 272
Ancistrus baudensis, 220
Ancistrus bodenhameri, 219
Ancistrus boliviensis, 219
Ancistrus Bovallii, 267
Ancistrus brachyurus, 234
Ancistrus brevifilis, 219
Ancistrus brevifilis bodenhameri, 219
Ancistrus brevipinnis, 219
Ancistrus bufonius, 219
Ancistrus caucanus, 219
Ancistrus centrolepis, 219
Ancistrus chagresi, 220, 222
Ancistrus cirrhosus, 220
Ancistrus cirrhosus dubius, 221
Ancistrus claro, 220
Ancistrus clementinae, 220
Ancistrus cryptophthalmus, 220
Ancistrus cuiabae, 220
Ancistrus damasceni, 220
Ancistrus dolichopterus, 220
Ancistrus dubius, 221
Ancistrus erinaceus, 221
Ancistrus eustictus, 221
Ancistrus formoso, 221
Ancistrus fulvus, 221
Ancistrus füsslii, 235
Ancistrus galani, 221
Ancistrus gibbiceps, 291
Ancistrus guacharote, 225
Ancistrus guentheri, 287
Ancistrus gymnorhynchus, 221
Ancistrus heterorhynchus, 221
Ancistrus hoplogenys, 221
Ancistrus jataiensis, 222
Ancistrus jelskii, 222
Ancistrus latifrons, 222
Ancistrus leucostictus, 222
Ancistrus lineolatus, 222
Ancistrus lithurgicus, 222
Ancistrus lituratus, 290, 291
Ancistrus longimanus, 291
Ancistrus macropthalmus, 222
Ancistrus maculatus, 222
Ancistrus malacops, 222
Ancistrus maracasae, 223
Ancistrus martini, 223
Ancistrus mattogrossensis, 300
Ancistrus mattogrossensis, 300
Ancistrus medians, 242, 243
Ancistrus megalostomus, 223
Ancistrus melas, 220
Ancistrus minutus, 223
Ancistrus montanus, 223
Ancistrus multiradiatus alternans, 291
Ancistrus multispinis, 223
Ancistrus multispinis, 265
Ancistrus mystacinus, 265
Ancistrus nationi, 223
Ancistrus nudiceps, 223
Ancistrus occidentalis, 223
Ancistrus occloii, 223
Ancistrus parecis, 224
Ancistrus pictus, 234, 265
Ancistrus pirareta, 224
Ancistrus piriformis, 224
Ancistrus planiceps, 264
Ancistrus ranunculus, 224
Ancistrus reisi, 224
Ancistrus salgadae, 300
Ancistrus scaphirhynchus, 234
Ancistrus snethlageae, 283, 285
Ancistrus spinosus, 224
Ancistrus stigmaticus, 224
Ancistrus tamboensis, 224
Ancistrus taunayi, 224
Ancistrus temminckii, 224
Ancistrus tombador, 225
Ancistrus trinitatis, 225
Ancistrus triradiatus, 225
Ancistrus triradiatus martini, 223
Ancistrus variolus, 225
Ancistrus verecundus, 225
Ancistrus vittatus var. *vermiculata*, 285
Ancistrus vittatus vermiculata, 285
andamanensis, *Arius*, 56
andersoni, *Liobagrus*, 19
andersoni, *Corymbophanes*, 232
Andersonia, 23
Andersonia brevior, 24
Andersonia leptura, 24
Andersonia pellegrini, 24
andersonii, *Exostoma*, 386
andersonii, *Glaridoglanis*, 386
Andinichthyidae, 30
Andinichthys, 30
Andinichthys bolivianensis, 30
Anduzedoras, 169
Anduzedoras arleoi, 169
Anduzedoras copei, 172
Anduzedoras oxyrhynchus, 169
anduzei, *Ernstichthys*, 60
anduzei, *Trachelyopterichthys*, 78
Aneistrus (Hemiancistrus) pulcher, 234
Anemanotus, 37
anfractus, *Clarias*, 139
angeli, *Pterodoras*, 177
angelica, *Synodontis*, 311
angelicus, *Otolithus (Arius)*, 434
angelicus zonatus, *Synodontis*, 311
angelis, *Ptedoras*, 178

- angipinnatus, Hypostomus**, 251
angipinnatus, Plecostomus, 251
angius, Pimelodus, 359
angolensis, Chiloglanis, 304
angolensis, Clarias, 139
angolensis, Doumea, 25
angolensis, Eutropius, 363
angolensis, Schilbe, 363
angolensis macronema, Chlarias, 141
angosturae, Farlowella, 240
anguilla, Ictalurus, 210
anguilla, Glanapteryx, 404
anguillaris, Clarias, 140, 144
anguillaris, Platystacus, 350, 351
anguillaris, Silurus, 138, 140, 143
anguillaris nigeriensis, Clarias, 140
anguillicauda, Liobagrus, 19
anguilloclarias, 139
angulatus, Arius, 55
angulatus, Otolithus (Arius), 435
angulicauda, Plecostomus, 235
angulicauda, Delturus, 234, 444
angusticeps, Adelopeltis, 107
angustifrons, Anoplopterus, 27
angustifrons, Tetracamphilius, 27
angustirostre, Scleronema, 413
angustirostris, Pygidium, 413
anisitsi, Liposarcus, 291
anisitsi, Pterygoplithys, 291
anisitsi, Homodiaetus, 406
anisura, Phenacorhamdia, 188
anisurus, Bagrus, 90
anisurus, Heptapterus, 188
anisurus, Pimelodus, 98
annae, Chaetostomus, 297
annae, Plecostomus, 297
annae, Squaliforma, 297
annandalei, Glyptothorax, 388
annectens, Acanthopoma, 403
annectens, Ancistrus, 242
annectens, Hemiancistrus, 242
annectens, Synodontis, 312
Anodontiglanis, 345
Anodontiglanis dahli, 345
anomala, Herklotsella, 375
anomala, Pterocryptis, 375
anomala sovichthys, Chaetostoma, 230
anomalopteryx, Parakysis, 15
anomalum, Chaetostoma, 227
anomalus, Chastostomus, 227
Anoplectropius, 165
Anoplectropius henrici, 165
Anoplopterus, 21
Anoplopterus angustifrons, 27
Anoplopterus longirostris, 22
Anoplopterus uranoscopus, 21, 23
anostomus, Silurus, 374
anoterus, Chiloglanis, 304
Ansorgia, 360
Ansorgia vittata, 360, 361
Ansorgia vittata bistrata, 361
ansorgii, Chrysichthys, 155
ansorgii, Eutropius, 363
ansorgii, Parauchenoglanis, 154
ansorgii, Phractura, 26
ansorgii, Physalia, 360
ansorgii, Synodontis, 312
Ansorgiichthys, 360
anteanalis, Pareuchiloglanis, 398
anthrax, Lasiancistrus, 288
anthrax, Pseudolithoxus, 288
antiquus, Astephus, 207
antiquus, Pimelodus, 207
antoniensis, Pimelodus, 205
anus, Loricaria, 269
anus, Loricariichthys, 269
Anyperistius, 347
Anyperistius perugiae, 347
aor, Pimelodus, 106
aor, Sperata, 106
aor sarwari, Aorichthys, 107
aorella, Sperata, 107
aorellus, Bagrus, 107
Aoria, 106
Aoria amemiyae, 91
Aoria amemiyai, 91
Aoria henryi, 100
Aoria hoi, 107
Aoria lacus, 108
Aoria rendahli, 108
Aoria virgatus, 101
Aorichthys, 106
Aorichthys aor sarwari, 107
arides, Bagrus, 106
aorinus, Bagrus, 107
apangi, Amblyceps, 17
apelto gaster, Loricaria, 268
apelto gaster amazonum, Loricaria, 288
apelto gaster var. amazonum, Loricaria, 288
Aphanotorulus, 225
Aphanotorulus ammophilus, 225
Aphanotorulus frankei, 225, 226
Aphanotorulus unicolor, 225
Aristotelia caria, 226
Aristotelia caria condei, 226
Aristotelia caria laani, 226
Aristotelia caria listrorhinos, 226
Aristotelia caria ommation, 226
apithanos, Pseudohemiodon, 288
Apodoglanis, 375
Apodoglanis furnessi, 375, 376
apogon, Phalacronotus, 374
apogon, Silurus, 374
Apomatoceros, 403
Apomatoceros allenii, 403
Aposturisoma, 226
Aposturisoma myriodon, 226
approuaguensis, Corydoras, 113
Apureadoras, 177
Apureadoras rivasi, 177
apurensis, Cephalosilurus, 353
apurensis, Pseudopimelodus, 353
apurensis, Sachsdoras, 177, 178
apus, Channallabes, 137
apus, Gymnallabes, 137
aquilus, Lophiobagrus, 161
arab, Plotosus, 351
arabi, Synodontis, 321
araurensis, Arius, 49
araguaiaensis, Corydoras, 113
araguayensis, Galeichthys, 339
aralensis, Silurus glanis, 379
arcana, Cetopsis, 131
arcifer, Callichthys, 111
arcuatus, Corydoras, 113
arcuatus, Gogo, 29
arcuatus, Rhineastes, 207
areio, Corydoras, 113
arekaima, Pimelodus, 334
arenaria, Peckoltia, 284
arenarius, Arius, 34
arenarius, Bagrus (Ariodes), 34
arenarius, Hemiancistrus, 284
arenatus, Arius, 39
arenatus, Cathorops, 39
areolatus, Trichomycterus, 415
Argenei, 63
argentata, Silurus (Clupisoma), 357
argenteus, Arius, 56
argenteus, Neosilurus, 345
argenteus, Pimelodus, 53
argenteus, Plotosus, 345
argenteus, Porochilus, 345
argentina, Corydoras punctatus, 122
argentina, Megalonema, 335
argentina, Perugia, 335
argentinus, Arius, 428
argentinus, Silurus, 209
argentivittatus, Macrones, 100
argentivittatus, Pelteobagrus, 100
Arges, 63
Arges boulegeri, 63
Arges brachycephalus, 63
Arges chotae, 63
Arges cirratus, 64
Arges eigenmanni, 64
Arges festae, 64
Arges fissidens, 64
Arges heterodon, 64
Arges homodon, 64
Arges longifilis, 65
Arges marmoratus, 65
Arges orientalis, 65
Arges peruanus, 66

- Arges regani*, 66
Arges retropinna, 66
Arges sabalo, 63, 67
Arges simonsii, 67
Arges stübeli, 67
Arges taczanowskii, 67
Arges theresiae, 67
Arges vaillanti, 67
Arges whymperi, 68
argus, Hypostomus, 251
argus, Plecostomus, 251
argyropleuron, Arius, 51
argyropleuron, Plicofollis, 51
argyrus, Pimelodus, 209
Arii, 30
Ariidae, 30
Ariodes, 34
Ariodes aeneus, 49
Ariodes macrocephalus, 51
Arius tonggol, 52
Arioida, 429, 432
arioides, Bagrus, 40, 54
Ariopsis, 32
Ariopsis, 428
Ariopsis aegyptiacus, 428
Ariopsis assimilis, 33
Ariopsis bonillai, 33
Ariopsis felis, 33
Ariopsis guatemalensis, 33
Ariopsis lentiginosa, 33
Ariopsis robertsi, 48
Ariopsis seemanni, 33
Ariopsis peyeria, 432
aripuanensis, Parotocinclus, 281
Aristommata, 249
Aristommata inexpectata, 249, 250
Aristotelis, Glanis, 378
aristotelis, Silurus, 378
aristotelis, Silurus (Parasilurus), 378
Arius, 30, 34
Arius (?) bartonensis, 56
Arius (Hemiarium) carinatus, 42
Arius (Hemiarium) danielsi, 42
Arius (Hemiarium) nudidens, 56
Arius acrocephalus, 47
Arius acutirostris, 31
Arius acutivelis, 155, 158
Arius acutus, 51
Arius aequibarbis, 55
Arius africana, 31
Arius alatus, 53
Arius albicans, 337
Arius albidus, 337
Arius andamanensis, 56
Arius angulatus, 55
Arius arafurensis, 49
Arius arenarius, 34
Arius arenatus, 39
Arius argenteus, 56
Arius argentinus, 428
Arius argyropleuron, 51
Arius arius, 34
Arius armiger, 46
Arius assimilis, 33, 434
Arius augustus, 46
Arius australis, 47
Arius baroni, 434
Arius Belangerii, 54
Arius bicolor, 34
Arius bleekeri, 46
Arius boakeii, 34
Arius bonneti, 36, 56
Arius borneensis, 56
Arius Brandtii, 54
Arius brevirostris, 31
Arius brunellii, 31
Arius Buchanani, 34
Arius burmanicus, 42
Arius cacharioides, 166
Arius caelatoides, 55
Arius cælatus, 46
Arius Capellonis, 32
Arius cavatus, 435
Arius cœrulescens, 33
Arius chondropterygioides, 55
Arius chondropterygius, 55
Arius clavispinosus, 36
Arius cleptolepis, 47
Arius clipeaster, 55
Arius clypeastroides, 55
Arius coatesi, 47
Arius cochinchinensis, 34
Arius cookei, 50
Arius couis, 31
Arius crassus, 435
Arius crossocheilos, 52
Arius cruciger, 106
Arius curtisi, 47
Arius dasyccephalus, 39
Arius dayi, 49
Arius despaxi, 36
Arius Dieperinki, 32
Arius digulensis, 47
Arius diocetes, 44
Arius dispar, 34
Arius doriae, 41
Arius dussumieri, 52
Arius Dutemplei, 56
Arius egertoni, 56
Arius egertoni belgicus, 56
Arius elatturus, 50
Arius equestris, 33
Arius falcarius, 34
Arius falcarius africana, 31
Arius falcarius var. africana, 31
Arius fangi, 34
Arius festae, 56
Arius festinus, 34
Arius fissus, 39
Arius Fraasi, 56
Arius froggatti, 42
Arius Fürthii, 39
Arius gagora, 34
Arius gageroides, 35
Arius gigas, 31, 56
Arius goniaspis, 52
Arius graeffei, 47
Arius grandicassis, 49, 50
Arius grandoculis, 53
Arius granducolis, 53
Arius granosus, 54
Arius granulatus, 32
Arius guatemalensis, 33
Arius hainesi, 32
Arius hamiltoni, 51
Arius hardenbergi, 36
Arius hastatus, 105
Arius Heckelii, 55
Arius heudelotii, 31
Arius heward-belli, 56
Arius hypophthalmus, 39, 40
Arius iheringi, 343
Arius insculptus, 50
Arius insidiator, 45
Arius jella, 36
Arius kanganamanensis, 39
Arius Kessleri, 50
Arius kirkii, 52
Arius kitsoni, 56
Arius kutchensis, 56
Arius laeviceps, 55
Arius lagoensis, 31
Arius laticeps, 41
Arius latirostris, 47
Arius latiscutatus, 31
Arius layardi, 51
Arius leiotetocephalus, 52
Arius lemoinei, 432
Arius leptonotacanthus, 34
Arius longibarbis, 334
Arius luniscutis, 36
Arius macracanthus, 36
Arius macrocephalus, 51
Arius macronotacanthus, 47
Arius macruopterygius, 55
Arius maculatus, 34, 35
Arius madagascariensis, 35
Arius magatensis, 52
Arius malabaricus, 35
Arius manillensis, 35
Arius manjong, 55
Arius mastersi, 45
Arius melanochir, 41
Arius melanoptyrygius, 56
Arius melanopus, 40
Arius mercatoris, 31
Arius microcephalus, 35

- Arius microgastropterygius*, 55
Arius micronotacanthus, 55
Arius micropterus, 166
Arius microstomus, 39
Arius micruropterygius, 36
Arius midgleyi, 47
Arius militaris, 51
Arius molliceps, 55
Arius multiradiatus, 40
Arius nasutus, 49
Arius neogranatensis, 50
Arius nigricans, 54
Arius nox, 38, 39
Arius nuchalis, 41
Arius nucleus, 57
Arius obesus, 81
Arius oetik, 35
Arius oncina, 75
Arius oncinus, 75
Arius osculus, 50
Arius papillosus, 165
Arius parkii, 32
Arius parmocassis, 50
Arius parvipinnis, 47
Arius paucus, 48
Arius pavementatus, 105
Arius pectoralis, 48
Arius phrygiatus, 32
Arius physacanthus, 36
Arius pidada, 55
Arius planiceps, 50
Arius platypogon, 54
Arius platystomus, 52
Arius pleurops, 39
Arius polystaphyłodon, 52
Arius proximus, 48, 49
Arius pumilus, 386
Arius puncticulatus, 41
Arius quadriscutis, 37
Arius ritoides, 106
Arius robertsi, 48
Arius rostratus, 55
Arius rugispinis, 32
Arius russi, 57
Arius rutschi, 435
Arius sagoroides, 45
Arius satparanus, 56
Arius schlegeli, 56
Arius sciurus, 35
Arius seemanni, 33
Arius serratus, 49
Arius similis, 434
Arius sinensis, 55
Arius solidus, 39
Arius spatula, 42, 43
Arius squalus, 166
Arius stauroforus, 32
Arius stirlingi, 46
Arius stricticassis, 50
Arius subrostratus, 35
Arius sumatrana, 36
Arius synodon, 166
Arius taylori, 40, 48
Arius tenuispinis, 52, 56
Arius tonggol, 52
Arius truncatus, 43
Arius tuyra, 40
Arius uncinatus, 36
Arius utarus, 48
Arius utik, 35
Arius vandeli, 50
Arius variolosus, 41
Arius venosus, 36
Arius verrucosus, 45
Arius villosus, 166
Arius viviparus, 55
arleoi, *Anduzedoras*, 169
arleoi, *Pygidium*, 415
arleoi, *Trichomycterus*, 415
armatulus, *Doras*, 176
armatulus*, *Platydoras, 176
armatus, *Ageneiosus*, 68, 69
armatus, *Aklysis*, 98
armatus, *Callichthys*, 113
armatus*, *Corydoras, 113
armatus, *Macrones*, 94
armatus*, *Microsynodontis, 309
armatus, *Mystus*, 94, 98
armatus*, *Nanobagrus, 98
armatus, *Silurus*, 177
armbrusteri*, *Notarius, 440
armeniaccum, *Glyptosternum*, 388
armeniacus*, *Glyptothonax, 388
armiger, *Arius*, 46
armiger*, *Mystus, 94
armiger*, *Nemapteryx, 46
armillatus*, *Pariolius, 188
arnoldi, *Otocinclus*, 271
arnoulti*, *Synodontis, 312
arnunachalensis*, *Amblyceps, 17
artedi, *Platystoma*, 341
ascita*, *Erethistoides, 384
ascita, *Mystus*, 177
ascita, *Silurus*, 54
asopos*, *Mastiglanis, 187
asotus, *Parasilurus*, 379
asotus*, *Silurus, 378, 381
asotus longus, *Parasilurus*, 379
asper, *Callichthys*, 110, 167
aspera*, *Rhinelepis, 293
asperatus*, *Hypostomus, 251
asperispinis*, *Lophiobagrus, 161
asperus, *Pimelodus*, 395
Aspidobagrus, 93
aspidolepis, *Chaetostomus*, 242
aspidolepis, *Hemiancistrus*, 242
Aspidoradidi, 108
Aspidoras, 108
Aspidoras albater, 108
Aspidoras belenos, 108
Aspidoras brunneus, 109
Aspidoras carvalhoi, 109
Aspidoras depinnai, 109
Aspidoras eurycephalus, 109
Aspidoras fuscoguttatus, 109
Aspidoras lakoi, 109
Aspidoras maculosus, 109
Aspidoras menezesi, 109
Aspidoras microgalaeus, 109
Aspidoras pauciradiatus, 109
Aspidoras poecilus, 109
Aspidoras psammatides, 110
Aspidoras raimundi, 110
Aspidoras rochai, 110
Aspidoras spilotus, 110
Aspidoras taurus, 110
Aspidoras velites, 110
Aspidoras virgulatus, 110
aspilogaster*, *Hypostomus, 251
aspilogaster, *Plecostomus*, 251
Aspistor, 36
Aspistor hardenbergi, 36
Aspistor luniscutis, 36
Aspistor parkeri, 36
Aspistor quadriscutis, 37
Aspredinichthys, 57
Aspredinichthys filamentosus, 57
Aspredinichthys tibicen, 58
Aspredinidae, 57
aspredinoides*, *Myoglanis, 187
Aspredo, 57, 58
Aspredo aspredo, 58
Aspredo batrachus, 58
Aspredo filamentosus, 57
Aspredo gronovii, 58, 60
Aspredo sexcirrhis, 61
Aspredo sicuephorus, 58
Aspredo sicyephorus, 58
Aspredo spectrum, 61
Aspredo tibicen, 57, 58
Aspredo verrucosa, 60
aspredo*, *Aspredo, 58
aspredo, *Silurus*, 58
assamensis, *Clarias*, 148
assamensis*, *Nangra, 396
assimilis, *Ariopsis*, 33
assimilis, *Arius*, 33
astatodon, *Atopochilus*, 308
astatodon*, *Euchilichthys, 308
Astemomycterus, 411
Astrophinae, 203
Astrophus, 203, 207
Astrophus antiquus, 207
Astrophus resimus, 207
asterifrons*, *Astrodonas, 170
asterifrons, *Doras*, 169, 170
Asterophysi, 68

- Asterophy whole, 68, 70
*Asterophy batrachus, 70
*Astroblepidae, 63
*Astroblepiformes, 63
*Astroblepus, 63
*Astroblepus boulegeri, 63
*Astroblepus brachycephalus, 63
*Astroblepus caquetae, 63
*Astroblepus chapmani, 63
*Astroblepus chimboraizoi, 63
*Astroblepus chotae, 63
*Astroblepus cirratus, 64
*Astroblepus cyclopus, 64
*Astroblepus cyclopus santanderensis, 67
*Astroblepus eigenmanni, 64
*Astroblepus festae, 64
*Astroblepus fissidens, 64
*Astroblepus formosus, 64
*Astroblepus frenatus, 64
*Astroblepus grimalvii, 64
*Astroblepus grimalvii micrescens, 65
*Astroblepus guentheri, 64
*Astroblepus heterodon, 64
*Astroblepus homodon, 64
*Astroblepus jurubidae, 65
*Astroblepus labialis, 65
*Astroblepus latidens, 65
*Astroblepus longiceps, 65
*Astroblepus longifilis, 65
*Astroblepus mancoi, 65
*Astroblepus mariae, 65
*Astroblepus marmoratus, 65
*Astroblepus micrescens, 65
*Astroblepus mindoensis, 65
*Astroblepus nicefori, 65
*Astroblepus nicéfori, 65
*Astroblepus orientalis, 65
*Astroblepus peruanus, 66
*Astroblepus phelpsi, 66
*Astroblepus pholeter, 66
*Astroblepus pirrensis, 66
*Astroblepus praelitorum, 66
*Astroblepus prenadiellus, 66
*Astroblepus regani, 66
*Astroblepus rengifoi, 66
*Astroblepus retropinnus, 66
*Astroblepus riberae, 66
*Astroblepus rosei, 66
*Astroblepus sabalo, 67
*Astroblepus santanderensis, 67
*Astroblepus simonsii, 67
*Astroblepus stuebeli, 67
*Astroblepus supramollis, 67
*Astroblepus taczanowskii, 67
*Astroblepus theresiae, 67
*Astroblepus trifasciatus, 67
*Astroblepus ubidai, 67
*Astroblepus unifasciatus, 67**
- Astroblepus vaillanti, 67
*Astroblepus vanceae, 67
*Astroblepus ventralis, 68
*Astroblepus whymperi, 68
*Astrodonas, 169
*Astrodonas asterifrons, 170
*Astrophysi, 68
*Astrophysus, 70
*asymetricalis, Chiloglanis, 304
*atahuap, Paracetopsis, 135
*atavus, Silurus glanis, 382
*ater, Cossyphus, 139, 148
*ater, Galeichthys, 43
*ater, Microglanis, 354
*ater, Neosilurus, 347
*ater sepikensis, Lambertichthys, 347
*atterima, Synodontis, 312
*atterimus, Synodontis, 312
*atesuensis, Amphilus, 21
*atherinoides, Neotropius, 359
*atherinoides, Silurus, 359
*atherinoides walkeri, Pseudeutropius, 359
*athiensis, Chiloglanis, 304
*athu, Silurus, 380
*atochae, Pygidium, 423
*Atopochilus, 303
*Atopochilus astatodon, 308
*Atopochilus chabanaudi, 303
*Atopochilus christyi, 303
*Atopochilus güntheri, 309
*Atopochilus macrocephalus, 303
*Atopochilus mandevillei, 304
*Atopochilus pachychilus, 304
*Atopochilus savorgnani, 304
*Atopochilus vogti, 304
*atra, Lambertia, 347
*atrarius, Pimelodus, 205
*atratoensis, Dolichancistrus, 235
*atratoensis, Spatularicaria, 296
*atratoensis, Pseudancistrus, 235
*atribanchus, Dinopterus, 148
*atrifasciatus, Mystus, 94
*atripes, Trachydoras, 179
*atripinnis, Hemipimelodus, 34
*atrizona, Hoplomyzon, 61
*atrobrunneus, Pimelodus, 337
*atronasus, Ageneiosus, 68
*atropersonatus, Corydoras, 113
*atropinnis, Hypostomus, 251
*atropumbeus, Tachysurus, 32
*atrorus, Schilbeodes marginatus, 213
*attemsi, Allabenchelys, 137
*attemsi, Clariallabes, 137
*attu, Silurus, 380
*attu, Wallago, 380
*attu, Wallagonia, 381
*attu valeya, Wallago, 381
*aubentoni, Mystus, 91**
- Auchenoglanis macrostom, 154
*Achenaspis, 154
*Achenipterichthys, 71
*Achenipterichthys coracoideus, 71
*Achenipterichthys dantei, 71
*Achenipterichthys longimanus, 71
*Achenipterichthys punctatus, 71
*Achenipterichthys thoracatus, 71
*Achenipteridae, 68
*Achenipterini, 68
*Achenipterus, 68, 71
*Achenipterus (Pseudauchenipterus) affinis, 76
*Achenipterus (Pseudauchenipterus) Jequitinhonhae, 76
*Achenipterus (Pseudauchenipterus) striatus, 80
*Achenipterus (Pseudepapterus) hasemani, 76
*Achenipterus ambyiacus, 71
*Achenipterus brachyurus, 72
*Achenipterus brevibarbis, 80
*Achenipterus brevior, 72
*Achenipterus britskii, 72
*Achenipterus ceratophysis, 79
*Achenipterus demerarae, 72
*Achenipterus dentatus, 72
*Achenipterus fordicei, 72
*Achenipterus furcatus, 76
*Achenipterus glaber, 80
*Achenipterus Heckelii, 73
*Achenipterus immaculatus, 80
*Achenipterus insignis, 79
*Achenipterus isacanthus, 80
*Achenipterus lacustris, 79
*Achenipterus longimanus, 71
*Achenipterus maculosus, 79
*Achenipterus Magdalena, 79
*Achenipterus menezesi, 72
*Achenipterus nigripinnis, 72
*Achenipterus nuchalis, 72
*Achenipterus obscurus, 81
*Achenipterus osteomystax, 72
*Achenipterus paysanduanus, 72
*Achenipterus punctatus, 71
*Achenipterus robustus, 80
*Achenipterus thoracatus, 71
*Achenipterus thoracicus, 71
*Achenipterus trachycorystes, 80, 81
*Achenoglanidinae, 153
*Achenoglanis, 153, 154
*Achenoglanis acuticeps, 154
*Achenoglanis ahli, 162
*Achenoglanis akiri, 154
*Achenoglanis altipinnis, 162
*Achenoglanis ballayi, 162
*Achenoglanis ballayi gravoti, 162
*Achenoglanis ballayi var. gravoti, 162**

- Auchenoglanis biscutatus*, 154
Auchenoglanis büttikoferi, 163
Auchenoglanis fasciatus, 163
Auchenoglanis grandis, 163
Auchenoglanis iturii, 163
Auchenoglanis longiceps, 163
Auchenoglanis macrostoma, 154
Auchenoglanis maculosus, 163
Auchenoglanis monkei, 163
Auchenoglanis ngamensis, 163
Auchenoglanis occidentalis, 154
Auchenoglanis occidentalis
 tanganicanus, 154
Auchenoglanis occidentalis
 tchadiensis, 154
Auchenoglanis occidentalis var.
 tanganicanus, 154
Auchenoglanis occidentalis var.
 tchadiensis, 154
Auchenoglanis pantherinus, 163
Auchenoglanis pietschmanni, 162
Auchenoglanis pulcher, 162
Auchenoglanis punctatus, 163
Auchenoglanis ubangensis, 162
Auchenoglanis Wittei, 154
Auchenopterus, 71
augierasi, *Synodontis*, 314
augusta, *Nemapteryx*, 46
augustus, *Arius*, 46
aulometopon, *Netuma*, 56
aulopygia, *Tatia*, 77
aulopygius, *Centromochlus*, 77
aurantiacus, *Bagrus*, 101
aurantiacus, *Hypostomus*, 278
aurantiacus, *Parancistrus*, 278
aurantiacus, *Pseudobagrus*, 101, 108
aurata, *Hemiloricaria*, 245
auratus, *Bagrus*, 155
auratus, *Chrysichthys*, 155, 158, 160
auratus, *Pimelodus*, 155
auratus, *Porcus*, 155
auratus, *Schilbe*, 364
aurea, *Loricaria*, 298
aureatus, *Scobinancistrus*, 296
aureum, *Sturisoma*, 298
auritus, *Siluranodon*, 366
auritus, *Silurus*, 366
aurofrenatus, *Corydoras*, 113
auroguttatus, *Hypostomus*, 251
auroguttatus, *Trichomycterus*, 415
australe, *Corydoras*, 60
australe, *Dysichthys*, 118
australis, *Amiurus*, 208
australis, *Arius*, 47
australis, *Ictalurus*, 208
australis, *Pimelodella*, 189
australis, *Neosilurus*, 347, 348
austriacus, *Heterobranchus*, 382
Austroglanididae, 81
Austroglanis, 81
Austroglanis barnardi, 81
Austroglanis gilli, 81
Austroglanis sclateri, 81
Autanadoras, 178
Autanadoras milesi, 178, 179
avanhandavae, *Pimelodella*, 189
axelrodi, *Corydoras*, 113
axillaris, *Ageniosus*, 69
Ayarnangra, 382
Ayarnangra estuarior, 382
aymarae, *Rhamdella*, 196
azureus, *Galeichthys*, 33
azygia, *Farlowella*, 238
azygolechis, *Hemipsilichthys*, 279
azygolechis, *Pareiorhaphis*, 279
B
bachi, *Chaetostomus*, 284
bachi, *Oxydoras*, 174
bachi, *Peckoltia*, 284
Bachmannia, 428
Bachmannia chabutensis, 428
baculum, *Encheloclarias*, 149
badeli, *Trachycorystes insignis*, 80
baderi, *Corydoras*, 113
Bagarina, 382
Bagarius, 382, 383
Bagarius bagarius, 383
Bagarius Buchananii, 383
Bagarius gigas, 383
Bagarius lica, 383
Bagarius Nieuwenhuisii, 383
Bagarius rutilus, 383
Bagarius suchus, 383
Bagarius yarrelli, 383
bagarius, *Bagarius*, 383
bagarius, *Pimelodus*, 383
Bagre, 30, 37
Bagre bagre, 37, 97
Bagre marinus, 37
Bagre panamensis, 38
Bagre pinnimaculatus, 38
bagre, *Bagre*, 37, 97
bagre, *Silurus*, 37, 38
bagre, *Stearopterus*, 37
Bagreidae, 30
Bagrichthyes, 81
Bagrichthyoidei, 81
Bagrichthys, 81, 82
Bagrichthys hypselopterus, 82
Bagrichthys macracanthus, 82
Bagrichthys macropterus, 82
Bagrichthys majusculus, 83
Bagrichthys micranodus, 83
Bagrichthys obscurus, 83
Bagrichthys vaillantii, 83
Bagridae, 81
Bagroides, 82, 83
Bagroides, 83
Bagroïdes macracanthus, 82
Bagroïdes macropterus, 82, 83
Bagroïdes melanopterus, 83
Bagroïdes melapterus, 83
Bagroïdes Vaillantii, 83
Bagroidinae, 82
Bagropsis, 329
Bagropsis reinhardti, 329
Bagrus, 81, 83
Bagrus (Ariodes) arenarius, 34
Bagrus (Ariodes) Meyenii, 52
Bagrus (Bagrus) ramentosus, 108
Bagrus (Sciades) emphysetus, 53, 54
Bagrus (Sciades) pictus, 334
Bagrus abbreviatus, 95
Bagrus Adansonii, 365
Bagrus affinis, 85, 107
Bagrus agricolus, 107
Bagrus albicans, 54
Bagrus albilabris, 95
Bagrus anisurus, 90
Bagrus aorellus, 107
Bagrus aorides, 106
Bagrus aorinus, 107
Bagrus arioides, 40, 54
Bagrus aurantiacus, 101
Bagrus auratus, 155
Bagrus bajad, 83
Bagrus barbatus, 44
Bagrus bayad macropterus, 83
Bagrus bayad var. *macropterus*, 84
Bagrus bilineatus, 49
Bagrus Birmannus, 95
Bagrus bouderius, 165
Bagrus buchanani, 361
Bagrus caeruleus, 84
Bagrus capensis, 43
Bagrus capito, 155
Bagrus carchariorhynchos, 49
Bagrus chinta, 431
Bagrus corsula, 89
Bagrus couma, 53
Bagrus crinalis, 55
Bagrus degeni, 84
Bagrus depressirostris, 364
Bagrus docmac niger, 84
Bagrus docmak, 84, 313
Bagrus doroides, 45
Bagrus exodon, 359
Bagrus filamentosus, 84
Bagrus flavicans, 344
Bagrus flavus, 90
Bagrus fuscus, 95
Bagrus gagorides, 35
Bagrus genidens, 43, 44
Bagrus goliath, 331
Bagrus guliooides, 95
Bagrus Halepensis, 93, 94, 96, 97

Bagrus heterurus, 97
Bagrus hoevenii, 88
Bagrus hypselopterus, 82
Bagrus Javensis, 45
Bagrus Keletius, 98
Bagrus koenigi, 84
Bagrus laevigatus, 49
Bagrus Lamarrii, 106, 107
Bagrus leucophasis, 96
Bagrus limbatus, 107
Bagrus lonah, 391
Bagrus lubosicus, 84
Bagrus macronemus, 37, 97
Bagrus macronemus, 93, 97
Bagrus Malabaricus, 96
Bagrus maurus, 158
Bagrus melas, 95
Bagrus meridionalis, 84
Bagrus mesops, 53
Bagrus micracanthus, 96
Bagrus micropogon, 92, 93
Bagrus montanus, 96
Bagrus nemurus, 87, 90
Bagrus netuma, 48, 49
Bagrus nigriceps, 96
Bagrus nigrita, 160
Bagrus nigropunctatus, 340
Bagrus oculatus, 96
Bagrus orientalis, 84
Bagrus passany, 54
Bagrus pemucus, 53
Bagrus planiceps, 90
Bagrus poecilopterus, 93
Bagrus proops, 54
Bagrus punctatus, 91
Bagrus punctulatus, 331, 339, 340
Bagrus reticulatus, 330, 331
Bagrus rhodonotus, 49
Bagrus rhodopterygius, 95
Bagrus rousseauxii, 331
Bagrus schilbeides, 362, 365
Bagrus Schlegelii, 36, 95
Bagrus shuwaiensis, 84
Bagrus Sieboldii, 90
Bagrus sinensis, 108
Bagrus singaringan, 97
Bagrus sondaicus, 45
Bagrus stenorhynchus, 103, 104
Bagrus sumatranaus, 36
Bagrus temminckianus, 55
Bagrus Temminckii, 55
Bagrus thalassinus, 49
Bagrus trachacanthus, 89
Bagrus trachipomus, 55
Bagrus ubangensis, 84
Bagrus urostigma, 84
Bagrus ussuriensis, 92, 100
Bagrus vachellii, 101
Bagrus venaticus, 55

Bagrus vertagus, 55
Bagrus wolffii, 98
Bagrus Wyckii, 91
Bagrus Yarrelli, 383
bahianus, *Corymbophanes*, 279
bahianus*, *Pareiorhaphis, 279
bahianus, *Pimelodus*, 195
bahianus*, *Trichomycterus, 415
bahiensis, *Galeichthys*, 38
bahiensis*, *Kalyptodoras, 172
bahiensis, *Microlepidogaster*, 281
bahiensis*, *Parotocinclus, 281
bahuaja*, *Crossoloricaria, 232
baileyi*, *Noturus, 211
bajad*, *Bagrus, 83
bajad, *Silurus*, 83
balayi*, *Parauchenoglanis, 162
balayi, *Pimelodus*, 162
ballayi, *Auchenoglanis*, 162
ballayi gravoti, *Auchenoglanis*, 162
ballayi var. *gravoti*, *Auchenoglanis*, 162
balsanus*, *Ictalurus, 208
balsanus, *Istlarius*, 208
balsanus occidentalis, *Istlarius*, 208
Balzanii, *Vandellia*, 427
bambui*, *Ituglanis, 406
banguela*, *Homodiaetus, 406
banguelensis, *Eutropius*, 363
banguelensis*, *Schilbe, 363
bankae, *Chaca*, 135
bankanensis*, *Chaca, 135
banneuai, *Pygidium*, 415
banneuai*, *Trichomycterus, 415
banneuai maracaiboensis, *Pygidium*, 420
barakensis*, *Pterocryptis, 441
barakensis*, *Sisor, 402
baramensis, *Akysis*, 16
baramensis*, *Hemibagrus, 87
baramensis*, *Kryptopterus, 369
baramensis, *Liocassis*, 93
baramensis, *Macrones*, 87
baramensis*, *Pseudobagarius, 16
barbancho, *Pimelodus*, 339
barbarmatus*, *Dentectus, 235
barbata, *Loricaria*, 298, 299
barbata*, *Microcambeva, 409
barbata, *Rhamdia*, 201
barbatula, *Trichomycterus*, 422
barbatum*, *Sturisoma, 299
barbatus, *Bagrus*, 44
barbatus, *Callichthys*, 129
barbatus, *Hypostomus*, 286, 287
barbatus*, *Malapterurus, 301
barbatus, *Platysilurus*, 340
barbatus*, *Pseudancistrus, 287
barbatus*, *Scleromystax, 129
barbatus*, *Xylipterus, 62
barbonica, *Laimumena*, 431, 432
barbouri, *Pygidium*, 415
barbouri*, *Trichomycterus, 415
barbus*, *Genidens, 44
barbus, *Pimelodus*, 44
barnardi*, *Astroglanis, 81
barnardi, *Gephyroglanis*, 81
baroni, *Arius*, 434
Baronis Müller, *Pimelodus*
(Rhamdia), 201
barrae, *Ancistrus*, 272
barrae*, *Megalancistrus, 272
barranquerensis, *Ageneiosus*, 69
bartonensis, *Arius bartonensis*, 56
bartonensis, *Otolithus (Arius) crassus*, 434
bartonensis, *Otolithus (Arius) danicus*, 434
bartoni, *Neosilurus*, 347
Baryancistrus, 226
Baryancistrus demantoides, 226
Baryancistrus longipinnis, 227
Baryancistrus niveatus, 227
basilewskii, *Silurichthys*, 381
bastari*, *Clarias, 357
bastiani*, *Synodontis, 312
batarensis, *Pseudeutropius murius*, 357
Batasinae, 81
Batasio, 81, 84
Batasio affinis, 85
Batasio batasio, 85, 86
Batasio buchanani, 84, 85
Batasio dayi, 85
Batasio elongatus, 85
Batasio fasciolatus, 442
Batasio havmoller, 85
Batasio macronotus, 85
Batasio merianensis, 85
Batasio niger, 445
Batasio pakistanicus, 85
Batasio sharavatiensis, 86
Batasio spilurus, 442
Batasio tengana, 86, 442
Batasio tigrinus, 86
Batasio travancoria, 86
batasio*, *Batasio, 85, 86
batasio, *Pimelodus*, 85
batasius, *Pimelodus*, 85, 98
batensoda*, *Brachysynodontis, 304
batensoda, *Synodontis*, 304
Batesi longispinis, *Synodontis*, 316
Batesi var. longispinis, *Synodontis*, 316
batesii*, *Chiloglanis, 304
batesii*, *Microsynodontis, 309
batesii*, *Synodontis, 312
Bathophilus, 412
Bathybagrus, 154
Bathybagrus tetranema, 154, 155

- Bathycetopsis*, 131
Bathycetopsis oliveirai, 131, 132
Bathyclarias, 136, 148
Bathyclarias euryodon, 136
Bathyclarias filicibarbis, 148
Bathyclarias gigas, 149
Bathyclarias illesi, 136
Bathyclarias longibarbis, 136
Bathyclarias loweae, 149
Bathyclarias rotundifrons, 136
Bathyclarias worthingtoni, 136
Bathypygidium, 412
bathyurus, *Pimelodus*, 201
Batrachocephalinae, 30
Batrachocephalus, 30, 38
Batrachocephalus ageneiosus, 38
Batrachocephalus micropogon, 38
Batrachocephalus mino, 38
Batrachoglanis, 352
batrachostoma, *Ochmacanthus*, 409
batrachus, *Aspredo*, 58
batrachus, *Asterophysus*, 70
batrachus, *Clarias*, 140
batrachus, *Silurus*, 140
Batrochoglanis, 352
Batrochoglanis acanthochiroides, 352
Batrochoglanis melanurus, 353
Batrochoglanis raninus, 353
Batrochoglanis transmontanus, 353
Batrochoglanis villosus, 353
batu, *Clarias*, 140
baudensis, *Ancistrus*, 220
baudoensis, *Cetopsis*, 131
baudoënsis, *Pseudocetopsis*, 131
baudoni, *Amphilius*, 21
baudoni uniformis, *Amphilius*, 21
baudoni var. *uniformis*, *Amphilius*, 21
bayad, *Porcus*, 83
bayad macropterus, *Porcus*, 83
bayad var. *macropterus*, *Porcus*, 84
Beadlei, *Synechoglanis*, 207, 210
beani, *Pangasius*, 328
beccarii, *Vandellia*, 427
bedado, *Pangasius*, 325
bedfordi, *Silurus*, 379
Belangerii, *Arius*, 54
beldti, *Cryptopterella*, 369, 371
belenos, *Aspidoras*, 108
belensis, *Trichomycterus*, 415
belgicus, *Arius egertoni*, 56
Belodontichthys, 368
Belodontichthys dinema, 368
Belodontichthys javanensis, 381
Belodontichthys macrochir, 368
Belodontichthys truncatus, 368
Belonoglanis, 24
Belonoglanis brieni, 24
Belonoglanis curvirostris, 24
Belonoglanis nudipectus, 24
Belonoglanis tenuis, 24
benderensis, *Ictalurus*, 206
benedettii, *Nannorhamdia*, 198
beninensis, *Malapterurus*, 301
bengalensis, *Malapterus (Ailia)*, 356
beni, *Hemiloricaria*, 245
beni, *Leptoplosternum*, 128
beni, *Loricaria*, 245
benjamini, *Entomocorus*, 74
benuensis, *Chiloglanis*, 305
berdmorei, *Exostoma*, 385
berdmorei, *Pterocryptis*, 376, 377
Berdmorei, *Silurichthys*, 376
Bergiaria, 329
Bergiaria platana, 330
Bergiaria westermannii, 330
Bergiella, 329
Bergiella platana, 330
berneyi, *Neoarius*, 47
berneyi, *Tachysurus (Pararius)*, 47
bernhardi, *Hemipimelodus*, 39
bertoni, *Branchioica*, 410
bertoni, *Corydoras*, 130
bertonii, *Branchioica*, 410
bicarinatus, *Chaenothorax*, 111, 125
bicirrhis, *Kryptopterus*, 369
bicirrhis, *Silurus*, 369
bicirrus, *Plecostomus*, 259
bicolor, *Arius*, 34
bicolor, *Bunocephalus*, 59
bicolor, *Corydoras*, 114
bicolor, *Hemipimelodus*, 34
bicolor, *Leiocassis*, 104
bicolor, *Phractocephalus*, 336
bicolor, *Pirarara*, 336
bidentatus, *Parotocinclus*, 282
bidorsalis, *Claroites*, 160
bidorsalis, *Heterobranchus*, 150
bifasciata, *Taunayia*, 203
bifasciatus, *Corydoras*, 114
bifasciatus, *Nannoglanis*, 203
biffi, *Notarius*, 49
bifidus, *Bunocephalus*, 59
bifurcus, *Chiloglanis*, 305
bilineata, *Netuma*, 49
bilineatus, *Bagrus*, 49
bilineatus, *Corydoras*, 114
bilineatus, *Epaionotus*, 236
bilineatus, *Pimelodus*, 107
bilobatum, *Hypoptopoma*, 250
bimaculatus, *Macrones*, 94
bimaculatus, *Mystus*, 94
bimaculatus, *Ompok*, 372
bimaculatus, *Silurus*, 372
binotata, *Synodontis notatus*, 318
binotatus, *Ompok*, 372
Birmannus, *Bagrus*, 95
biscutatus, *Auchenoglanis*, 154
biscutatus, *Pimelodus*, 154
biseriata, *Squaliforma*, 297
biseriatus, *Plecostomus*, 297
bistriata, *Ansorgia vittata*, 361
biwaensis, *Parasilurus*, 379
biwaensis, *Silurus*, 379
bleekeri, *Arius*, 46
bleekeri, *Cryptopterus*, 374
bleekeri, *Heptapterus*, 184
bleekeri, *Macrones*, 89, 94
bleekeri, *Micronema*, 374
bleekeri, *Mystus*, 94
bleekeri, *Nemapteryx*, 46
bleekeri, *Paracetopsis*, 134
bleekeri, *Phalacronotus*, 374
blochi, *Corydoras*, 114
blochi vittatus, *Corydoras*, 126
Blochii, *Cotylephorus*, 61
blockii, *Doras*, 167
Blochii, *Galeichthys*, 38
Blochii, *Osteogeneiosus*, 51
blochii, *Pimelodus*, 336, 337
blohmi, *Epapterus*, 74
Blythii, *Exostoma*, 396
Blythii, *Macrones*, 85
blythii, *Myersglanis*, 396
bo, *Macrones*, 88
boakeii, *Arius*, 34
boalis, *Silurus*, 380
bocagii, *Chrysichthys*, 155
Bocagii, *Eutropius*, 363
bocagii, *Schilbe*, 363
bockmanni, *Glanidium*, 75
bocourtii, *Heterobagrus*, 94
bocourtii, *Mystus*, 94
bocourtii, *Pangasius*, 325
Bocourtii, *Pangasius*
(Pseudopangasius), 325
bodenhameri, *Ancistrus*, 219
boehlkei, *Corydoras*, 114
boehlkei, *Rhinodoras*, 178
boemia, *Tatia*, 77
boesemani, *Corydoras*, 114
bogotense, *Pygidium*, 415
bogotensis, *Trichomycterus*, 415
bokorensis, *Penesilurus*, 376
bokorensis, *Pterocryptis*, 376
bolivarensis, *Amblydoras*, 168
bolivarensis, *Hildadoras*, 168
boliviana, *Farlovella*, 238
boliviana, *Phenacorhamdia*, 188
boliviana, *Pimelodella*, 189
boliviana, *Xenocara*, 219
bolivianensis, *Andinichthys*, 30
bolivianus, *Ancistrus*, 219
bolivianus, *Corydoras*, 117
bolivianus, *Hypostomus*, 252
bolivianus, *Imparfinis*, 188
bolivianus, *Plecostomus*, 252

bollii, *Amiurus*, 205
bolteni, *Callichthys callichthys*, 111
bomae, *Eutropius*, 363
bomboides, *Pseudomystus*, 103
bomboizanum, *Pygidium*, 416
bomboizanus, *Trichomycterus*, 416
bondi, *Acanthopoma*, 413
bondi, *Corydoras*, 114
bondi, *Schultzichthys*, 413
bondi coppenamensis, *Corydoras*, 115
bongan, *Hemibagrus*, 87
bongan, *Macrones*, 87
bonillai, *Ariopsis*, 33
bonillai, *Galeichthys*, 33
bonneti, *Arius*, 36, 56
boquillae, *Cetopsorhamdia*, 181
borealis, *Silurus (Pimelodus)*, 210
borellii, *Hypostomus*, 252
borellii, *Plecostomus*, 252
borelli, *Trichomycterus*, 416
borneensis, *Callichrous (Silurodes)*, 372
borneensis, *Cephalocassis*, 41, 42
borneensis, *Ompok*, 372
borneensis, *Arius*, 56
borneensis, *Pimelodus*, 41
borodini, *Imparfinis*, 185
bororo, *Otocinclus*, 274
börressoni, *Chrysichthys*, 156
bortayro, *Silvinichthys*, 413
boschmai, *Pimelodella*, 190
bostocki, *Tandanus*, 352
bostockii, *Cnidoglanis*, 346
boticario, *Listrura*, 408
botius, *Glyptothorax*, 388
botius, *Pimelodus*, 388
Boucardi, *Pimelodus*, 201
bouderius, *Bagrus*, 165
bouderius, *Cranoglanis*, 165
bouilloni, *Leptoglanis*, 164
boujardi, *Lithoxus*, 267
boulengeri, *Arges*, 63
boulengeri, *Astroblepus*, 63
boulengeri, *Euchilichthys*, 309
boulengeri, *Hemidoras*, 175
boulengeri, *Hemidoras (Leptodoras)*, 175
boulengeri, *Heterobranchus*, 151
boulengeri, *Hypostomus*, 252
boulengeri, *Opsodoras*, 172, 175
boulengeri, *Plecostomus*, 252
bourguyi, *Microlepidogaster*, 300
boutchangai, *Anaspidoglanis*, 154
boutchangai, *Parauchenoglanis*, 154
bouvieri, *Schilbe*, 364
bovallii, *Ancistrus*, 267
bovallii, *Lithoxus*, 267
bovei, *Peltura*, 26
bovei, *Phractura*, 26

boylei, *Pygidium*, 416
boylei, *Trichomycterus*, 416
brachiatus, *Centrodoras*, 170
brachiatus, *Doras*, 170
Brachyspondylus cretaceus, 432
brachyacanthus, *Amiurus*, 205
brachybarbatus, *Akysis*, 13
brachycephalus, *Arges*, 63
brachycephalus, *Astroblepus*, 63
brachycephalus, *Pimelodus*, 198
Brachyglanis, 180
Brachyglanis frenata, 180
Brachyglanis frenatus, 180
Brachyglanis magoi, 180
Brachyglanis metas, 181
Brachyglanis microphthalmus, 181
Brachyglanis nocturnus, 181
Brachyglanis phalacra, 181
Brachymystus, 87
brachynema, *Chrysichthys*, 155
brachynema, *Chasmocranus*, 182
brachynemus, *Chasmocranus*, 182
Brachyplatystoma, 329, 330
Brachyplatystoma capapretum, 330
Brachyplatystoma filamentosum, 330
Brachyplatystoma goeldii, 330
Brachyplatystoma juruense, 331
Brachyplatystoma paraense, 331
Brachyplatystoma parnahybae, 331
Brachyplatystoma platynema, 331
Brachyplatystoma platynemum, 331
Brachyplatystoma promagdalena, 331
Brachyplatystoma rousseauxii, 331
Brachyplatystoma tigrinum, 331
Brachyplatystoma vaillantii, 331
Brachyplatystomatini, 329
brachyopterus, *Eutropius*, 362
brachyopterus, *Pseudeutropius*, 362
brachypterus, *Pimelodus (Rhamdia)*, 198
Brachyrhamdia, 181
Brachyrhamdia heteropleura, 181
Brachyrhamdia imitator, 181
Brachyrhamdia marthae, 181
Brachyrhamdia meesi, 181
Brachyrhamdia rambarrani, 181
brachyrhyncha, *Pareiorhina*, 281
brachysoma, *Clarias*, 140
brachysoma, *Horabagrus*, 429
brachysoma, *Pseudobagrus*, 429
brachysoma, *Pseudexostoma*, 401
Brachyspondylus, 432
Brachyspondylus indicus, 326
Brachyspondylus saropterix, 82
Brachystacus, 135
Brachysynodontis, 304
Brachysynodontis batensoda, 304
brachyura, *Dekeyseria*, 234
brachyurus, *Ancistrus*, 234

brachyurus, *Auchenipterus*, 72
brachyurus, *Euanemus*, 72
Branchioica, 410
Branchioica bertoni, 410
Branchioica bertonii, 410
Branchioica magdaleneae, 411
Branchioica phaneronema, 411
Branchiosteus, 17
Brandtii, *Arius*, 54
branickii, *Chaetostoma*, 228
branickii, *Chaetostomus*, 228
branneri, *Rhamdia*, 201
branneri voulezi, *Rhamdia*, 201
bransfordi, *Loricaria*, 240
bransfordii, *Rhamdia*, 201
brashnikowi, *Leiocassis*, 100
brashnikowi, *Macrones (Leiocassis)*, 100
brashnikowi, *Pelteobagrus*, 100
brasiliensis, *Caecorhamdella*, 189, 190
brasiliensis, *Pimelodella*, 190, 192
brasiliensis, *Pimelodus*
(*Pseudorhamdia*), 192
brasiliensis, *Plecostomus*, 259
brasiliensis, *Trichomycterus*, 416
brasiliensis itatiayae, *Trichomycterus*, 419
brasiliensis tristis, *Trichomycterus*, 416
braueri, *Hemiancistrus*, 284
braueri, *Peckoltia*, 284
breei, *Corydoras*, 114
Breitensteinia, 15
Breitensteinia cessator, 15
Breitensteinia hypselurus, 15
Breitensteinia insignis, 15
breve, *Chaetostoma*, 228
brevianalis, *Eutropius*, 363
brevianalis, *Pseudobagrus*, 101
brevianalis, *Schilbe*, 363
brevibarbis, *Ancharius*, 29
brevibarbis, *Auchenipterus*, 80
brevibarbis, *Chiloglanis*, 305
brevibarbis, *Chrysichthys*, 156
brevibarbis, *Chrysobagrus*, 155, 156
brevibarbis, *Clariablabes*, 137
brevibarbis, *Gogo*, 29
brevibarbus, *Plotosus*, 351
brevicauda, *Hypostomus*, 252
brevicauda, *Phractura*, 26
brevicauda, *Plecostomus*, 252
brevicaudatus, *Leiocassis*, 102
brevicaudatus, *Pseudobagrus*, 102
Brevicephalooides, 139
Breviceps, 37
Breviceps, *Clarias*, 141
Breviceps, *Liocassis*, 103
Breviceps, *Pimelodella*, 190
Breviceps, *Pimelodus*, 190

breviceps, *Pseudomystus*, 103
brevicorpus, *Coreobagrus*, 86
brevidorsalis, *Amphilius*, 23
brevidorsalis, *Copidoranis*, 347
brevidorsalis, *Neosilurus*, 347
brevifilis, *Ageneiosus*, 68, 69
brevifilis, *Ancistrus*, 219
brevifilis, *bodenhameri*, *Ancistrus*, 219
Breviglanis, 180
brevilabiatum, *Chaetostoma*, 228
brevilabiatus, *Chaetostomus*, 228
brevinuchalis, *Clarias*, 140
brevior, *Allabenchelys*, 137
brevior, *Andersonia*, 24
brevior, *Auchenipterus*, 72
brevior, *Chasmocranus*, 182
brevipinna, *Amphilius*
 grammatophorus, 22
brevipinna, *Amphilius*
 grammatophorus var., 23
brevipinnis, *Ancistrus*, 219
brevipinnis, *Glyptothorax*, 388
brevipinnis, *Xenocara*, 219
brevipinnis alaknandi, *Glyptothorax*, 388
brevirostre, *Sturisoma*, 299
brevirostris, *Arius*, 31
brevirostris, *Corydoras*, 114
brevirostris, *Loricaria*, 299
brevirostris, *Loricaria* (*Rineloricaria*), 298
brevis, *Ageneiosus*, 69
brevis, *Amphilius*, 21
brevis, *Chaetostomus*, 228
brevis, *Delturus*, 443
brevis, *Doras* (*Corydoras*), 179
brevis, *Hemiancistrus*, 284
brevis, *Hypostomus*, 252
brevis, *Leptoglanis*, 28
brevis, *Mochocus*, 310
brevis, *Mochokus*, 310
brevis, *Peckoltia*, 284
brevis, *Pimelodus*, 337
brevis, *Plecostomus*, 252
brevis, *Trachydoras*, 179
brevis, *Tridens*, 426
brevis, *Tridensimilis*, 426
brevis, *Zaireichthys*, 28
brevispinis, *Lasiancistrus*, 287
brevispinis, *Lophiobagrus*, 161
brevispinis, *Pseudancistrus*, 287
brevitentaculatus, *Hypostomus*, 291
brichardi, *Phyllonemus*, 164
brichardi, *Synodontis*, 312
brieni, *Belonoglanis*, 24
britskii, *Auchenipterus*, 72
britskii, *Brochis*, 114
britskii, *Corydoras*, 114
britskii, *Parotocinclus*, 282

broadbenti, *Tachysurus*, 51
Brochiloricaria, 227
Brochiloricaria chauliodon, 227
Brochiloricaria macrodon, 227
Brochis, 111
Brochis britskii, 114
Brochis coeruleus, 111, 125
Brochis dipteris, 111, 125
Brochis multiradiatus, 120
Brochis splendens, 125
Brontes, 63
Brontes prenadilla, 63, 66
brunellii, *Arius*, 31
brunnea, *Loricaria*, 269
brunnea, *Tatia*, 77
brunneascens, *Doras*, 168
brunneus, *Ameiurus*, 204
brunneus, *Amiurus*, 204
brunneus, *Aspidoras*, 109
brunneus, *Loricariichthys*, 269
Brustiarius, 38
Brustiarius nox, 38
Brustiarius solidus, 39
buccata, *Pterocryptis*, 376
bucculenta, *Nangra*, 396
Buchanani, *Arius*, 34
Buchanani, *Bagarius*, 383
buchanani, *Bagrus*, 361
Buchanani, *Batasio*, 84, 85
buchanani, *Chaca*, 136
buchanani, *Glyptothorax*, 388
buchanani, *Hara*, 395
Buchanani, *Nangra*, 397
Buchanani, *Pangasius*, 325, 327
buchanani, *Proeutropiichthys*, 361
buchanani, *Rama*, 105
Buchanani, *Rita*, 105, 106
Bucklandium, 432
Bucklandium diluvii, 432, 433
buckleyi, *Pimelodella*, 190
buckleyi, *Pimelodus*, 190
budgetti, *Clarias*, 140
budgetti, *Synodontis*, 312
buettikoferi, *Clarias*, 140
buettikoferi, *Parauchenoglanis*, 163
buffei, *Eutropiellus*, 361
buffei, *Pareutropius* 360
bufonia, *Xenocara*, 219
bufonius, *Ancistrus*, 219
bufonius, *Pimelodus*, 355
bufonius, *Pseudopimelodus*, 355
bufonius, *Xenocara*, 219
bullocki, *Hatcheria*, 404
Bullockia, 403
Bullockia maldonadoi, 404
buluma, *Clarias*, 140
Bunocephalichthys, 58
Bunocephalichthys gronovii, 60
Bunocephalini, 57
Bunocephalus, 57, 58
Bunocephalus albifasciatus, 62
Bunocephalus aleuropsis, 58
Bunocephalus amaurus, 58
Bunocephalus amaurus aloikae, 58
Bunocephalus amaurus sipaliwini, 58
Bunocephalus amazonicus, 59
Bunocephalus bicolor, 59
Bunocephalus bifidus, 59
Bunocephalus carvalhoi, 59
Bunocephalus chamaizelus, 59
Bunocephalus colombianus, 59
Bunocephalus coracoideus, 59
Bunocephalus depressus, 62
Bunocephalus dolichurus, 62
Bunocephalus doriae, 59
Bunocephalus haggini, 59
Bunocephalus hypsiurus, 57
Bunocephalus iheringii, 59
Bunocephalus knerii, 59
Bunocephalus larai, 59
Bunocephalus melas, 58
Bunocephalus minutus, 59
Bunocephalus quadriradiatus, 60
Bunocephalus retropinnis, 59
Bunocephalus rugosus, 60
Bunocephalus salathei, 59
Bunocephalus scabriceps, 58, 60
Bunocephalus verrucosus, 60
burgessi, *Corydoras*, 114
burgini, *Pangasius*, 326
burmanensis, *Pterocryptis*, 376
burmanensis, *Silurus*, 376
burmanica, *Cochlefelis*, 42
burmanica, *Olyra*, 99
burmanicus, *Arius*, 42
burmanicus, *Glyptothorax*, 388
burmanicus, *Proeutropiichthys taakree*, 362
burmannicus, *Eutropiichthys*, 358
Burmeisteri, *Pygidium*, 405
burmensis, *Chaca*, 135
butantanis, *Plecostomus margaritifer*, 256
butcheri, *Pseudopimelodus villosus*, 352
buthupogon, *Clarias*, 141
butleri, *Paraplotosus*, 349
Büttikoferi, *Acrochordonichthys*, 13
Büttikoferi, *Auchenoglanis*, 163
Büttikoferi, *Clarias*, 141
Büttikoferi, *Chrysichthys*, 158
C
cabrerae, *Rhamdia*, 198
cacerensis, *Hemiloricaria*, 245
cacerensis, *Loricaria*, 245
cachariooides, *Arius*, 166
cadeae, *Loricaria*, 294
cadeae, *Rineloricaria*, 293

- Caecorhamdella*, 189
Caecorhamdella brasiliensis, 189, 192
Caecorhamdia, 197
Caecorhamdia urichi, 197, 202
caecutiens, Amblyceps, 17, 18
Caelatoglanis, 382, 383
Caelatoglanis zonatus, 383
caelata, Nemapteryx, 46
cælatus, Arius, 46
caelatoides, Arius, 55
caerulescens, Pimelodus, 81
caeruleus, Bagrus, 84
caerulorostris, Platypogon, 339, 340
caesius, Plotosus, 350
caesius, Propimelodus, 443
cahualli, Tridentopsis, 426
cahyensis, Ituglanis, 444
caíapo, Cetopsis, 131
Cainosilurus, 347
calamita, Hypostomus, 219
Calichthys, 110
caliense, Pygidium, 416
caliensis, Trichomycterus, 416
callarias, Silurus, 313
Callichrous, 371
Callichrous (Silurodes) borneensis, 372
Callichrous ceylonensis, 372
Callichrous egertonii, 374
Callichrous eugeneiatus, 369
Callichrous nigrescens, 374
Callichrous notatus, 374
Callichrous Sindensis, 373
Callichrous weberi, 373
Callichthini, 108
Callichthyidae, 108
Callichthys, 108, 110
Callichthys adspersus, 127
Callichthys affinis, 111
Callichthys albidus, 128
Callichthys arcifer, 111
Callichthys armatus, 113
Callichthys asper, 110, 167
Callichthys barbatus, 129
Callichthys callichthys, 110
Callichthys callichthys bolteni, 111
Callichthys callichthys demararae, 111
Callichthys chiquitos, 128
Callichthys exaratus, 129
Callichthys fabricioi, 111
Callichthys hemiphractus, 111
Callichthys kneri, 111
Callichthys laevigatus, 127
Callichthys littoralis, 127
Callichthys longifilis, 129
Callichthys loricatus, 111
Callichthys melampterus, 127, 128
Callichthys oibaensis, 111
Callichthys paleatus, 122
Callichthys pectoralis, 128
Callichthys pictus, 129
Callichthys serralabium, 111
Callichthys splendens, 125
Callichthys sulcatus, 129
Callichthys taiosh, 125
Callichthys tamoata, 111
Callichthys thoracatus, 129
callichthys, Callichthys, 110
callichthys bolteni, Callichthys, 111
callichthys demararae, Callichthys, 111
Callomystax, 385
Callomystax Schmidti, 392
callopterus, Glyptothorax, 388
calmoni, Hemipsilichthys, 279
Calophysinae, 329
Calophysus, 329, 332
Calophysus macropterus, 332
calvarius, Silurus, 99, 100
calvus, Isbrueckerichthys, 441
calvus, Rhineastes, 207
camaronensis, Chrysichthys, 158
camelopardalis, Synodontis, 312
cameronensis, Chiloglanis, 305
cameronensis, Chrysichthys, 158
cameronensis, Clarias, 144
cameronensis, Eutropius, 365
cameroni, Hemipsilichthys, 279
cameroni, Pareiorhaphis, 279
cameroni, Psilichthys, 247, 279
camerunensis, Clarias, 139, 141, 144
camerunensis, Leptoglanis, 28
camerunensis, Zaireichthys, 28
campensis, Diplomystes, 165
camposi, Eremophilus, 408
camposi, Listrura, 408
canaliferus, Otothyris, 275, 276
canarensis, Mystus, 96
cancellatus, Ameiurus, 211
candidus, Eremophilus, 416
candidus, Trichomycterus, 416
candiru, Cetopsis, 131
candombe, Hisonotus, 440
canensis, Acanthicus, 266
canensis, Leptoancistrus, 266
canio, Silurus, 373, 374
canius, Plotosus, 350
Canthopomus, 289
Canthopomus montebelloi, 263
Cantonensis, Pimelodus, 431
cantorisi, Osteogeneiosus, 51
capapretum, Brachyplatystoma, 330
caparary, Sorubim, 341
Capellonis, Arius, 32
capensis, Bagrus, 43
capensis, Clarias, 143
capetensis, Dasyloricaria, 233
capetensis, Loricaria, 233
capito, Bagrus, 155
capitulum, Macrones fortis, 88
caquetae, Astroblepus, 63
caquetae, Corydoras, 119
caquetae, Euacanthagenys, 296
caquetae, Harttia, 300
caquetae, Hemiancistrus, 265
caquetae, Loricaria, 296
caquetae, Spatuloricaria, 296
caquetae, Sturisomatichthys, 300
caracasensis, Hemiloricaria, 244, 245
caracassensis, Hemiloricaria, 245
carachama, Monistiancistrus, 289
carcharhinoides, Nangra, 396
carchariorhynchos, Bagrus, 49
carcio, Pimelodus, 85, 98
carinata, Farlowella, 237
carinata, Loricaria, 268
carinata, Oxyropsis, 276
carinata, Trewavasia, 433
carinatum, Amblyceps, 18
carinatum, Hypoptopoma, 276
carinatus, Arius (Hemiarius), 42
carinatus, Cinetodus, 42
carinatus, Doras, 171, 173
carinatus, Hypostomus, 252
carinatus, Plecostomus, 252
carinatus, Silurus, 170, 171
carinatus, Xenopholis, 433
Carinotus, 234
carinotus, Delturus, 235
carinotus, Plecostomus (Carinotus), 234, 235
carlae, Corydoras, 115
Carnaticus, Pimelodus, 383
carnegiei, Dolichancistrus, 235
carnegiei, Pseudancistrus, 235
carnosus, Chiloglanis, 305
carnosus, Pseudomystus, 103
carolinensis, Mystus, 37
carrancas, Pareiorhina, 281
carrioni, Lipopterichthys, 266, 267
carsonii, Clarias, 145
carvalhoi, Aspidoras, 109
carvalhoi, Bunocephalus, 59
carvalhoi, Harttia, 240
carvalhoi, Hypostomus, 252
carvalhoi, Plecostomus, 252
Cascadura, 108, 127
Cascadura maculocephala, 127, 128
Cascaduridi, 108
cashibo, Loricaria, 270
cashibo, Loricariichthys, 270
castanea, Loricaria, 270
castaneoides, Plotosus, 351
castaneo-ventris, Doras, 168
castaneus, Helogenes, 134
castaneus, Leyvaichthys, 134, 407
castaneus, Loricariichthys, 270

castaneus, *Mystus*, 94
castaneus, *Plotosus*, 351
castelnaui, *Hemiancistrus*, 265
castroi, *Hemiloricaria*, 245
castroi, *Rineloricaria*, 245
castroi, *Trichomycterus*, 416
catamarcensis, *Loricaria*, 294
catamarcensis, *Rineloricaria*, 294
catamarcensis, *Trichomycterus*, 416
catanai, *Leptodoras*, 172
cataphracta, *Loricaria*, 268, 270
Cataphractus, 127
Cataphractus, 110, 167
Cataphractus americanus, 167
Cataphractus callichthys, 111
Cataphractus depressus, 111
Cataphractus punctatus, 111, 123
cataphractus, *Plecostomus*, 270
cataphractus, *Silurus*, 167, 168
cataractus, *Clarias*, 141
cataractus, *Phagorus*, 141
Catastoma, 48
catharinensis, *Glanidium*, 75
Cathorops, 39
Cathorops agassizii, 39
Cathorops aguadulce, 39
Cathorops arenatus, 39
Cathorops dasycephalus, 39
Cathorops fuerthii, 39
Cathorops hypophthalmus, 40
Cathorops mapale, 40
Cathorops melanopus, 40
Cathorops multiradiatus, 40
Cathorops spixii, 40
Cathorops steindachneri, 40
Cathorops tuyra, 40
catulus, *Pimelodus*, 205
catus, *Ameiurus*, 204
catus, *Silurus*, 204
caucanus, *Ageneiosus*, 70
caucanus, *Ancistrus*, 219
caucanus, *Lasiancistrus*, 264
caudalis, *Synodontis*, 312
cauda-furcatus, *Pimelodus*, 209
caudimaculatus, *Corydoras*, 115
caudofasciatus, *Trichomycterus*, 416
caudivittatus, *Paradoxoglanis*, 302
caudovittata, *Synodontis*, 312
caudovittatus, *Synodontis*, 313
cavalliensis, *Malapterurus*, 301
cavasius, *Pimelodus*, 95
cavasius, *Mystus*, 95
cavatica, *Peckoltia*, 284
cavatura, *Erethistoides*, 384
cavatus, *Arius*, 435
caveatus, *Hemibagrus*, 87
cavernicola, *Clarias*, 141
cavia, *Glyptothorax*, 388
cavia, *Pimelodus*, 388

caxarari, *Otocinclus*, 274
cayennae, *Hoplosternum thoracatum*, 128
cæcutiens, *Amblyceps*, 18
cæcutiens, *Silurus*, 131
cælatus, *Arius*, 46
cælatus, *Callichthys*, 111
cærulescens, *Arius*, 33
cærulescens, *Pimelodus*, 81
cearensis, *Parotocinclus*, 282
celsae, *Trichomycterus*, 416
cenia, *Gagata*, 386
cenia, *Pimelodus*, 386
centralis, *Clariallabes*, 137
centralis, *Clarias*, 137
centralis, *Synodontis*, 313
centralus, *Hemibagrus*, 87
Centranodon japonicus, 379
Centrochir, 170
Centrochir crocodili, 170
Centrochirinae, 166
Centrodoras, 170
Centrodoras brachiatus, 170
Centrodoras hasemani, 170
centrolepis, *Ancistrus*, 219
Centromochli, 68
Centromochlus, 68, 73
Centromochlus (Gephyromochlus) leopardus, 75
Centromochlus altae, 73
Centromochlus aulopygius, 77
Centromochlus concolor, 73
Centromochlus creutzbergi, 77
Centromochlus dunni, 77
Centromochlus existimatus, 73
Centromochlus gyrinus, 77
Centromochlus heckelii, 73
Centromochlus intermedius, 77
Centromochlus macracanthus, 73
Centromochlus megalops, 73
Centromochlus perugiae, 73
Centromochlus punctatus, 73
Centromochlus reticulatus, 73
Centromochlus romani, 73
Centromochlus schultzi, 73
Centromochlus Steindachneri, 73
Centrophorus, 411
cephalacanthus, *Otocinclus*, 276
cephalaspis, *Crossoloricaria*, 232
Cephalocassis, 41
Cephalocassis borneensis, 41, 42
Cephalocassis jatia, 41
Cephalocassis manillensis, 41
Cephalocassis melanochir, 41
Cephalocassis Stormii, 44, 45
Cephalosilurus, 353
Cephalosilurus albomarginatus, 353
Cephalosilurus apurensis, 353
Cephalosilurus fowleri, 353

Cephalosilurus nigricaudus, 353
Ceratocheilus, 71
Ceratocheilus osteomystax, 71, 72
Ceratoglanis, 368
Ceratoglanis pachynema, 368
Ceratoglanis pachynemus, 368
Ceratoglanis scleronemus, 368
ceratophysus, *Auchenipterus*, 79
ceratophysus, *Trachelyopterus*, 79
Ceratorhynchus, 68
cerosus, *Hemipsilichthys*, 279
cerosus, *Pareiorhaphis*, 279
Cerulescens, *Pimelodus*, 211
Cerulescens, *Silurus*, 207, 208, 211
cerulescens melanurus, *Silurus*, 209
cerulescens var. *melanurus*, *Silurus*, 209
cervinus, *Corydoras*, 115
cesarpintoi, *Glanidium*, 75
cesarpintoi, *Paraotocinclus*, 282
cesarpintoi, *Parotocinclus*, 282
cessator, *Breitensteinia*, 15
Cetopagansius, 324
Cetopangasius, 324
Cetopangasius chaetobranchus, 324
Cetopsidae, 130
Cetopsidium, 130
Cetopsidium ferreira, 130
Cetopsidium minutum, 130
Cetopsidium morenoi, 130
Cetopsidium orientale, 130
Cetopsidium pemon, 131
Cetopsidium roae, 131
Cetopsini, 130
Cetopsis, 130, 131
Cetopsis amphioxo, 131
Cetopsis arcana, 131
Cetopsis baudoensis, 131
Cetopsis caiapo, 131
Cetopsis candiru, 131
Cetopsis chalmersi, 132
Cetopsis coecutiens, 131
Cetopsis fimbriata, 132
Cetopsis gobiooides, 131, 132
Cetopsis jurubidae, 132
Cetopsis macroteronema, 133
Cetopsis montana, 132
Cetopsis motatanensis, 132
Cetopsis occidentalis, 134, 135
Cetopsis oliveirai, 132
Cetopsis orinoco, 132
Cetopsis othonops, 132
Cetopsis parma, 132
Cetopsis pearsoni, 132
Cetopsis plumbea, 133
Cetopsis plumbeus, 133
Cetopsis sandrae, 133
Cetopsis sarcodes, 133
Cetopsis Spixii, 131

- Cetopsis starnesi*, 133
Cetopsis umbrosa, 133
Cetopsis ventralis, 135
Cetopsogiton, 135
Cetopsorhamdia, 181
Cetopsorhamdia boquillae, 181
Cetopsorhamdia filamentosa, 182
Cetopsorhamdia iheringi, 182
Cetopsorhamdia insidiosa, 182
Cetopsorhamdia moliniae, 182
Cetopsorhamdia nasus, 181, 182
Cetopsorhamdia orinoco, 182
Cetopsorhamdia phantasia, 182
Cetopsorhamdia picklei, 182
Cetopsorhamdia pijpersi, 186
Cetopsorhamdia shermani, 182
ceylonensis, *Callichrous*, 372
chabanaudi, *Atopochilus*, 303
chaberti, *Trichomycterus*, 417
chabutensis, *Bachmannia*, 428
Chaca, 135
Chaca bankae, 135
Chaca bankanensis, 135
Chaca burmensis, 135
Chaca chaca, 136
Chaca hamiltonii, 135, 136
Chaca lophioides, 135, 136
chaca, *Chaca*, 136
chaca, *Platystacus*, 135, 136
Chacidae, 135
Chacini, 135
Chaenothorax, 111
Chaenothorax bicarinatus, 111, 125
Chaenothorax eigenmanni, 125
Chaenothorax multiradiatus, 120
chaetobranchus, *Cetopangasius*, 324
Chaetostoma, 227
Chaetostoma aburrense, 227
Chaetostoma aequinoctiale, 227
Chaetostoma alternifasciatum, 227
Chaetostoma anale, 227
Chaetostoma anomala sovichthys, 230
Chaetostoma anomalum, 227
Chaetostoma branickii, 228
Chaetostoma breve, 228
Chaetostoma brevilabiatum, 228
Chaetostoma changeae, 444
Chaetostoma daidalmatos, 444
Chaetostoma dermorhynchum, 228
Chaetostoma dorsale, 228
Chaetostoma dupouii, 228
Chaetostoma fischeri, 228
Chaetostoma greeni, 228
Chaetostoma guairensis, 228
Chaetostoma jegui, 228
Chaetostoma lepturum, 229
Chaetostoma leucomelas, 229
Chaetostoma lineopunctata, 229
Chaetostoma lineopunctatum, 229
Chaetostoma loborhynchos, 227, 229
Chaetostoma machiquense, 229
Chaetostoma machiquensis, 229
Chaetostoma marcapatae, 229
Chaetostoma marginatum, 229
Chaetostoma marmorescens, 229
Chaetostoma microps, 229
Chaetostoma milesi, 229
Chaetostoma mollinasum, 230
Chaetostoma niveum, 230
Chaetostoma nudirostre, 230
Chaetostoma palmeri, 230
Chaetostoma patiae, 230
Chaetostoma paucispinis, 230
Chaetostoma pearsei, 230
Chaetostoma sericeum, 230
Chaetostoma sovichthys, 230
Chaetostoma stannii, 230
Chaetostoma stroumpoulos, 444
Chaetostoma tachiraense, 230
Chaetostoma tachiraensis, 230
Chaetostoma taczanowskii, 231
Chaetostoma thomsoni, 231
Chaetostoma vagum, 231
Chaetostoma vasquezi, 231
Chaetostoma venezuelae, 231
Chaetostoma yurubiene, 231
Chaetostomidi, 217
Chaetostomus, 217
Chaetostomus (Ancistrus) cirrhosus maculatus, 222
Chaetostomus (Ancistrus) cirrhosus punctata, 222
Chaetostomus (Ancistrus) cirrhosus var. maculatus, 222
Chaetostomus (Ancistrus) cirrhosus var. punctata, 222
Chaetostomus aculeatus, 272
Chaetostomus aequinoctialis, 227
Chaetostomus alga, 221
Chaetostomus annae, 297
Chaetostomus anomalus, 227
Chaetostomus aspidolepis, 242
Chaetostomus bachi, 284
Chaetostomus branickii, 228
Chaetostomus brevilabiatus, 228
Chaetostomus brevis, 228
Chaetostomus cirrhosus var. *maculata*, 228
Chaetostomus cochliodon, 278
Chaetostomus dentex, 277
Chaetostomus depressus, 287
Chaetostomus dermorhynchus, 228
Chaetostomus dorsalis, 228
Chaetostomus eptingi, 253
Chaetostomus Fischeri, 228
Chaetostomus Fordii, 286
Chaetostomus furcatus, 284
Chaetostomus gibbosus, 278
Chaetostomus gigas, 272
Chaetostomus guairensis, 228
Chaetostomus heteracanthus, 264, 265
Chaetostomus hoplogenys, 221
Chaetostomus Jelskii, 222
Chaetostomus latifrons, 219, 222
Chaetostomus lepturus, 229
Chaetostomus leucomelas, 229
Chaetostomus leucostictus, 222
Chaetostomus macrops, 243
Chaetostomus maculatus, 228
Chaetostomus malacops, 222
Chaetostomus marcapatae, 229
Chaetostomus marginatus, 229
Chaetostomus megacephalus, 244
Chaetostomus microps, 229
Chaetostomus milesi, 229
Chaetostomus mollinasus, 230
Chaetostomus nigrolineatus, 277, 278
Chaetostomus nudirostris, 230
Chaetostomus oligospilus, 284
Chaetostomus palmeri, 230
Chaetostomus paucispinis, 230
Chaetostomus platycephalus, 231
Chaetostomus punctatissimus, 274
Chaetostomus schomburgkii, 265
Chaetostomus sericeus, 230
Chaetostomus setosus, 235
Chaetostomus Stannii, 230
Chaetostomus Taczanowskii, 231
Chaetostomus tectirostris, 221
Chaetostomus thomsoni, 231
Chaetostomus trinitatis, 225
Chaetostomus undecimalis, 292
Chaetostomus vagus, 231
Chaetostomus variolus, 225
Chaetostomus vittatus, 283, 285
chagresi, *Ancistrus*, 220, 222
chagresi, *Pimelodella*, 190
Chagresi, *Pimelodus*
(Pseudorhamdia), 190
chagresi odynea, *Pimelodella*, 193
chalmersi, *Cetopsis*, 132
chamaeleon, *Acrochordonichthys*, 12
chamaeleon, *Sosia*, 12
Chamaigenes, 57
chamaizelus, *Bunocephalus*, 59
Chandramara, 86
Chandramara chandramara, 86, 105
chandramara, *Chandramara*, 86, 105
chandramara, *Pimelodus*, 86
chandramara, *Rama*, 86
changae, *Chaetostoma*, 444
changae, *Panaqolus*, 277
changae, *Panaque*, 277
changi, *Pseudobagrus*, 103
chanjoo, *Loricariichthys*, 270
chanjoo, *Parahemiodon*, 270

- Channallabes*, 137
Channallabes apus, 137
chantrei, *Silurus*, 379
chaparae, *Pimelodella*, 190
chaparae, *Plecostomus*, 226
Chaperi, *Diastatomycter*, 368
chapini, *Acanthocheilichthron*, 303
chapmani, *Astroblepus*, 63
chapmani, *Cyclopium*, 63
chapmani, *Pygidium*, 417
chapmani, *Trichomycterus*, 417
chappuisi, *Doumea*, 25
chaquensis, *Epapterus*, 74
charmuth, *Macropteronotus*, 138, 148
charrua, *Hisonotus*, 440
charus, *Pimelodus*, 355
charus, *Pseudopimelodus*, 355
chaseni, *Leiocassis*, 93
Chasmcephalus, 182
Chasmocranes rosae, 183
Chasmocranus, 182
Chasmocranus brachynema, 182
Chasmocranus brachynemus, 182
Chasmocranus brevior, 182
Chasmocranus chimantanus, 183
Chasmocranus longior, 182, 183
Chasmocranus lopezi, 183
Chasmocranus peruanus, 183
Chasmocranus quadrizonatus, 183
Chasmocranus rosae, 183
Chasmocranus surinamensis, 183
Chasmocranus truncatorostris, 183
chaudhurii, *Glyptosternum*, 385
chauliodon, *Brochiloricaria*, 227
chechra, *Silurus*, 374
Cheiridodus, 250
Cheirocerus, 332
Cheirocerus abuelo, 332
Cheirocerus eques, 332
Cheirocerus goeldii, 332
chennuah, *Sisor*, 402
Cheveyi, *Cryptopterus*, 371
cheveyi, *Micronema*, 371
childreni, *Ageneiosus*, 366
childreni, *Silonia*, 366
chilensis, *Diplomystes*, 165
chilensis, *Silurus*, 165
Chiloglanidinae, 303
Chiloglanis, 303, 304
Chiloglanis angolensis, 304
Chiloglanis anoterus, 304
Chiloglanis asymetricaudalis, 304
Chiloglanis athiensis, 304
Chiloglanis batesii, 304
Chiloglanis benuensis, 305
Chiloglanis bifurcus, 305
Chiloglanis brevibarbis, 305
Chiloglanis cameronensis, 305
Chiloglanis carnosus, 305
Chiloglanis congicus, 305
Chiloglanis deckenii, 304, 305
Chiloglanis disneyi, 305
Chiloglanis dybowskii, 309
Chiloglanis elisabethianus, 305
Chiloglanis emarginatus, 305
Chiloglanis engiops, 308
Chiloglanis fasciatus, 306
Chiloglanis harbinger, 306
Chiloglanis kalambo, 306
Chiloglanis lamottei, 306
Chiloglanis lufirae, 306
Chiloglanis lukugae, 306
Chiloglanis macropterus, 306
Chiloglanis marlieri, 306
Chiloglanis mbozi, 306
Chiloglanis micropogon, 305
Chiloglanis microps, 306
Chiloglanis modjensis, 306
Chiloglanis neumannii, 307
Chiloglanis niger, 307
Chiloglanis niloticus, 307
Chiloglanis niloticus waterloti, 307
Chiloglanis normani, 307
Chiloglanis occidentalis, 307
Chiloglanis paratus, 307
Chiloglanis pojeri, 307
Chiloglanis polyodon, 307
Chiloglanis polypogon, 307
Chiloglanis pretoriae, 307
Chiloglanis productus, 443
Chiloglanis pumilus, 307
Chiloglanis reticulatus, 307
Chiloglanis rukwaensis, 308
Chiloglanis ruziziensis, 308
Chiloglanis sanagaensis, 308
Chiloglanis sardinhai, 308
Chiloglanis somereni, 308
Chiloglanis swierstrai, 308
Chiloglanis trilobatus, 308
Chiloglanis voltae, 308
chiltoni, *Pygidium*, 417
chiltoni, *Trichomycterus*, 417
chimantanus, *Chasmocranus*, 183
Chimarrhoglanis, 21
Chimarrhoglanis Leroyi, 21, 23
Chimarrichthys, 385
Chimarrichthys Davidi, 385
chimborazoi, *Astroblepus*, 63
chimborazoi, *Cyclopium*, 63
chinensis, *Macrones*, 88
chinensis, *Pseudobagrus*, 101
chinensis, *Tachisurus*, 431
chinta, *Bagrus*, 431
chiquitos, *Callichthys*, 128
Chlarias, 138
Chlarias angolensis macronema, 141
Chlarias olivaceus, 146
chlorostictus, *Hemiancistrus*, 243
Choeroplotosus, 346
Choeroplotosus decemfilis, 346
Choeroplotosus limbatus, 346
chondropterygioides, *Arius*, 55
chondropterygius, *Arius*, 55
chotae, *Arges*, 63
chotae, *Astroblepus*, 63
christyi, *Atopochilus*, 303
Christyi, *Microsynodontis*, 309
chrysea, *Rita*, 105
chryseus, *Pseudobagrus*, 429
Chrysichthyinae, 153
Chrysichthys, 153, 155
Chrysichthys acutirostris, 158
Chrysichthys aluvensis, 155
Chrysichthys ansorgii, 155
Chrysichthys auratus, 155, 158, 160
Chrysichthys bocagii, 155
Chrysichthys börressoni, 156
Chrysichthys brachynema, 155
Chrysichthys brevibarbis, 156
Chrysichthys büttikoferi, 158
Chrysichthys camaronensis, 158
Chrysichthys cameronensis, 158
Chrysichthys coriscanus, 160
Chrysichthys cranchii, 156
Chrysichthys cyclurus, 161
Chrysichthys dageti, 156
Chrysichthys delhezi, 156
Chrysichthys delhezi thomasi, 156
Chrysichthys duttoni, 156
Chrysichthys eaglesomei, 160
Chrysichthys filamentosus, 155
Chrysichthys furcatus, 156
Chrysichthys grandis, 156
Chrysichthys graueri, 156
Chrysichthys habereri, 156
Chrysichthys helicophagus, 156
Chrysichthys hildae, 153
Chrysichthys johnelsi, 156
Chrysichthys kingsleyae, 160
Chrysichthys lagoensis, 160
Chrysichthys laticeps, 157
Chrysichthys levequei, 157
Chrysichthys longibarbis, 157
Chrysichthys longidorsalis, 157
Chrysichthys longidorsalis nyongensis, 158
Chrysichthys longifilis, 155
Chrysichthys longipinnis, 157
Chrysichthys mabusi, 157
Chrysichthys macropogon, 160
Chrysichthys macrops, 155
Chrysichthys macropterus, 157
Chrysichthys macrotis, 157
Chrysichthys magnus, 157
Chrysichthys mahengeensis, 157
Chrysichthys maurus, 157, 158, 160
Chrysichthys myriodon, 155

- Chrysichthys nigrodigitatus*, 158, 160
Chrysichthys nyongensis, 158
Chrysichthys ogooensis, 158
Chrysichthys ogowensis, 158
Chrysichthys okae, 158
Chrysichthys ornatus, 158
Chrysichthys persimilis, 158
Chrysichthys pitmani, 160
Chrysichthys platycephalus, 158
Chrysichthys polli, 159
Chrysichthys punctatus, 159
Chrysichthys rueppelli, 159
Chrysichthys sharpii, 159
Chrysichthys sianenna, 159
Chrysichthys stappersii, 159
Chrysichthys teugelsi, 159
Chrysichthys theobaldi, 105
Chrysichthys thonneri, 159
Chrysichthys thysi, 159
Chrysichthys uniformis, 159
Chrysichthys velifer, 157, 158
Chrysichthys wagenaari, 159
Chrysichthys walkeri, 159, 160
Chrysobagrus, 155
Chrysobagrus brevibarbis, 155, 156
Chrysobagrus longibarbis, 157
chrysops, Hemibagrus, 87
chungaraensis, Trichomycterus, 417
cibelae, Microglanis, 354
cinerascens, Pimelodella, 201
cinerascens, Pimelodus, 200
cinereus, Silurus, 379
Cinetodus, 42
Cinetodus carinatus, 42
Cinetodus conorrhynchus, 42
Cinetodus crassilabris, 42
Cinetodus froggatti, 42
cirratus, Arges, 64
cirratus, Astroblepus, 64
cirrhosa, Loricaria, 268, 296
cirrhosa, Vandellia, 427
cirrhosus, Ancistrus, 220
cirrhosus, Hypostomus, 219, 220
cirrhosus dubius, Ancistrus, 221
cirrhosus maculatus, Chaetostomus (Ancistrus), 222
cirrhosus punctata, Chaetostomus (Ancistrus), 222
cirrhosus var. maculata, Chaetostomus, 228
cirrhosus var. maculatus, Chaetostomus (Ancistrus), 222
cirrhosus var. punctata, Chaetostomus (Ancistrus), 222
cisternarum, Phreatobius, 429, 430
citatus, Silurichthys, 377
Citerñii, Synodontis, 314
citurensis, Oxylicaria, 300
citurensis, Sturisomatichthys, 300
Claibornichthys, 434
Claibornichthys troelli, 434, 435
clandestinus, Tetracanthphilius, 27
Clariallabes, 137
Clariallabes attenuata, 137
Clariallabes brevibarbis, 137
Clariallabes centralis, 137
Clariallabes dumerili, 138
Clariallabes heterocephalus, 137
Clariallabes laticeps, 137
Clariallabes longicauda, 137
Clariallabes longicaudatus, 150
Clariallabes manyangae, 137
Clariallabes melas, 137
Clariallabes melas lembesseensis, 137
Clariallabes mutsindziensis, 137
Clariallabes petricola, 138
Clariallabes pietschmanni, 138
Clariallabes platyprosopos, 138
Clariallabes simeoni, 138
Clariallabes teugelsi, 138
Clariallabes uelensis, 138
Clariallabes variabilis, 138
Clarias, 136, 138
Clarias (Allabenchelys) dumerili longibarbis, 138
Clarias (Allabenchelys) submarginatus thysvillensis, 141
Clarias (Allabenchelys) uelensis, 138
Clarias (Clariooides) aboinensis, 144
Clarias (Clariooides) agboyiensis, 139
Clarias (Clariooides) curtus, 140
Clarias (Clariooides) isheriensis, 139
Clarias abbreviatus, 148
Clarias agboyiensis, 139
Clarias albopunctatus, 139
Clarias alluaudi, 139
Clarias amplexicauda, 148
Clarias anfractus, 139
Clarias angolensis, 139
Clarias anguillaris, 140, 144, 148
Clarias anguillaris nigeriensis, 140
Clarias Assamensis, 148
Clarias batrachus, 140
Clarias batu, 140
Clarias brachysoma, 140
Clarias breviceps, 141
Clarias brevinuchalis, 140
Clarias budgetti, 140
Clarias buettikoferi, 140
Clarias bulumae, 140
Clarias buthupogon, 141
Clarias büttikoferi, 141
Clarias cameronensis, 144
Clarias camerunensis, 139, 141, 144
Clarias capensis, 143
Clarias carsonii, 145
Clarias cataractus, 141
Clarias cavernicola, 141
Clarias centralis, 137
Clarias confluentus, 140
Clarias congicus, 142
Clarias dahomeyensis, 142
Clarias dayi, 141
Clarias depressus, 143
Clarias dhonti, 142
Clarias dialonensis, 142
Clarias dolloi, 142
Clarias dorsimarmoratus, 141
Clarias duchaillui, 141
Clarias dumerili, 142
Clarias dussumieri, 142
Clarias ebriensis, 142
Clarias ekibondoi, 147
Clarias engelseni, 142
Clarias esamesae, 146
Clarias eupogon, 153
Clarias falconeri, 142
Clarias fouloni, 148
Clarias foveolatus, 149
Clarias fuscus, 142
Clarias gabonensis, 142
Clarias gariepinus, 140, 143, 148
Clarias gilli, 146
Clarias gracilis, 144
Clarias guentheri, 143
Clarias guineensis, 141
Clarias hasselquistii, 140
Clarias hilgendorfi, 139
Clarias hilli, 144
Clarias hollyi, 144, 147
Clarias insolitus, 144
Clarias intermedius, 144
Clarias jaensis, 144
Clarias kapuasensis, 144
Clarias kingsleyae, 144
Clarias laeviceps, 142, 144
Clarias laeviceps dialonensis, 142
Clarias lamottei, 144
Clarias lazera, 143
Clarias leiacanthus, 145
Clarias liacanthus, 145
Clarias liberiensis, 141
Clarias lindicus, 141
Clarias liocephalus, 145
Clarias loangwensis, 151
Clarias longibarbis, 136, 138
Clarias longiceps, 143
Clarias longior, 145
Clarias lualae, 142
Clarias maclareni, 145
Clarias macracanthus, 143
Clarias macrocephalus, 145
Clarias macromystax, 145
Clarias macrurus, 148
Clarias malaris, 143
Clarias marpus, 148
Clarias megapogon, 141

Clarias meladerma, 145
Clarias melanoderma, 146
Clarias melanopogon, 145
Clarias melas, 137
Clarias melasoma, 146
Clarias Mellandi, 146
Clarias microphthalmus, 143
Clarias microstomus, 146
Clarias monkei, 141
Clarias moorii, 143
Clarias mossambicus, 143
Clarias mülleri, 143
Clarias N'gamensis, 146
Clarias nebulosus, 148
Clarias neumanni, 145
Clarias ngamensis, 146
Clarias ngola, 142
Clarias nieuhofii, 139, 146
Clarias nigeriae, 141
Clarias nigricans, 146
Clarias nigromarmoratus, 146
Clarias notozygurus, 143
Clarias nyasensis, 149
Clarias obscurus, 141
Clarias olivaceus, 146
Clarias ornatus, 145
Clarias orontis, 143
Clarias oxycephalus, 146
Clarias pachynema, 146
Clarias parvimanus, 140
Clarias pentapterus, 146
Clarias phillipsi, 145
Clarias planiceps, 147
Clarias platycephalus, 139, 147
Clarias plioacaenius, 432
Clarias poensis, 141
Clarias pseudoleiacanthus, 147
Clarias pseudonieuhoefii, 147
Clarias pulcher, 147
Clarias pulicaris, 142
Clarias punctatus, 140
Clarias robecchii, 143
Clarias salae, 147
Clarias sauteri, 148
Clarias Senegalensis, 140
Clarias smithii, 143
Clarias stappersii, 147
Clarias submarginatus, 147
Clarias sulcatus, 147
Clarias Syriacus, 143
Clarias teijsmanni, 147
Clarias tenuis, 148
Clarias theodorae, 139, 147
Clarias thienemanni, 148
Clarias tsanensis, 143
Clarias vandenhouwei, 144
Clarias varispinis, 147
Clarias vinciguerreae, 143
Clarias walkeri, 141

Clarias wernerii, 148
Clarias xenodon, 143
Clarias youngicus, 145
Clarias zygouron, 141
clarias, *Silurus*, 311, 313, 321, 336, 337
clarias, *Pimelodus*, 320
clarias, *Pseudariodes*, 336
clarias*, *Synodontis, 313
clarias coprophagus, *Pimelodus*, 337
Clariidae, 136
Clariini, 136
Clariooides, 139
Clarislurus, 151
Clarislurus kemratensis, 151, 152
claro*, *Ancistrus, 220
Claroteidae, 153
Claroteini, 153
Clarotes, 153, 160
Clarotes bidorsalis, 160
Clarotes heuglini, 160
Clarotes laticeps, 160
Clarotes macrocephalus, 160
clauseni*, *Phractura, 26
clavipinna*, *Loricaria, 268
clavispinosus, *Arius*, 36
clavulus*, *Akysis, 14
clementinae*, *Ancistrus, 220
cleptolepis, *Arius*, 47
clijsenaar*, *Arius, 55
clinatus*, *Akysis, 14, 357
Clupisoma bastari, 357
Clupisoma garua, 357
Clupisoma longianalis, 357
Clupisoma montana, 357
Clupisoma naziri, 357
Clupisoma nuijangense, 357
Clupisoma prateri, 357
Clupisoma roosae, 357
Clupisoma sinense, 358
clypeastroides, *Arius*, 55
Cnidoglanis, 345
Cnidoglanis bostockii, 346
Cnidoglanis lepturus, 346, 442
Cnidoglanis macrocephalus, 346
Cnidoglanis microcephalus, 346
Cnidoglanis mülleri, 349
Cnidoglanis nudiceps, 346
coatesi, *Arius*, 47
coatesi*, *Neoarius, 47
coatesi*, *Neosilurus, 348
coatesi, *Tandanus*, 348
Cobitiglanis, 405
cobreensis*, *Dolichancistrus, 235
cobreensis, *Pseudancistrus pediculatus*, 235
cocama*, *Otocinclus, 274
cochabambae*, *Imparfinis, 185
cochabambae, *Pimelodella*, 185
cochinchinensis, *Arius*, 34
cochinchinensis*, *Pterocryptis, 376
cochinchinensis, *Silurus*, 376
cochlearis, *Hemipimelodus*, 43
Cochlefelis, 42
Cochlefelis burmanica, 42
Cochlefelis danielsi, 42
Cochlefelis spatula, 43
Cochliodon, 250
Cochliodon hypostomus, 252
Cochliodon plecostomoides, 258
Cochliodon pospisili, 254
Cochliodon pyrineusi, 259
Cochliodon taphorni, 261
cochliodon, *Chaetostomus*, 278
cochliodon*, *Hypostomus, 250, 252, 253-262
cochliodon*, *Panaque, 277
cochui*, *Corydoras, 115
coecutiens*, *Cetopsis, 131
coecutiens*, *Silurus, 131
***coelestinus*, *Bagrus* (*Bagrus*)**, 53
coerulescens*, *Pimelodus, 81
coeruleus*, *Brochus, 111, 125
coheni*, *Glyptothorax, 388, 389
coila*, *Ailia, 356
coila*, *Malapterurus, 356
colcloughi*, *Hemipimelodus, 51
Collettii*, *Heptapterus, 99
collinsae*, *Parotocinclus, 282
collinus*, *Leiocassis, 443
colombianus*, *Bunocephalus, 59
colombiense*, *Acetridium, 218
colombiensis*, *Acetridium, 218
colombiensis*, *Farlowella, 237
colvillii*, *Macrones, 97
colyeri*, *Synodontis, 318
colymbetes*, *Euanemus, 71, 72
commersoni*, *Hypostomus, 252
Commersonii*, *Pimelodus, 43, 44
commersonii scabriceps, *Plecostomus*, 260
commersonoides*, *Loricaria, 278
commersonoides*, *Paraloricaria, 278
commersonoides, *Plecostomus*, 255
comoensis*, *Synodontis, 313
conchophilus*, *Pangasius, 325
concolor*, *Centromochlus, 73
concolor*, *Corydoras, 115
concolor*, *Tatia, 73
concolor*, *Trichomycterus, 417
condei*, *Aristoloricaria, 226
condiscipulus*, *Corydoras, 115
confinis*, *Pimelodus, 205
confluentus*, *Clarias, 140
congensis*, *Schilbe, 363
congensis*, *Silurus, 362, 363
congica*, *Parailia, 360
congica*, *Synodontis, 313

conicus, *Chiloglanis*, 305
conicus, *Clarias*, 142
conicus, *Gephyroglanis*, 160, 161
conicus, *Synodontis*, 313
congolensis, *Eutropius*, 363
congolensis, *Schilbe*, 364
conirostre, *Glyptosternum*, 389
conirostre poonaensis, *Glyptothonax*, 395
conirostris, *Conorhynchos*, 428
conirostris, *Glyptothonax*, 389, 395
conirostris, *Pimelodus*, 428
conirostris punjabensis, *Glyptothonax*, 393
Conorhynchichthys, 428
Conorhynchos, 53, 428
Conorhynchos conirostris, 428
Conorhynchos nelsoni, 53
Conorhynchus, 428
conorhynchus*, *Cinetodus, 42
conorhynchus, *Tetramesodon*, 42
Conorhynchus glaber, 428
Conostome, 428
conquetaensis, *Pimelodella*, 190
conquistador, *Gladioglanis*, 183
conradi, *Pygidium*, 417
conradi, *Trichomycterus*, 417
Conta, 382, 383
Conta conta, 384
Conta pectinata, 384
conta*, *Conta, 384
conta, *Pimelodus*, 383, 384
Continae, 382
contracta*, *Synodontis, 313
contractus, *Synodontis*, 313
cookei, *Arius*, 50
cookei*, *Notarius, 50
cooperensis, *Neosiluroides*, 347
copei, *Anduzedoras*, 172
copei*, *Corydoras, 115
copei*, *Leptodoras, 172
copei, *Pimelodella*, 190
Copidoglanis, 351
Copidoglanis brevidorsalis, 347
Copidoglanis curtus, 345
Copidoglanis equinus, 348
Copidoglanis gjellerupi, 348
Copidoglanis glencoensis, 348
Copidoglanis idenburgi, 348
Copidoglanis labiosus, 345
Copidoglanis labrosus, 345
Copidoglanis levis, 345
Copidoglanis longifilis, 349
Copidoglanis meraukensis, 351
Copidoglanis novae-guineae niger, 349
Copidoglanis obscurus, 345, 350
Copidoglanis papuensis, 348
Copidoglanis rendahli, 345

Copionodon, 403, 404
Copionodon lianae, 404
Copionodon orthiocarinatus, 404
Copionodon pecten, 404
Copionodontinae, 403
coppenameensis, *Corydoras*, 115
coppenameensis, *Corydoras bondi*, 115
coppenameensis, *Hypostomus*, 253
coprophagus*, *Pimelodus, 337
coprophagus, *Pimelodus clariss*, 337
coquenani, *Ancistrus (Pseudancistrus)*, 287
coquenani*, *Pseudancistrus, 287
coracoideus*, *Auchenipterichthys, 71
coracoideus*, *Bunocephalus, 59
coracoideus, *Dysichthys*, 58, 59
coracoideus, *Trachycorystes*, 71
Coraglanis, 385
Coraglanis kishinouyei, 385
corantijni*, *Hypostomus, 253
cordobensis*, *Leptoancistrus, 266
Cordorinus, 111
cordovae, *Plecostomus*, 258
corduvensis*, *Trichomycterus, 417
Cordylancistrus, 231, 443
Cordylancistrus daguae, 231
Cordylancistrus nephelion, 443
Cordylancistrus perijae, 231
Cordylancistrus platycephalus, 231
Cordylancistrus platyrhynchus, 232
Cordylancistrus torbesensis, 232
Coreobagrus, 86, 100
Coreobagrus brevicorpus, 86
Coreobagrus ichikawai, 86
Coreobagrus okadai, 86
coriaceus*, *Trachelyopterus, 79
coriaceus maculosus, *Trachelyopterus*, 79
coriatae*, *Corydoras, 115
coriscanus, *Chrysichthys*, 160
corruscans, *Platystoma*, 341
corruscans*, *Pseudoplatystoma, 341
corsula, *Bagrus*, 89
corsula, *Macrones*, 90
corsula, *Mugil*, 89
corsula, *Pimelodus*, 89
cortesi, *Corydoras*, 124
Corumbataia, 232
Corumbataia cuestae, 232
Corumbataia tocantinensis, 232
coruscans, *Pimelodes*, 37
coruscans, *Platystoma*, 341
Corydoradinae, 108
Corydoras, 108, 111
Corydoras acrensis, 112
Corydoras acutus, 112
Corydoras adolfoi, 112
Corydoras aeneus, 112
Corydoras agassizii, 112

Corydoras albolineatus, 112
Corydoras amandajanea, 112
Corydoras amapaensis, 113
Corydoras ambiacus, 113
Corydoras amphibelus, 113
Corydoras approuaguensis, 113
Corydoras araguaiaensis, 113
Corydoras arcuatus, 113
Corydoras areio, 113
Corydoras armatus, 113
Corydoras atropersonatus, 113
Corydoras aurofrenatus, 113
Corydoras australis, 118
Corydoras axelrodi, 113
Corydoras baderi, 113
Corydoras bertoni, 130
Corydoras bicolor, 114
Corydoras bifasciatus, 114
Corydoras bilineatus, 114
Corydoras blochi, 114
Corydoras blochi vittatus, 126
Corydoras boehlkei, 114
Corydoras boesemani, 114
Corydoras boliviensis, 117
Corydoras bondi, 114
Corydoras bondi coppenamensis, 115
Corydoras breei, 114
Corydoras brevirostris, 114
Corydoras britskii, 114
Corydoras burgessi, 114
Corydoras caquetae, 119
Corydoras carlcae, 115
Corydoras caudimaculatus, 115
Corydoras cervinus, 115
Corydoras cochui, 115
Corydoras concolor, 115
Corydoras condiscipulus, 115
Corydoras copei, 115
Corydoras coppenamensis, 115
Corydoras coriatae, 115
Corydoras cortesi, 124
Corydoras crimeni, 115
Corydoras cruzeiensis, 115
Corydoras crypticus, 115
Corydoras davidsandsi, 116
Corydoras delphax, 116
Corydoras difluviatilis, 116
Corydoras diphyes, 116
Corydoras dubius, 126
Corydoras duplicareus, 116
Corydoras edentatus, 176
Corydoras ehrhardti, 116
Corydoras eigenmanni, 129
Corydoras elegans, 111, 116
Corydoras elegans njisseni, 121
Corydoras ellisae, 116
Corydoras ephippifer, 116
Corydoras episcopi, 126
Corydoras eques, 112, 116

- Corydoras esperanzae*, 117
Corydoras evelynae, 117
Corydoras filamentosus, 117
Corydoras flaveolus, 117
Corydoras fowleri, 117
Corydoras funnelli, 119
Corydoras garbei, 117
Corydoras geoffroy, 111, 117
Corydoras geryi, 117
Corydoras gomezi, 117
Corydoras gossei, 118
Corydoras gracilis, 118
Corydoras grafi, 113
Corydoras griseus, 118
Corydoras guapore, 118
Corydoras guianensis, 118
Corydoras habrosus, 118
Corydoras haraldschultzi, 118
Corydoras hastatus, 112, 118
Corydoras heteromorphus, 118
Corydoras imitator, 118
Corydoras incolicana, 118
Corydoras isbrueckeri, 119
Corydoras julii, 119
Corydoras juquiaae, 121
Corydoras juquiae, 121
Corydoras kanei, 119
Corydoras kronei, 129
Corydoras lacerdai, 119
Corydoras lambererti, 119
Corydoras latus, 119
Corydoras leopardus, 119
Corydoras leucomelas, 119
Corydoras loretoensis, 119
Corydoras loxozonus, 119
Corydoras macropterus, 130
Corydoras macrosteus, 112
Corydoras maculatus, 122
Corydoras maculifer, 119
Corydoras mamore, 120
Corydoras mamoré, 120
Corydoras marmoratus, 122
Corydoras melanistius, 120
Corydoras melanistius brevirostris, 114
Corydoras melanistius longirostris, 113
Corydoras melanotaenia, 120
Corydoras melini, 120
Corydoras meridionalis, 116
Corydoras metae, 120
Corydoras micracanthus, 120
Corydoras microcephalus, 122
Corydoras microps, 112
Corydoras multimaculatus, 120
Corydoras multiradiatus, 120
Corydoras myersi, 123
Corydoras nanus, 120
Corydoras napoensis, 120
Corydoras narcissus, 120
Corydoras nattereri, 121
Corydoras nattereri triseriatus, 121
Corydoras negro, 121
Corydoras njsseni, 121
Corydoras noelkempffi, 121
Corydoras octocirrus, 117
Corydoras oelemariensis, 114
Corydoras oiapoquensis, 121
Corydoras ornatus, 121
Corydoras orphnopterus, 121
Corydoras osteocarus, 121
Corydoras ourastigma, 121
Corydoras oxyrhynchus, 121
Corydoras paleatus, 122
Corydoras pantanalensis, 122
Corydoras paragua, 122
Corydoras parallelus, 122
Corydoras pastazensis, 122
Corydoras pastazensis orcesi, 122
Corydoras paucerna, 122
Corydoras pauciradiatus, 109
Corydoras pestai, 116
Corydoras pinheiroi, 122
Corydoras polystictus, 123
Corydoras potaroensis, 123
Corydoras prionotos, 130
Corydoras pulcher, 123
Corydoras punctatus, 123
Corydoras punctatus argentina, 122
Corydoras punctatus sipaliwini, 125
Corydoras pygmaeus, 123
Corydoras rabauti, 123
Corydoras raimundi, 110
Corydoras reticulatus, 123
Corydoras revelatus, 123
Corydoras reynoldsi, 123
*Corydoras robinea*e, 124
Corydoras robustus, 124
Corydoras sanches, 124
Corydoras saramaccensis, 124
Corydoras sarareensis, 124
Corydoras schultzei, 112
Corydoras schwartzii, 124
Corydoras schwartzii surinamensis, 126
Corydoras semiaquilus, 124
Corydoras semiscutatus, 125
Corydoras septentrionalis, 124
Corydoras serratus, 124
Corydoras seussi, 124
Corydoras similis, 124
Corydoras simulatus, 124
Corydoras sipaliwini, 125
Corydoras sodalis, 125
Corydoras solox, 125
Corydoras spectabilis, 125
Corydoras spilurus, 125
Corydoras splendens, 125
Corydoras steindachneri, 125
Corydoras stenocephalus, 125
Corydoras sterbai, 125
Corydoras surinamensis, 126
Corydoras sychri, 126
Corydoras treitlii, 126
Corydoras trilineatus, 126
Corydoras tukano, 126
Corydoras undulatus, 126
Corydoras venezuelanus, 112
Corydoras virescens, 123
Corydoras virginiae, 126
Corydoras vittatus, 126
Corydoras weitzmani, 126
Corydoras wotroi, 120
Corydoras xinguensis, 126
Corydoras zygatus, 126
Corymbophanes, 217, 232
Corymbophanes andersoni, 232
Corymbophanes bahianus, 279
Corymbophanes kaiei, 232
Corymbophanes venezuelae, 231
Cossyphus, 139
Cossyphus ater, 139, 148
costatus, *Platydoras*, 176
costatus, *Silurus*, 176
cottoides, *Microglanis*, 354
cottoides, *Pimelodus*
 (*Pseudopimelodus*), 354
Cotylephorus, 61
Cotylephorus blochii, 61
cotylephorus, *Platystacus*, 61
couma, *Bagrus*, 53
couma, *Sciades*, 53
courteti, *Synodontis*, 313
cous, *Arius*, 31
cous, *Glyptothorax*, 31, 389
cous, *Silurus*, 31, 387, 389
cragini, *Amiurus*, 205
cranchii, *Chrysichthys*, 156
cranchii, *Pimelodus*, 156
Cranoglanididae, 164
Cranoglanidae, 164
Cranoglanis, 164
Cranoglanis boudierius, 165
Cranoglanis henrici, 165
Cranoglanis multiradiatus, 165
Cranoglanis sinensis, 164, 165
crassicauda, *Harttia*, 242
crassicauda, *Harttiella*, 242
crassicauda, *Hypostomus*, 253
crassicauda, *Pseudecheneis*, 400
crassilabris, *Cinetodus*, 42
crassilabris, *Hemipimelodus*, 42
crassilabris, *Leiocassis*, 92
crassilabris, *Liocassis*, 92
crassilabris macrops, *Leiocassis*, 92
crassirostris, *Leiocassis*, 92

crassirostris, *Liocassis*, 92
crassus, *Arius*, 435
crassus, *Otolithus* cf., 434
crassus, *Otolithus (incertae sedis)*, 434
crassus, *Tachysurus*, 434, 435
cratensis, *Trachycorystes*, 80
crenula, *Pterocryptis*, 376
cretaceus, *Brachyspondylus*, 432
kreutzbergi, *Centromochlus*, 77
kreutzbergi, *Tatia*, 77
crimmeni, *Corydoras*, 115
crinalis, *Bagrus*, 55
cristata, *Pimelodella*, 190
cristata, *Physopyxis*, 176
cristatus, *Parotocinclus*, 282
cristatus, *Pimelodus*, 189, 190
crocodili, *Centrochir*, 170
Crocodili, *Doras*, 170
crossocheilos, *Arius*, 52
Crossoloricaria, 232
Crossoloricaria bahuaja, 232
Crossoloricaria cephalaspis, 232
Crossoloricaria rhami, 232
Crossoloricaria variegata, 233
Crossoloricaria venezuelae, 233
cruciger, *Arius*, 106
Cruciglanis, 443
Cruciglanis pacifici, 443
cruxenti, *Pimelodella*, 190
cruziensis, *Corydoras*, 115
Cryptarius, 43
Cryptarius daugueti, 43
Cryptarius truncatus, 43
crypticus, *Corydoras*, 115
crypticus, *Noturus*, 211, 442
cryptobullatus, *Amphilus*, 21
cryptodon, *Planiloricaria*, 285
cryptodon, *Pseudohemiodon*
(*Planiloricaria*), 285
cryptopthalmus, *Ancistrus*, 220
Cryptopterella, 369
Cryptopterella beldti, 369, 371
Cryptopterus, 369
Cryptopterus amboinensis, 369
Cryptopterus bleekeri, 374
Cryptopterus cheveyi, 371
Cryptopterus latovittatus, 374
Cryptopterus lumholzii, 370
Cryptopterus urbaini, 373
cryptopterus, *Kryptopterus*, 369
cryptopterus, *Silurus*, 369
cryptus, *Paradoxoglanis*, 303
Cteniloricaria, 233
Cteniloricaria fowleri, 233
Cteniloricaria maculata, 233
Cteniloricaria platystoma, 233
ctenodus, *Pimelodus*, 332
cuangoana, *Synodontis*, 313
cuangoanus, *Synodontis*, 313

cuao, *Pygidianops*, 412
cubangoensis, *Amphilus platychir*, 23
cubataonis, *Loricaria*, 294
cubataonis, *Rineloricaria*, 294
cubataonis, *Trichomycterus*, 425
cucphuongensis, *Pterocryptis*, 376
cucphuongensis, *Silurus*, 376
cucuyensis, *Pseudepapterus*, 76
cuestae, *Corumbataia*, 232
cuiabae, *Ancistrus*, 220
cuiyi, *Nematogenys*, 324
culturatus, *Pangasius*, 328
cunaguaro, *Ginesia*, 330, 331
cunningtoni, *Dinotopterus*, 148
cupido, *Trachymochlus*, 431, 432
cuprea, *Pimelodus (Rhamdia) Queleni*,
201
cupreoides, *Pimelodus*, 205
cupreus, *Silurus*, 215
curtirostra, *Farlowella*, 237
curtisi, *Arius*, 47
curtisoma, *Encheloclarias*, 149
curtus, *Clarias (Clarioides)*, 140
curtus, *Copidoglanis*, 345
curvirostris, *Belonoglanis*, 24
curvispina, *Loricaria*, 296
curvispina, *Spatuloricaria*, 296
cuspicaudus, *Sorubim*, 342
cuvieri, *Genidens*, 44
Cuvieri, *Silurus (Acanthonotus)*, 356
Cuyabae, *Pimelodus (Rhamdia)*, 201
cuyanus, *Diplomystes viedmensis*, 166
cuyanus, *Olivaichthys*, 166
cyanochloros, *Pimelodus*, 392
cyanostigma, *Pimelodella*, 190
cyanostigma, *Rhamdia*, 190
Cyclopidae, 63
Cyclopium, 63
Cyclopium chapmani, 63
Cyclopium chimborazoi, 63
Cyclopium humboldtii, 63, 64
Cyclopium mariae, 65
Cyclopium mindoense, 65
Cyclopium pirrense, 66
Cyclopium trifasciatum, 67
Cyclopium ubidai, 67
Cyclopium unifasciatum, 67
Cyclopium vanceae, 67
Cyclopium ventrale, 68
cyclopium, *Pimelodus*, 64
cyclopium, *Pimelodus*, 64
cyclopus, *Astroblepus*, 64
cyclopus santanderensis, *Astroblepus*,
67
cyclurus, *Chrysichthys*, 161
cyclurus, *Lophiobagrus*, 161
D
dagetii, *Chrysichthys*, 156
dagetii, *Synodontis*, 312

daguae, *Cordylancistrus*, 231
daguae, *Hemiancistrus*, 231
dahli, *Anodontiglanis*, 345
dahomeyensis, *Clarias*, 142
daidalmatos, *Chaetostoma*, 444
daillyi, *Hoplosternum littorale*, 128
dakpathari, *Glyptothorax*, 389
dalungshanensis, *Mystus*, 95
damasceni, *Ancistrus*, 220
damasceni, *Xenocara*, 220
danicus, *Otolithus (Arius)*, 434
danielsi, *Arius (Hemiarthus)*, 42
danielsi, *Cochlefelis*, 42
dantei, *Auchenipterichthys*, 71
dariense, *Sturisoma*, 299
dariensis, *Oxyloricaria*, 299
dasycephalus, *Arius*, 39
dasycephalus, *Cathorops*, 39
Dasyloricaria, 233
Dasyloricaria capetensis, 233
Dasyloricaria filamentosa, 233
Dasyloricaria latiura, 233
Dasyloricaria seminuda, 234
Dasyloricaria tuyensis, 234
daugueti, *Cryptarius*, 43
Daugueti, *Hemipimelodus*, 43
dauuricus, *Silurus*, 379
Davalla, 68
Davalla Schomburgkii, 68, 69
davalla, *Hypothalmus*, 69
Davidi, *Chimarrichthys*, 385
davidi, *Euchiloglanis*, 385
davidi, *Synodontis*, 313
davidsandsi, *Corydoras*, 116
davisi, *Pygidium*, 417
davisi, *Trichomycterus*, 417
davissinghi, *Glyptothorax*, 389
dawalla, *Hypothalmus*, 69
dayi, *Arius*, 49
dayi, *Batasio*, 85
dayi, *Clarias*, 141
dayi, *Hemipimelodus*, 46
Dayi, *Macrones*, 85
dayi, *Nedystoma*, 46
de Zwaanii, *Pangasius*, 328
debauwi, *Eutropius*, 361
debauwi, *Pareutropius*, 361
Decapogon, 127
Decapogon urostriatum, 127
Decapogon urotriatum, 127
Decapogon verissimi, 127
decaradiatus, *Trachelyichthys*, 78
decemfilis, *Trachelyichthys*, 346
decipiens, *Otolithus (Sciaenidarum)*,
434
deckenii, *Chiloglanis*, 304, 305
decolor, *Rhamdia guatemalensis*, 202
decora, *Synodontis*, 314
decorus, *Amiurus*, 210

- decorus*, *Synodontis*, 314
degeni, *Bagrus*, 84
deignani, *Kryptopterus*, 375
dekayi, *Pimelodus*, 206
Dekeyseria, 234
Dekeyseria amazonica, 234
Dekeyseria brachyura, 234
Dekeyseria niveata, 234
Dekeyseria picta, 234
Dekeyseria pulchra, 234
Dekeyseria scaphirhyncha, 234
dekimpei, *Synodontis*, 314
dekkanense, *Glyptosternum*, 391
delacouri, *Oreoglanis*, 397
delacouri, *Paroreoglanis*, 397
delhezi, *Chrysichthys*, 156
delhezi thomasi, *Chrysichthys*, 156
delicata, *Psammophiletria*, 27
delicatissimus, *Pangasius*, 328
delphax, *Corydoras*, 116
Deltadoras, 173
Deltadoras guayoensis, 173, 174
Delturinae, 444
Delturus, 234, 444
Delturus angulicauda, 234, 444
Delturus brevis, 443
Delturus carinotus, 235
Delturus parahybae, 234, 235, 444
demanoides, *Baryancistrus*, 226
demararae, *Callichthys callichthys*, 111
demerarae, *Auchenipterus*, 72
dendrophorus, *Rheoglanis*, 164
Dentata, *Loricaria*, 268
dentatus, *Ageneiosus*, 70
dentatus, *Auchenipterus*, 72
dentatus, *Doras*, 177
Dentectus, 235
Dentectus barbarmatus, 235
dentex, *Chaetostomus*, 277
dentex, *Panaqolus*, 277
dentex, *Panaque*, 277
Denticetopsis, 133
Denticetopsis epa, 133
Denticetopsis iwokrama, 133
Denticetopsis macilenta, 133
Denticetopsis praecox, 133
Denticetopsis royeroi, 133
Denticetopsis sauli, 133, 134
Denticetopsis seducta, 134
depauwi, *Synodontis*, 314
depierrei, *Platyglanis*, 164
depinnai, *Aspidoras*, 109
Deportator, 350
Deppei, *Pimelodus*, 200
depressa, *Harttia*, 240
depressa, *Hemiodon*, 293
depressa, *Reganella*, 293
depressa, *Rhamdia*, 201
depressa, *Tocantinsia*, 78
depressicauda, *Hisonotus*, 248
depressicauda, *Otocinclus*, 248
depressinotus, *Hisonotus*, 248
depressinotus, *Microlepidogaster*, 248
depressirostris, *Bagrus*, 364
depressirostris, *Eutropius*, 364
depressus, *Acanthodoras*, 167
depressus, *Bunocephalus*, 62
depressus, *Cataphractus*, 111
depressus, *Chaetostomus*, 287
depressus, *Clarias*, 143
depressus, *Doras (Rhinodoras)*, 167
depressus, *Gnathobagrus*, 161
depressus, *Hemiodon*, 292, 293
depressus, *Pseudancistrus*, 287
depressus, *Pterobunocephalus*, 62
dequinensis, *Glyptothorax*, 389
derbyi, *Hypostomus*, 253
derbyi, *Loricariichthys*, 270
derbyi, *Plecostomus*, 253
Dermocassis, 92
dermorhynchum, *Chaetostoma*, 228
dermorhynchus, *Chaetostomus*, 228
despaxi, *Arius*, 36
devall, *Hypophthalmus*, 334
devincenzi, *Loricaria*, 288
devincenzi, *Pseudohemiodon*, 288
deyi, *Amblyceps*, 17
dhonti, *Allabenchelys*, 142
dhonti, *Clarias*, 142
dhonti, *Synodontis*, 314
diabolicus, *Plectrochilus*, 411
diabolicus, *Urinophilus*, 411
diabolus, *Trichomycterus*, 417
dialonensis, *Clarias*, 142
dialonensis, *Clarias laeviceps*, 142
Dianema, 108, 127
Dianema longibarbis, 127
Dianema urostriatum, 127
Dianemidi, 108
Diapeltoplites, 249
diaphanus, *Ammoglanis*, 403
diaphina, *Ageniosus (Silonia)*, 86
Diastatomyste, 368
Diastatomyste Chaperi, 368
Diasternum, 127
dibrugarensis, *Macrones montanus*, 96
dichromum, *Aestridium*, 218
dicra, *Scolopla*, 367
Dieperinki, *Arius*, 32
difluviatilis, *Corydoras*, 116
digulensis, *Arius*, 47
 diluvii, *Bucklandium*, 432, 433
dinema, *Belodontichthys*, 368
dinema, *Wallago*, 368
Dinotopteroides, 139
Dinotopteroides prentissgrayi, 139, 146
Dinopterus, 148
Dinopterus atribranchus, 148
Dinopterus cunningtoni, 148
Dinopterus euryodon, 136
Dinopterus filicabarbis, 148
Dinopterus foveolatus, 149
Dinopterus gigas, 149
Dinopterus ilesi, 136
Dinopterus jacksoni, 149
Dinopterus jallae, 146
Dinopterus longibarbis, 136
Dinopterus loweae, 149
Dinopterus nyasensis, 149
Dinopterus rotundifrons, 136
Dinopterus worthingtoni, 136
diocles, *Arius*, 44
diocles, *Hemiarius*, 44
diphyes, *Corydoras*, 116
Diplomystax, 165
Diplomyste, 165
Diplomystes, 165
Diplomystes campensis, 165
Diplomystes chilensis, 165
Diplomystes mesembrinus, 166
Diplomystes nahuelbutaensis, 166
Diplomystes rudis, 435
Diplomystes viedmensis, 166
Diplomystes viedmensis cuyanus, 166
Diplomystes viedmensis mesembrinus, 166
Diplomystidae, 165
dipterus, *Brochus*, 111, 125
discus, *Aestridium*, 218
disjunctivus, *Liposarcus*, 291
disjunctivus, *Pterygoplichthys*, 291
disneyi, *Chiloglanis*, 305
dispar, *Arius*, 34
dispar, *Pygidium*, 417
dispar, *Trichomycterus*, 417
displurus, *Epapterus*, 74
dissidens, *Harttia*, 241
dissitus, *Kryptopterus*, 369
distolothrix, *Scolopla*, 367
djambal, *Pangasius* 325
djemer, *Eutropius*, 363
djemer, *Schilbe*, 363
dlouhyi, *Hypostomus*, 253
doceana, *Steindachneridion*, 343
doceanus, *Microlepidogaster*, 282
doceanus, *Parotocinclus*, 282
docmac niger, *Bagrus*, 84
docmak, *Bagrus*, 84, 313
docmak, *Silurus*, 84
Doiichthyidae, 30
Doiichthys, 30, 46
Doiichthys novaeguineae, 46
Doiichthys novae-guineae, 46
Dolichallabes, 149

- Dolichallabes microphthalmus*, 149
Dolichamphilius, 24
Dolichamphilius brieni, 24
Dolichamphilius longiceps, 24
Dolichancistrus, 235
Dolichancistrus atratoensis, 235
Dolichancistrus carnegiei, 235
Dolichancistrus cobrensis, 235
Dolichancistrus fuesslii, 235
Dolichancistrus pediculatus, 235
Dolichancistrus setosus, 235
dolichophia, *Scolopax*, 367
dolichopterus, *Ancistrus*, 220
dolichurus, *Bunocephalus*, 62
dolichurus, *Pterobunocephalus*, 62
dolloi, *Clarias*, 142
Doradidae, 166
Doradini, 166
dorae, *Leptoglanis*, 28
dorae, *Zaireichthys*, 28
Doraops, 170
Doraops zuloagai, 170
Doras, 166, 170
Doras (Agamyxis) flavopictus, 168
Doras (Corydoras) brevis, 179
Doras (Corydoras) ophthalmicus, 169
Doras (Corydoras) punctatus, 167
Doras (Oxydoras) d'Orbignyi, 178
Doras (Oxydoras) lipophthalmus, 169
Doras (Oxydoras) niger, 175
Doras (Oxydoras) stenopeltis, 171, 172
Doras (Rhinodoras) depressus, 167
Doras affinis, 168
Doras albomaculatus, 168
Doras armatus, 176
Doras asterifrons, 169, 170
Doras Blochii, 167
*Doras brachiatu*s, 170
Doras brunnescens, 168
Doras Calderonensis, 179
Doras carinatus, 171, 173
Doras castaneo-ventris, 168
Doras cataphractus, 168
Doras Crocodili, 170
Doras dentatus, 177
Doras dorsalis, 173
Doras fimbriatus, 167
Doras granulosus, 177
Doras grypus, 169
Doras Hancockii, 177
Doras Heckelii, 178
Doras helicophilus, 177
Doras humboldti, 176
Doras humeralis, 174
Doras insculptus, 179
Doras laevigatus, 177
Doras lentiginosus, 177
Doras libertatis, 174
Doras lithogaster, 173
Doras longispinis, 170
Doras loricatus, 167
Doras maculatus, 177
Doras marmoratus, 171
Doras micropoeus, 171
Doras murica, 177
Doras nebulosus, 178
Doras niger, 175
Doras Oxyrhynchus, 169
Doras papilionatus, 173
Doras pectinifrons, 168
Doras polygramma, 168
Doras punctatus, 167
Doras regani, 169
Doras spinosissimus, 168
Doras uranoscopus, 173, 174
Doras weddelli, 169
d'Orbigny, *Doras (Oxydoras)*, 178
d'Orbignyi, *Doras (Oxydoras)*, 178
dorbignyi, *Doras (Oxydoras)*, 178
dorbigny, *Rhinodoras*, 178
dorbignyi, *Rhinodoras*, 178
doriae, *Arius*, 41
doriae, *Bunocephalus*, 59
doriae, *Liocassis*, 93
doroides, *Bagrurus*, 45
dorsale, *Chaetostoma*, 228
dorsalis, *Chaetostomus*, 228
dorsalis, *Doras*, 173
dorsalis, *Glyptothorax*, 389
dorsalis, *Lithodoras*, 173
dorsalis, *Rhamdia*, 201
dorseyi, *Pimelodella*, 190
dorsimarmoratus, *Clarias*, 141
dorsomaculata, *Synodontis*, 314
dorsomaculatus, *Synodontis*, 314
dorsostriatum, *Pygidium*, 417
dorsotriatum, *Pygidium*, 417
dorsotriatus, *Trichomycterus*, 417
Doumea, 20, 24
Doumea alula, 24
Doumea angolensis, 25
Doumea chappuisi, 25
Doumea scaphyrhynchura, 27
Doumea thysi, 25
Doumea typica, 24, 25
Doumeinae, 20
dowii, *Leptarius*, 53
dowii, *Sciades*, 53
Duanensis, 378
duanensis, *Silurus*, 379
dubia, *Netuma*, 54
dubius, *Ancistrus*, 221
dubius, *Corydoras*, 126
duchaillui, *Clarias*, 141
duda, *Silurus*, 374
duellmani, *Trichomycterus*, 418
dugesii, *Ictalurus*, 208, 209
dugèsii, *Amiurus*, 208
dukai, *Silurus*, 377
dumerili, *Clariallabes*, 138
dumerili, *Leiocassis*, 93
dumerili, *Rhinobagrus*, 92, 93
dumerili longibarbis, *Clarias (Allabenchelys)*, 138
dumerili, *Clarias*, 142
dumus, *Lasiancistrus*, 289
dumus, *Pseudolithoxus*, 289
dunni, *Centromochlus*, 77
dunni, *Tatia*, 77
duodecimalis, *Hypostomus*, 290, 291
Duopalatinus, 332
Duopalatinus emarginatus, 332
Duopalatinus olallae, 340
Duopalatinus peruanus, 332
Duoplatinus goeldii, 340
duplicareus, *Corydoras*, 116
dupouii, *Chaetostoma*, 228
Dupouyichthys, 60
Dupouyichthys sapito, 60
duquei, *Rhamdia*, 202
dura, *Loricaria*, 268
duriceps, *Ostophycephalus*, 346
durinii, *Eutropius*, 363
durinii, *Schilbe*, 363
duriventris, *Hartia*, 241
duseni, *Hemipsilichthys*, 262, 279
duseni, *Isbrueckerichthys*, 262
Dussumieri, *Arius*, 52
dussumieri, *Clarias*, 142
dussumieri, *Plicofollis*, 52
Dutemplei, *Arius*, 56
duttoni, *Chrysichthys*, 156
dybowskii, *Chiloglanis*, 309
dybowskii, *Euchilichthys*, 309
Dysichthys, 58
Dysichthys amazonicus, 59
Dysichthys australe, 60
Dysichthys coracoideus, 58, 59
Dysichthys quadriradiatus, 60
E
eaglesomei, *Eaglesomia*, 160
Eaglesomia, 160
Eaglesomia eaglesomei, 160
ebriensis, *Clarias*, 142
eburnensis, *Synodontis*, 312
echinatus, *Ictalurus*, 208
echthrus, *Megalocentor*, 409
eclipsis, *Eutropius*, 363
eddsi, *Pseudecheneis*, 442
edentatus, *Corydoras*, 176
edentatus, *Hypophthalmus*, 333
edentatus, *Loricariichthys*, 270
egertoni, *Arius*, 56
Egertoni, *Silurus*, 56
egertoni belgicus, *Arius*, 56
egertonii, *Callichrous*, 374
ehrhardtii, *Corydoras*, 116

- eichorniarum, Ituglanis*, 407
eichorniarum, Trichomycterus, 407
Eidouxii, Galeichthys, 38
eigenmanni, Arges, 64
eigenmanni, Astroblepus, 64
eigenmanni, Chaenothorax, 125
eigenmanni, Corydoras, 129
eigenmanni, Galeichthys, 33
eigenmanni, Heptapterus, 184
eigenmanni, Hemiloricaria, 245
Eigenmanni, Loricaria, 245
eigenmanni, Orinocodoras, 175
eigenmanni, Oxydoras, 167
eigenmanni, Pimelodella, 191, 340
eigenmanni, Pimelodella, 193
eigenmanni, Pimelodus, 340
eigenmanni, Pimelodus (Pimelodella), 191
eigenmanni, Propimelodus, 340
eigenmanni, Pygidianops, 412
eigenmanni, Trichomycterus, 423
eigenmanniorum, Pimelodella, 191
eigenmanniorum, Rhamdia, 191
ekibondoi, Clarias, 147
elassochir, Noturus, 212
elatturus, Arius, 50
electricus, Malapterurus, 301
electricus, Silurus, 301
electricus ogooensis, Malapterurus, 302
electricus oguensis, Malapterurus, 302
elegans, Corydoras, 111, 116
elegans, Noturus, 212
elegans njsseni, Corydoras, 121
eleutherus, Noturus, 212
elisabethianus, Chiloglanis, 305
Ellenriederii, Leiocassis, 104
Elliops, 207
ellisae, Corydoras, 116
Ellisichthys, 127
elongata, Eumeda, 347, 348
elongata, Hara, 384
elongata, Olyra, 99
elongata, Pimelodella, 191
elongatus, Batasio, 73
elongatus, Macrones, 89
elongatus, Nemadoras, 174
elongatus, Oxydoras, 174
elongatus, Pangasius, 326
elongatus, Pimelodus, 191
elongatus, Plotosus, 346
elongatus, Sorubim, 342
elongatus hongus, Hemibagrus, 89
emanueli, Trichomycterus, 418
emanueli emanueli, Pygidium, 418
emanueli motatanensis, Pygidium, 421
emarginata, Microsynodontis, 309
emarginata, Squaliforma, 297
emarginatum, Platystoma, 332
emarginatus, Chiloglanis, 305
emarginatus, Duopalatinus, 332
emarginatus, Hypostomus, 297
emarginatus, Liocassis, 102
emarginatus, Microsynodontis, 309
emarginatus, Pseudobagrus, 100
Emini, Schilbe, 364
emmeline, Tachysurus, 40
emphysetus, Bagrus (Sciades), 53, 54
empousa, Scolopax, 367
Encheloclarias, 149
Encheloclarias baculum, 149
Encheloclarias curtisoma, 149
Encheloclarias keliooides, 149
Encheloclarias prolatus, 150
Encheloclarias tapeinopterus, 150
Encheloclarias velatus, 150
Endorrhis, 349
enfurnada, Rhamdia, 197
engelseni, Allabenchelys, 142
engelseni, Clarias, 142
engiops, Chiloglanis, 308
enigmatica, Lacantunia, 216
enochi, Pimelodella, 191
Entomocorus, 74
Entomocorus benjamini, 74
Entomocorus gameroi, 74
Entomocorus melaphareus, 74
Entomocorus radiosus, 443
Eobagrus, 87
Eobagrus hoerdzanicus, 87
Eomacrones, 87
Eomacrones wilsoni, 87
Eopeyeria, 428
Eopeyeria aegyptiaca, 428
epa, Denticetopsis, 133
Epactionotus, 236
Epactionotus aky, 236
Epactionotus bilineatus, 236
Epactionotus gracilis, 236
Epactionotus itaimbezinho, 236
Epactionotus yasi, 236
epakmos, Isbrueckerichthys, 262
Epapterus, 74
Epapterus blohmi, 74
Epapterus chaquensis, 74
Epapterus dispilurus, 74
ephippiata, Amphilus pulcher, 23
ephippifer, Akysis, 14
ephippifer, Corydoras, 116
epikarsticus, Ituglanis, 407
episcopi, Corydoras, 126
eppleyi, Parotocinclus, 282
eptingi, Chaetostomus, 253
eptingi, Hypostomus, 253
equatorialis, Tachysurus, 40
eques, Cheirocerus, 332
eques, Corydoras, 112, 116
eques, Goeldiella, 184
eques, Pimelodus, 184
equestris, Arius, 33
equinus, Copidoglanis, 348
equinus, Neosilurus, 348
erebennus, Amiurus, 205
Eremophilini, 403
Eremophilus, 403, 404
Eremophilus camposi, 408
Eremophilus candidus, 416
Eremophilus mutisii, 404
Erethistes, 382, 384, 444
Erethistes maesotensis, 384
Erethistes pusillus, 384
Erethistes pussilus, 384
Erethistidae, 382
Erethistides, 382
Erethistoides, 382
Erethistoides ascita, 384
Erethistoides cavatura, 384
Erethistoides infuscatus, 442
Erethistoides montana, 384
Erethistoides montana pipri, 384
Erethistoides pipri, 384
Erethistoides sicula, 384
eriarcha, Rhamdella, 196
eriarcha, Rhamdia, 196
ericae, Hypostomus, 253
ericius, Hypostomus, 253
erinaceus, Ancistrus, 221
erinaceus, Hypostomus, 221
Ernstichthys, 60
Ernstichthys anduzei, 60
Ernstichthys intonsus, 60
Ernstichthys megistus, 60
erythrogaster, Silurus (Callichrus), 373
Erythroptera, Silurus maculatus, 209
erythrurus, Urinophilus, 427
esamesae, Clarias, 146
esmeraldas, Paracetopsis, 135
esperanzae, Corydoras, 117
espiritosantensis, Neoplecostomus, 273
essequibensis, Leptoglanis, 187
essequibensis, Leptorhamdia, 187
estuarius, Ayarnangra, 382
etentaculatum, Hypostoma, 291
etentaculatus, Pterygoplichthys, 291
Euacanthagenys, 296
Euacanthagenys caquetae, 296
euacanthagenys, Spatularicaria, 296
Euanemini, 68
Euanemus, 68, 71
Euanemus brachyurus, 72
Euanemus colymbetes, 71, 72
Euanemus longipinnis, 74
Euanemus nigripinnis, 72
Euchilichthys, 308
Euchilichthys astatodon, 308

- Euchilichthys boulengeri*, 309
Euchilichthys dybowskii, 309
Euchilichthys guentheri, 309
Euchilichthys habereri, 309
Euchilichthys royauxi, 309
Euchiloglanis, 384
Euchiloglanis davidi, 385
Euchiloglanis gracilicaudata, 398
Euchiloglanis hodgarti, 398
Euchiloglanis kamenensis, 399
Euchiloglanis kishinouyei, 385
Euchiloglanis longicauda, 399
Euchiloglanis macrotrema, 399
Euchiloglanis myzostoma, 399
Euchiloglanis sinensis, 399
Euclyptosternum, 387
eugeneiatus, *Callichrous*, 369
eugeneiatus, *Kryptopterus*, 369
Euglyptosternum lineatum, 388
Eumeda, 347
Eumeda elongata, 347, 348
eupogoides, *Pseudobagrus*, 107
eupogon, *Clarias*, 153
eupogon, *Pelteobagrus*, 100
eupogon, *Pseudobagrus*, 100
eupogon, *Xenoclarias*, 153
euptera, *Synodontis*, 314
eupterus, *Synodontis*, 314
Euristhmus, 346
Euristhmus lepturus, 346
Euristhmus microceps, 346
Euristhmus microphthalmus, 442
Euristhmus nudiceps, 346
Euristhmus sandrae, 442
eurycephalus, *Aspidoras*, 109
eurycephalus, *Hemipsilichthys*, 279
eurycephalus, *Pareiorhaphis*, 279
Eurycheilichthys, 236
Eurycheilichthys limulus, 236
Eurycheilichthys pantherinus, 236
Eurycheilus, 236
Eurycheilus pantherinus, 236
euryodon, *Bathyclarias*, 136
euryodon, *Dinotopterus*, 136
eurystoma, *Microglanis*, 354
eurystomus, *Satan*, 216
eurystomus, *Synodontis*, 305, 319
eustictus, *Ancistrus*, 221
eustictus, *Pristiancistrus*, 219, 221
eutaenia, *Pimelodella*, 191
Eutropiellus, 360
Eutropiellus buffei, 361
Eutropiellus kasai, 360, 361
Eutropiellus vandeweyeri, 361
Eutropiichthys, 358
Eutropiichthys burmannicus, 358
Eutropiichthys murius, 358
Eutropiichthys vacha, 358
Eutropius, 362
Eutropius altipinnis, 365
Eutropius angolensis, 363
Eutropius ansorgii, 363
Eutropius banguelensis, 363
Eutropius bocagii, 363
Eutropius bomae, 363
Eutropius brachyopterus, 362
Eutropius brevianalis, 363
Eutropius cameronensis, 365
Eutropius congolensis, 363, 364
Eutropius debauwi, 361
Eutropius depressirostris, 364
Eutropius djemeru, 363
Eutropius durinii, 363
Eutropius eclipsis, 363
Eutropius gastratus, 363
Eutropius grenfelli, 363
Eutropius laticeps, 364
Eutropius lemairii, 364
Eutropius liberiensis, 364
Eutropius longifilis, 361
Eutropius macrophthalmos, 361, 362
Eutropius mandibularis, 364
Eutropius mentalis, 364
Eutropius micropogon, 364
Eutropius möbiusii, 364, 365
Eutropius moebii, 365
Eutropius multilineatus, 365
Eutropius multitaeniatus, 365
Eutropius nasalis, 363
Eutropius nyongensis, 365
Eutropius obtusirostris, 365
Eutropius seraui, 363
Eutropius tumbanus, 365
Eutropius yangambianus, 365
Evansii, *Loricaria*, 296
evansii, *Spatuloricaria*, 296
evelynae, *Corydoras*, 117
evermanni, *Tachysurus*, 40
Exallodontus, 333
Exallodontus aguanai, 333
exaratus, *Callichthys*, 129
Exastilithoxus, 236
Exastilithoxus fimbriatus, 237
Exastilithoxus hoedemani, 237
Exilichthys, 346
exilis, *Noturus*, 212
exilis, *Trachelyichthys*, 78
existimatus, *Centromochlus*, 73
exodon, *Bagrus*, 359
exodon, *Glyptothorax*, 389
Exostoma, 382, 385
Exostoma andersonii, 386
Exostoma berdmorei, 385
Exostoma Blythii, 396
Exostoma Feae, 398
Exostoma gracile, 387
Exostoma labiatum, 385
Exostoma labrax, 387
Exostoma macropterum, 398
Exostoma Oschanini, 402
Exostoma stoliczkae, 387
Exostoma Vinciguerrae, 385
Exostomatina, 382
exsudans, *Pimelodus*, 196
exsudans, *Rhamdella*, 196
Eydouxii, *Galeichthys*, 38
F
fabricioi, *Callichthys*, 111
Fajumia, 428
Fajumia menoni, 429
Fajumia misrai, 429
Fajumia schweinfurthi, 428, 429
Fajumia stromeri, 429
falcarius, *Arius*, 34
falcarius, *Mystus*, 95
falcarius africana, *Arius*, 31
falcarius var. *africana*, *Arius*, 31
falcifer, *Acrochordonichthys*, 12
falconeri, *Clarias*, 143
fallax, *Hemiloricaria*, 245
fallax, *Loricaria* (*Loricariichthys*), 245
fangi, *Arius*, 34
fangi, *Pseudobagrus*, 101
Farlowella, 217, 237
Farlowella acestrichthys, 238
Farlowella acus, 237
Farlowella acus venezuelensis, 239
Farlowella agustini, 240
Farlowella altocorpus, 444
Farlowella amazona, 237
Farlowella amazonum, 237
Farlowella angosturae, 240
Farlowella azygia, 238
Farlowella boliviiana, 238
Farlowella carinata, 237
Farlowella colombiensis, 237
Farlowella curtirostra, 237
Farlowella gladiolus, 237
Farlowella gracilis, 237
Farlowella guaricensis, 240
Farlowella hahni, 238
Farlowella hargreavesi, 238
Farlowella hasemani, 238
Farlowella henriquei, 238
Farlowella isbruckeri, 238
Farlowella jauruensis, 238
Farlowella jauruensis, 238
Farlowella knerii, 238
Farlowella latisoma, 239
Farlowella mariae, 238
Farlowella martini, 238
Farlowella nattereri, 238
Farlowella odontotumulus, 238
Farlowella oliveirae, 237
Farlowella oxyrryncha, 239
Farlowella paraguayensis, 239
Farlowella paranaense, 237

Farlowella parvicarinata, 239
Farlowella platyynchus, 239
Farlowella pleurotaenia, 237
Farlowella pseudoglaadiolus, 237
Farlowella reticulata, 239
Farlowella roncallii, 240
Farlowella rugosa, 239
Farlowella schreitmüllerii, 239
Farlowella schreitmüllerii, 239
Farlowella smithi, 239
Farlowella taphorni, 239
Farlowella venezuelensis, 239
Farlowella vittata, 239
Farlowellidi, 217
farsonensis, *Hypsidoris*, 203
fasciata, *Phractura*, 26
fasciata, *Schilbe senegalensis*, 364
fasciatum, *Pseudoplatystoma*, 341
fasciatum brevifile, *Pseudoplatystoma*, 341
fasciatum intermedium,
Pseudoplatystoma, 341
fasciatum nigricans,
Pseudoplatystoma, 341
fasciatum reticulatum,
Pseudoplatystoma, 341
fasciatus, *Auchenoglanis*, 163
fasciatus, *Chiloglanis*, 306
fasciatus, *Nannoglanis*, 187, 188
fasciatus, *Noturus*, 212
fasciatus, *Silurus*, 340, 341
fasciolatus, *Batasio*, 442
fascipinna, *Synodontis*, 314
fassli, *Pygidium*, 418
fassli, *Trichomycterus*, 418
fasslii, *Pygidium*, 418
feae, *Exostoma*, 398
feae, *Pareuchiloglanis*, 398
feliceps, *Galeichthys*, 43
Felichthys, 37
Felichthys amblops, 79
Felichthys filamentosus, 37
Felichthys flavescens, 76
Felichthys stauroforus, 32
felinus, *Pimelodus*, 205
felipponei, *Loricaria*, 294
felipponei, *Rineloricaria*, 294
felis, *Ariopsis*, 33
felis, *Pimelodus*, 206
felis, *Silurus*, 33
ferox, *Silurus*, 431
ferrarii, *Niobichthys*, 274
ferreira, *Cetopsidium*, 130
ferula, *Pseudolaguvia*, 442
festae, *Arges*, 64
festae, *Arius*, 56
festae, *Astroblepus*, 64
festae, *Plecostomus*, 263
festinus, *Arius*, 34

festivum, *Sturisoma*, 299
figueroai, *Pimelodella*, 191
filamentissima, *Harttia*, 264
filamentosa, *Cetopsorhamdia*, 182
filamentosa, *Dasyloricaria*, 233
filamentosa, *Hara*, 395
filamentosa, *Harttia*, 263, 264
filamentosa, *Loricaria*, 233
filamentosa, *Synodontis*, 314
filamentosa latiura, *Loricaria*, 233
filamentosa seminuda, *Loricaria*, 234
filamentosum, *Brachyplatystoma*, 330
filamentosus, *Aspredinichthys*, 57
filamentosus, *Aspredo*, 57
filamentosus, *Bagrus*, 84
filamentosus, *Chrysichthys*, 155
filamentosus, *Corydoras*, 117
filamentosus, *Felichthys*, 37
filamentosus, *Lamontichthys*, 263
filamentosus, *Pimelodes*, 330
filamentosus, *Synodontis*, 314
filamentus, *Hemibagrus*, 88
filamentus, *Macrones (Hemibagrus)*, 88
filicaudata, *Peckoltia*, 284
filicaudatus, *Peckoltichthys*, 283, 284
filicarbis, *Bathyclarias*, 148
filicarbis, *Dinotopterus*, 148
filifer, *Akysis*, 16
filifer, *Pseudobagarius*, 16
filinemus, *Phyllonemus*, 164
fimbriata, *Cetopsis*, 132
fimbriata, *Loricaria*, 297
fimbriata, *Spatuloricaria*, 297
fimbriatus, *Doras*, 167
fimbriatus, *Exastilithoxus*, 237
fimbriatus, *Hypophthalmus*, 333
fimbriatus, *Otocinclus*, 271
fimbriatus, *Pseudacanthicus*
(Lithoxus), 236, 237
fimbriatus, *Pseudocanthicus*
(Lithoxus), 237
firestonei, *Paramphilius*, 25
fisadoha, *Plotosus*, 350
fischeri, *Chaetostoma*, 228
Fischeri, *Chaetostomus*, 228
fisheri, *Trachelyopterus*, 79
fisheri, *Trachycorystes*, 79
fissidens, *Arges*, 64
fissidens, *Astroblepus*, 64
fissipinnis, *Heptapterus*, 184
fissus, *Arius*, 39
flabelliferus, *Ochmacanthus*, 409
flagellaris, *Plecostomus*, 268
flava, *Loricaria*, 259
flavater, *Noturus*, 212
flaveolus, *Corydoras*, 117
flavescens, *Bagrus*, 54
flavescens, *Felichthys*, 76

flavescens, *Pseudauchenipterus*, 76
flavicans, *Bagrus*, 344
flavipinnis, *Noturus*, 212
flavipinnis, *Pimelodina*, 336
flavipinnis, *Pimelodus (Pimelodina)*, 336
flavipinnis, *Pseudomystus*, 103
flavitaeniata, *Synodontis*, 314
flavitaeniatus, *Synodontis*, 314
flavolineatus, *Plotosus*, 345
flavomaculatus, *Leptoglanis*, 28
flavomaculatus, *Zaireichthys*, 28
flavopictus, *Doras (Agamyxis)*, 168
flavus, *Bagrus*, 90
flavus, *Hyalobagrus*, 92
flavus, *Noturus*, 211, 212
flexilis, *Macrotocinclus*, 271
flexilis, *Otocinclus*, 271, 272
floreense, *Pygidium*, 414
fluviatilis, *Hypostomus*, 253
fluviatilis, *Leiocassis*, 85
fluviatilis, *Mystus*, 164
fluviatilis, *Plecostomus*, 253
Fluvidraco, 101
foina, *Pimelodus*, 197
foina, *Rhamdia*, 197
fokiensis, *Glyptosternum*, 389
fokiensis, *Glyptothorax*, 389
fonchii, *Hypostomus*, 253
Fonchiichthys, 240
Fonchiichthys rupestris, 240
Fonchiichthys uracanthus, 240
foratum, *Amblyceps*, 18
fordicei, *Auchenipterus*, 72
Fordii, *Chaetostomus*, 286
fordii, *Pseudacanthicus*, 286
forficulatus, *Hypodoras*, 172
formosa, *Hemiloricaria*, 245
formosa, *Rineloricaria*, 245
formosanus, *Liobagrus*, 19
formoso, *Ancistrus*, 221
formosus, *Astroblepus*, 64
fortis, *Hemibagrus*, 88
fortis, *Macrones*, 88
fortis capitulum, *Macrones*, 88
fortis var. *capitulum*, *Macrones*, 88
fossilis, *Heteropneustes*, 151
fossilis, *Silurus*, 151
fossor, *Pimelodes*, 431
Fouloni, *Clarias*, 148
foveolata, *Pseudolaguvia*, 401
foveolatus, *Clarias*, 149
foveolatus, *Dinotopterus*, 149
fowleri, *Cephalosilurus*, 353
fowleri, *Corydoras*, 117
fowleri, *Cteniloricaria*, 233
Fowleri, *Oxyloricaria*, 233
fowleri, *Pangasius*, 328
Fraasi, *Arius*, 56

- francirochai, Hisonotus*, 248
francirochai, Otocinclus, 248
francisci, Hypostomus, 254
francisci, Plecostomus, 254
Franciscodoras, 171
Franciscodoras marmoratus, 171
franciscoensis, Neoplecostomus, 273
frankei, Aphanotorulus, 225, 226
freiei, Ageneiosus, 70
frenata, Brachyglanis, 180
frenata, Loricaria, 299
frenatum, Sturisoma, 299
frenatus, Astroblepus, 64
frenatus, Brachyglanis, 180
frenatus, Oreoglanis, 397
frenatus, Rhamdioglanis, 202
froggatti, Arius, 42
froggatti, Cinetodus, 42
frontosa, Synodontis, 314
frontosus, Synodontis, 314
fuelleborni, Synodontis, 315
fuerthii, Cathorops, 39
fuesslii, Dolichancistrus, 235
fugleri, Hemiancistrus, 243
fui, Pseudobagrus, 107
fukiensis, Glyptothorax, 389
fukiensis fukiensis, Glyptothorax, 389
fukiensis hainanensis, Glyptothorax, 390
fukiensis honghensis, Glyptothorax, 390
fukiensis honghenensis, Glyptothorax, 390
fuliginatus, Akysis, 14
fuliginosus, Hemiancistrus, 243
fulva, Xenocara, 221
fulvidraco, Pelteobagrus, 100
fulvi-draco, Pimelodus, 100
fulvus, Ancistrus, 221
fumidus, Ompok, 372
fumosus, Pseudomystus, 103
funebris, Noturus, 212
funnelli, Corydoras, 119
fur, Pimelodus, 337
fur, Pseudorhamdia, 337
furcata, Peckoltia, 284
furcatus, Auchenipterus, 76
furcatus, Chaetostomus, 284
furcatus, Chrysichthys, 156
furcatus, Hemibagrus, 88
furcatus, Ictalurus, 208, 211
furcatus, Nemasiluroides, 361
furcatus, Pimelodus, 208, 209
furcifer, Pimelodus, 210
Furcodontichthys, 240
Furcodontichthys novaesi, 240
furious, Noturus, 211, 212
furnessi, Apodoglanis, 375, 376
furnessi, Pterocryptis, 376
- Fürthii, Arius*, 39
Fuscatus, Silurus lividus, 205
fuscoguttatus, Aspidoras, 109
fuscum, Pygidium, 427
fuscus, Akysis, 16
fuscus, Ancharius, 29
fuscus, Bagrus, 95
fuscus, Clarias, 142
fuscus, Glyptothorax, 389
fuscus, Leiocassis, 103
fuscus, Macropterodonotus, 142
fuscus, Neobagrus, 19, 20
fuscus, Pseudobagarius, 16
fuscus, Pseudomystus, 103
Fusiloricaria, 268
füsslii, Ancistrus, 235
- G**
- gabardinii, Ageneiosus*, 69
gabonensis, Clarias, 142
gabrieli, Pygidium, 418
gabrieli, Trichomycterus, 418
gadense, Nigerium, 98
Gagata, 385
Gagata cenia, 386
Gagata dolichonema, 386
Gagata gagata, 386
Gagata gasawayuh, 386
Gagata itchkeea, 386
Gagata melanoptera, 386
Gagata melanopterus, 386
Gagata pakistanica, 386
Gagata sexualis, 386
Gagata typus, 386
Gagata youssoufi, 386
gagata, Gagata, 385, 386
gagora, Arius, 34
gagora, Pimelodus, 34
gagorides, Bagrus, 35
gagoroides, Arius, 35
galani, Ancistrus, 221
galaxias, Leporacanthicus, 265
galaxias, Tatia, 77
galeatus, Silurus, 79
galeatus, Trachelyopterus, 79
Galeichthys, 43
Galeichthys aequus, 435
Galeichthys aguadulce, 39
Galeichthys angelicus, 434
Galeichthys araguayensis, 339
Galeichthys ater, 43
Galeichthys azureus, 33
Galeichthys bahiensis, 38
Galeichthys Blochii, 38
Galeichthys bonillai, 33
Galeichthys eigenmanni, 33
Galeichthys Eidouxii, 38
Galeichthys Eydouxii, 38
Galeichthys feliceps, 43
Galeichthys gilberti, 33
- Galeichthys Gronovii*, 37
Galeichthys guentheri, 33
Galeichthys ocellatus, 43
Galeichthys Parrae, 38
Galeichthys peruvianus, 43
Galeichthys simonsi, 33
Galeichthys stanneus, 49
Galeichthys xenauchen, 33
galinae, Synodontis, 323
gallowayi, Schilbeodes, 212
gambensis, Pimelodus, 54
gambiensis, Synodontis, 315
gambiensis latifrons, Synodontis, 315
gameroi, Entomocorus, 74
gangelica, Pterocryptis, 375, 376, 377
gangetica, Silundia, 366
Gangra, 395
garavelloii, Harttia, 241
garavelloii, Microglanis, 354
garbei, Corydoras, 117
garbei, Hemipsilichthys, 279
garbei, Pareiorhaphis, 279
garbei, Pimelodela, 189
garciabarrigai, Pimelodus, 337
garcia-barrigai, Pimelodus, 337
garhwali, Glyptothorax, 389
gariepinus, Clarias, 140, 143, 148
gariepinus, Silurus (Heterobranchus), 143
- garmani, Hypostomus*, 254
garmani, Plecostomus, 254
garua, Clupisoma, 357
garua, Silurus, 357
gasawayuh, Gagata, 386
gastratus, Eutropius, 363
Gastrodermus, 111
gaudryi, Silurus, 432
Gelanoglanis, 73, 74
Gelanoglanis nanonocticolus, 74
Gelanoglanis stroudi, 74
gelatinosus, Malacoglanis, 408
geledensis, Synodontis, 315
geminus, Kryptopterus, 369
genibarbis, Pseudorinelepis, 289
genibarbis, Rinelepis, 289
Genidens, 32, 43
Genidens barbus, 44
Genidens cuvieri, 44
Genidens genidens, 44
Genidens granulosus, 44
Genidens machadoi, 44
Genidens planifrons, 44
Genidens valenciennesii, 44
genidens, Genidens, 44
genidens, Pimelodus, 43, 44
genisetiger, Pseudancistrus, 287
geoffroy, Corydoras, 111, 117
Geoffroyi, Heterobranchus, 150
Gephyroglanidini, 153

- Gephyroglanis*, 153, 160
Gephyroglanis barnardi, 81
Gephyroglanis congicus, 160, 161
Gephyroglanis gigas, 157
Gephyroglanis gilli, 81
Gephyroglanis gymnorhynchus, 161
Gephyroglanis habereri, 161
Gephyroglanis longipinnis, 157
Gephyroglanis lowei, 158
Gephyroglanis ogoensis, 158
Gephyroglanis ogooensis, 158
Gephyroglanis rotundiceps, 28
Gephyroglanis sclateri, 81
Gephyroglanis tilhoi, 155
Gephyroglanis velifer, 157
Gephyromochlus, 75
germanicus, *Otolithus* (*Arius*), 434
geryi, *Corydoras*, 117
geryi, *Pimelodella*, 191
gibbiceps, *Ancistrus*, 291
gibbiceps, *Pterygoplichthys*, 291
gibbiceps, *Silurichthys*, 377
gibbosa, *Lampiella*, 264
gibbosus, *Chaetostomus*, 278
gibbosus, *Otocinclus*, 264
gigantea, *Vandellia*, 427
giganteus, *Trichomycterus*, 418
gigas, *Arius*, 31, 56
gigas, *Bagarius*, 383
gigas, *Bathyclarias*, 149
gigas, *Chaetostomus*, 272
gigas, *Dinotopterus*, 149
gigas, *Gephyroglanis*, 157
gigas, *Pangasianodon*, 325
gigas, *Platystoma*, 330
gigas, *Xiurenbagrus*, 20
gilberti, *Galeichthys*, 33
gilberti, *Noturus*, 213
gilberti, *Silurus*, 376
gilli, *Astroglanis*, 81
gilli, *Clarias*, 146
gilli, *Gephyroglanis*, 81
gilli, *Rhamdella*, 196
gilli, *Rhamdia*, 196
Ginesia, 330
Ginesia cunaguardo, 330, 331
gjellerupi, *Copidoglanis*, 348
gjellerupi, *Neosilurus*, 348
glaber, *Auchenipterus*, 80
glaber, *Conorhynchus*, 428
glaber, *Otolithus* (*Arius* ?), 435
gladiator, *Noturus*, 213
Gladioglanis, 183
Gladioglanis conquistador, 183
Gladioglanis machadoi, 183
gladiolus, *Acestra*, 237
gladius, *Acestra*, 239
gladysae, *Phractura*, 26
Glanapteryxinae, 403
- Glanapteryx*, 403, 404
Glanapteryx anguilla, 404
Glanapteryx niobium, 405
Glanidium, 75
Glanidium albescens, 75
Glanidium bockmanni, 75
Glanidium catharinensis, 75
Glanidium cesarpintoi, 75
Glanidium leopardum, 75
Glanidium melanopterum, 75
Glanidium neivai, 77
Glanidium piresi, 78
Glanidium ribeiroi, 75
Glanis, 37, 378
Glanis Aristotelis, 378
glanis aralensis, *Silurus*, 379
glanis, *Silurus*, 378, 379
Glyaphropoma, 405
Glyaphropoma rodriguesi, 405
Glaridoglanis, 386
Glaridoglanis andersonii, 386
glencoensis, *Copidoglanis*, 348
gloveri, *Neosilurus*, 348
Glyptocephalus, 432
Glyptocephalus radiatus, 432, 433
Glyptoperichthys, 290
Glyptoperichthys joselimaianus, 291
Glyptoperichthys parnaibae, 292
Glyptoperichthys xinguensis, 292
Glyptosterni, 382
Glyptosternon, 382, 387
Glyptosternon akhtari, 387
Glyptosternon hainanensis, 390
Glyptosternon interspinatum, 390
Glyptosternon labiatum, 385
Glyptosternon maculatum, 387
Glyptosternon malaisei, 387
Glyptosternon minutum, 393
Glyptosternon Nieuwenhuisi, 392
Glyptosternon pectinopterus, 392
Glyptosternon punctatum, 389
Glyptosternon quadriocellatum, 393
Glyptosternon reticulatum, 387
Glyptosternon reticulatus, 387
Glyptosternon reticulatus, 387
Glyptosternon striatus, 387, 394
Glyptosternon sulcatus, 400
Glyptosternum, 387
Glyptosternum akhtari, 387
Glyptosternum armeniacum, 388
Glyptosternum chaudhurii, 385
Glyptosternum conirostre, 389
Glyptosternum dekkanense, 391
Glyptosternum fokiensis, 389
Glyptosternum gracile, 390
Glyptosternum hodgarti, 398
Glyptosternum kükenthali, 391
Glyptosternum kurdistanicum, 390
Glyptosternum laak, 395
- Glyptosternum madraspatanum*, 391
Glyptosternum modestum, 389
Glyptosternum pallozonum, 388, 392
Glyptosternum saisi, 393
Glyptosternum sinense, 393
Glyptosternum Stoličkae, 393
Glyptosternum steindachneri, 393
Glyptosternum stuarti, 385
Glyptosternum sykesi, 394
Glyptosternum tiong, 395
Glyptosternum yunnanensis, 401
Glyptocephalini, 382
Glyptothorax, 382, 387
Glyptothorax alaknandi, 388
Glyptothorax anamalaiensis, 388
Glyptothorax annandalei, 388
Glyptothorax armeniacus, 388
Glyptothorax botius, 388
Glyptothorax brevipinnis, 388
Glyptothorax brevipinnis alaknandi, 388
Glyptothorax buchanani, 388
Glyptothorax burmanicus, 388
Glyptothorax callopterus, 388
Glyptothorax cavia, 388
Glyptothorax coheni, 388, 389
Glyptothorax conirostre poonaensis, 395
Glyptothorax conirostris, 389, 395
Glyptothorax conirostris punjabensis, 393
Glyptothorax couesi, 31, 389
Glyptothorax dakpathari, 389
Glyptothorax davissinghi, 389
Glyptothorax deqinensis, 389
Glyptothorax dorsalis, 389
Glyptothorax exodon, 389
Glyptothorax fokiensis, 389
Glyptothorax fukiensis fukiensis, 389
Glyptothorax fukiensis hainanensis, 390
Glyptothorax fukiensis honghensis, 390
Glyptothorax fukiensis honghenensis, 390
Glyptothorax fuscus, 389
Glyptothorax garhwali, 389
Glyptothorax gracilis, 389, 390
Glyptothorax hainanensis, 390
Glyptothorax honghensis, 390
Glyptothorax horai, 390
Glyptothorax housei, 390
Glyptothorax indicus, 390
Glyptothorax interspinatus, 390
Glyptothorax jalalensis, 390
Glyptothorax kashmirensis, 390, 393
Glyptothorax kurdistanicus, 390
Glyptothorax lampris, 391
Glyptothorax laosensis, 391

- Glyptothorax lonah*, 391, 394
Glyptothorax longicauda, 391
Glyptothorax longinema, 394
Glyptothorax longjiangensis, 391
Glyptothorax macromaculatus, 391
Glyptothorax madraspatanus, 391
Glyptothorax major, 391
Glyptothorax manipurensis, 391
Glyptothorax merus, 390
Glyptothorax minimaculatus, 391
Glyptothorax minutus, 391, 392, 393
Glyptothorax naziri, 392
Glyptothorax nelsoni, 392
Glyptothorax nieuwenhuisi, 392
Glyptothorax obscura, 392
Glyptothorax obscurus, 392
Glyptothorax pallozonus, 392
Glyptothorax panda, 392
Glyptothorax pectinopterus, 392
Glyptothorax platypogon, 392
Glyptothorax platypogonides, 392
Glyptothorax platypogonoides, 392
Glyptothorax prashadi, 393
Glyptothorax punjabensis, 393
Glyptothorax quadriocellatus, 393
Glyptothorax rubermentus, 394
Glyptothorax saisi, 393
Glyptothorax siamensis, 393
Glyptothorax silviae, 393
Glyptothorax sinensis, 393
Glyptothorax spectrum, 393
Glyptothorax steindachneri, 393
Glyptothorax stocki, 393
Glyptothorax stolickae, 393
Glyptothorax striatus, 394
Glyptothorax sufii, 394
Glyptothorax sykesi, 394
Glyptothorax telchitta, 394
Glyptothorax telchitta sufii, 394
Glyptothorax trewavasae, 394
Glyptothorax trilineatoides, 394
Glyptothorax trilineatus, 394
Glyptothorax tuberculatus, 401, 402
Glyptothorax ventrolineatus, 394
Glyptothorax zainaensis, 394
Glyptothorax zanaensis, 394
Glyptothorax zhuijangensis, 394
Gnathobagrus, 161
Gnathobagrus depressus, 161
gnomus, *Panaqolus*, 277
gnomus, *Panaque*, 277
goae, *Silurus*, 374
Gobibagrus, 87
Gobibagrus hoerdzanicus, 87
gobio, *Hemipsilichthys*, 247
gobio, *Xenomystus*, 247
gobiooides, *Cetopsis*, 131, 132
gobroni, *Synodontis*, 315
godavarii, *Pangasius pangasius*, 327
godfreyi, *Tachysurus (Pararius)*, 45
godmanni, *Pimelodus*, 200
Goeldiella, 184
Goeldiella eques, 184
goeldii, *Brachyplatystoma*, 330
goeldii, *Cheirocerus*, 332
goeldii, *Duoplatinus*, 340
goeldii, *Pimelodina*, 332
goeldii, *Trichomycterus*, 418
Gogangra, 395
Gogangra laevis, 395
Gogangra viridescens, 395
Gogo, 29
Gogo arcuatus, 29
Gogo brevibarbis, 29
Gogo ornatus, 29
gogra, *Phractocephalus*, 105
gogra, *Rita*, 105
Gogrius, 105
Gogrius sykesii, 105
Goliath, *Bagrus*, 331
omesi, *Plecostomus*, 298
omesi, *Squaliforma*, 298
gomezi, *Corydoras*, 117
gongshanensis, *Pareuchiloglanis*, 398
goniaspis, *Arius*, 52
Gonocephalus, 160
gonzalezi, *Amblydoras*, 168
gonzalezi, *Zatorax*, 168
goodi, *Paramphilius*, 25
goongwaree, *Hypophthalmus*, 362
goongwaree, *Proeutropiichthys*, 362
gorgona, *Trichomycterus*, 418
Goslinia, 330
gossei, *Corydoras*, 118
gossei, *Malapterurus*, 302
gouldingi, *Helogenes*, 134
gouldingi, *Stauroglanis*, 413
goyazensis, *Hypostomus*, 254
goyazensis, *Plecostomus*, 254
gracile, *Exostoma*, 387
gracile, *Glyptosternum*, 390
gracilicaudata, *Euchiloglanis*, 398
gracilicaudatus, *Pareuchiloglanis*, 398
graciliformis, *Schmidelia*, 432
gracilior, *Ituglanis*, 407
gracilior, *Pygidium*, 407
gracilis, *Clarias*, 144
gracilis, *Corydoras*, 118
gracilis, *Epactionotus*, 236
gracilis, *Farlowella*, 237
gracilis, *Glyptothorax*, 389, 390
gracilis, *Harttia*, 241
gracilis, *Hemibagrus*, 88
gracilis, *Pimelodella*, 191
gracilis, *Pimelodus*, 191, 196, 210
gracilis, *Osteogeneiosus*, 51
gracilis, *Pseudepapterus*, 76
gracilis, *Pseudobagrus*, 102
gracilis, *Schultzichthys*, 413
gracilis, *Trichomycterus*, 422
graciosa, *Homodiaetus*, 406
graciosus, *Pimelodus*, 210
Graeffei, *Arius*, 47
graeffei, *Neoarius*, 47
grafii, *Corydoras*, 113
grahami, *Silurus*, 379
grammatophorus, *Amphilus*, 22, 23
grammatophorus brevipinna, *Amphilus*, 22
grammatophorus inaequalis, *Amphilus*, 23
grammatophorus marmoratus, *Amphilus*, 23
grandicassis, *Arius*, 49, 50
grandicassis, *Notarius*, 50
grandiops, *Synodontis*, 445
grandis, *Amphilus*, 23
grandis, *Auchenoglanis*, 163
grandis, *Chrysichthys*, 156
grandis, *Parakysis* 15
grandis, *Socnopaea*, 430
grandiscutata, *Rita*, 105
grandoculis, *Arius*, 53
grandoculis, *Potamarius*, 53
granducolis, *Arius*, 53
grangeri, *Rhineastes*, 432
granosus, *Arius*, 54
granosus, *Hypostomus*, 273
granosus, *Neoplecostomus*, 273
granulosa, *Synodontis*, 315
granulatus, *Arius*, 32
granulosus, *Doras*, 177
granulosus, *Genidens* 44
granulosus, *Pterodoras*, 177
granulosus, *Synodontis*, 315
graueri, *Chrysichthys*, 156
gravoti, *Auchenoglanis ballayi*, 162
greeni, *Chaetostoma*, 228
grenfelli, *Eutropius*, 363
grenfelli, *Schilbe*, 363
greshoffi, *Synodontis*, 315
griffini, *Pimelodella*, 191
grisea, *Pimelodella*, 191
griseus, *Ancharius*, 29
griseus, *Corydoras*, 118
griseus, *Limatulichthys*, 266
griseus, *Loricaria*, 266
griseus, *Pimelodus (Pimelodella)*, 191
grixalvii, *Astroblepus*, 63, 64
grixalvii micrescens, *Astroblepus*, 65
Gronias, 204
Gronias nigrilabris, 204, 206
Gronovii, *Galeichthys*, 37
gronovii, *Aspredo*, 58, 60
grosskopfii, *Pimelodus*, 338

grosskopfii, *Pimelodus* (*Pimelodus*), 338
grosskopfii navarroi, *Pimelodus*, 338
grunniens, *Pimelodus*, 336
grypus, *Anadoras*, 169
grypus, *Doras*, 169
guacamaya, *Typhlobelus*, 426
guacari, *Hypostomus*, 250, 258
guacharote, *Ancistrus*, 225
guacharote, *Hypostomus*, 265
guacharote, *Lasiancistrus*, 265
guahiborum, *Hemiancistrus*, 243
guairense, *Chaetostoma*, 228
guairenensis, *Chaetostomus*, 228
guairenensis, *Rhamdia*, 198
guapore, *Corydoras*, 118
guapore, *Lasiancistrus*, 265
guaraquessaba, *Trichomycterus*, 418
guaricensis, *Farlowella*, 240
guasarensis, *Rhamdia*, 197
guatemalensis, *Ariopsis*, 33
guatemalensis, *Arius*, 33
guatemalensis, *Pimelodus*, 200
guatemalensis decolor, *Rhamdia*, 202
guatemalensis murieei, *Rhamdia*, 202
guatemalensis stygaea, *Rhamdia*, 202
guayaberensis, *Medemichthys*, 188
guayaberensis, *Ituglanis*, 407
guayaberensis, *Pygidium metae*, 407
guayoensis, *Deltadoras*, 173
guayoensis, *Megalodoras*, 173
guentheri, *Ancistrus*, 287
guentheri, *Astroblepus*, 64
guentheri, *Atopochilus*, 308
guentheri, *Clarias*, 143
guentheri, *Euchilichthys*, 309
guentheri, *Galeichthys*, 33
guentheri, *Hypoptopoma*, 250
guentheri, *Microlepidogaster*, 296
guentheri, *Pseudancistrus*, 287
guentheri, *Oxyloricaria*, 299
guentheri, *Sturisoma*, 299
guentheri, *Stygogenes*, 64
Guentheri, *Synodontis*, 309
guianense, *Hypoptopoma*, 250
guianense, *Pygidium*, 418
guianensis, *Ageneiosus* 70
guianensis, *Corydoras*, 118
guianensis, *Harttia*, 241
guianensis, *Pygidium*, 418
guianensis, *Trichomycterus*, 418
guineensis, *Clarias*, 141
Guirali, *Pimelodus*, 162
Guiritinga, 43
gulare, *Hypoptopoma*, 249, 250
gulio, *Bagrus*, 93
gulio, *Mystus*, 95
gulio, *Pimelodus*, 93, 95
guliooides, *Bagrus*, 95

gulosus, *Tachisurus*, 40
guntheri, *Microlepidogaster*, 295
guntheri, *Schizolecis*, 295
güntheri, *Atopochilus*, 309
güntheri, *Ancistrus*, 287
güntheri, *Microlepidogaster*, 296
guppyi, *Pseudauchenipterus*, 76
gurgu, *Silurus*, 431
guttata, *Synodontis*, 315
guttatus, *Acrochordonichthys*, 12
guttatus, *Hemibagrus*, 88
guttatus, *Hypostomus*, 287
guttatus, *Imparfinis*, 185
guttatus, *Nannorhamdia*, 185
guttatus, *Parauchenoglanis*, 163
guttatus, *Pimelodus*, 88, 162, 163
guttatus, *Synodontis*, 315
Guyanancistrus, 287
Gymnallabes, 137, 150
Gymnallabes alvarezi, 150
Gymnallabes apus, 137
Gymnallabes nops, 150
Gymnallabes tihoni, 152
Gymnallabes typus, 150
Gymnallabes typus forma heterocercalis, 150
Gymnallabes typus heterocercalis, 150
gymnogaster, *Loricaria*, 297
gymnogaster, *Spatuloricaria*, 297
gymnogaster lagoichthys, *Loricaria*, 297
gymnorhynchus, *Ancistrus*, 221
gymnorhynchus, *Gephyroglanis*, 161
gymnorhynchus, *Hypostomus*, 254
gymnorhynchus, *Plecostomus*, 254
gymnorhynchus occidentalis,
 Hypostomus, 257
gymnorhynchus tapanahoniensis,
 Hypostomus, 261
gyrina, *Tatia*, 77
Gyrinurus, 409
Gyrinurus batrachostoma, 409
gyrinus, *Acrochordonichthys* 12
gyrinus, *Centromochlus*, 77
gyrinus, *Noturus*, 213
gyrinus, *Silurus*, 211, 213, 214
H
habereri, *Chrysichthys*, 156
habereri, *Euchilichthys*, 309
habereri, *Gephyroglanis*, 161
habrosus, *Corydoras*, 118
Haemomaster, 405
Haemomaster venezuelae, 405
haemomyzon, *Homodiaetus*, 412
haemomyzon, *Pseudostegophilus*, 412
haggini, *Bunocephalus*, 59
hahni, *Farlowella*, 238
hainanensis, *Glyptosternon*, 390
hainanensis, *Glyptothorax*, 390
hainanensis, *Glyptothorax fukiensis*, 390
hainanensis, *Hemibagrus*, 88
hainanensis, *Leiocassis*, 88
hainesi, *Amissidens*, 32
hainesi, *Arius*, 32
Halepensis, *Bagrus*, 93, 94, 96, 97
hamiltonii, *Chaca*, 135, 136
hamiltonii, *Plotosus (Clarias)*, 140
hamiltonis, *Arius*, 51
hammarlundi, *Hemiancistrus*, 243
Hammondii, *Pimelodus*, 210
Hancockii, *Doras*, 177
Haplodoras, 173
Hara, 382, 395, 444
Hara buchanani, 395
Hara elongata, 384
Hara filamentosa, 395
Hara hara, 395, 396
Hara horai, 396
Hara jerdoni, 396
Hara Malabarica, 96
Hara saharsai, 396
Hara serrata, 396
Hara serratus, 396
hara, *Hara*, 396
hara, *Pimelodus*, 395, 396
haraldschultzi, *Corydoras*, 118
harbinger, *Chiloglanis*, 306
hardenbergi, *Arius*, 36
hardenbergi, *Aspistor*, 36
Hardwickii, *Acanthonotus*, 356
Hargeri, *Amphilius*, 23
hargreavesi, *Farlowella*, 238
Harmandi, *Hemiarius*, 31
haroldoi, *Parotocinclus*, 282
hartii, *Pimelodella*, 191
Hartii, *Pimelodus (Pseudorhamdia)*, 191
Harttia, 217, 240, 440
Harttia caquetae, 300
Harttia carvalhoi, 240
Harttia crassicauda, 242
Harttia depressa, 240
Harttia dissidens, 241
Harttia duriventris, 241
Harttia filamentissima, 264
Harttia filamentosa, 263, 264
Harttia garavello, 241
Harttia gracilis, 241
Harttia guianensis, 241
Harttia kronei, 241
Harttia leiopleura, 240, 241
Harttia longipinna, 241
Harttia loricariformis, 240, 241
Harttia merevari, 241
Harttia microps, 290
Harttia njsseni, 272
Harttia novalimensis, 241

- Harttia punctata*, 241
Harttia rhombocephala, 242
Harttia surinamensis, 242
Harttia torrenticola, 242
Harttia trombetensis, 242
Harttia uatumensis, 242
Harttiella, 242
Harttiella crassicauda, 242
Harttiinae, 217
hartwelli, Pimelodella, 192
hasemani, Auchenipterus (Pseudepapterus), 76
hasemani, Centrodoras, 170
hasemani, Farlowella, 238
hasemani, Hemidoras, 172
hasemani, Hemiloricaria, 245
hasemani, Imparfinis, 185
hasemani, Leptodoras, 172
hasemani, Otocinclus, 274
hasemani, Oxydoras (Rhinodoras) amazonum, 170
hasemani, Pimelodella, 192
hasemani, Pseudepapterus 76
hasemani, Pygidium, 418
hasemani, Rineloricaria, 245
hasemani, Trichomycterus, 418
hasemani, Vandellia, 411
Hassar, 171
Hassar affinis, 171
Hassar iheringi, 171
Hassar orestis, 171
Hassar praelongus, 173
Hassar ucayalensis, 171
Hassar wilderi, 171
Hassar woodi, 171
Hasselquistii, Clarias, 140
Hasselquistii, Schilbe, 365
hasseltii, Silurichthys, 377
hassleriana, Netuma, 56
hastatus, Arius, 105
hastatus, Corydoras, 112, 118
Hatcheria, 405
Hatcheria bullocki, 404
Hatcheria macraei, 405
Hatcheria Maldonadoi, 403, 404
Hatcheria patagoniensis, 405
Hatcheria pique, 405
Hatcheria titcombi, 405
haugi, Synodontis, 315
Haustor, 208
Haustor ochoterenai, 209
hauxwelli, Loricariichthys, 270
havmollerii, Batasio, 85
havmollerii, Mystus, 85
hazenensis, Ameiurus, 204
hazenensis, Ictalurus (Amiurus), 204
Heckelii, Arius, 55
heckelii, Auchenipterus, 73
heckelii, Centromochlus, 73
Heckelii, Doras, 178
heckelii, Scorpiodoras, 178
Helicophagus, 324
Helicophagus hypophthalmus, 325
Helicophagus leptorhynchus, 324
Helicophagus typus, 324, 325
Helicophagus waandersii, 325
helicophagus, Chrysichthys 156
helophilus, Doras, 177
Helogenes, 130, 134
Helogenes amazonae, 134
Helogenes castaneus, 134
Helogenes gouldingi, 134
Helogenes marmoratus, 134
Helogenes marmoratus uruyensis, 134
Helogenes unidorsalis, 134
Helogenes uruyensis, 134
Helogenidae, 130
hematophaga, Vandellia, 410
Hemiancistrus, 218, 242
Hemiancistrus albocinctus, 223
Hemiancistrus annectens, 242
Hemiancistrus arenarius, 284
Hemiancistrus aspidolepis, 242
Hemiancistrus braueri, 284
Hemiancistrus brevis, 284
Hemiancistrus caquetae, 265
Hemiancistrus castelnau, 265
Hemiancistrus chlorostictus, 243
Hemiancistrus daguae, 231
Hemiancistrus fugleri, 243
Hemiancistrus fuliginosus, 243
Hemiancistrus guahiborum, 243
Hemiancistrus hammarlundi, 243
Hemiancistrus holostictus, 243
Hemiancistrus landoni, 243
Hemiancistrus longipinnis, 227
Hemiancistrus macrops, 243
Hemiancistrus maracaiboensis, 243
Hemiancistrus mayoloi, 264
Hemiancistrus medians, 243
Hemiancistrus megacephalus, 244
Hemiancistrus megalopteryx, 244
Hemiancistrus meizospilos, 244
Hemiancistrus micrommatos, 244
Hemiancistrus niceforoi, 257
Hemiancistrus niger, 287
Hemiancistrus platyrhynchus, 232
Hemiancistrus punctulatus, 244
Hemiancistrus spilomma, 244
Hemiancistrus spinosissimus, 244
Hemiancistrus subviridis, 244
Hemiancistrus ucayalensis, 283, 285
Hemiancistrus votouro, 244
Hemiancistrus wilsoni, 244
Hemiarrius, 44
Hemiarrius dioctes, 44
Hemiarrius harmandi, 31
Hemiarrius insidiator, 44
Hemiarrius sona, 44
Hemiarrius stormi, 45
Hemiarrius stormii, 45
Hemiarrius verrucosus, 45
Hemibagrus, 87
Hemibagrus baramensis, 87
Hemibagrus bongan, 87
Hemibagrus caveatus, 87
Hemibagrus centralis, 87
Hemibagrus chrysops, 87
Hemibagrus elongatus hongus, 89
Hemibagrus filamentus, 88
Hemibagrus fortis, 88
Hemibagrus furcatus, 88
Hemibagrus gracilis, 88
Hemibagrus guttatus, 88
Hemibagrus hainanensis, 88
Hemibagrus hoevenii, 88
Hemibagrus hongus, 89
Hemibagrus imbrifer, 89
Hemibagrus johorensis, 89
Hemibagrus macropterus, 87, 89, 91
Hemibagrus major, 89
Hemibagrus maydelli, 89
Hemibagrus menoda, 89
Hemibagrus microphthalmus, 90
Hemibagrus nemurus, 90
Hemibagrus olyroides, 90
Hemibagrus peguensis, 90
Hemibagrus planiceps, 90
Hemibagrus pluriradiatus, 90
Hemibagrus punctatus, 91
Hemibagrus sabanus, 91
Hemibagrus spilopterus, 91
Hemibagrus taphrophilus, 108
Hemibagrus variegatus, 91
Hemibagrus velox, 91
Hemibagrus vietnamicus, 91
Hemibagrus vietnammicus, 91
Hemibagrus wyckii, 91
Hemibagrus wyckioides, 91
Hemicetopsis, 131
Hemicetopsis amphiloxus, 131
Hemicetopsis macilentus, 133
Hemicetopsis minutus, 130
Hemicetopsis morenoi, 130
Hemicetopsis othonops, 132
hemicochliodon, Hypostomus, 254
Hemidoradinae, 166
Hemidoras, 171
Hemidoras (Leptodoras) boulengeri, 175
Hemidoras boulengeri, 175
Hemidoras hasemani, 172
Hemidoras leporinus, 174
Hemidoras micropoeus, 171
Hemidoras microstomus, 179
Hemidoras morrisi, 172
Hemidoras notospilus, 171

Hemidoras paraguayensis, 179
Hemidoras stenopeltis, 172
Hemileiocassis, 91
Hemileiocassis panjang, 91
Hemiloricaria, 244
Hemiloricaria altipinnis, 245
Hemiloricaria aurata, 245
Hemiloricaria beni, 245
Hemiloricaria cacerensis, 245
Hemiloricaria caracasensis, 244, 245
Hemiloricaria caracassensis, 245
Hemiloricaria castroi, 245
Hemiloricaria eigenmanni, 245
Hemiloricaria fallax, 245
Hemiloricaria formosa, 245
Hemiloricaria hasemani, 245
Hemiloricaria hoehnei, 245
Hemiloricaria jubata, 246
Hemiloricaria konopickyi, 246
Hemiloricaria lanceolata, 246
*Hemiloricaria magdalena*e, 246
Hemiloricaria melini, 246
Hemiloricaria morrowi, 246
Hemiloricaria nigricauda, 246
Hemiloricaria parva, 246
Hemiloricaria phoxocephala, 246
Hemiloricaria platyura, 246
Hemiloricaria sneiderni, 246
Hemiloricaria stewarti, 246
Hemiloricaria teffiana, 247
Hemiloricaria wolfei, 247
Hemiodon, 292
Hemiodon acipenserinus, 247
Hemiodon depressus, 292, 293
Hemiodon platycephalus, 288
Hemiodontichthysina, 217
Hemiodontichthys, 217, 247
Hemiodontichthys acipenserinus, 247
hemiolipterus, *Phractocephalus*, 336
hemiolipterus, *Silurus*, 336
hemipeltis, *Nemadoras*, 174
hemipeltis, *Opsodoras*, 174
hemiphractus, *Callichthys*, 111
Hemipimelodinae, 30
Hemipimelodus, 30, 41
Hemipimelodus aaldereni, 32
Hemipimelodus atripinnis, 34
Hemipimelodus bernhardi, 39
Hemipimelodus bicolor, 34
Hemipimelodus cochlearis, 43
Hemipimelodus colcloughi, 51
Hemipimelodus crassilabris, 42
Hemipimelodus daugueti, 43
Hemipimelodus dayi, 46
Hemipimelodus intermedius, 42
Hemipimelodus macrocephalus, 41
Hemipimelodus macrorhynchus, 32
Hemipimelodus papillifer, 48
Hemipimelodus siamensis, 41

Hemipimelodus sundanensis, 56
Hemipimelodus taylori, 48
Hemipimelodus velutinus, 48
Hemiplatystoma, 341
Hemipsilichthys, 247, 444
Hemipsilichthys azygolechis, 279
Hemipsilichthys calmoni, 279
Hemipsilichthys cameroni, 279
Hemipsilichthys cerosus, 279
Hemipsilichthys duseni, 262, 279
Hemipsilichthys eurycephalus, 279
Hemipsilichthys garbei, 279
Hemipsilichthys gobio, 247
Hemipsilichthys hypselurus, 280
Hemipsilichthys hystrix, 280
Hemipsilichthys mutuca, 280
Hemipsilichthys nimius, 247, 444
Hemipsilichthys nudulus, 280
Hemipsilichthys papillatus, 248, 444
Hemipsilichthys regani, 280
Hemipsilichthys splendens, 280
Hemipsilichthys steindachneri, 280
Hemipsilichthys stephanus, 280
Hemipsilichthys stomias, 281
Hemipsilichthys vestigipinnis, 281
Hemisilurus, 368
Hemisilurus heterorhynchus, 368
Hemisilurus mekongensis, 368
Hemisilurus moolenburghi, 369
Hemisilurus schilbeides, 371
Hemisilurus sclerонema, 368
Hemisorubim, 333
Hemisorubim platyrhynchos, 333
Hemisynodontis, 309
Hemisynodontis membranacea, 309
hemiurus, *Hypostomus*, 254
hemiurus, *Plecostomus*, 254
hendricksoni, *Akyris*, 14
henni, *Hexanematicthys*, 56
Henonemus, 405
Henonemus intermedius, 405
Henonemus macrops, 406
Henonemus panzeri, 414
Henonemus punctatus, 406
Henonemus taxistigma, 406
Henonemus triacanthopomus, 440
henrici, *Anoplectropius*, 165
henrici, *Cranoglanis*, 165
henriquei, *Farlowella*, 238
henryi, *Aoria*, 100
henselii, *Loricaria*, 294
henselii, *Rineloricaria*, 294
Heptapteridae, 180
Heptapterinae, 180
Heptapterus, 180, 184
Heptapterus anisurus, 188
Heptapterus bleekeri, 184
Heptapterus collettii, 99
Heptapterus eigenmanni, 184
Heptapterus fissipinnis, 184
Heptapterus multiradiatus, 184
Heptapterus mustelinus, 184
Heptapterus nissipinnis, 184
Heptapterus ornaticeps, 185
Heptapterus somnians, 189
Heptapterus stewarti, 184
Heptapterus surinamensis, 183
Heptapterus sympterygium, 184
Heptapterus tapanahoniensis, 184
Heptapterus tenuis, 184
heraldoi, *Pimelodus*, 338
herberti, *Ituglanis*, 407
Herberti, *Trichomycterus*, 407
HerklotSELLA, 375
HerklotSELLA anomala, 375
hermanni, *Hypostomus*, 254
Hermannii, *Plecostomus*, 254
herzbergii, *SciaDES*, 53
Herzbergii, *Silurus*, 53
herzensteini, *Leiocassis*, 92
herzensteini, *Macrones*, 92
hesperius, *Kryptopterus*, 370
heteracantha, *Rhamdia*, 201
heteracanthus, *Chaetostomus*, 264, 265
heteracanthus, *Lasiancistrus*, 265
Heterobagrus, 94
Heterobagrus bocourti, 94
Heterobranchoides, 139
Heterobranchus, 136, 150
Heterobranchus 5-tentaculatus, 197
Heterobranchus 6-tentaculatus, 197, 200
Heterobranchus austriacus, 382
Heterobranchus bidorsalis, 150
Heterobranchus boulengeri, 151
Heterobranchus Geoffroyi, 150
Heterobranchus intermedius, 150
Heterobranchus isopterus, 151
Heterobranchus laticeps, 151
Heterobranchus longifilis, 151
Heterobranchus macronema, 151
Heterobranchus palaeindicus, 151
Heterobranchus platycephalus, 151
Heterobranchus Senegalensis, 150
Heterobranchus sextentaculatus, 197, 200
Heterobranchus tapeinopterus, 149, 150
heterocephalus, *Clariallabes*, 137
heterocercalis, *Gymnallabes typus*, 150
heterodon, *Arges*, 64
heterodon, *Astroblepus*, 64
heterodon, *Leporacanthicus*, 266
heterodontum, *Pygidium*, 418
heterodontus, *Trichomycterus*, 418
heteromorphus, *Corydoras*, 118

- heteropleura**, *Brachyrhamdia*, 181
heteropleurus, *Pimelodus*, 181
Heteropneustes, 136, 151
Heteropneustes fossilis, 151
Heteropneustes kemratensis, 152
Heteropneustes longipectoralis, 152
Heteropneustidae, 136
heteroptera, *Rineloricaria*, 293, 294
heterorhynchus, *Ancistrus*, 221
heterorhynchus, *Hemisilurus*, 368
heterorhynchus, *Wallago*, 368
heterorhynchus, *Xenocara*, 221
heterurus, *Akysis*, 14
heterurus, *Bagrurus*, 97
heterurus, *Zaireichthys*, 28
Heudelotii, *Arius*, 31
heuglini, *Clarotes*, 160
eward-belli, *Arius*, 56
hexacicinnus, *Macropteronotus*, 148
hexadactylus, *Silurus*, 61
hexanema, *Laides*, 358
hexanema, *Pangasius*, 358
hexanema, *Silurodon*, 380, 381
Hexanematicichthys, 45
Hexanematicichthys henni, 56
Hexanematicichthys hymenorrhinos, 54
Hexanematicichthys leptaspis, 47
Hexanematicichthys leptocassis, 45
Hexanematicichthys mastersi, 45
Hexanematicichthys sagor, 45
Hexanematicichthys sundaicus, 45
Hexanematicichthys surinamensis, 33
hexapterus, *Micronema*, 371
hexapterus, *Silurus* 371
heylandi, *Kronichthys*, 263
heylandi, *Plecostomus*, 263
Hilarii, *Pimelodus*, 200
Hildadoras, 175
Hildadoras bolivarensis, 168
Hildadoras orinocensis, 175, 176
hildae, *Amarginops*, 153
hildae, *Chrysichthys*, 153
hildebrandi, *Noturus*, 213
hildebrandi, *Schilbeodes*, 213
hildebrandi laetus, *Noturus*, 213
hilgendorfi, *Clarias*, 139
hilli, *Clarias*, 144
hirsuta, *Microsynodontis*, 310
hirsutus, *Liocassus*, 107
hirsutus, *Microsynodontis*, 310
Hisonotus, 248
Hisonotus candombe, 440
Hisonotus charrua, 440
Hisonotus depressicauda, 248
Hisonotus depressinotus, 248
Hisonotus francirochai, 248
Hisonotus insperatus, 248
Hisonotus laevior, 248
Hisonotus leptochilus, 248
Hisonotus leucofrenatus, 248
Hisonotus maculipinnis, 248
Hisonotus nigricauda, 248
Hisonotus notatus, 248
Hisonotus paulinus, 249
Hisonotus ringueleti, 249
Hisonotus taimensis, 249
histrix, *Pseudacanthicus*, 286
histrix, *Rinelepis*, 218, 286
Hito, 375
Hito taytayensis, 375, 377
Hitoichthys, 375
Hitoichthys taytayensis, 375, 377
hodgarti, *Euchiloglanis*, 398
hodgarti, *Glyptosternum*, 398
hodgarti, *Parachiloglanis*, 398
hoedemani, *Exastilithoxus*, 237
hoehnei, *Hemiloricaria*, 245
hoehnei, *Loricaria*, 245
hoehnei, *Nannoglanis*, 188
hoehnei, *Phenacorhamdia*, 188
hoeksi, *Pangasius*, 328
hoerdzanicus, *Eobagrus*, 87
hoerdzanicus, *Gobibagrus*, 87
hoevenii, *Bagrurus*, 88
hoevenii, *Hemibagrus*, 88
Hoffstetterichthys pucae, 30
hoi, *Aoria*, 107
holdeni, *Oxydoras*, 176
hollandi, *Imparfinis*, 185
Hollyi, *Clarias*, 144
hollyi, *Clarias*, 147
Hollyi, *Synodontis*, 323
Hollyi ntemensis, *Synodontis*, 322
holobranchus, *Xenoclarias*, 153
holomelas, *Pimelodus*, 199
holomelas rupununi, *Rhamdia*, 197
holopercnus, *Synodontis*, 315
holostictus, *Hemiancistrus*, 243
Homodiaetus, 406
Homodiaetus anisitsi, 406
Homodiaetus banguela, 406
Homodiaetus graciosa, 406
Homodiaetus haemomyzon, 412
Homodiaetus passarelli, 406
Homodiaetus vazferreira, 406
homodon, *Arges*, 64
homodon, *Astroblepus*, 64
hondae, *Hypostomus*, 254
hondae, *Plecostomus*, 250, 254
honghenensis, *Glyptothorax fukiensis*, 390
honghensis, *Glyptothorax*, 390
honghensis, *Glyptothorax fukiensis*, 390
hongus, *Hemibagrus*, 89
hongus, *Hemibagrus elongatus*, 89
Hopladelus, 215
Hoplancistrini, 217
Hoplancistrus, 217, 249
Hoplancistrus tricornis, 249
Hoplisoma, 111
Hoplodaros, 173
Hoplodoras, 173
Hoplodoras ramirezi, 174
hoplogenys, *Ancistrus*, 221
hoplogenys, *Chaetostomus*, 221
Hoplomizoninae, 57
Hoplomyzon, 57, 61
Hoplomyzon atrizona, 61
Hoplomyzon atrizona petroleus, 61
Hoplomyzon megistus, 60
Hoplomyzon papillatus, 61
Hoplomyzon sexpapilostoma, 61
Hoplomyzontinae, 57
hoplonites, *Hypostomus*, 254
Hoplosoma aeneum, 112
Hoplosterninae, 108
Hoplosternum, 108, 127
Hoplosternum littorale, 127
Hoplosternum littorale daillyi, 128
Hoplosternum magdalena, 128
Hoplosternum oronocoi, 129
Hoplosternum punctatum, 127, 128
Hoplosternum schreineri, 128
Hoplosternum shirui, 128
Hoplosternum steverdii, 128
Hoplosternum thoracatum cayennae, 128
Hoplosternum thoracatum surinamensis, 129
hoppei, *Otocinclus*, 275
Horabagrinae, 441
Horabagrus, 356, 429, 441
Horabagrus brachysoma, 429
Horabagrus nigricollaris, 429
horae, *Amblyceps*, 99
horae, *Olyra*, 99
Horaglanidinae, 441
Horaglanis, 152, 441
Horaglanis alikunhi, 152
Horaglanis krishnai, 152
horai, *Glyptothorax*, 390
horai, *Glyptothorax*, 390
horai, *Hara*, 396
horai, *Mystus*, 95
horai, *Mystus (Mystus) vittatus*, 95
horai, *Olyra*, 99
horai, *Pteroglanis*, 387, 390
horai, *Socnopaea*, 431
Horiomyzon, 185
Horiomyzon retropinnatus, 185
horrida, *Squaliforma*, 298
horridus, *Hypostomus*, 297, 298
horridus, *Plotosus*, 350
Hosii, *Liocassis*, 93
houghi, *Pimelodus*, 210
housei, *Glyptothorax*, 390

- howesi*, *Pimelodella*, 192
howong, *Macrones*, 88
hoysi, *Pimelodus*, 206
huaorani, *Otocinclus*, 275
huberi, *Oxydoras (Rhinodoras)*, 174
humboldti, *Doras*, 176
Humboldtii, *Cyclopium*, 63, 64
humboldtii, *Zungaro*, 344
humeralis, *Doras*, 174
humeralis, *Nemadoras*, 174
humeralis, *Pangasius*, 326
humeratus, *Synodontis*, 321
humilis, *Pimelodus*, 198
humilis, *Rhamdia*, 198
hwanghoensis, *Leiocassis*, 107
Hyalobagrus, 92
Hyalobagrus flavus, 92
Hyalobagrus leiacanthus, 92
Hyalobagrus ornatus, 92
hydrostaticus, *Pleurophysus*, 410
hymenorhinos, *Hexanematicthys*, 54
Hypancistrus, 249
Hypancistrus inspector, 249
Hypancistrus zebra, 249
Hypocolpterus, 227
Hypocolpterus analis, 227
Hypodoras, 172
Hypodoras forciculatus, 172
Hypophthalmini, 329
Hypophthalmus, 329, 333
Hypophthalmus devall, 334
Hypophthalmus edentatus, 333
Hypophthalmus fimbriatus, 333
Hypophthalmus goongwaree, 362
Hypophthalmus longifilis, 334
Hypophthalmus marginatus, 333
Hypophthalmus niloticus, 362, 365
Hypophthalmus nuchalis, 71, 72
Hypophthalmus oremaculatus, 333
Hypophthalmus perporosus, 334
Hypophthalmus Spixii, 333
Hypophthalmus taakree, 362
hypophthalmus, *Helicophagus*, 325
hypophthalmus, *Arius*, 39, 40
hypophthalmus, *Cathorops*, 40
hypophthalmus, *Pangasianodon*, 325
hypophthalmus, *Ompok*, 371, 372
hypophthalmus, *Silurus*, 372
Hypoptopoma, 217, 249
Hypoptopoma bilobatum, 250
Hypoptopoma carinatum, 276
Hypoptopoma guentheri, 250
Hypoptopoma guianense, 250
Hypoptopoma gulare, 249, 250
Hypoptopoma inexpectata, 250
Hypoptopoma inexpectatum, 250
Hypoptopoma joberti, 250
Hypoptopoma psilogaster, 250
Hypoptopoma steindachneri, 250
Hypoptopoma thoracatum, 249, 250
Hypoptopomatinae, 217
Hypoptopominae, 217
Hypostoma ententaculatum, 291
Hypostoma punctatum, 220
Hypostoma squalinum, 298
Hypostomiden, 216
Hypostomus, 216, 217, 250
Hypostomus aburrensis, 227
Hypostomus affinis, 251
Hypostomus agna, 251
Hypostomus alatus, 251
Hypostomus albopunctatus, 251
Hypostomus ancistroides, 251
Hypostomus angipinnatus, 251
Hypostomus argus, 251
Hypostomus asperatus, 251
Hypostomus aspilogaster, 251
Hypostomus atropinnis, 251
Hypostomus aurantiacus, 278
Hypostomus auroguttatus, 251
Hypostomus barbatus, 286, 287
Hypostomus boliviensis, 252
Hypostomus borellii, 252
Hypostomus boulegeri, 252
Hypostomus brevicauda, 252
Hypostomus brevis, 252
Hypostomus brevitentaculatus, 291
Hypostomus bufonius, 219
Hypostomus calamita, 219
Hypostomus carinatus, 252
Hypostomus carvalhoi, 252
Hypostomus cirrhosus, 219, 220
Hypostomus cochliodon, 250, 252, 253-262
Hypostomus commersoni, 252
Hypostomus coppenamensis, 253
Hypostomus corantijni, 253
Hypostomus crassicauda, 253
Hypostomus derbyi, 253
Hypostomus dlowhyi, 253
Hypostomus duodecimalis, 290, 291
Hypostomus emarginatus, 297
Hypostomus eptingi, 253
Hypostomus ericae, 253
Hypostomus ericius, 253
Hypostomus erinaceus, 221
Hypostomus fluviatilis, 253
Hypostomus fonchii, 253
Hypostomus francisci, 254
Hypostomus garmani, 254
Hypostomus goyazensis, 254
Hypostomus granosus, 273
Hypostomus guacari, 250, 258, 259
Hypostomus guacharote, 265
Hypostomus guttatus, 287
Hypostomus gymnorhynchus, 254
Hypostomus gymnorhynchus occidentalis, 257
Hypostomus gymnorhynchus tapanahoniensis, 261
Hypostomus hemicochliodon, 254
Hypostomus hemiurus, 254
Hypostomus hermanni, 254
Hypostomus hondae, 254
Hypostomus hoplonites, 254
Hypostomus horridus, 297, 298
Hypostomus iheringii, 255
Hypostomus interruptus, 255
Hypostomus isbrueckeri, 255
Hypostomus itacua, 217
Hypostomus jaguribensis, 255
Hypostomus johnii, 255
Hypostomus laplateae, 255
Hypostomus latifrons, 255
Hypostomus latirostris, 255
Hypostomus levis, 255
Hypostomus lexi, 255
Hypostomus lima, 256
Hypostomus longiradiatus, 256
Hypostomus luteomaculatus, 256
Hypostomus luteus, 256
Hypostomus macrophthalmus, 256
Hypostomus macrops, 256
Hypostomus macushi, 256
Hypostomus margaritifer, 256
Hypostomus meleagris, 256
Hypostomus micromaculatus, 256
Hypostomus microstomus, 250, 257
Hypostomus multiradiatus, 290, 291
Hypostomus mutucae, 257
Hypostomus myersi, 257
Hypostomus nematopterus, 257
Hypostomus niceforoi, 257
Hypostomus nickeriensis, 257
Hypostomus niger, 257
Hypostomus nigricans, 278
Hypostomus nigromaculatus, 257
Hypostomus niveatus, 226, 227
Hypostomus nudiceps, 223
Hypostomus nudiventralis, 257
Hypostomus occidentalis, 257
Hypostomus oculeus, 257
Hypostomus pagei, 257
Hypostomus pantherinus, 258
Hypostomus papariae, 258
Hypostomus paranensis, 258
Hypostomus pardalis, 292
Hypostomus paucimaculatus, 258
Hypostomus paucipunctatus, 258
Hypostomus paulinus, 258
Hypostomus pictus, 265
Hypostomus piratatu, 258
Hypostomus plecostomoides, 258
Hypostomus plecostomus, 258
Hypostomus pseudohemiurus, 259
Hypostomus pseudohemiurus macrophthalmus, 256

Hypostomus punctatus, 220, 259
Hypostomus pusarum, 259
Hypostomus pyrineusi, 259
Hypostomus regani, 259
Hypostomus robini, 259
Hypostomus rondoni, 259
Hypostomus roseopunctatus, 259
Hypostomus saramaccensis, 259
Hypostomus scabriceps, 260
Hypostomus scaphiceps, 260
Hypostomus sculpodon, 260
Hypostomus seminudus, 260
Hypostomus serratus, 286
Hypostomus simios, 260
Hypostomus sipaliwini, 260
Hypostomus soniae, 260
Hypostomus spinosus, 286
Hypostomus strigaticeps, 260
Hypostomus subcarinatus, 260
Hypostomus surinamensis, 260
Hypostomus tapanahoniensis, 260
Hypostomus taphorni, 261
Hypostomus tapijara, 261
Hypostomus Temminckii, 224
Hypostomus tenuis, 298
Hypostomus ternetzi, 261
Hypostomus tietensis, 261
Hypostomus topavae, 261
Hypostomus unae, 261
Hypostomus uruguayensis, 261
Hypostomus vaillanti, 261
Hypostomus variipictus, 261
Hypostomus varimaculosus, 261
Hypostomus variostictus, 261
Hypostomus ventromaculatus, 262
Hypostomus vermicularis, 262
Hypostomus verres, 262
Hypostomus vicinus, 278
Hypostomus waiampi, 262
Hypostomus watwata, 262
Hypostomus winzi, 262
Hypostomus wuchereri, 262
Hypothalmus dawalla, 69
Hypselobagrus, 93
Hypselobagrus macronema, 97
hypselopterus, *Bagrichthys*, 82
hypselopterus, *Bagrus*, 82
hypselurus, *Breitensteinia*, 15
hypselurus, *Hemipsilichthys*, 280
hypselurus, *Pareiorhaphis*, 280
hypselurus, *Pimelodus*, 198
Hypsidoridae, 203
Hypsidoris, 203
Hypsidoris farsonensis, 203
Hypsidoris oregonensis, 203
hpsiura, *Amaralia*, 57
hpsiurus, *Bunocephalus*, 57
hpsiurus, *Oreoglanis*, 397
hyrtlii, *Neosilurus*, 347, 348

hystrix, *Acanthicus*, 218
hystrix, *Hemipsilichthys*, 280
hystrix, *Pareiorhaphis*, 280
I
ichikawai, *Coreobagrus*, 86
ichikawai, *Pseudobagrus*, 86
ichneumon, *Coreobagrus*, 431
Ichthaelurinae, 203
Ichthaelurus, 203, 208
Ichthaelurus Kevinskii, 204
Ichthaelurus McCaskei, 204
Ichthaelurus robustus, 210
Ichthyaelurus, 208
Ictaluri, 203
Ictaluridae, 203
Ictalurus, 203, 207
Ictalurus (Amiurus) hazenensis, 204
Ictalurus anguilla, 210
Ictalurus australis, 208
Ictalurus balsanus, 208
Ictalurus benderensis, 206
Ictalurus dugesii, 208, 209
Ictalurus echinatus, 208
Ictalurus furcatus, 208, 211
Ictalurus lambda, 209
Ictalurus lavetti, 204
Ictalurus leidyi, 204
Ictalurus lupus, 209, 211
Ictalurus macgrewi, 205
Ictalurus mexicanus, 209
Ictalurus nebulosus pannonicus, 206
Ictalurus ochoterenai, 209
Ictalurus okeechobeensis, 210
Ictalurus pectinatus, 206
Ictalurus peregrinus, 207
Ictalurus pricei, 209
Ictalurus punctatus, 208, 209, 211
Ictalurus rhaeas, 210
Ictalurus sawrockensis, 206
Ictalurus serracanthus, 206
Ictalurus simpsonii, 210
Ictalurus spodus, 211
Ictalurus vespertinus, 206
idenburgi, *Copidoglanis*, 348
idenburgi, *Neosilurus*, 348
ignobilis, *Rhamdella*, 196
iheringi, *Arius*, 343
iheringi, *Cetopsorhamdia*, 182
iheringi, *Hassar*, 171
iheringi, *Microglanis*, 354
iheringi, *Plecostomus*, 298
iheringi, *Pygidium*, 419
iheringi, *Steindachneridion*, 343
iheringi, *Trichomycterus*, 419
Iheringichthys, 334
Iheringichthys labrosus, 334
Iheringichthys megalops, 334
iheringii, *Bunocephalus*, 59
iheringii, *Hypostomus*, 255
iheringii, *Plecostomus*, 255, 298
ikapor, *Plotoseus*, 350
ikiensis, *Pseudobagrus*, 102
ilebrevis, *Synodontis*, 445
ilesi, *Bathyclarrias*, 136
ilesi, *Dinotopterus*, 136
Ilictis, 215
imberbis, *Silurus*, 70, 379
imbrifer, *Hemibagrus*, 89
imitator, *Brachyramdia*, 181
imitator, *Corydoras*, 118
immaculata, *Pseudecheneis*, 400
immaculatum, *Pygidium*, 419
immaculatus, *Auchenipterus*, 80
immaculatus, *Pseudecheneis*, 400
immaculatus, *Silurus (Callichrus)*, 374
immaculatus, *Trichomycterus*, 419
Imparales, 180, 188
Imparales mariai, 188
Imparales panamensis, 180
Imparfinis, 185
Imparfinis boliviensis, 188
Imparfinis borodini, 185
Imparfinis cochabambae, 185
Imparfinis guttatus, 185
Imparfinis hasemani, 185
Imparfinis hollandi, 185
Imparfinis insidiosus, 182
Imparfinis lineatus, 185
Imparfinis longicauda, 185
Imparfinis longicauda, 185
Imparfinis microps, 186
Imparfinis minutus, 186
Imparfinis mirini, 186
Imparfinis nemacheir, 186
Imparfinis pijpersi, 186
Imparfinis piperatus, 185, 186
Imparfinis pristos, 186
Imparfinis pseudonemacheir, 186
Imparfinis schubarti, 186
Imparfinis spurrelli, 186
Imparfinis stictonotus, 186
Imparfinis tenebrosus, 189
impluviatus, *Mystus*, 95
inaequalis, *Amphilius*
 grammatophorus, 23
incae, *Trichomycterus*, 423
Incaichthys suarezi, 30
inolicana, *Corydoras*, 118
indicus, *Brachyspondylus*, 326
indicus, *Glyptothorax*, 390
indicus, *Kryptopterus*, 367
indicus, *Pangasius*, 326
indicus, *Pimelodus*, 19
indicus, *Silurus*, 374
indragiriensis, *Silurichthys*, 378
ineac, *Phractura*, 28
ineac, *Trachyglanis*, 28
inermis, *Ageneiosus*, 69

inermis, *Akysis*, 16
inermis, *Amblyceps*, 17
inermis, *Nematogenys*, 324
inermis, *Pseudobagarius*, 16
inermis, *Silurus*, 69, 379
inermis, *Trichomycterus*, 324
inexpectata, *Aristommata*, 249, 250
inexpectata, *Hypoptopoma*, 250
inxpectatum*, *Hypoptopoma, 250
infraoculare, *Sorubim*, 342
infulatus*, *Oreoglanis, 397
infuscatus*, *Erethistoides, 442
ingluvies, *Osteogeneiosus*, 51
inornata*, *Pseudolaguvia, 401
inornatus, *Liocassis*, 103
inornatus*, *Pseudomystus, 103
inpai*, *Aguarunichthys, 329
insculptus, *Arius*, 50
insculptus, *Doras*, 179
insculptus*, *Notarius, 50
insidiator, *Arius*, 45
insidiator*, *Hemiarius, 44
insidiosa*, *Cetopsorhamdia, 182
insidiosus, *Imparfinis*, 182
insidiosus*, *Stegophilus, 414
insignarius, *Pimelodon*, 211, 213
insigne, *Pimelodus*, 213
insignis, *Auchenipterus*, 79
insignis*, *Breitensteinia, 15
insignis*, *Noturus, 213
insignis*, *Oreoglanis, 397
insignis, *Pimelodella*, 190
insignis, *Pimelodus*, 339
insignis*, *Trachelyopterus, 79
insignis badeli, *Trachycorystes*, 80
insignis peloichthys, *Trachycorystes*, 80
insolitus*, *Clarias, 144
inspector*, *Hypancistrus, 249
insperatus*, *Hisonotus, 248
insularum, *Netuma*, 50
intermedia*, *Phractura, 26
intermedia*, *Tatia, 77
intermedium, *Pseudoplatystoma fasciatum*, 341
intermedius, *Centromochlus*, 77
intermedius*, *Clarias, 144
intermedius, *Hemipimelodus*, 42
intermedius*, *Henonemus, 405
intermedius, *Heterobranchus*, 150
intermedius*, *Pelteobagrus, 100
intermedius, *Pseudecheneis*, 400
intermedius, *Pseudobagrus*, 100
intermedius, *Pseudotocinclus*, 290
intermedius*, *Schilbe, 363
intermedius, *Stegophilus*, 405
intermedius*, *Trachyglanis, 28
interruptus*, *Hypostomus, 255
interruptus, *Plecostomus*, 255

interspinatum, *Glyptosternon*, 390
interspinatus*, *Glyptothorax, 390
intonsus*, *Ernstichthys, 60
inusitata*, *Pterocryptis, 377
irsacae, *Synodontis*, 314
Irvineia, 358
Irvineia orientalis, 358
Irvineia voltae, 358
irwini, *Megalodoras*, 173, 174
isacanthus, *Auchenipterus*, 80
isbrueckeri, *Farlowella*, 238
isbrueckeri*, *Corydoras, 119
isbrueckeri*, *Farlowella, 238
isbrueckeri, *Hypostomus*, 255
Isbrueckerichthys, 262, 441
Isbrueckerichthys alipionis, 262
Isbrueckerichthys calvus, 441
Isbrueckerichthys duseni, 262
Isbrueckerichthys epakmos, 262
Isbrueckerichthys saxicola, 441
ischnosoma*, *Acrochordonichthys, 12
isheriensis, *Clarias (Clarioides)*, 139
Isidori, *Schilbe*, 365
isopterus*, *Heterobranchus, 151
Isorineloricaria, 263
Isorineloricaria spinosissima, 263
isthmenensis, *Aelurichthys*, 38
Istlarius, 208
Istlarius balsanus, 208
Istlarius balsanus occidentalis, 208
itacaiunas*, *Rhamdia, 198
itacambirussu*, *Trichomycterus, 419
itacarambiensis*, *Trichomycterus, 419
itacua, *Hypostomus*, 217
itaimbezinho*, *Epauctionotus, 236
itapicuruensis*, *Pimelodella, 192
itatiayae*, *Trichomycterus, 419
itchkeea*, *Gagata, 386
itchkeea, *Phractocephalus*, 386
Ituglanis, 406
Ituglanis amazonicus, 406
Ituglanis bambui, 406
Ituglanis cahyensis, 444
Ituglanis eichorniarum, 407
Ituglanis epikarsticus, 407
Ituglanis gracilior, 407
Ituglanis guayaberensis, 407
Ituglanis herberti, 407
Ituglanis laticeps, 407
Ituglanis macunaima, 407
Ituglanis metae, 407
Ituglanis nebulosus, 407
Ituglanis parahybae, 407
Ituglanis parkoi, 407
Ituglanis passensis, 408
Ituglanis proops, 408
Ituglanis ramiroi, 408
ituriiensis, *Synodontis*, 315
iturii, *Auchenoglanis*, 163

iturii*, *Synodontis, 315
iwokrama*, *Denticetopsis, 133
Ixinandria, 263
Ixinandria montebelloi, 263
Ixinandria steinbachi, 263
izabalensis*, *Potamarius, 53
J
jacksonensis, *Netuma thalassina*, 49
jacksoni*, *Dinopterous, 149
jacksonii*, *Amphilus, 21
jacupiranga*, *Trichomycterus, 419
jaekeli, *Otolithus (Arius)*, 435
jaensis*, *Clarias, 144
jagur, *Macropterodonotus*, 140
jaguribensis*, *Hypostomus, 255
jaguribensis, *Plecostomus*, 255
jahu, *Paulicea*, 344
jahu*, *Zungaro, 344
jalalensis*, *Glyptothorax, 390
jallae*, *Dinopterous, 146
jallae*, *Synodontis, 316
jandia*, *Sorubim, 342, 343
janeirensis*, *Pseudotothyris, 290
japonicus, *Centranodon*, 379
japonicus, *Silurus*, 378, 379
jaraguensis, *Loricaria*, 294
jaraguensis*, *Rineloricaria, 294
jataiensis*, *Ancistrus, 222
jatia*, *Cephalocassis, 41
jatius*, *Pimelodus, 41
jauruensis*, *Farlowella, 238
jauruensis, *Farlowella*, 238
javanensis, *Belodontichthys*, 381
javanica, *Rhamdia*, 431
Javensis, *Bagrus*, 45
javus, *Pimelodus*, 431
jayarami*, *Myersglanis, 396
jaynei*, *Ompok, 372
jeanesianus, *Liposarcus*, 292
jegui*, *Chaetostoma, 228
jella, *Arius*, 36
jelskii*, *Ancistrus, 222
Jelskii, *Chaetostomus*, 222
jenynsii, *Pimelodus*, 196
jenynsii*, *Rhamdella, 196
jequitinhonha*, *Rhamdia, 198
Jequitinhonhae, *Auchenipterus (Pseudauchenipterus)*, 76
jequitinhonhae*, *Pseudauchenipterus, 76
jequitinhonhae*, *Trichomycterus, 419
jerdoni, *Hara*, 396
jimi*, *Parotocinclus, 282
jivaro, *Pimelodus*, 338
joberti*, *Hypoptopoma, 250
joberti, *Otocinclus*, 250
johannae, *Septobranchus*, 42
johnelsi*, *Chrysichthys, 156
johnii*, *Hypostomus, 255

- Johnii*, *Plecostomus*, 255
johsoni, *Pygidium*, 419
johsoni, *Trichomycterus*, 419
johorensis, *Hemibagrus*, 89
johorensis, *Mystus*, 89
jokeannae, *Trachycorystes*, 80
jordani, *Tachisurus*, 33
joselimaiai, *Leporacanthicus*, 266
joselimaianus, *Glyptoperichthys*, 291
joselimaianus, *Pterygoplichthys*, 291
juaro, *Pangasius*, 327
jubata, *Hemiloricaria*, 246
jubata, *Loricaria*, 246
julii, *Corydoras*, 119
jumbo, *Parotocinclus*, 282
juquiae, *Corydoras*, 121
juquiae, *Corydoras*, 121
juquiae, *Otothyris*, 276
juquiae, *Pseudotocinclus*, 290
jurubidae, *Astroblepus*, 65
jurubidae, *Cetopsis*, 132
jurubidae, *Pseudocetopsis*, 132
juruense, *Brachyplatystoma*, 331
juruense, *Platystoma*, 331
juruensis, *Leptodoras*, 172
juvens, *Pterygoplichthys*, 291
K
kaiei, *Corymbophanes*, 232
kaifenensis, *Leiocassis*, 107
kajan, *Macrones*, 88
kakrimensis, *Amphilinus*, 21
kalambo, *Chiloglanis*, 306
Kalyptodoras, 172
Kalyptodoras bahiensis, 172
kamengensis, *Euchiloglanis*, 399
kamengensis, *Pareuchiloglanis*, 399
kanei, *Corydoras*, 119
kanganamanensis, *Arius*, 39
kapurensis, *Pinniwallago*, 375
kapuasensis, *Clarias*, 144
kapuri, *Laguvia ribeiroi*, 401
kapuri, *Pseudolaguvia*, 401
kasai, *Eutropiellus*, 360, 361
kashmirensis, *Glyptothorax*, 390, 393
katangae, *Synodontis*, 315
Keletius, *Bagrus*, 98
kelioides, *Encheloclarias*, 149
kempi, *Olyra*, 99
kemratensis, *Clarislurus*, 151, 152
kemratensis, *Heteropneustes*, 152
kessleri, *Arius*, 50
kessleri, *Notarius*, 50
Ketengus, 45
Ketengus typus, 45
Kevinskii, *Ichthaelurus*, 204
khartoumensis, *Synodontis*, 316
khavalchor, *Neotropius*, 359
kinabatanganensis, *Pangasius*, 326
kingi, *Liobagrus*, 19
kingsleyae, *Clarias*, 144
kingsleyae, *Chrysichthys*, 160
kirkii, *Arius*, 52
kishinouyei, *Coraglanis*, 385
kishinouyei, *Euchiloglanis*, 385
kitsoni, *Arius* 56
kivuensis, *Amphilinus*, 21
kneri, *Callichthys*, 111
kneri, *Loricaria*, 299
kneri, *Oxydoras*, 175
kneri, *Sturisoma*, 299
Knerii, *Acestra*, 238
knerii, *Bunocephalus*, 59
knerii, *Farlowella*, 238
knerii, *Pimelodus (Rhamdia)*, 199
knerii, *Trichomycterus*, 419
koenigi, *Bagrus*, 84
koensis, *Synodontis*, 316
koepckeii, *Myoglanis*, 187
konopickyi, *Hemiloricaria*, 246
konopickyi, *Loricaria*, 246
koreanus, *Pseudobagrus*, 102
krattensis, *Wallago*, 374
krefftii, *Amphilinus*, 23
krempfi, *Pangasius*, 326
krishnai, *Horaglanis*, 152
krishnensis, *Mystus*, 89
kronei, *Corydoras*, 129
kronei, *Harttia*, 241
kronei, *Loricaria*, 294
kronei, *Pimelodella*, 192
kronei, *Rineloricaria*, 294
kronei, *Typhlobagrus*, 189, 192
Kronichthys, 263
Kronichthys heylandi, 263
Kronichthys lacerta, 263
Kronichthys subteres, 263
Kryptopterichthys, 369
Kryptopterichthys macrocephalus, 370
Kryptopterini, 367
Kryptopterus, 367, 369
Kryptopterus baramensis, 369
Kryptopterus bicirrhos, 369
Kryptopterus cheveyi, 371
Kryptopterus cryptopterus, 369
Kryptopterus deignani, 375
Kryptopterus dissitus, 369
Kryptopterus eugeneiatus, 369
Kryptopterus geminus, 369
Kryptopterus hesperius, 370
Kryptopterus indicus, 367
Kryptopterus lais, 370
Kryptopterus limpop, 370
Kryptopterus lumholtzi, 370
Kryptopterus macrocephalus, 370
Kryptopterus micropus, 369
Kryptopterus minimus, 370
Kryptopterus minor, 370
Kryptopterus mononema, 370
Kryptopterus moorei, 371
Kryptopterus palembangensis, 370
Kryptopterus parascilbeides, 370
Kryptopterus parvanalis, 375
Kryptopterus piperatus, 370
Kryptopterus platypogon, 371
Kryptopterus sabanus, 370
Kryptopterus schilbeides, 371
kryptos, *Xiliphius*, 62
kuhlmanni, *Peckoltia*, 284
kuhlmanni, *Peckoltichthys*, 284
kükenthali, *Glyptosternum*, 391
kunyit, *Pangasius*, 326
kurdistanicum, *Glyptosternum*, 390
kurdistanicus, *Glyptothorax*, 390
kurzii, *Akysis*, 18
kutchensis, *Arius*, 56
kuturnee, *Phractocephalus*, 105
kuturnee, *Rita*, 105
kyphus, *Pelteobagrus*, 82
kyphus, *Pseudobagrus*, 82
L
laak, *Glyptosternum*, 395
laani, *Apistoloricaria*, 226
labeo, *Synodontis*, 323
labialis, *Astroblepus*, 65
labialis, *Loricaria*, 270
labialis, *Loricarichthys*, 270
labiatum, *Exostoma*, 385
labiatus, *Glyptosternon*, 385
labiosus, *Copidoglanis*, 345
labrax, *Exostoma*, 387
labrosus, *Copidoglanis*, 345
labrosus, *Iheringichthys*, 334
labrosus, *Pimelodus*, 334
labrosus, *Xyliphius*, 62
Lacantunia, 216
Lacantunia enigmatica, 216
Lacantuniidae, 216
lacerdai, *Corydoras*, 119
lacerta, *Kronichthys*, 263
lacerta, *Plecostomus*, 263
lachneri, *Noturus*, 213
lacus, *Aoria*, 108
lacustricolus, *Synodontis*, 322
lacustris, *Auchenipterus*, 79
lacustris, *Trachelyopterus*, 79
lacustris, *Gadus*, 208
laessoei, *Synodontis*, 316
laeviceps, *Arius*, 55
laeviceps, *Clarias*, 144
laeviceps dialonensis, *Clarias*, 142
laevigata, *Microsynodontis*, 310
laevigatus, *Doras*, 177
laevigatus, *Bagrus*, 49
laevigatus, *Callichthys*, 127
laevigatus, *Microsynodontis*, 310
laevior, *Hisonotus*, 248
laevis, *Gogangra*, 395

- laevis*, *Platystacus*, 58
laeviuscula, *Loricaria*, 289
laeviuscula, *Pseudoloricaria*, 289
lagoensis, *Arius*, 31
lagoensis, *Chrysichthys*, 160
lagoichthys, *Loricaria gymnotogaster*, 297
lagoichthys, *Spatuloricaria*, 297
Laguvia, 382, 395
Laguvia manipurensis, 382
Laguvia ribeiroi, 401
Laguvia ribeiroi kapuri, 401
Laguvia shawi, 395, 402
Laguviini, 382
Laises, 358
Laises hexanema, 358
Laises longibarbis, 359
Laimumena, 431
Laimumena barbonica, 431, 432
Lais, 358
lais, *Kryptopterus*, 370
Laïs, 358
laïs, *Silurus*, 370
lakoi, *Aspidoras*, 109
laluchensis, *Rhamdia*, 198
lamani, *Amphilinus*, 22
Lamarrii, *Bagrus*, 106, 107
lambda, *Ictalurus*, 209
lamberti, *Corydoras*, 119
lamberti, *Microsynodus*, 310
Lambertia, 347
Lambertia atra, 347
Lambertichthys, 347
Lambertichthys ater sepikensis, 347
lamghur, *Silurus*, 373
lamina, *Loricaria*, 288
laminus, *Pseudohemiodon*, 288
Lamontichthys, 263
Lamontichthys filamentosus, 263
Lamontichthys llanero, 264
Lamontichthys maracaibero, 264
Lamontichthys stibaros, 264
lamottei, *Chiloglanis*, 306
lamottei, *Clarias*, 144
lampei, *Amphilinus*, 22
Lampiella, 217, 264
Lampiella gibbosa, 264
Lampiellini, 217
lampris, *Glyptothorax*, 391
lanceolata, *Hemiloricaria*, 246
lanceolata, *Loricaria*, 246
lanceolata, *Rineloricaria*, 246
lanceolatus, *Nemuroglanis*, 188
landanensis, *Tachysurus*, 57
landinga, *Trichomycterus*, 419
landoni, *Hemiancistrus*, 243
lanzhouensis, *Silurus*, 380
laosensis, *Glyptothorax*, 391
laplatae, *Hypostomus*, 255
laplatae, *Plecostomus*, 255
larai, *Bunocephalus*, 59
larnaudii, *Pangasius*, 326
Lasiancistrus, 264
Lasiancistrus anthrax, 288
Lasiancistrus brevispinis, 287
Lasiancistrus caucanus, 264
Lasiancistrus dumus, 289
Lasiancistrus guacharote, 265
Lasiancistrus guapore, 265
Lasiancistrus heteracanthus, 265
Lasiancistrus longispinis, 287
Lasiancistrus maracaiboensis, 265
Lasiancistrus mayoloi, 265
Lasiancistrus nationi, 223
Lasiancistrus nicoi, 289
Lasiancistrus saetiger, 265
Lasiancistrus schomburgkii, 265
Lasiancistrus scolymus, 265
Lasiancistrus tentaculatus, 265
Lasiancistrus tigris, 288, 289
Lasiancistrus volcanensis, 264
lata, *Loricaria*, 268
lateralis, *Pimeletropis*, 332
Lateralis, *Pimelodus pallidus* var., 210
Lateralis, *Silurus pallidus*, 210
lateristriga, *Pimelodella*, 192
lateristrigus, *Pimelodes*, 192
lateristrigus, *Pimelodus*, 192
laticauda, *Rhamdia*, 198, 199, 202
laticauda typhla, *Rhamdia*, 198
laticaudatus, *Amphilinus*, 22
laticaudus, *Pimelodus*, 198
laticeps, *Adelopeltis*, 107
laticeps, *Allabenchelys*, 137
laticeps, *Amblyceps*, 17
laticeps, *Arius*, 41
laticeps, *Chrysichthys*, 157
laticeps, *Clariallabes*, 137
laticeps, *Clarotes*, 160
laticeps, *Eutropius*, 364
laticeps, *Heterobranchus*, 151
laticeps, *Ituglanis*, 407
laticeps, *Loricaria*, 288
laticeps, *Olyra*, 17, 18
laticeps, *Pimelodella*, 192
laticeps, *Pimelodus*, 160
laticeps, *Plotosus*, 345
laticeps, *Pseudohemiodon*, 288
laticeps, *Schilbe*, 364
laticeps, *Silurus*, 151
laticeps, *Trichomycterus*, 407
laticeps australis, *Pimelodella*, 189
latidens, *Astroblepus*, 65
latidens, *Pygidium*, 420
latidens, *Trichomycterus*, 419
latifrons, *Ancistrus*, 222
latifrons, *Chaetostomus*, 219, 222
latifrons, *Hypostomus*, 255
latifrons, *Noturus*, 211
latifrons, *Synodontis gabonensis*, 315
latirostris, *Arius*, 47
latirostris, *Hypostomus*, 255
latirostris, *Loricaria*, 294
latirostris, *Neoarius*, 47
latirostris, *Plecostomus*, 255
latirostris, *Rineloricaria*, 294
latirostris, *Sorubim*, 342
latiscutatus, *Arius*, 31
latisoma, *Farlowella*, 239
latistriatum, *Pygidium*, 420
latistriatus, *Trichomycterus*, 420
latiura, *Dasyloricaria*, 233
latiura, *Loricaria filamentosa*, 233
latovittatus, *Cryptopterus*, 374
latus, *Corydoras*, 119
laucaensis, *Trichomycterus*, 420
laukidi, *Rhamdia*, 199
laurenti, *Pimelodella*, 192
lautus, *Noturus hildebrandi*, 213
lavetti, *Ameiurus*, 204
lavetti, *Ictalurus*, 204
layardi, *Arius*, 51
lazera, *Clarias*, 143
laeviuscula, *Loricaria*, 289
leerii, *Wallago*, 380, 381
lehmanni, *Rhamdia*, 202
leiakanthus, *Clarias*, 145
leiakanthus, *Hyalobagrus*, 92
leiakanthus, *Leiocassis*, 104
leiakanthus, *Ompok*, 371, 372
leiakanthus, *Pseudomystus*, 104
leiakanthus, *Wallago*, 371, 372
Leiarius, 334
Leiarius marmoratus, 334
Leiarius pictus, 334
leidyi, *Ameiurus*, 204
leidyi, *Ictalurus*, 204
leightonii, *Oxyloricaria*, 300
leightonii, *Sturisomatichthys*, 300
Leiocassis, 92
Leiocassis (Dermocassis) analis, 101
Leiocassis aculeatus, 92
Leiocassis albicularis, 104
Leiocassis albomarginatus, 101
Leiocassis bicolor, 104
Leiocassis brashnikowi, 100
Leiocassis brevicaudatus, 102
Leiocassis chaseni, 93
Leiocassis collinus, 443
Leiocassis crassilabris, 92
Leiocassis crassilabris macrops, 92
Leiocassis crassirostris, 92
Leiocassis ellenriederii, 104
Leiocassis fluviatilis, 85
Leiocassis fuscus, 103
Leiocassis hainanensis, 88
Leiocassis herzensteini, 92

Leiocassis hwanghoensis, 107
Leiocassis kaifensis, 107
Leiocassis leiacanthus, 104
Leiocassis longibarbus, 92
Leiocassis longirostris, 93
Leiocassis lui, 102
Leiocassis macropterus, 83
Leiocassis mahakamensis, 104
Leiocassis micropogon, 93
Leiocassis microps, 100
Leiocassis moeschii, 105
Leiocassis myersi, 104
Leiocassis nitidus, 82
Leiocassis omeihensis, 102
Leiocassis poecilopterus, 93
Leiocassis regani, 93
Leiocassis robustus, 104
Leiocassis saravacensis, 93
Leiocassis similis, 108
Leiocassis sinyanensis, 108
Leiocassis tenebris, 443
Leiocassis tenuifurcatus, 93
Leiocassis trilineatus, 103
Leiocassis ussuriensis, 101
Leiocassis virgatus, 101
leiopleura*, *Harttia, 240, 241
Leiosynodontis, 311
leiotetracephalus, *Arius*, 52
Leliella, 293, 294
lemai, *Rhamdella*, 196
lemairii, *Eutropius*, 364
lembesseensis, *Clariallabes melas*, 137
lemniscatus, *Pimelodus*, 213
lemoinei, *Arius*, 432
lentiginosa*, *Ariopsis, 33
lentiginosa, *Loricaria prolixa*, 286
lentiginosa*, *Proloricaria, 286
lentiginosus*, *Amphilius, 22
lentiginosus, *Doras*, 177
lentiginosus, *Tachisurus*, 33
leonensis*, *Malapterurus, 301
leoparda*, *Synodontis, 316
leopardina*, *Synodontis, 316
leopardinus, *Synodontis*, 316
leopardinus*, *Trachelyopterus, 80
leopardinus, *Trachycorystes*, 80
leopardum*, *Glanidium, 75
leopardus, *Centromochlus*
 (*Gephyromochlus*), 75
leopardus*, *Corydoras, 119
leopardus*, *Pseudacanthicus, 286
leopardus, *Stoneiella*, 286
leopardus, *Synodontis*, 316
Leporacanthicus, 265
Leporacanthicus galaxias, 265
Leporacanthicus heterodon, 266
Leporacanthicus joselimai, 266
Leporacanthicus triactis, 266
leporhinus, *Hemidoras*, 174

leporhinus, *Nemadoras*, 174
leptacanthus*, *Noturus, 213
Leptarius, 53
Leptarius dowii, 53
leptaspis, *Hexanematicthys*, 47
leptaspis*, *Neoarius, 47
Leptoplosternum, 128
Leptoplosternum altamazonicum,
 128
Leptoplosternum beni, 128
Leptoplosternum pectorale, 128
Leptoplosternum stellatum, 128
Leptoplosternum tordilho, 129
Leptoplosternum ucamara, 129
Leptoancistrus, 266
Leptoancistrus canensis, 266
Leptoancistrus cordobensis, 266
leptocassis, *Hexanematicthys*, 45
leptochilus*, *Hisonotus, 248
Leptodoras, 172
Leptodoras acipenserinus, 172
Leptodoras cataniai, 172
Leptodoras copei, 172
Leptodoras hasemani, 172
Leptodoras juriensis, 172
Leptodoras linnelli, 173
Leptodoras myersi, 173
Leptodoras nelsoni, 173
Leptodoras praelongus, 173
Leptodoras rogersae, 173
Leptodoras trimaculatus, 174
Leptoglanidinae, 20
Leptoglaninae, 20
Leptoglanis, 20, 25
Leptoglanis, 187
Leptoglanis bouilloni, 164
Leptoglanis brevis, 28
Leptoglanis brieni, 24
Leptoglanis camerunensis, 28
Leptoglanis dorae, 28
Leptoglanis essequibensis, 187
Leptoglanis flavomaculatus, 28
Leptoglanis mandevillei, 28
Leptoglanis wamiensis, 29
Leptoglanis xenognathus, 25
leptonema, *Silurus*, 374
leptonotacanthus*, *Arius, 34
Leptops, 215
Leptorhamdia, 187
Leptorhamdia essequibensis, 187
Leptorhamdia marmorata, 187
Leptorhamdia schultzi, 187
leptorhynchus*, *Helicophagus, 324
leptos, *Acentronichthys*, 180
leptosoma*, *Rhamdella, 196
leptostriatus*, *Microglanis, 441
leptura*, *Andersonia, 24
lepturum*, *Chaetostoma, 229
lepturus, *Chaetostomus*, 229
lepturus, *Cnidoglanis*, 346, 442
lepturus*, *Euristhmus, 346
lepturus*, *Oreoglanis, 397
lepturus*, *Xyliphius, 62
leptus, *Pimelodus*, 332
***Lerichei*, *Otolithus* (*Arius*)**, 434
leroyi, *Chimarrhoglanis*, 21, 23
lestradei, *Lophiobagrus*, 161, 162
leucofrenatus*, *Hisonotus, 248
leucofrenatus, *Otocinclus*, 248
leucomelas*, *Chaetostoma, 229
leucomelas, *Chaetostomus*, 229
leucomelas*, *Corydoras, 119
leucophasis, *Bagrus*, 96
leucophasis*, *Mystus, 96
leucopodus, *Silurichthys*, 378
Leucoptera*, *Pimelodus pallidus, 210
Leucoptera*, *Silurus pallidus, 210
leucorhynchus, *Akysis*, 16
leucorhynchus*, *Pseudobagarius, 16
leucostictus*, *Ancistrus, 222
leucostictus, *Chaetostomus*, 222
levequei*, *Chrysichthys, 157
levequei*, *Synodontis, 316
levi, *Copidoglanis*, 345
levi*, *Hypostomus, 255
levi, *Rhinelepis*, 255
lewi*, *Trichomycterus, 420
lexi*, *Hypostomus, 255
lexi*, *Plecostomus, 255
Leyvaichthys, 134
Leyvaichthys, 134
Leyvaichthys castaneus, 134, 407
liacanthus, *Clarias*, 145
lianae*, *Copionodon, 404
Liauchenoglanis, 161
Liauchenoglanis maculatus, 161
liberiensis, *Clarias*, 141
liberiensis, *Eutropius*, 364
libertatis, *Doras*, 174
lica, *Bagarius*, 383
lima*, *Hypostomus, 256
lima, *Loricaria*, 294
lima, *Plecostomus*, 256
lima*, *Rineloricaria, 294
lima, *Silurus*, 342
lima*, *Sorubim, 342
lima atropinnis, *Plecostomus*, 251
lima microlepidota, *Loricaria*, 295
lima var. *microlepidota*, *Loricaria*, 295
Limatulichthys, 266
Limatulichthys griseus, 266
Limatulichthys petleyi, 266
limbatus, *Bagrus*, 107
limbatus, *Choeroplotosus*, 346
limbatus*, *Plotosus, 108, 346, 350
limosus, *Pimelodus*, 215
limosus, *Plecostomus*, 253
limosus, *Pylodictis*, 215

limosus, *Silurus*, 215
limpok, *Kryptopterus*, 370
limpok, *Silurus*, 370
limulus, *Eurycheilichthys*, 236
linami, *Pimelodella*, 192
lindica, *Phractura*, 26
lindicus, *Clarias*, 141
lineata, *Nannorhamdia*, 185
lineatum, *Euglyptosternum*, 388
lineatus, *Imparfinis*, 185
lineatus, *Plotosus*, 350
lineatus, *Plotosus*, 351, 352
lineatus, *Silurus*, 350
lineolatus, *Ancistrus*, 222
lineopunctata, *Chaetostoma*, 229
lineopunctatum, *Chaetostoma*, 229
linnelli, *Leptodoras*, 173
Liobagrus, 19
Liobagrus andersoni, 19
Liobagrus anguillicauda, 19
Liobagrus formosanus, 19
Liobagrus kingi, 19
Liobagrus marginatoides, 19
Liobagrus marginatus, 19
Liobagrus mediadiposalis, 19
Liobagrus nantoensis, 19
Liobagrus nantoensis, 19
Liobagrus nigricauda, 20
Liobagrus obesus, 20
Liobagrus reinii, 19, 20
Liobagrus styani, 20
Liobagrus sugubii, 20
Liobagrus xiurenensis, 20
Liocassis, 92
Liocassis baramensis, 93
Liocassis breviceps, 103
Liocassis crassilabris, 92
Liocassis crassirostris, 92
Liocassis doriae, 93
Liocassis emarginatus, 102
Liocassis hosii, 93
Liocassis inornatus, 103
Liocassis longirostris, 92, 93
Liocassis merabensis, 93
Liocassis moeschii, 104
Liocassis naso, 108
Liocassis rugosus, 104
Liocassis saravakensis, 93
Liocassis siamensis, 104
Liocassis torosilabris, 108
Liocassis truncatus, 103
Liocassis vaillanti, 105
Liocassis hirsutus, 107
liocephalus, *Clarias*, 145
Liosomadoras, 75
Liosomadoras morrowi, 75
Liosomadoras oncinus, 75
lipophthalmus, *Doras (Oxydoras)*, 169
Lipopterichthys, 266

Lipopterichthys carrioni, 266, 267
Liposarcus, 290
Liposarcus altipinnis, 291
Liposarcus ambrosettii, 291
Liposarcus anisitsi, 291
Liposarcus disjunctivus, 291
Liposarcus jeansesianus, 292
Liposarcus scrophus, 292
Liposarcus varius, 292
liropus, *Tachysurus*, 41
listrorhinos, *Apistoloricaria*, 226
Listrura, 408
Listrura boticario, 408
Listrura campani, 408
Listrura nematopteryx, 408
Listrura picinguabae, 444
Listrura tetraradiata, 408
Lithodoras, 173
Lithodoras dorsalis, 173
lithogaster, *Doras*, 173
Lithogeneinae, 217
Lithogenes, 217, 267
Lithogenes valencia, 267
Lithogenes villosum, 267
lithoides, *Lithoxus*, 267
lithophilus, *Parasilurus*, 380
lithophilus, *Silurus*, 380
lithostoma, *Pangasius*, 326
Lithoxancistrus, 286
Lithoxancistrus orinoco, 286, 287
Lithoxina, 217
Lithoxus, 217, 267
Lithoxus (Paralithoxus) pallidimaculatus, 267
Lithoxus (Paralithoxus) planquettei, 267
Lithoxus (Paralithoxus) surinamensis, 268
Lithoxus boujardi, 267
Lithoxus bovallii, 267
Lithoxus lithoides, 267
Lithoxus pallidimaculatus, 267
Lithoxus planquettei, 267
Lithoxus stocki, 268
Lithoxus surinamensis, 268
lithurgicus, *Ancistrus*, 222
littoralis, *Hoplosternum*, 127
littoralis, *Hoplosternum*, 128
littoralis, *Callichthys*, 127
lituratus, *Ancistrus*, 290, 291
lituratus, *Pterygoplichthys*, 291
lividus, *Silurus*, 204, 205
lividus, *Fuscatus*, *Silurus*, 205
lividus var. *Fuscatus*, *Silurus*, 205
livrée, *Pimelode*, 213
llanero, *Lamontichthys*, 264
loangwensis, *Clarias*, 151
loborhynchos, *Chaetostoma*, 227, 229

lombarderoi, *Xyliphius*, 62
lonah, *Bagrus*, 391
lonah, *Glyptothorax*, 391, 394
longianalis, *Clupisoma*, 357
longianalis, *Platytropius*, 357
longibarbus, *Trichomycterus*, 420
longibarbis, *Arius*, 334
longibarbis, *Bathyclarias*, 136
longibarbis, *Chrysichthys*, 157
longibarbis, *Chrysobagrus*, 157
longibarbis, *Clarias*, 136, 138
longibarbis, *Clarias (Allabenchelys) dumerili*, 138
longibarbis, *Dianema*, 127
longibarbis, *Dinotopterus*, 136
longibarbis, *Laides*, 359
longibarbis, *Pangasius*, 359
longibarbus, *Leiocassis*, 92
longicauda, *Allabenchelys*, 137
longicauda, *Clariallabes*, 137
longicauda, *Euchiloglanis*, 399
longicauda, *Glyptothorax*, 391
longicauda, *Imparfinis*, 185
longicauda, *Pareuchiloglanis*, 399
longicauda, *Phractura*, 26
longicauda, *Pimelodus (Rhamdia)*, 185
longicauda, *Rineloricaria*, 294
longicaudata, *Olyra*, 99
longicaudatus, *Olyra*, 99
longicaudatus, *Clariallabes*, 150
longicephalus, *Tachisurus*, 39
longicephalus, *Tachisurus*, 39
longiceps, *Astroblepus*, 65
longiceps, *Auchenoglanis*, 163
longiceps, *Clarias*, 143
longiceps, *Dolichamphilius*, 24
longiceps, *Osteogeneiosus*, 51
longiceps, *Parauchenoglanis*, 163
longidorsalis, *Chrysichthys*, 157
longidorsalis nyongensis,
Chrysichthys, 158
longifilis, *Akysis*, 442
longifilis, *Arges*, 65
longifilis, *Astroblepus*, 65
longifilis, *Callichthys*, 129
longifilis, *Chrysichthys*, 155
longifilis, *Copidoglanis*, 349
longifilis, *Eutropius*, 361
longifilis, *Heterobranchus*, 151
longifilis, *Hypophthalmus*, 334
longifilis, *Parailia*, 360
longifilis, *Pareutropius*, 361
longifilis, *Pimelodus*, 338
longimanus, *Ancistrus*, 291
longimanus, *Auchenipterichthys*, 71
longimanus, *Auchenipterus*, 71
longimanus, *Pseudeutropius*, 362
longinema, *Glyptothorax*, 394
longior, *Chasmocranus*, 182, 183

- longior, Clarias**, 145
longipectoralis, *Heteropneustes*, 152
longipinna, Harttia, 241
longipinnis, Bariancistrus, 227
longipinnis, Chrysichthys, 157
longipinnis, *Euanemus*, 74
longipinnis, *Gephyroglanis*, 157
longipinnis, *Hemiancistrus*, 227
longipinnis, Rhamdella, 196
longipinnis, Trichogenes, 414
longiradiatus, Hypostomus, 256
longiradiatus, *Plecostomus*, 256
longirostris, Amphilius, 22
longirostris, *Anoplopterus*, 22
longirostris, *Corydoras melanistius*, 113
longirostris, Leiocassis, 93
longirostris, *Liocassis*, 92, 93
longirostris, Parakysis, 16
longirostris, Parotocinclus, 282
longirostris, Synodontis, 316
longispinis, *Elurichthys*, 38
longispinis, *Doras*, 170
longispinis, *Lasiancistrus*, 287
longispinis, Pseudancistrus, 287
longispinis, Synodontis, 316
longispinis, *Synodontis Batesi*, 316
longirostris, Nkondobagrus, 99
longiuscula, Rhamdella, 196
longiangensis, Glyptothorax, 391
longus, *Parasilurus asotus*, 379
lopezi, Chasmocranus, 183
Lophiobagrus, 161
Lophiobagrus aquilus, 161
Lophiobagrus asperispinus, 161
Lophiobagrus brevispinis, 161
Lophiobagrus cyclurus, 161
Lophiobagrus lestradei, 161, 162
lophioides, *Chaca*, 135, 136
Lophiosilurus, 353
Lophiosilurus alexandri, 353
lophius, *Amiurus*, 204
lophophanes, Otothyris, 276
lophophanes, *Rhinelepis*, 276
loppei, *Synodontis*, 318
loretoensis, Corydoras, 119
Loricaria, 216, 268
Loricaria (Loricariichthys) fallax, 245
Loricaria accipenser, 270
Loricaria acuta, 269
Loricaria altipinnis, 245
Loricaria amazonica, 270
Loricaria anus, 269
Loricaria apeltogaster, 268
Loricaria apeltogaster amazonum, 288
Loricaria apeltogaster var. *amazonum*, 288
Loricaria aurea, 298
Loricaria barbata, 298, 299
Loricaria beni, 245
Loricaria bransfordi, 240
Loricaria brevirostris, 299
Loricaria brunnea, 269
Loricaria cacerensis, 245
Loricaria cadeae, 294
Loricaria capetensis, 233
Loricaria caquetae, 296
Loricaria carinata, 268
Loricaria cashibo, 270
Loricaria castanea, 270
Loricaria catamarcensis, 294
Loricaria cataphracta, 268, 270
Loricaria cirrhosa, 268, 296
Loricaria clavipinna, 268
Loricaria commersonoides, 278
Loricaria cubataonis, 294
Loricaria curvispina, 296
Loricaria Dentata, 268
Loricaria devincenzi, 288
Loricaria dura, 268
Loricaria Eigenmanni, 245
Loricaria Evansii, 296
Loricaria felipponei, 294
Loricaria filamentosa, 233, 234
Loricaria filamentosa latiura, 233
Loricaria filamentosa seminuda, 234
Loricaria fimbriata, 297
Loricaria flava, 259
Loricaria frenata, 299
Loricaria griseus, 266
Loricaria gymnogaster, 297
Loricaria gymnogaster lagoichthys, 297
Loricaria henselii, 294
Loricaria hoehnei, 245
Loricaria jaraguensis, 294
Loricaria jubata, 246
Loricaria kneri, 299
Loricaria konopickyi, 246
Loricaria kronei, 294
Loricaria labialis, 270
Loricaria laeviuscula, 289
Loricaria laeviuscula, 289
Loricaria lamina, 288
Loricaria lanceolata, 246
Loricaria lata, 268
Loricaria laticeps, 288
Loricaria latirostris, 294
Loricaria lima, 293, 294
Loricaria lima microlepidota, 295
Loricaria lima var. *microlepidota*, 295
Loricaria macrodon, 227
Loricaria macromystax, 293
Loricaria macrops, 293
Loricaria maculata, 269, 270
Loricaria magdalena, 246
Loricaria melanoptera, 252
Loricaria microdon, 271
Loricaria microlepidogaster, 295
Loricaria nickeriensis, 269
Loricaria nigricauda, 246
Loricaria nudirostris, 271
Loricaria nudiventris, 297
Loricaria panamensis, 299
Loricaria parahemiodon, 270
Loricaria pareiacantha, 295
Loricaria parnahybae, 269
Loricaria parva, 246
Loricaria paulina, 294
Loricaria phoxocephala, 246
Loricaria piauhiae, 269
Loricaria piracicabae, 269
Loricaria platystoma, 233
Loricaria platyura, 246
Loricaria prolixa, 286
Loricaria prolixa lentiginosa, 286
Loricaria puganensis, 297
Loricaria punctata, 266, 292
Loricaria rostrata, 245, 298, 300
Loricaria scolopacina, 237
Loricaria setifera, 268
Loricaria simillima, 269
Loricaria sneiderni, 246
Loricaria spinosae, 286
Loricaria Spixii, 270
Loricaria Steinbachi, 263
Loricaria steindachneri, 295
Loricaria stewarti, 247
Loricaria strigilata, 295
Loricaria Stübelii, 271
Loricaria submarginatus, 246
Loricaria tefficana, 247
Loricaria thrissoceps, 295
Loricaria tucumanensis, 269
Loricaria tuyrensis, 234
Loricaria uracantha, 240
Loricaria uracantha rupestris, 240
Loricaria Valenciennesii, 247
Loricaria variegata, 232, 233
Loricaria variegata venezuelae, 233
Loricaria vetula, 278
loricariformis, Harttia, 240, 241
Loricariichthyna, 217
Loricariichthys, 217, 269
Loricariichthys acutus, 269
Loricariichthys anus, 269
Loricariichthys brunneus, 269
Loricariichthys cashibo, 270
Loricariichthys castaneus, 270
Loricariichthys chanjoo, 270
Loricariichthys derbyi, 270
Loricariichthys edentatus, 270
Loricariichthys hauxwelli, 270
Loricariichthys labialis, 270
Loricariichthys maculatus, 270
Loricariichthys melanocheilus, 271
Loricariichthys melini, 246

Loricariichthys microdon, 271
Loricariichthys nudirostris, 271
Loricariichthys parnahybae, 266
Loricariichthys platymetopon, 271
Loricariichthys rostratus, 271
Loricariichthys stuebelii, 271
Loricariichthys ucayalensis, 271
Loricariidae, 216
loricatus, *Callichthys*, 111
loricatus, *Doras*, 167
loweae, *Bathyclarias*, 149
loweae, *Dinotopterus*, 149
lowei, *Gephyroglanis*, 158
loxozonus, *Corydoras*, 119
lualae, *Clarias*, 142
lubosicus, *Bagrus*, 84
lucenai, *Trachelyopterus*, 80
Luceri, *Platystoma*, 342
Luciopimelodinae, 329
Luciopimelodus, 329, 334
Luciopimelodus pati, 334
lucipinnis, *Synodontis*, 445
luderwaldti, *Pseudancistrus*, 280
lufirae, *Chiloglanis*, 306
lufirae, *Synodontis*, 316
lui, *Leiocassis*, 102
luigiana, *Rhamdia*, 199
lukugae, *Chiloglanis*, 306
lukugae, *Phractura*, 26
lumholzti, *Cryptopterus*, 370
lumholzti, *Kryptopterus*, 370
lundbergi, *Prietella*, 215
lundbergi, *Typhlobelus*, 426
luniscutis, *Arius*, 36
luniscutis, *Aspistor*, 36
lupus, *Ictalurus*, 209, 211
lupus, *Pimelodus*, 209
lurida, *Ageniosus (Silonia)*, 366
luridus, *Macrones*, 90
luridus, *Pachypterus*, 327
luteomaculatus, *Hypostomus*, 256
luteomaculatus, *Plecostomus*, 256
lutescens, *Pimelodus*, 215
luteus, *Hypostomus*, 256
luteus, *Noturus*, 212
luteus, *Oloplotosus*, 349
luteus, *Plecostomus*, 256
Lütkeni, *Platystoma*, 344
Lütkeni, *Plecostomus*, 251
luvur, *Silurus*, 431
lynx, *Pimelodus*, 204
lyra, *Oxylicaria*, 299
lyra, *Physopyxis*, 176
lyra, *Sturisoma*, 299
lyriformis, *Agmus*, 60
M
mabusi, *Chrysichthys*, 157
macarenensis, *Phenacorhamdia*, 188,
 189

maccus, *Panaqolus*, 277
maccus, *Panaque*, 277
macgrewi, *Ameiurus*, 204
macgrewi, *Ictalurus*, 205
machadoi, *Genidens*, 44
machadoi, *Gladioglanis*, 183
machadoi, *Platyclarias*, 152
machadoi, *Plectrochilus*, 411
machadoi, *Tachysurus*, 44
machiquense, *Chaetostoma*, 229
machiquensis, *Chaetostoma*, 229
macilenta, *Denticetopsis*, 133
macilentus, *Hemicetopsis*, 133
maclareni, *Clarias*, 145
macconnelli, *Ameiurus*, 211
macracanthus, *Arius*, 36
macracanthus, *Bagrichthys*, 82
macracanthus, *Bagroides*, 82
macracanthus, *Centromochlus*, 73
macracanthus, *Clarias*, 143
macracanthus, *Pseudobagrichthys*, 82
macracanthus, *Rita*, 106
macraei, *Hatcheria*, 405
macraei, *Thrichomycterus*, 405
macrepipterus, *Synodontis*, 314
macrocephala, *Nannorhamdia*, 192
macrocephala, *Pimelodella*, 192
macrocephalus, *Ariodes*, 51
macrocephalus, *Arius*, 51
macrocephalus, *Atopochilus*, 303
macrocephalus, *Clarias*, 145
macrocephalus, *Clarotes*, 160
macrocephalus, *Cnidoglanis*, 346
macrocephalus, *Hemipimelodus*, 41
macrocephalus, *Kryptopterichthys*, 370
macrocephalus, *Kryptopterus*, 370
macrocephalus, *Osteogeneiosus*, 51
macrocephalus, *Plotosus*, 346
macrocephalus, *Silurus*, 341
macrochir, *Belodontichthys*, 368
macrodon, *Brochiloricaria*, 227
macrodon, *Loricaria*, 227
macrodon, *Synodontis*, 313
macromaculatus, *Glyptothorax*, 391
macromycter, *Typhlobelus*, 426
macromystax, *Clarias*, 145
macromystax, *Loricaria*, 293
macromystax, *Rhadinoloricaria*, 293
macronema, *Akysis*, 16
macronema, *Chlarias angolensis*, 141
macronema, *Heterobranchus*, 151
macronema, *Hypselobagrus*, 97
macronema, *Pangasius*, 327
macronema, *Pseudorhamdia*, 337
macronema, *Silurodes*, 372
macronema, *Silurus*, 371, 372
macronemus, *Bagrus*, 37, 97
macronemus, *Bagrus*, 93, 97
macronemus, *Oreoglanis*, 397
macronemus, *Pseudobagarius*, 16
Macrones, 85, 106
Macrones (Hemibagrus) filamentus, 88
Macrones (Hemibagrus) wyckiooides, 91
Macrones (Leiocassis) brashnikowi, 100
Macrones (Liocassis) taeniatus, 102
Macrones (Pseudobagrus) tenuis, 103
Macrones aleppensis, 97
Macrones argentivittatus, 100
Macrones armatus, 94
Macrones baramensis, 87
Macrones bimaculatus, 94
Macrones bleekeri, 89, 94
Macrones Blythii, 85
Macrones bo, 88
Macrones bongan, 87
Macrones chinensis, 88
Macrones colvillii, 97
Macrones corsula, 90
Macrones Dayi, 85
Macrones elongatus, 88
Macrones fortis, 88
Macrones fortis capitulum, 88
Macrones fortis var. capitulum, 88
Macrones herzensteini, 92
Macrones howong, 88
Macrones kajan, 88
Macrones luridus, 90
Macrones medianalis, 102
Macrones merianiensis, 85
Macrones microphthalmus, 90
Macrones montanus dibrugarensis, 96
Macrones montanus var.
 dibrugarensis, 96
Macrones peguensis, 90
Macrones pluriradiatus, 90
Macrones pratti, 102
Macrones pulcher, 97
Macrones rufescens, 94, 97
Macrones sinensis, 165
Macrones vittata, 106
Macronichthys, 106
Macronoides, 85, 87
Macronoides wilsoni, 87
macronotacanthus, *Arius*, 47
macronotus, *Batasio*, 85
macrophthalmalma, *Synodontis*, 316
macrophthalmalma, *Xenocara*, 222
macrophthalmalus, *Ancistrus*, 222
macrophthalmalus, *Hypostomus*, 256
macrophthalmalus, *Hypostomus*
 pseudohemiurus, 256
macrophthalmalus, *Plotosus*, 349
macrophthalmalus, *Synodontis*, 316
macropogon, *Chrysichthys*, 160
macrops, *Chaetostomus*, 243

- macrops*, *Chrysichthys*, 155
macrops, *Hemiancistrus*, 243
macrops, *Henonemus*, 406
macrops, *Hypostomus*, 256
macrops, *Leiocassis crassilabris*, 92
macrops, *Loricaria*, 293
macrops, *Plecostomus*, 256
macrops, *Ricola*, 293
macrops, *Stegophilus*, 406
macrops, *Synodontis*, 316
Macropterobagrus, 87
Macropteronotus, 138
Macropteronotus charmuth, 138, 148
Macropteronotus fuscus, 142
Macropteronotus hexacicinus, 148
Macropteronotus jagur, 140
Macropteronotus magur, 140
macropterus, *Exostoma*, 398
macropterus, *Amblyceps*, 18
macropterus, *Bagrichthys*, 82
macropterus, *Bagroides*, 82, 83
macropterus, *Bagroides*, 82
macropterus, *Bagrus bayad*, 83
macropterus, *Calophysus*, 332
macropterus, *Chiloglanis*, 306
macropterus, *Chrysichthys*, 157
macropterus, *Corydoras*, 130
macropterus, *Hemibagrus*, 87, 89, 91
macropterus, *Leiocassis*, 83
macropterus, *Pareuchiloglanis*, 399
macropterus, *Pimelodes*, 332
macropterus, *Porcus bayad*, 83
macropterus, *Oreoglanis*, 398
macropterus, *Scleromystax*, 130
macropthalmos, *Eutropius*, 361, 362
macropthalmos, *Proeutropiichthys*, 362
macropthalmos, *Pseudosilurus*, 374
macrorhynchus, *Hemipimelodus*, 32
macrospila, *Piramutana*, 339
macrospilus, *Otocinclus*, 275
macrosteus, *Corydoras*, 112
macrostigma, *Synodontis*, 317
macrostoma, *Auchenoglanis*, 154
macrostoma, *Synodontis*, 317
macrostomus, *Anaspidoglanis*, 154
macrostomus, *Silurus (Callichirus)*, 381
macroteronema, *Cetopsis*, 133
macrotis, *Chrysichthys*, 157
Macrotocinclus, 271
Macrotocinclus affinis, 271
Macrotocinclus flexilis, 271
macrotrema, *Euchiloglanis*, 399
macrotremus, *Pareuchiloglanis*, 399
macrura, *Phractura*, 27
macruropterygius, *Arius*, 55
macrurus, *Clarias*, 148
macturki, *Pimelodella*, 192
maculata, *Cteniloricaria*, 233
maculata, *Loricaria*, 269, 270
maculata, *Parasturisoma*, 233
maculata, *Wertheimeria*, 179
maculatum, *Glyptosternon*, 387
maculatum, *Parexostoma*, 387
maculatus, *Ancistrus*, 222
maculatus, *Arius*, 34, 35
maculatus, *Chaetostomus*, 228
maculatus, *Chaetostomus (Ancistrus) cirrhosus*, 222, 228
maculatus, *Chaetostomus (Ancistrus) cirrhosus* var., 222, 228
maculatus, *Corydoras*, 122
maculatus, *Doras*, 177
maculatus, *Liauchenoglanis*, 161
maculatus, *Loricariichthys*, 270
maculatus, *Nothoglanidium*, 161
maculatus, *Parastegophilus*, 410
maculatus, *Pimelodus*, 336, 338, 345
maculatus, *Pimelodus*, 209
maculatus, *Pimelodus (Bagrus)*, 345
maculatus, *Silurus*, 35, 207, 209
maculatus, *Stegophilus*, 410
maculatus, *Synodontis*, 318
maculatus, *Trichomycterus*, 415
maculatus, *Wallago*, 381
maculatus, *Erythroptera*, *Silurus*, 209
maculatus var. *Erythroptera*, *Silurus*, 209
maculicauda, *Otocinclus*, 281, 283
maculicauda, *Parotocinclus*, 283
maculifer, *Corydoras*, 119
maculipinna, *Synodontis*, 320
maculipinnis, *Akysis*, 14
maculipinnis, *Hisonotus*, 248
maculipinnis, *Otocinclus*, 248
maculocephala, *Cascadura*, 127, 128
maculosus, *Aspidoras*, 109
maculosus, *Auchenipterus*, 79
maculosus, *Auchenoglanis*, 163
maculosus, *Synodontis*, 311, 318, 321
maculosus, *Trachelyopterus coriaceus*, 79
macunaima, *Ituglanis*, 407
macushi, *Hypostomus*, 256
macuspanensis, *Rhamdia*, 199
madagascaricensis, *Arius*, 35
madeirae, *Plecostomus*, 225
madeirensis, *Ageneiosus*, 69
madraspatanum, *Glyptosternum*, 391
madraspatanus, *Glyptothorax*, 388, 391
maesi, *Amphilinus*, 22
maesotensis, *Eretistes*, 384
magatensis, *Arius*, 52
Magdalena, *Auchenipterus*, 79
magdalena, *Branchioica*, 411
magdalena, *Hemiloricaria*, 246
magdalena, *Hoplosternum*, 128
Magdalena, *Loricaria*, 246
magdalena, *Trachycorystes*, 79
magdalena, *Xyliphius*, 62
magnus, *Chrysichthys*, 157
magoi, *Ageneiosus*, 69
magoi, *Brachyglanis*, 180
magoi, *Pygidianops*, 412
magur, *Macropteronotus*, 140
mahakamensis, *Acrochordonichthys*, 12
mahakamensis, *Leiocassis*, 104
mahakamensis, *Pangasius*, 327
mahakamensis, *Pseudomystus*, 104
mahengeensis, *Chrysichthys*, 157
major, *Akysis*, 391
major, *Glyptothorax*, 391
major, *Hemibagrus*, 89
majusculus, *Bagrichthys*, 83
malabarbai, *Microglanis*, 354
Malabarica, *Hara*, 96
malabaricus, *Arius*, 35
Malabaricus, *Bagrus*, 96
malabaricus, *Mystus*, 96
malabaricus, *Ompok*, 372
malabaricus, *Silurus*, 372
Malacobagrus, 330
Malacoglanis, 408
Malacoglanis gelatinosus, 408
malacops, *Ancistrus*, 222
malacops, *Chaetostomus*, 222
Malacofterurus, 301
malaisei, *Glyptosternon*, 387
Malapteruridae, 300
Malapterurini, 300
Malapterurus, 300, 301
Malapterurus affinis, 301
Malapterurus barbatus, 301
Malapterurus beninensis, 301
Malapterurus cavalliensis, 301
Malapterurus coila, 356
Malapterurus electricus, 301
Malapterurus electricus oguensis, 302
Malapterurus electricus, var.
ogooensis, 302
Malapterurus electricus var. *oguensis*, 302
Malapterurus gossei, 302
Malapterurus leonensis, 301
Malapterurus melanochir, 301
Malapterurus microstoma, 301
Malapterurus microstomus, 301
Malapterurus minjiriya, 302
Malapterurus monsembeensis, 302
Malapterurus murrayi, 302
Malapterurus occidentalis, 302
Malapterurus oguensis, 302
Malapterurus polli, 302
Malapterurus punctatus, 302
Malapterurus shirensis, 302

- Malapterurus stiassnyae*, 301
Malapterurus tanganyikaensis, 302
Malapterurus tanoensis, 302
Malapterurus teugelsi, 302
Malapterurus thysi, 301
Malapterurus zambeziensis, 302
Malapterus, 301
Malapterus, 301, 356
Malapterus (Ailia) bengalensis, 356
Malapturus, 301
malaris, *Clarias*, 143
malarimo, *Platysilurus*, 340
maldonadoi, *Bullockia*, 404
Maldonadoi, *Hatcheria*, 403, 404
malignus, *Plotosus*, 351
Malopterus, 301
mamore, *Corydoras*, 120
mamoré, *Corydoras*, 120
managuensis, *Pimelodus*, 199
mancoi, *Astroblepus*, 65
mandevillei, *Atopochilus*, 304
mandevillei, *Leptoglanis*, 28
mandevillei, *Pareutropius*, 361
mandevillei, *Zaireichthys*, 28
mandibularis, *Eutropius*, 364
mandibularis, *Schilbe*, 364
mangois, *Amblyceps*, 18
mangois, *Pimelodus*, 18
mangrullo, *Silurus*, 341
mangurus, *Pimelodus*, 355
mangurus, *Pseudopimelodus*, 355
manillensis, *Arius*, 35
manillensis, *Cephalocassis*, 41
Manillensis, *Pimelodus*, 41
manipurensis, *Glyptothorax*, 391
manipurensis, *Laguvia*, 382
maniradii, *Sorubim*, 342
manjong, *Arius*, 55
manni, *Pteroglanis*, 343
manni, *Synodontis*, 317
manyangae, *Allabenchelys*, 137
manyangae, *Clariallabes*, 137
mapale, *Cathorops*, 40
maquinensis, *Rineloricaria*, 294
maracaibero, *Lamontichthys*, 264
maracaiboensis, *Hemiancistrus*, 243
maracaiboensis, *Lasiancistrus*, 265
maracaiboensis, *Pygidium banneui*, 420
maracaiboensis, *Trichomycterus*, 420
maracasae, *Ancistrus*, 223
marapoama, *Otothyropsis*, 276
marcapatae, *Chaetostoma*, 229
marcapatae, *Chaetostomus*, 229
margaritifer, *Hypostomus*, 256
margaritifer, *Plecostomus*, 256
margaritifer butantanis, *Plecostomus*, 256
marginata, *Pimelodus pallidus* var., 210
marginata, *Taunayia*, 203
marginatoides, *Amblyceps*, 19
marginatoides, *Liobagrus*, 19
marginatum, *Chaetostoma*, 229
marginatus, *Amblyceps*, 19
marginatus, *Chaetostomus*, 229
marginatus, *Hypophthalmus*, 333
marginatus, *Liobagrus*, 19
marginatus, *Noturus*, 213
marginatus, *Pimelodus*, 206
marginatus, *Plotosus*, 350
marginatus, *Silurus pallidus*, 210
marginatus, *Silurus pallidus* var., 210
marginatus atrorus, *Schilbeodes*, 213
mariae, *Astroblepus*, 65
mariae, *Cyclopium*, 65
mariae, *Oloplotosus*, 349
mariae, *Otocinclus*, 275
mariaelenae, *Farlowella*, 238
mariai, *Imparales*, 188
mariai, *Nemuroglanis*, 188
marinus, *Bagre*, 37
marinus, *Silurus*, 37, 38, 44
marlieri, *Chiloglanis*, 306
marmorata, *Leptorhamdia*, 187
marmorata, *Synodontis*, 317
marmoratum, *Trachypoma*, 404
marmoratus, *Ageneiosus*, 69
marmoratus, *Amphilius*
grammatophorus, 23
marmoratus, *Arges*, 65
marmoratus, *Astroblepus*, 65
marmoratus, *Corydoras*, 122
marmoratus, *Helogenes*, 134
marmoratus, *Doras*, 171
marmoratus, *Franciscodoras*, 171
marmoratus, *Leiarius*, 334
marmoratus, *Pimelodus*, 206
marmoratus, *Schilbe*, 364
marmoratus, *Sciades*, 334
marmoratus, *Silurichthys*, 378
marmoratus, *Synodontis*, 317
marmoratus, *Trichomycterus*, 415
marmoratus truncatus, *Synodontis*, 317
marmorescens, *Chaetostoma*, 229
marpus, *Clarias*, 148
marquesi, *Ageneiosus*, 70
marthae, *Brachyrhamdia*, 181
marthae, *Rhamdia*, 181
martinezii, *Pimelodella*, 193
martini, *Acestridium*, 218
martini, *Ancistrus*, 223
martini, *Ancistrus triradiatus*, 223
martini, *Farlowella*, 238
Martyi, *Rhamdia sebae*, 202
mastersi, *Arius*, 45
mastersi, *Hexanematichthys*, 45
Mastiglanis, 187
Mastiglanis asopos, 187
mathisoni, *Zungaro*, 356
matthesi, *Synodontis*, 317
mattogrossensis, *Ancistrus*, 300
mattogrossensis, *Ancistrus*, 300
maurus, *Bagrus*, 158
maurus, *Chrysichthys*, 157, 158, 160
maydelli, *Hemibagrus*, 89
maydelli, *Mystus (Mystus)*, 89
maydeni, *Noturus*, 441
mayoloi, *Hemiancistrus*, 264
mayoloi, *Lasiancistrus*, 265
mazatlana, *Netuma*, 54
mboycy, *Trichomycterus*, 420
mbozi, *Chiloglanis*, 306
McCaskei, *Ichthaelurus*, 204
Medemichthys, 188
Medemichthys guayaberensis, 188
mediadiposalis, *Liobagrus*, 19
medianalis, *Macrones*, 102
medianalis, *Pseudobagrus*, 102
medians, *Ancistrus*, 242, 243
medians, *Hemancistrus*, 243
mediobarbis, *Neosilurus*, 347
meeki, *Amiurus*, 209
meeki, *Pimelodella*, 193
meesi, *Brachyrhamdia*, 181
megacephalus, *Chaetostomus*, 244
megacephalus, *Hemiancistrus*, 244
Megalancistrus, 272
Megalancistrus barrae, 272
Megalancistrus parananus, 272
Megalechis, 129
Megalechis picta, 129
Megalechis thoracata, 129
Megalocentor, 409
Megalocentor echthrus, 409
Megalodoras, 173
Megalodoras guayoensis, 173
Megalodoras irwini, 173, 174
Megalodoras paucisquamatus, 173
Megalodoras uranoscopus, 174
Megalonema, 335
Megalonema argentina, 335
Megalonema pauciradiatum, 335
Megalonema platanum, 335
Megalonema platycephalum, 335
Megalonema platycephalum
psammium, 335
Megalonema psammium, 335
Megalonema punctatum, 339
Megalonema rhabdostigma, 338
Megalonema robustum, 339
Megalonema xanthum, 335
megalops, *Centromochlus*, 73
megalops, *Iheringichthys*, 334
megalops, *Pimelodella*, 193
megalops, *Pimelodus*, 210

- megalops*, *Pseudeutropius*, 362
megaloptyx, *Hemiancistrus*, 244
meglostomus, *Ancistrus*, 223
meglura, *Pimelodella*, 193
megapogon, *Clarias*, 141
megastomus, *Plotosus*, 345, 346
megistus, *Ernstichthys*, 60
megistus, *Hoplomyzon*, 60
meizospilos, *Hemiancistrus*, 244
mekongensis, *Hemisilurus*, 368
mekongensis, *Pangasius*, 327
meladerma, *Clarias*, 145
melampterus, *Callichthys*, 127, 128
melanistius, *Corydoras*, 120
melanistius brevirostris, *Corydoras*, 114
melanistius longirostris, *Corydoras*, 113
melanocheilus, *Loricariichthys*, 271
melanochir, *Arius*, 41
melanochir, *Cephalocassis*, 41
melanochir, *Malapterurus*, 301
Melanodactylus, 155
melanoderma, *Clarias*, 146
melanodermatum, *Steindachneridion*, 343
melanogaster, *Acrochordonichthys*, 13
melanogaster, *Pimelodus*, 12, 13
melanopogon, *Ageneiosus*, 68
melanopogon, *Clarias*, 145
melanops, *Tridens*, 426
melanoptera, *Gagata*, 386
melanoptera, *Loricaria*, 252
melanoptera, *Synodontis*, 317
melanopterum, *Glanidium*, 75
melanopterus, *Bagroides*, 83
melanopterus, *Gagata*, 386
melanopterus, *Synodontis*, 317
melanopterus, *Xyliphius*, 62
melanopterygius, *Arius*, 56
melanopus, *Arius*, 40
melanopus, *Cathorops*, 40
melanostictus, *Synodontis*, 318
melanostictus ituri, *Synodontis*, 315
melanostictus var. *ituri*, *Synodontis*, 315
melanotaenia, *Corydoras*, 120
melanurus, *Batrochoglanis*, 353
melanurus, *Pachypterus*, 358
melanurus, *Silurus cerulescens*, 209
melanurus, *Silurus cerulescens*, var., 209
melaphareus, *Entomocorus*, 74
melapterus, *Bagroides*, 83
melapterus, *Bagroides*, 83
melas, *Ameiurus*, 205
melas, *Ancistrus*, 220
melas, *Bagrus*, 95
melas, *Brachyglanis*, 181
melas, *Bunocephalus*, 58
melas, *Clariallabes*, 137
melas, *Clarias*, 137
melas, *Pimelodus*, 205
melas, *Silurus*, 205
melas lembesseensis, *Clarias*, 137
melasoma, *Clarias*, 146
meleagris, *Hypostomus*, 256
meleagris, *Plecostomus*, 256
melini, *Corydoras*, 120
melini, *Hemiloricaria*, 246
melini, *Loricariichthys*, 246
Mellandi, *Clarias*, 146
membranacea, *Hemisynodontis*, 309
membranaceus, *Pimelodus*, 309
mendozensis, *Silvinichthys*, 413
mendozensis, *Trichomycterus*, 413
menezesi, *Aspidoras*, 109
menezesi, *Auchenipterus*, 72
menezesi, *Pyxiloricaria*, 292
menoda, *Hemibagrus*, 89
menoda, *Pimelodus*, 89
menoni, *Fajumia*, 429
mentalis, *Eutropius*, 364
mento, *Silurus*, 380
merabensis, *Liocassis*, 93
meraukensis, *Copidoglanis*, 351
meraukensis, *Porochilus*, 351
mercatoris, *Arius*, 31
merevari, *Harttia*, 241
merianensis, *Batasio*, 85
merianensis, *Macrones*, 85
meridae, *Trichomycterus*, 420
meridionalis, *Akysis*, 17
meridionalis, *Amiurus*, 208
meridionalis, *Bagrus*, 84
meridionalis, *Corydoras*, 116
meridionalis, *Pseudobagarius*, 17
meridionalis, *Silurus*, 380
meridionalis, *Silurus soldatovi*, 380
Merodontotus, 330
Merodontotus tigrinus, 330, 331
merus, *Glyptothonax*, 390
mesembrinus, *Diplomystes*, 166
mesembrinus, *Diplomystes viedmensis*, 166
mesembrinus, *Olivaichthys*, 166
mesops, *Bagrus*, 53
metae, *Corydoras*, 120
metae, *Ituglanis*, 407
metae, *Pimelodella*, 193
metae, *Pygodium*, 407
metae guayaberensis, *Pygodium*, 407
Metaloricaria, 217, 272
Metaloricaria nijsseni, 272
Metaloricaria paucidens, 272
Metaloricariina, 217
mexicanus, *Amiurus*, 209
mexicanus, *Ictalurus*, 209
meyenii, *Bagrus (Ariodes)*, 52
mica, *Mystus*, 100
mica, *Pelteobagrus*, 100
micai, *Rhamdia*, 197
micracanthus, *Bagrus*, 96
micracanthus, *Corydoras*, 120
micranodus, *Bagrichthys*, 83
micriscens, *Astroblepus*, 65
micriscens, *Astroblepus grixalvii*, 65
micristius, *Pareutropius*, 360, 361
Microcambeva, 409
Microcambeva barbata, 409
Microcambeva ribeirae, 409
microcephala, *Rhamdia*, 202
microcephala, *Rhamdiopsis*, 202
microcephalus, *Arius*, 35
microcephalus, *Cnidoglanis*, 346
microcephalus, *Corydoras*, 122
microcephalus, *Pseudarius*, 35
microcephalus, *Saccobranchus*, 151
microcephalus, *Silurus*, 374
microceps, *Euristhmus*, 346
microceps, *Plotosus*, 346
Microcorydoras, 112
microdon, *Loricaria*, 271
microdon, *Loricariichthys*, 271
microdorsalis, *Parasilurus*, 380
microdorsalis, *Silurus*, 380
microgalaeus, *Aspidoras*, 109
microgastropterygius, *Arius*, 55
Microglanis, 354
Microglanis ater, 354
Microglanis cibelae, 354
Microglanis cottooides, 354
Microglanis eurystoma, 354
Microglanis garavelloii, 354
Microglanis iheringi, 354
Microglanis leptostriatus, 441
Microglanis malabarbai, 354
Microglanis nigripinnis, 354
Microglanis parahybae, 354
Microglanis pataxo, 444
Microglanis pellopterygius, 354
Microglanis poecilus, 354
Microglanis secundus, 355
Microglanis variegatus, 355
Microglanis zonatus, 355
Microlepidogaster, 272
Microlepidogaster bahiensis, 281
Microlepidogaster bourguyi, 300
Microlepidogaster depressinotus, 248
Microlepidogaster doceanus, 282
Microlepidogaster guentheri, 296
Microlepidogaster guntheri, 295
Microlepidogaster güntheri, 296
Microlepidogaster perforated, 272
Microlepidogaster perforatus, 272
Microlepidogaster taimensis, 249
microlepidogaster, *Loricaria*, 295

- microlepidogaster*, *Rineloricaria*, 295
microlepidota, *Loricaria lima*, 295
microlepidota, *Rineloricaria*, 295
micromaculatus, *Hypostomus*, 256
micrommatus, *Hemiancistrus*, 244
Micromyzon, 61
Micromyzon akamai, 61
Micronema, 371
Micronema bleekeri, 374
Micronema cheveyi, 371
Micronema hexapterus, 371
Micronema moorei, 371
Micronema platypogon, 371
Micronema typus, 375
micronemus, *Pangasius*, 328
micronemus, *Phalacronotus*, 375
micronemus, *Pseudolais*, 328
micronemus, *Silurus*, 375
micronotacanthus, *Arius*, 55
microphthalmus, *Brachyglanis*, 181
microphthalmus, *Clarias*, 143
microphthalmus, *Dolichallabes*, 149
microphthalmus, *Euristhmus*, 442
microphthalmus, *Hemibagrus*, 90
microphthalmus, *Macrones*, 90
micropoeus, *Doras*, 171
micropoeus, *Hemidoras*, 171
micropogon, *Bagrus*, 92, 93
micropogon, *Batrachocephalus*, 38
micropogon, *Chiloglanis*, 305
micropogon, *Eutropius*, 364
micropogon, *Leiocassis*, 93
micropogon, *Schilbe*, 364
micropogon, *Silurus*, 374
micropogon, *Wallago*, 381
microps, *Akysis*, 14
microps, *Chaetostoma*, 229
microps, *Chaetostomus*, 229
microps, *Chiloglanis*, 306
microps, *Corydoras*, 112
microps, *Harttia*, 290
microps, *Imparfinis*, 186
microps, *Leiocassis*, 100
microps, *Neoplecostomus*, 273
microps, *Pareiodon*, 411
microps, *Pelteobagrus*, 100
microps, *Plecostomus*, 273, 281
microps, *Plecostomus (Rhinelepis)*, 281
microps, *Pterosturisoma*, 290
microps, *Rhamdia*, 201
microps, *Saccobranchus*, 151
micropterus, *Arius*, 166
micropterus, *Pimelodus*, 200
micropunctatus, *Plecostomus*, 225
micropus, *Kryptopterus*, 369
microstoma, *Malapterurus*, 301
microstoma, *Pimelodus*, 338
microstomus, *Arius*, 39
microstomus, *Clarias*, 146
microstomus, *Hemidoras*, 179
microstomus, *Hypostomus*, 250, 257
microstomus, *Malapterurus*, 301
microstomus, *Trachydoras*, 179
Microsynodontis, 309
Microsynodontis armatus, 309
Microsynodontis batesii, 309
Microsynodontis Christyi, 309
Microsynodontis emarginata, 309
Microsynodontis emarginatus, 309
Microsynodontis hirsuta, 310
Microsynodontis hirsutus, 310
Microsynodontis laevigata, 310
Microsynodontis laevigatus, 310
Microsynodontis lamberti, 310
Microsynodontis nannoculus, 310
Microsynodontis nasutus, 310
Microsynodontis notata, 310
Microsynodontis notatus, 310
Microsynodontis polli, 310
Microsynodontis vigilis, 310
micruropterus, *Phalacronotus*, 375
micruropterygius, *Arius*, 36
midgleyi, *Arius*, 47
midgleyi, *Neoarius*, 47
migrans, *Pygidium*, 420
migrans, *Trichomycterus*, 420
milesi, *Autanadoras*, 178, 179
milesi, *Chaetostoma*, 229
milesi, *Chaetostomus*, 229
militaris, *Ageneiosus*, 68, 69
militaris, *Ageniosus*, 68
militaris, *Arius*, 51
militaris, *Osteogeneiosus*, 51
militaris, *Silurus*, 51, 68, 69
mimonha, *Trichomycterus*, 420
mimulus, *Otocinclus*, 275
mindoense, *Cyclopium*, 65
mindoensis, *Astroblepus*, 65
minimaculatus, *Glyptothonax*, 391
minimus, *Kryptopterus*, 370
minjirya, *Malapterurus*, 302
mino, *Ageneiosus*, 38
mino, *Batrachocephalus*, 38
minor, *Kryptopterus*, 370
minuta, *Rhamdia*, 186
minutum, *Cetopsidium*, 130
minutum, *Glyptosternon*, 393
minutum, *Scleronema*, 413
minutus, *Ancistrus*, 223
minutus, *Bunocephalus*, 59
minutus, *Hemicetopsis*, 130
minutus, *[Hisonotus]*, 249
minutus, *Imparfinis*, 186
minutus, *Glyptothonax*, 391, 392, 393
minutus, *Parotocinclus*, 283
minutus, *Trachyglanis*, 28
minutus, *Silurus*, 431
minutus, *Trichomycterus*, 413
miostoma, *Wallago*, 372
miostomus, *Ompok*, 372
mirini, *Imparfinis*, 186
mirissumba, *Trichomycterus*, 420
misionera, *Rineloricaria*, 295
misilliensis, *Amiurus*, 206
misrai, *Fajumia*, 429
misrai, *Mystus*, 97
mitchelli, *Pseudeutropius*, 362
Miuroglanis, 409
Miuroglanis platycephalus, 409
miurus, *Noturus*, 214
möbiusii, *Eutropius*, 364
Mochocus, 310
Mochocus brevis, 310
Mochokidae, 303
Mochokiella, 310
Mochokiella paynei, 310
Mochokus, 303, 310
Mochokus brevis, 310
Mochokus niloticus, 310, 311
modesta, *Pimelodella*, 193
modestum, *Glyptosternum*, 389
modestus, *Pimelodus*, 193
modjensis, *Chiloglanis*, 306
moebii, *Eutropius*, 365
moebiusii, *Schilbe*, 364
moeschii, *Leiocassis*, 105
moeschii, *Liocassis*, 104
moeschii, *Pseudomystus*, 104
molinae, *Cetopsorhamdia*, 182
mollespiculum, *Neosilurus*, 348
molliceps, *Arius*, 55
mollinasum, *Chaetostoma*, 230
mollinasus, *Chaetostomus*, 230
mollis, *Silurus*, 214
mondolfi, *Pygidium*, 421
mondolfi, *Trichomycterus*, 421
mong, *Pimelodus*, 55
mongallensis, *Slatinia*, 24
Monistiancistrus, 289
Monistiancistrus carachama, 289
monitor, *Amblydoras*, 169
monitor, *Zatorax*, 168, 169
monkei, *Auchenoglanis*, 163
monkei, *Clarias*, 141
monkei, *Parauchenoglanis*, 163
mononema, *Kryptopterus*, 370
mononema, *Silurus*, 370
monopelte, *Sturisoma*, 299
monsembeensis, *Malapterurus*, 302
montana, *Cetopsis*, 132
montana, *Clupisoma*, 357
montana, *Erethistoides*, 384
montana, *Pimelodella*, 193
montana, *Rhamdella*, 196
montana, *Xenocara*, 223
montana pipri, *Erethistoides*, 384

- montanus*, *Ancistrus***, 223
montanus*, *Bagrus, 96
montanus*, *Mystus, 96
montanus dibrugarensis, *Macrones*, 96
montebelloi, *Canthopomus*, 263
montebelloi, *Ixinandria*, 263
moolenburghae, *Pseudeutropius*, 362
moolenburghi, *Hemisilurus*, 369
moorei, *Kryptopterus*, 371
moorei, *Micronema*, 371
moorii, *Clarias*, 143
moravicus, *Otolithus (Arius ?)*, 434
moreensis, *Silurus*, 376
morei, *Opsodoras*, 175
Morei, *Oxydoras*, 175
moreirai, *Rhamdiopsis*, 202, 203
morenoi, *Cetopsidium*, 130
morenoi, *Hemicetopsis*, 130
Mormyrostoma, 170
morrisi, *Hemidoras*, 172
morrowi, *Hemiloricaria*, 246
morrowi, *Liosomadoras*, 75
morrowi, *Rhineloricaria*, 246
mortiauxi, *Tanganikallabes*, 152
mortoni, *Neosilurus*, 348
mossambicus, *Clarias*, 143
motaguensis, *Pimelodus*, 198
motatanensis, *Cetopsis*, 132
motatanensis, *Pseudocetopsis plumbeus*, 132
motatanensis, *Pygidium emanueli*, 421
motatanensis, *Trichomycterus*, 421
mounseyi, *Rhamdia*, 201
mucosa, *Pimelodella*, 193
mucosa, *Platystoma*, 340
mucusos, *Platysilurus*, 340
mucronatum, *Amblyceps*, 18
muelleri, *Paraplotosus*, 349
muelleri, *Rhamdia*, 199
Mugil corsula, 89
mukherjii, *Mystus*, 95
mülleri, *Clarias*, 143
mülleri, *Cnidoglanis*, 349
mülleri, *Pimelodus*, 199
Mülleri, *Silurus*, 380, 381
multilineatus, *Eutropius*, 365
multimaculata, *Synodontis*, 317
multimaculatus, *Corydoras*, 120
multimaculatus, *Synodontis*, 317
multimaculatus, *Zungaropsis*, 344
multipunctata, *Amphilophus nigricaudatus*, 21
multipunctata, *Synodontis*, 317
multipunctatus, *Synodontis*, 317
multiradiatus, *Arius*, 40
multiradiatus, *Brochis*, 120
multiradiatus, *Cathorops*, 40
multiradiatus, *Chaenothorax*, 120
multiradiatus, *Corydoras*, 120
multiradiatus, *Cranoglanis*, 165
multiradiatus, *Heptapterus*, 184
multiradiatus, *Hypostomus*, 290, 291
multiradiatus, *Mystus*, 96
multiradiatus, *Pimelodus*, 334
multiradiatus, *Plotosus*, 350
multiradiatus, *Pseudeutropichthys*, 165
multiradiatus, *Pterygoplichthys*, 291
multiradiatus alternans, *Ancistrus*, 291
multispinis, *Ancistrus*, 223
multispinis, *Ancistrus*, 265
multispinis, *Xenocara*, 223
multitaeniatus, *Eutropius*, 365
multitaeniatus, *Schilbe*, 365
munitus, *Noturus*, 214
mura, *Otocinclus*, 275
muriaensis, *Parotocinclus*, 283
murica, *Doras*, 177
muricata, *Pseudolaguvia*, 401
murieci, *Rhamdia guatemalensis*, 202
murinus, *Spectracanthicus*, 297
murius, *Eutropiichthys*, 357
murius, *Pimelodus*, 357
murius batarensis, *Pseudeutropius*, 357
murrayi, *Malapterurus*, 302
murraystuarti, *Amblyceps*, 18
murray-stuarti, *Amblyceps*, 18
musaica, *Tatia*, 77
musculus, *Pimelodus*, 200
mustelinus, *Heptapterus*, 184
mustelinus, *Pimelodus*, 184
muticus, *Silurus*, 337
mutisi, *Eremophilus*, 404
mutsindoziensis, *Clariallabes*, 137
mutuca, *Hemipsilichthys*, 280
mutuca, *Pareiorhaphis*, 280
mutucae, *Hypostomus*, 257
myanmar, *Pangasius*, 327
Myersglanis, 396
Myersglanis blythii, 396
Myersglanis jayarami, 396
myersi, *Corydoras*, 123
myersi, *Hypostomus*, 257
myersi, *Leptodoras*, 173
myersi, *Plecostomus*, 257
myersi, *Pseudomystus*, 104
Myoglanis, 187
Myoglanis aspredinoides, 187
Myoglanis koepckeae, 187
Myoglanis potaroensis, 187
Myoglanis potaroënsis, 187
myriodon, *Aposturisoma*, 226
myriodon, *Chrysichthys*, 155
Mysoricus, *Silurus*, 374
mystacinus, *Ancistrus*, 265
mysterious, *Pimelodus*, 338
mysticetus, *Mystus*, 96
Mystus, 93
Mystus (Mystus) maydelli, 89
Mystus (Mystus) vittatus horai, 95
Mystus alasensis, 94
Mystus albolineatus, 94
Mystus armatus, 94, 98
Mystus armiger, 94
Mystus ascita, 177
Mystus atrifasciatus, 94
Mystus aubentoni, 91
Mystus bimaculatus, 94
Mystus bleekeri, 94
Mystus bocourti, 94
Mystus canarensis, 96
Mystus carolinensis, 37
Mystus castaneus, 94
Mystus cavasius, 95
Mystus dalungshanensis, 95
Mystus falcarius, 95
Mystus fluviatilis, 164
Mystus gulio, 95
Mystus havmollerii, 85
Mystus horai, 95
Mystus impluviatus, 95
Mystus johorensis, 89
Mystus krishnensis, 89
Mystus leucophasis, 96
Mystus malabaricus, 96
Mystus mica, 100
Mystus misrai, 97
Mystus montanus, 96
Mystus mukherjii, 95
Mystus multiradiatus, 96
Mystus mysticetus, 96
Mystus nigriceps, 96
Mystus oculatus, 96
Mystus olyroides, 90
Mystus pahangensis, 90
Mystus pelusi, 96
Mystus pulcher, 97
Mystus punctifer, 97
Mystus rhegma, 97
Mystus rufescens, 94, 97
Mystus sabanus, 91
Mystus seengtee, 97
Mystus singaringan, 97
Mystus spinipectoralis, 97
Mystus stigmaturus, 85
Mystus tengara, 97, 98
Mystus vittatus, 98
Mystus wolffii, 98
mystus, *Schilbe*, 365
mystus, *Silurus*, 362, 365
myzostoma, *Euchiloglanis*, 399
myzostomus, *Pareuchiloglanis*, 399
N
N'gamensis, *Ciaras*, 146
N'gamensis, *Clarias*, 146
nahuelbutaensis, *Diplomystes*, 166

- naipi*, *Trichomycterus*, 421
namdia, *Pimelodus*, 200
Nangra, 396
Nangra assamensis, 396
Nangra bucculenta, 396
Nangra Buchanani, 397
Nangra carcharhinoides, 396
Nangra nangra, 397
Nangra ornata, 397
Nangra punctata, 395
Nangra robusta, 397
nangra, *Nangra*, 397
nangra, *Pimelodus*, 396, 397
nannoculus, *Microsynodontis*, 310
Nannoglanis, 187
Nannoglanis bifasciatus, 203
Nannoglanis fasciatus, 187, 188
Nannoglanis hoehnei, 188
Nannocephalum, 272
Nannocephalum spectabile, 272
Nannocephalum sternoptychum, 273
Nannorhamdia, 180, 185
Nannorhamdia benedettii, 198
Nannorhamdia guttatus, 185
Nannorhamdia lineata, 185
Nannorhamdia macrocephala, 192
Nannorhamdia nemacheir, 186
Nannorhamdia schubarti, 186
Nannorhamdia spurrellii, 185, 186
Nannorhamdia stictonotus, 186
Nanobagrus, 98
Nanobagrus armatus, 98
Nanobagrus nebulosus, 98
Nanobagrus stellatus, 98
nanonocticulus, *Gelanoglanis*, 74
nantoensis, *Liobagrus*, 19
nantoënsis, *Liobagrus* 19
nanus, *Corydoras*, 120
napoensis, *Corydoras*, 120
narcissus, *Corydoras*, 120
nasalis, *Eutropius*, 363
naso, *Liocassis*, 108
Nasocassis, 92
nassi, *Phractocephalus*, 336
nasus, *Cetopsorhamdia*, 181, 182
nasus, *Pimelodina*, 336
nasuta, *Psammophiletria*, 27
nasuta, *Rhamdia*, 201
nasutus, *Arius*, 49
nasutus, *Microsynodontis*, 310
nasutus, *Pangasius*, 327
nasutus, *Pseudopangasius*, 327
natalensis, *Amphilios*, 22
natalis, *Ameiurus*, 205
natalis, *Pimelodus*, 205
natalis analis, *Amiurus*, 205
nationi, *Ancistrus*, 223
nationi, *Lasiancistrus*, 223
nattereri, *Corydoras*, 121
nattereri, *Farlowella*, 238
Nattereri, *Oxydoras*, 179
nattereri, *Trachydoras*, 179
nattereri triseriatus, *Corydoras*, 121
nauticus, *Amblydoras*, 169
nauticus, *Zatherax*, 169
navarroi, *Pimelodus*, 338
navarroi, *Pimelodus grosskopfii*, 338
naziri, *Clarias*, 357
naziri, *Glyptothorax*, 392
Neblinichthys, 273
Neblinichthys pilosus, 273
Neblinichthys roraima, 273
Neblinichthys yaravi, 273
nebulifer, *Pareuchiloglanis*, 399
nebulosa, *Synodontis*, 317
nebulosus, *Ameiurus*, 205
nebulosus, *Clarias*, 148
nebulosus, *Doras*, 178
nebulosus, *Ituglanis*, 407
nebulosus, *Nanobagrus*, 98
nebulosus, *Pimelodus*, 205, 215
nebulosus, *Silurus*, 215
nebulosus, *Silurus (Callichrus)*, 374
nebulosus, *Synodontis*, 317
nebulosus, *Wallago*, 381
nebulosus pannonicus, *Ictalurus*, 206
Nedystoma, 46
Nedystoma dayi, 46
Nedystoma novaeguineae, 46
negro, *Corydoras*, 121
neivai, *Glanidium*, 77
neivai, *Tatia*, 77
nella, *Pimelodus*, 52
nella, *Plicofollis*, 52
nelsoni, *Conorhynchus*, 53
nelsoni, *Glyptothorax*, 392
nelsoni, *Leptodoras*, 173
nelsoni, *Potamarius*, 53
nemacheir, *Imparfinis*, 186
nemacheir, *Nannorhamdia*, 186
Nemadoras, 174
Nemadoras elongatus, 174
Nemadoras hemipeltis, 174
Nemadoras humeralis, 174
Nemadoras leporinus, 174
Nemadoras trimaculatus, 174
Nemapteryx, 46
Nemapteryx armiger, 46
Nemapteryx augusta, 46
Nemapteryx bleekeri, 46
Nemapteryx caelata, 46
Nemapteryx macronotacantha, 46
Nemapteryx nenga, 46, 47
Nemasiluroides, 361
Nemasiluroides furcatus, 361
Nematogenyidae, 323
Nematogenyini, 323
Nematogenys, 323, 324
Nematogenys cuivi, 324
Nematogenys inermis, 324
Nematogenys nigricans, 324
Nematogenys pallidus, 324
nematorphorus, *Platystacus*, 61
nematopterus, *Hypostomus*, 257
nematopteryx, *Listrura*, 408
Nemuroglanis, 180, 188
Nemuroglanis lanceolatus, 188
Nemuroglanis mariae, 188
Nemuroglanis pauciradiatus, 188
nemurus, *Bagrus*, 87, 90
nemurus, *Hemibagrus*, 90
nemurus, *Pseudostegophilus*, 412
nemurus, *Stegophilus*, 412
nenga, *Nemapteryx*, 46, 47
nenga, *Pimelodus*, 47
Neoarius, 33, 47
Neoarius berneyi, 47
Neoarius coatesi, 47
Neoarius graeffei, 47
Neoarius latirostris, 47
Neoarius leptaspis, 47
Neoarius midgleyi, 47
Neoarius paucus, 48
Neoarius pectoralis, 48
Neoarius taylori, 48
Neoarius utarus, 48
Neoarius velutinus, 48
Neobagrus, 19
Neobagrus fuscus, 19, 20
neogrammatensis, *Arius*, 50
neogrammatensis, *Notarius*, 50
Neopangasius, 325
Neopangasius Nieuwenhuisii, 325, 327
Neoplecostominae, 217
Neoplecostomus, 217, 273
Neoplecostomus espiritosantensis, 273
Neoplecostomus franciscoensis, 273
Neoplecostomus granosus, 273
Neoplecostomus microps, 273
Neoplecostomus paranensis, 273
Neoplecostomus ribeirensis, 274
Neoplecostomus variipictus, 274
Neoplotosus, 346
Neoplotosus waterhousii, 346
Neosiluroides, 347
Neosiluroides cooperensis, 347
Neosilurus, 347
Neosilurus ater, 347
Neosilurus argenteus, 345
Neosilurus australis, 347, 348
Neosilurus bartoni, 347
Neosilurus brevidorsalis, 347
Neosilurus coatesi, 348
Neosilurus equinus, 348
Neosilurus gjellerupi, 348
Neosilurus gloveri, 348

- Neosilurus hyrtlii*, 347, 348
Neosilurus idenburgi, 348
Neosilurus mediobarbis, 347
Neosilurus mollespiculum, 348
Neosilurus mortoni, 348
Neosilurus novaeguineae, 348
Neosilurus pseudospinosus, 349
Neosilurus rendahli, 345
Neosilurus robustus, 348
Neotropius, 356, 359
Neotropius acutirostris, 359
Neotropius atherinoides, 359
Neotropius khavalchor, 359
nephelion, *Cordylancistrus*, 443
Netuma, 48
Netuma aulometopon, 56
Netuma bilineata, 49
Netuma dubia, 54
Netuma hassleriana, 56
Netuma insularum, 50
Netuma mazatlana, 54
Netuma osakae, 49
Netuma planifrons, 44
Netuma proxima, 48
Netuma radiata, 435
Netuma thalassina, 48
Netuma thalassina jacksonensis, 49
netuma, *Bagrus*, 49
neumannii, *Chiloglanis*, 307
neumannii, *Clarias*, 145
newtoni, *Otolithus (Arius)*, 435
ngamensis, *Clarias*, 146
ngamensis, *Auchenoglanis*, 163
ngamensis, *Parauchenoglanis*, 163
ngola, *Clarias*, 142
nicaraguensis, *Pimelodus*, 199
nicaraguensis, *Rhamdia*, 199
nicefori, *Astroblepus*, 65
nicéfori, *Astroblepus*, 65
niceforoi, *Hemiancistrus*, 257
niceforoi, *Hypostomus*, 257
nickeriensis, *Hypostomus*, 257
nickeriensis, *Loricaria*, 269
nicoi, *Acanthobunocephalus*, 57
nicoi, *Lasiancistrus*, 289
nicoi, *Pseudolithoxus*, 289
nieuhofii, *Clarias*, 139, 146
Nieuwenhuisi, *Glyptosternon*, 392
nieuwenhuisi, *Glyptothorax*, 392
Nieuwenhuisii, *Bagarius*, 383
Nieuwenhuisii, *Neopangasius*, 325, 327
nieuwenhuisii, *Pangasius*, 327
niger, *Bagrus docmac*, 84
niger, *Batasio*, 445
niger, *Chiloglanis*, 307
niger, *Copidoglanis novae-guineae*, 349
niger, *Doras*, 175
niger, *Doras (Oxydoras)*, 175
niger, *Hemiancistrus*, 287
niger, *Hypostomus*, 257
niger, *Oxydoras*, 175
niger, *Plecostomus*, 257
niger, *Pseudancistrus*, 287
nigeriae, *Clarias*, 141
nigeriensis, *Clarias anguillaris*, 140
Nigerium, 98
Nigerium gadense, 98
Nigerium wurnoense, 98, 99
nigrescens, *Callichrous*, 374
nigrescens, *Pseudancistrus*, 287
nigrescens, *Silurus (Pimelodus)*, 210
nigribarbis, *Parapimelodus*, 335
nigribarbis, *Pimelodus (Pseudorhamdia)*, 335
nigricans, *Arius*, 54
nigricans, *Clarias*, 146
nigricans, *Hypostomus*, 278
nigricans, *Nematogenys*, 324
nigricans, *Pimelodus*, 208, 209, 210
nigricans, *Plotosus*, 352
nigricans, *Pseudoplatystoma fasciatum*, 341
nigricans, *Trichomycterus*, 414, 421
nigricauda, *Hemiloricaria*, 246
nigricauda, *Hisonotus*, 248
nigricauda, *Liobagrus*, 20
nigricauda, *Loricaria*, 246
nigricauda, *Otocinclus*, 248
nigricauda, *Pseudopimelodus*, 353
nigricaudatus, *Amphilinus*, 21
nigricaudatus multipunctata, *Amphilinus*, 21
nigricaudus, *Cephalosilurus*, 353
nigriceps, *Bagrus*, 96
nigriceps, *Mystus*, 96
nigricollaris, *Horabagrus*, 429
nigricollis, *Tympanopleura*, 68
nigrilabris, *Gronias*, 204, 206
nigripinnis, *Auchenipterus*, 72
nigripinnis, *Euanemus*, 72
nigripinnis, *Microglanis*, 354
nigrirostrum, *Sturiosoma*, 299
nigrita, *Bagrus*, 160
nigrita, *Synodontis*, 317
nigriventris, *Synodontis*, 318
nigrodigitatus, *Chrysichthys*, 158, 160
nigrodigitatus, *Pimelodus*, 158
nigrofasciata, *Pimelodella*, 193
nigrofasciatus, *Pimelodus*, 193
nigrolineata, *Phenacorhamdia*, 189
nigrolineatus, *Chaetostomus*, 277, 278
nigrolineatus, *Panaque*, 278
nigrolineatus, *Panaque*, 278
nigromaculata, *Synodontis*, 318
nigromaculatus, *Hypostomus*, 257
nigromaculatus, *Plecostomus*, 257
nigromaculatus, *Synodontis*, 318
nigromaculatus, *Trichomycterus*, 421
nigromarmoratus, *Clarias*, 146
nigropunctatus, *Bagrus*, 340
nijsseni, *Corydoras*, 121
nijsseni, *Corydoras elegans*, 121
nijsseni, *Harttia*, 272
nijsseni, *Metaloricaria*, 272
niloticus, *Chiloglanis*, 307
niloticus, *Hypophthalmus*, 362, 365
niloticus, *Mochokus*, 310, 311
niloticus, *Silurus schilbe*, 431
niloticus, *waterloti*, *Chiloglanis*, 307
nimius, *Hemipsilichthys*, 247, 444
Niobicthys, 274
Niobicthys ferrarisi, 274
niobium, *Glanapteryx*, 405
nitidus, *Akysis*, 17, 82
nitidus, *Leiocassis*, 82
nitidus, *Pelteobagrus*, 82
nitidus, *Pseudobagarius*, 17
nitidus, *Pseudobagrus*, 82
niveata, *Dekeyseria*, 234
niveatus, *Baryancistrus*, 227
niveatus, *Hypostomus*, 226, 227
niveatus, *Plecostomus*, 234
niveiventris, *Amiurus*, 204
niveum, *Chaetostoma*, 230
njassae, *Synodontis*, 318
Nkondobabrus, 99
Nkondobagrus longisrostris, 99
nkunga, *Plotosus*, 351
nocturnus, *Brachyglanis*, 181
nocturnus, *Noturus*, 214
nocturnus, *Panaqolus*, 277
nocturnus, *Panaque*, 277
nodosus, *Pseudauchenipterus*, 76
nodosus, *Silurus*, 75, 76
noelkempffi, *Corydoras*, 121
nops, *Gymnallabes*, 150
normani, *Chiloglanis*, 307
Notarius, 49, 440
Notarius armbrusteri, 440
Notarius biffl, 49
Notarius cookei, 50
Notarius grandicassis, 50
Notarius insculptus, 50
Notarius kessleri, 50
Notarius neogranatensis, 50
Notarius osculus, 50
Notarius planiceps, 50
notata, *Microsynodontis*, 310
notata, *Synodontis*, 318
notatus, *Amphilinus*, 27
notatus, *Callichrous*, 374
notatus, *Hisonotus*, 248
notatus, *Microsynodontis*, 310
notatus, *Pimelodus*, 210, 339
notatus, *Platynematicthys*, 339

- notatus*, *Synodontis*, 318
notatus*, *Tetracamphilius, 27
notatus binotata, *Synodontis*, 318
notatus ocellatus, *Synodontis*, 318
notatus var. *binotata*, *Synodontis*, 318
notatus var. *ocellatus*, *Synodontis*, 318
notialis*, *Parakysis, 16
Notoglanidium, 161, 162
Notoglandium maculatum, 161
Notoglanidium pallidum, 162
Notoglanidium thomasi, 162
Notoglanidium walkeri, 162
notomelas*, *Pimelodella, 193
Notophthalmus, 333
notospilus, *Hemidoras*, 171
notozygurus, *Clarias*, 143
Noturus, 211
Noturus albater, 211
Noturus baileyi, 211
Noturus crypticus, 211, 442
Noturus elassochir, 212
Noturus elegans, 212
Noturus eleutherus, 212
Noturus exilis, 212
Noturus fasciatus, 212
Noturus flavater, 212
Noturus flaviginnis, 212
Noturus flavus, 211, 212
Noturus funebris, 212
Noturus furiosus, 211, 212
Noturus giberti, 213
Noturus gladiator, 213
Noturus gyrinus, 213
Noturus hildebrandi, 213
Noturus hildebrandi laetus, 213
Noturus insignis, 213
Noturus lachneri, 213
Noturus latifrons, 211
Noturus leptacanthus, 213
Noturus luteus, 212
Noturus marginatus, 213
Noturus maydeni, 441
Noturus miurus, 214
Noturus munitus, 214
Noturus nocturnus, 214
Noturus occidentalis, 212
Noturus phaeus, 214
Noturus placidus, 214
Noturus platycephalus, 212
Noturus sialis, 213
Noturus stanauli, 214, 442
Noturus stigmosus, 214
Noturus taylori, 214
Noturus trautmani, 214
novaeguineae*, *Nedystoma, 46
novaeguineae*, *Neosilurus, 348
novaesi*, *Furcodontichthys, 240
novaeguineae, *Copidoglanis*, 348
novaeguineae, *Doiichthys*, 46
novae-guineae niger, *Copidoglanis*, 349
novalimensis*, *Harttia, 241
nox, *Arius*, 38, 39
nox*, *Brustiarius, 38
ntemensis, *Synodontis Hollyi*, 322
nuchalis, *Ælurichthys*, 38
nuchalis, *Arius*, 41
nuchalis*, *Auchenipterus, 72
nuchalis, *Hypophthalmus*, 71, 72
nucleus, *Arius*, 57
nudiceps*, *Ancistrus, 223
nudiceps, *Cnidoglanis*, 346
nudiceps*, *Euristhmus, 346
nudiceps, *Hypostomus*, 223
nudiceps*, *Pelteobagrus, 100
nudiceps, *Pseudobagrus*, 100
nudidens, *Arius (Hemiarius)*, 56
nudipectus, *Belonoglanis*, 24
nudirostre*, *Chaetostoma, 230
nudirostris, *Chaetostomus*, 230
nudiventris*, *Hypostomus, 257
nudirostris, *Loricaria*, 271
nudirostris*, *Loricariichthys, 271
nudiventris, *Plecostomus*, 257
nudiventris, *Loricaria*, 297
nudiventris*, *Parancistrus, 279
nudiventris*, *Spatularicaria, 297
nudulus, *Hemipsilichthys*, 280
nudulus, *Pareiorhaphis*, 280
nuijangense, *Clupisoma*, 357
nummifer*, *Synodontis, 318
nyasensis, *Clarias*, 149
nyasensis*, *Dinotopterus, 149
nyongensis*, *Chrysichthys, 158
nyongensis, *Chrysichthys*
 longidorsalis, 158
nyongensis, *Eutropius*, 365
nyongensis*, *Schilbe, 365
ñssipinnis, *Heptapterus*, 184
O
oaxacae, *Rhamdia*, 201
obbesi*, *Porochilus, 351
obesa, *Rhamdia*, 199
obesus, *Amiurus*, 205
obesus, *Arius*, 81
obesus*, *Liobagrus, 20
obesus*, *Synodontis, 318, 323
oblongus, *Tachysurus*, 435
obscura, *Glyptothorax*, 392
obscurum*, *Pogonopoma, 285
obscurus, *Acrochordonichthys*, 13
obscurus, *Auchenipterus*, 81
obscurus*, *Bagrichthys, 83
obscurus, *Clarias*, 141
obscurus, *Copidoglanis*, 345, 350
obscurus*, *Glyptothorax, 392
obtusa*, *Pseudotothyris, 290
obtusirostris, *Eutropius*, 365
obtusirostris, *Plecostomus*, 218
obtusos, *Otocinclus*, 290
obtusus, *Otocinclus*, 290
occidentalis, *Ailia*, 360
occidentalis, *Ancistrus*, 223
occidentalis, *Auchenoglanis*, 154
occidentalis, *Cetopsis*, 134, 135
occidentalis, *Chiloglanis*, 307
occidentalis, *Hypostomus*, 257
occidentalis, *Hypostomus*
 gymnorhynchus, 257
occidentalis, *Istlarius balsanus*, 208
occidentalis*, *Malapterurus, 302
occidentalis, *Noturus*, 212
occidentalis*, *Parailia, 360
occidentalis, *Pimelodus*, 154
occidentalis, *Xenocara*, 223
occidentalis tanganicus,
 Auchenoglanis, 154
occidentalis tchadiensis,
 Auchenoglanis, 154
occidentalis var. *tanganicanus*,
 Auchenoglanis, 154
occidentalis var. *tchadiensis*,
 Auchenoglanis, 154
occlai*, *Ancistrus, 223
ocellatus, *Galeichthys*, 43
ocellatus, *Silurus*, 35
ocellatus, *Synodontis notatus*, 318
ocellifer*, *Synodontis, 318
Ochmacanthus, 409
Ochmacanthus alternus, 409
Ochmacanthus batrachostoma, 409
Ochmacanthus flabelliferus, 409
Ochmacanthus orinoco, 409
Ochmacanthus reinhardtii, 410
Ochmacanthus taxistigma, 405, 406
ochoterenai, *Haustor*, 209
ochoterenai*, *Ictalurus, 209
octocirrus, *Pimelodus*, 431
octocirrus, *Corydoras*, 117
Octonematicthys, 160
oculatus, *Bagrus*, 96
oculatus*, *Mystus, 96
oculeus*, *Hypostomus, 257
oculeus, *Panaque*, 257
odontotumulus*, *Farlowella, 238
odynea*, *Pimelodella, 193
odynea, *Pimelodella chagresi*, 193
oelemariensis, *Corydoras*, 114
oetik*, *Arius, 35
ogilviei, *Ageneiosus*, 69
ogilviei, *Opsodoras*, 174, 175
ogoensis, *Gephyroglanis*, 158
ogooensis*, *Chrysichthys, 158
ogooensis, *Gephyroglanis*, 158
ogooensis, *Malapterurus electricus*,
 var., 302
ogowensis, *Chrysichthys*, 158

- oguensis, Malapterurus*, 302
oguensis, Malapterurus electricus, 302
oguensis, Malapterurus electricus var., 302
oiapoquensis, Corydoras, 121
obiaensis, Callichthys, 111
okadai, Coreobagrus, 86
okae, Chrysichthys, 158
okeechoeensis, Ictalurus, 210
olallae, Duopalatinus, 340
olallae, Platysilurus, 340
Oligancistrus, 274
Oligancistrus punctatissimus, 274
oligospila, Peckoltia, 284
oligospilus, Chaetostomus, 284
olivaceus, Chlarias, 146
oliveaceus, Clarias, 146
olivaceus, Pimelodus, 210
Olivaichthys, 166
Olivaichthys cuyanus, 166
Olivaichthys mesembrinus, 166
Olivaichthys viedmensis, 166
olivarisi, Pygocentrus, 215
olivarisi, Silurus, 215
oliveirae, Farlowella, 237
oliveirai, Bathycetopsis, 131, 132
oliveirai, Cetopsis, 132
Oloplotosus, 349
Oloplotosus luteus, 349
Oloplotosus mariae, 349
Oloplotosus torobo, 349
Olyra, 81, 99
Olyra burmanica, 99
Olyra elongata, 99
Olyra horae, 99
Olyra horai, 99
Olyra kempfi, 99
Olyra laticeps, 17, 18
Olyra longicaudata, 99
Olyra longicaudatus, 99
Olyrinæ, 81
olyroides, Hemibagrus, 90
olyroides, Mystus, 90
omeihensis, Leiocassis, 102
omeihensis, Pseudobagrus, 102
omias, Synodontis, 319
ommation, Apistoricaria, 226
Ompok, 371
Ompok bimaculatus, 372
Ompok binotatus, 372
Ompok borneensis, 372
Ompok fumidus, 372
Ompok hypophthalmus, 371, 372
Ompok jaynei, 372
Ompok leiacanthus, 371, 372
Ompok malabaricus, 372
Ompok miostomus, 372
Ompok pabda, 372
Ompok pabo, 373
Ompok pinnatus, 373
Ompok platyrhynchus, 373
Ompok pluriradiatus, 373
Ompok rhadinurus, 373
Ompok sabanus, 371
Ompok siluroides, 371
Ompok siluroïdes, 373
Ompok sindensis, 373
Ompok urbaini, 373
Ompok weberi, 373
oncina, Arius, 75
oncinus, Arius, 75
oncinus, Liosomadoras, 75
ondon, Pseudobagrus, 102
operculatum, Scleronema, 413
ophthalmica, Pimelodella, 193
ophthalmicus, Pimelodus, 194
ophthalmus, Doras (Corydoras), 169
opistophthalmus, Amphilius, 22
Opladelus, 215
Opsodoras, 175
Opsodoras boulegeri, 172, 175
Opsodoras hemipeltis, 174
Opsodoras morei, 175
Opsodoras ogilviei, 174, 175
Opsodoras orthacanthus, 175
Opsodoras parallelus, 174
Opsodoras steindachneri, 169
Opsodoras stuebelii, 175
Opsodoras ternetzi, 175
Opsodoras trimaculatus, 174
orbignianus, Platystoma, 341
orbignyanum, Platystoma, 341
orcesi, Corydoras pastazensis, 122
oregonensis, Hypsidoris, 203
oremaculatus, Hypophthalmus, 333
Oreoglanis, 397
Oreoglanis delacouri, 397
Oreoglanis frenatus, 397
Oreoglanis hypsiurus, 397
Oreoglanis inflatus, 397
Oreoglanis insignis, 397
Oreoglanis lepturus, 397
Oreoglanis macronemus, 397
Oreoglanis macropterus, 398
Oreoglanis setiger, 398
Oreoglanis siamensis, 397, 398
orestes, Oxydoras, 171
Orestis, Oxydoras, 171
orientale, Cetopsidium, 130
orientale, Pseudocetopsis, 130
orientalis, Arges, 65
orientalis, Astroblepus, 65
orientalis, Bagrus, 84
orientalis, Irvineia, 358
orinocensis, Hildadoras, 175, 176
orinoco, Cetopsis, 132
orinoco, Cetopsorhamdia, 182
orinoco, Lithoxancistrus, 286, 287
orinoco, Ochmacanthus, 409
orinoco, Pseudancistrus, 287
orinoco, Pseudocetopsis plumbeus, 132
Orinocodoras, 175
Orinocodoras eigenmanni, 175
ornata, Nangra, 397
ornaticeps, Heptapterus, 185
ornatipinnis, Synodontis, 319
ornatissima, Synodontis, 319
ornatissimus, Synodontis, 319
ornatus, Chrysichthys, 158
ornatus, Clarias, 145
ornatus, Corydoras, 121
ornatus, Gogo, 29
ornatus, Hyalobagrus, 92
ornatus, Pimelodus, 338
ornatus, Pseudobagrus, 92
ornatus, Pseudomystus, 104
ornatus, Synodontis, 317, 319
oronocoi, Hoplosternum, 129
orontis, Clarias, 143
oroyae, Pygidium, 423
orophopterus, Corydoras, 121
orthacanthus, Opsodoras, 175
orthiocarinatus, Copionodon, 404
ortmanni, Pimelodus, 338
ortoni, Rhamdia, 201
ortoni, Sorubimichthys, 343
osakae, Netuma, 49
Oschanini, Exostoma, 402
osculus, Arius, 50
osculus, Notarius, 50
Osteobagrus, 106
osteocarus, Corydoras, 121
Osteogaster, 112
Osteogeneiosus, 51
Osteogeneiosus blanchardi, 51
Osteogeneiosus cantoris, 51
Osteogeneiosus gracilis, 51
Osteogeneiosus ingluvies, 51
Osteogeneiosus longiceps, 51
Osteogeneiosus macrocephalus, 51
Osteogeneiosus militaris, 51
Osteogeneiosus valenciennesi, 51
Osteogeniosus stenocephalus, 51
Osteogeniosus stenocephalus, 51
Osteomystax, 71
osteomystax, Auchenipterus, 72
osteomystax, Ceratocheilus, 71, 72
Ostophycephalus, 346
Ostophycephalus duriceps, 346
othonops, Cetopsis, 132
othonops, Hemicetopsis, 132
Otocinclini, 217
Otocinclus, 217, 264, 271, 274
Otocinclus (Microlepidogaster) tietensis, 290
Otocinclus affinis, 271

Otocinclus arnoldi, 271
Otocinclus bororo, 274
Otocinclus caxarari, 274
Otocinclus cephalacanthus, 276
Otocinclus cocama, 274
Otocinclus depressicauda, 248
Otocinclus fimbriatus, 271
Otocinclus flexilis, 271, 272
Otocinclus francirochai, 248
Otocinclus gibbosus, 264
Otocinclus hasemani, 274
Otocinclus hoppei, 275
Otocinclus huaoranii, 275
Otocinclus joberti, 250
Otocinclus leucofrenatus, 248
Otocinclus macrospilus, 275
Otocinclus maculicauda, 281, 283
Otocinclus maculipinnis, 248
Otocinclus mariae, 275
Otocinclus mimulus, 275
Otocinclus mura, 275
Otocinclus nigricauda, 248
Otocinclus obtusos, 290
Otocinclus obtusus, 290
Otocinclus paulinus, 249
Otocinclus spectabilis, 273
Otocinclus tapirape, 275
Otocinclus vestitus, 274, 275
Otocinclus vittatus, 275
Otocinclus xakriaba, 275
Otolithus (Arius) aequus, 435
Otolithus (Arius) africanus, 435
Otolithus (Arius) amekiensis, 435
Otolithus (Arius) angelicus, 434
Otolithus (Arius) angulatus, 435
Otolithus (Arius) crassus bartonensis, 434
Otolithus (Arius) danicus, 434
Otolithus (Arius) danicus bartonensis, 434
Otolithus (Arius) decipiens, 434
Otolithus (Arius) germanicus, 434
Otolithus (Arius ?) glaber, 435
Otolithus (Arius) jaekeli, 435
Otolithus (Arius) Lerichei, 434
Otolithus (Arius ?) moravicus, 434
Otolithus (Arius) newtoni, 435
Otolithus (Arius ?) parvus, 435
Otolithus (Arius) planus, 435
Otolithus (Arius) rotundatus, 435
Otolithus (Arius) tenius, 435
Otolithus (Arius) temuis, 435
Otolithus (Arius) vanigonis, 434
Otolithus (incertae sedis) crassus, 434
Otolithus (Sciaenidarum) decipiens, 434
Otolithus (Siluridarum ?) incertus, 434
Otothyridini, 217, 440
Otothyris, 217, 275

Otothyris canaliferus, 275, 276
Otothyris juquiae, 276
Otothyris lophophanes, 276
Otothyris rostrata, 276
Otothyris travassosi, 276
Otothyropsis, 276
Otothyropsis marapoama, 276
ourastigma, *Corydoras*, 121
ovidius, *Synodontis*, 316
oxycephalus, *Clarias*, 146
Oxydoras, 175
Oxydoras (Rhinodoras) amazonum
hasemani, 170
Oxydoras (Rhinodoras) huberi, 174
Oxydoras acipenserinus, 172
Oxydoras affinis, 171
Oxydoras bachi, 174
Oxydoras eigenmanni, 167
Oxydoras elongatus, 174
Oxydoras holdeni, 176
Oxydoras kneri, 175
Oxydoras Morei, 175
Oxydoras Nattereri, 179
Oxydoras niger, 175
Oxydoras orestes, 171
Oxydoras Orestis, 171
Oxydoras sifontesi, 176
Oxydoras steindachneri, 179
Oxydoras Stübelii, 175
Oxydoras trachyparia, 179
Oxydoras trimaculatus, 174
Oxyglanis, 154
Oxyglanis sacchii, 154
Oxyloricaria, 298
Oxyloricaria citurensis, 300
Oxyloricaria dariensis, 299
Oxyloricaria Fowleri, 233
Oxyloricaria guentheri, 299
Oxyloricaria leightoni, 300
Oxyloricaria lyra, 299
Oxyloricaria robusta, 299
Oxyloricaria tamanae, 300
Oxyloricaria tenuirostris, 300
oxyptera, *Paravandellia*, 410
oxyrhinus, *Amphilinus*, 23
oxyrynchus, *Anduzedoras*, 169
oxyrynchus, *Corydoras*, 121
Oxyrynchus Doras, 169
Oxyropsis, 276
Oxyropsis acutirostris, 276
Oxyropsis carinata, 276
Oxyropsis wrightiana, 276
Oxyropsis wrightii, 276
oxyrryncha, *Acestra*, 239
oxyrryncha, *Acestra*, 239
oxyrryncha, *Farlowella*, 239
P
pabda, *Ompok*, 372
pabda, *Silurus*, 371, 372
pabo, *Ompok*, 373
pabo, *Silurus*, 373
pachychilus, *Atopochilus*, 304
pachyderma, *Acrochordonichthys*, 13
pachynema, *Ceratoglanis*, 368
pachynemus, *Ceratoglanis*, 368
pachynema, *Clarias*, 146
Pachypterus, 359
Pachypterus luridus, 327
Pachypterus melanurus, 358
Pachypterus punctatus, 358
Pachypterus trifasciatus, 359
Pachyula, 42
pacifici, *Cruciglanis*, 443
pagei, *Hypostomus*, 257
pahangensis, *Mystus*, 90
pakistanica, *Gagata*, 386
pakistanicus, *Batasio*, 85
palavanensis, *Penelirus*, 375, 377
palaeindicus, *Heterobranchus*, 151
paleatus, *Callichthys*, 122
paleatus, *Corydoras*, 122
palembangensis, *Kryptopterus*, 370
palembangensis, *Silurus*, 369, 370
palleus, *Trichomycterus*, 415
pallida, *Pimelodella*, 194
pallida, *Sosia chamaeleon*, 13
pallidimaculatus, *Lithoxus*, 267
pallidimaculatus, *Lithoxus*
(*Paralithoxus*), 267
pallidum, *Notoglanidium*, 162
pallidus, *Nematogenys*, 324
pallidus, *Silurus*, 210
pallidus Lateralis, *Silurus*, 210
pallidus Leucoptera, 210
pallidus Marginatus, *Silurus*, 210
pallidus* var. *Lateralis*, *Pimelodus, 210
pallidus* var. *Lateralis*, *Silurus, 210
pallidus* var. *Leucoptera*, *Pimelodus, 210
pallidus* var. *Leucoptera*, *Silurus, 210
pallidus* var. *Marginata*, *Silurus, 210
pallidus* var. *Marginatus*, *Pimelodus, 210
pallidus* var. *Marginatus*, *Silurus, 210
pallozonum, *Glyptosternum*, 388, 392
pallozonus, *Glyptothorax*, 392
palmeri, *Chaetostoma*, 230
palmeri, *Chaetostomus* 230
palmeri, *Schilbe*, 364
panamense, *Sturisoma*, 299
panamensis, *Aelurichthys*, 37, 38
panamensis, *Bagre*, 38
panamensis, *Imparales*, 180
panamensis, *Loricaria*, 299
panamensis, *Plecostomus plecostomus*, 243
Panaqolus, 277
Panaqolus albomaculatus, 277

- Panaqolus changae*, 277
Panaqolus dentex, 277
Panaqolus gnomus, 277
Panaqolus maccus, 277
Panaqolus nocturnus, 277
Panaqolus purusiensis, 277
Panaque, 277
Panaque albomaculatus, 277
Panaque changae, 277
Panaque cochliodon, 277
Panaque dentex, 277
Panaque gnomus, 277
Panaque maccus, 277
Panaque nigrolineatus, 278
Panaque nocturnus, 277
Panaque ocaleus, 257
Panaque purusiensis, 277
Panaque suttoni, 278
Panaque suttonorum, 278
panda, *Corydoras*, 122
panda, *Glyptothorax*, 392
Pangasianodon, 324
Pangasianodon gigas, 325
Pangasianodon hypophthalmus, 325
Pangasianodonidi, 324
Pangasiidae, 324
Pangasini, 324
Pangasius, 324, 325
Pangasius (Pseudopangasius) bocourti, 325
Pangasius aequilabialis, 327
Pangasius altifrons, 325
Pangasius beani, 328
Pangasius bedado, 325
Pangasius bocourti, 325
Pangasius Buchanani, 325, 327
Pangasius burgini, 326
Pangasius conchophilus, 325
Pangasius cultratus, 328
Pangasius de Zwaani, 328
Pangasius delicatissimus, 328
Pangasius djambal, 325, 326
Pangasius elongatus, 326
Pangasius fowleri, 328
Pangasius hexanema, 358
Pangasius hoeksi, 328
Pangasius humeralis, 326
Pangasius indicus, 326
Pangasius juaro, 327
Pangasius kinabatanganensis, 326
Pangasius krempfi, 326
Pangasius kunyit, 326
Pangasius larnaudii, 326
Pangasius lithostoma, 326
Pangasius longibarbis, 359
Pangasius macronema, 327
Pangasius mahakamensis, 327
Pangasius mekongensis, 327
Pangasius micronemus, 328
Pangasius myanmar, 327
Pangasius nasutus, 327
Pangasius nieuwenhuisii, 327
Pangasius pangasius, 327
Pangasius pangasius godavarii, 327
Pangasius pangasius upiensis, 327
Pangasius paucidens, 325
Pangasius pleurotaenia, 328
Pangasius polyuranodon, 325, 327
Pangasius ponderosus, 327
Pangasius rheophilus, 328
Pangasius rios, 328
Pangasius sabahensis, 328
Pangasius sanitwongsei, 328
Pangasius siamensis, 327
Pangasius sutchi, 325
Pangasius taeniura, 326
Pangasius tubbi, 328
panjang, *Hemileiocassis*, 91
pannonicus, *Ictalurus nebulosus*, 206
pantanensis, *Corydoras*, 122
panthale, *Platystoma*, 341, 342
pantherinus, *Auchenoglanis*, 163
pantherinus, *Erycheilichthys*, 236
pantherinus, *Eurycheilus*, 236
pantherinus, *Hypostomus*, 258
pantherinus, *Parauchenoglanis*, 163
pantherinus, *Pseudarioides*, 339
pantherinus, *Synodontis*, 315
pantherinus, *Trichomycterus*, 421
panzeri, *Henonemus*, 414
panzeri, *Stegophilus*, 414
paolence, *Pygidium*, 421
paolencis, *Trichomycterus*, 421
papariae, *Hypostomus*, 258
papariae, *Plecostomus plecostomus*, 258
papariae, *Pseudancistrus*, 287
papariae, *Rhamdella*, 197
papilionatus, *Doras*, 173
papillatus, *Hemipilichthys*, 248, 444
papillatus, *Hoplomyzon*, 61
papillifer, *Hemipimelodus*, 48
papilliferus, *Trichomycterus*, 421
papillosus, *Arius*, 165
pappenheimi, *Pimelodella*, 194
papuensis, *Copidoglanis*, 348
papuensis, *Plotosus*, 351
paquequerense, *Pygidium*, 421
paquequerensis, *Trichomycterus*, 421
Parabranchioica, 410
Parabranchioica teaguei, 411
Paracanthopoma, 410
Paracanthopoma parva, 410
Paracetopsis, 134
Paracetopsis atahualpa, 135
Paracetopsis bleekeri, 134, 135
Paracetopsis esmeraldas, 135
Parachiloglanis, 398
Parachiloglanis hodgarti, 398
Paradiplomystes, 37
Paradoxoglanis, 302
Paradoxoglanis caudivittatus, 302
Paradoxoglanis cryptus, 303
Paradoxoglanis parvus, 303
paraense, *Brachyplatystoma*, 331
Paraglyptothorax, 388
paragua, *Corydoras*, 122
paraguayensis, *Farlowella*, 239
paraguayensis, *Hemidoras*, 179
paraguayensis, *Trachydoras*, 179
paraguensis, *Rhinolepis*, 275
Parahemiodon, 269
Parahemiodon chanjoo, 270
Parahemiodon typus, 269, 270
parahemiodon, *Loricaria*, 270
parahybae, *Delturus*, 234, 235, 444
parahybae, *Ituglanis*, 407
parahybae, *Microglanis*, 354
parahybae, *Pimelodus (Rhamdia)*, 201
Parahybae, *Platystoma*, 344
parahybae, *Pogonopoma*, 285
parahybae, *Pseudopimelodus*, 354
parahybae, *Pseudotocinclus*, 290
parahybae, *Pygidium proops*, 406, 407
Parahybae, *Rhinelepis*, 285
parahybae, *Steindachneridion*, 344
Parailia, 360
Parailia congica, 360
Parailia longifilis, 360
Parailia occidentalis, 360
Parailia pellucida, 360
Parailia somalensis, 360
Parailia spiniserrata, 360
Parakysisidae, 12
Parakysis, 12, 15
Parakysis anomalopteryx, 15
Parakysis grandis, 15
Parakysis longirostris, 16
Parakysis notialis, 16
Parakysis verrucosa, 15, 16
Parakysis verrucosus, 16
Paralithoides, 267
Paralithoxus, 267
parallelus, *Corydoras*, 122
parallelus, *Opsodoras*, 174
Paraloricaria, 278
Paraloricaria agastor, 278
Paraloricaria commersonoides, 278
Paraloricaria vetula, 278
Paramphilius, 25
Paramphilius firestonei, 25
Paramphilius goodi, 25
Paramphilius teugelsi, 25
Paramphilius trichomycteroidea, 25
paranaënsis, *Farlowella*, 237
paranaensis, *Pimelodus*, 338
parananus, *Megalancistrus*, 272

- parananus*, *Pterygoplichthys*
(Ancistrus), 272
- Parancistrus*, 278
- Parancistrus aurantiacus*, 278
- Parancistrus nudiventris*, 279
- paranensis*, *Hypostomus*, 258
- paranensis*, *Neoplecostomus*, 273
- paranensis*, *Parapterodoras*, 177
- Paraotocinclus cesarpintoi*, 282
- Paraphractura*, 26
- Paraphractura tenuicauda*, 26, 27
- Parapimelodus*, 335
- Parapimelodus nigribarbis*, 335
- Parapimelodus valenciennis*, 335
- Paraplotosus*, 349
- Paraplotosus albilabris*, 349
- Paraplotosus butleri*, 349
- Paraplotosus muelleri*, 349
- Parapseudecheneis*, 400
- Parapterodoras*, 177
- Parapterodoras paranensis*, 177
- Pararius*, 48
- paraschilbeides*, *Kryptopterus*, 370
- Parasilurus*, 378
- Parasilurus asotus longus*, 379
- Parasilurus biwaensis*, 379
- Parasilurus lithophilus*, 380
- Parasilurus microdorsalis*, 380
- Parastegophilus*, 410
- Parastegophilus maculatus*, 410
- Parastegophilus paulensis*, 410
- Parasturisoma*, 298
- Parasturisoma maculata*, 233
- paratus*, *Chiloglanis*, 307
- Parauchenipterus*, 79
- Parauchenipterus paseae*, 79
- Parauchenoglanis*, 162
- Parauchenoglanis ahli*, 162
- Parauchenoglanis altipinnis*, 162
- Parauchenoglanis ansorgii*, 154
- Parauchenoglanis balayi*, 162
- Parauchenoglanis boutchangai*, 154
- Parauchenoglanis buettikoferi*, 163
- Parauchenoglanis longiceps*, 163
- Parauchenoglanis monkei*, 163
- Parauchenoglanis ngamensis*, 163
- Parauchenoglanis pantherinus*, 163
- Parauchenoglanis punctatus*, 163
- Paravandellia*, 410
- Paravandellia oxyptera*, 410
- Paravandellia phaneronema*, 411
- pardale*, *Platystoma*, 341
- pardalis*, *Ageneiosus*, 70
- pardalis*, *Hypostomus*, 292
- pardalis*, *Platystoma*, 341, 342
- pardalis*, *Pterygoplichthys*, 292
- pardalis*, *Synodontis*, 319
- Pardiglanis*, 163
- Pardiglanis tarabinii*, 163
- pardus*, *Trichomycterus*, 423
- parecis*, *Ancistrus*, 224
- pareiacantha*, *Loricaria*, 295
- pareiacantha*, *Rineloricaria*, 295
- Pareiodon*, 403, 411
- Pareiodon microps*, 411
- Pareiodontinae*, 403
- Pareiorhaphis*, 247, 279
- Pareiorhaphis alipionis*, 262
- Pareiorhaphis azygolechis*, 279
- Pareiorhaphis bahianus*, 279
- Pareiorhaphis cameroni*, 279
- Pareiorhaphis cerosus*, 279
- Pareiorhaphis eurycephalus*, 279
- Pareiorhaphis garbei*, 279
- Pareiorhaphis hypselurus*, 280
- Pareiorhaphis hystrix*, 280
- Pareiorhaphis mutuca*, 280
- Pareiorhaphis nudulus*, 280
- Pareiorhaphis parmula*, 280
- Pareiorhaphis regani*, 280
- Pareiorhaphis splendens*, 280
- Pareiorhaphis steindachneri*, 280
- Pareiorhaphis stephanus*, 280
- Pareiorhaphis stomias*, 280
- Pareiorhaphis vestigipinnis*, 281
- Pareiorhina*, 281
- Pareiorhina brachyrhyncha*, 281
- Pareiorhina carrancas*, 281
- Pareiorhina rudolphi*, 281
- Pareuchiloglanis*, 398
- Pareuchiloglanis anteanalisis*, 398
- Pareuchiloglanis feae*, 398
- Pareuchiloglanis gongshanensis*, 398
- Pareuchiloglanis gracilicaudatus*, 398
- Pareuchiloglanis kamengensis*, 399
- Pareuchiloglanis longicauda*, 399
- Pareuchiloglanis macropterus*, 399
- Pareuchiloglanis macrotremus*, 399
- Pareuchiloglanis myzostomus*, 399
- Pareuchiloglanis nebulifer*, 399
- Pareuchiloglanis poilanei*, 398, 399
- Pareuchiloglanis rhabdurus*, 399
- Pareuchiloglanis robusta*, 399
- Pareuchiloglanis robustus*, 399
- Pareuchiloglanis sichuanensis*, 399
- Pareuchiloglanis sinensis*, 399
- Pareuchiloglanis songdaensis*, 400
- Pareuchiloglanis songmaensis*, 400
- Pareuchiloglanis tianquanensis*, 400
- Pareutropius*, 360
- Pareutropius buffei*, 360
- Pareutropius debauwi*, 361
- Pareutropius longifilis*, 361
- Pareutropius mandevillei*, 361
- Pareutropius micristius*, 360, 361
- Parexostoma*, 387
- Parexostoma maculatum*, 387
- pariolispos*, *Scobinancistrus*, 296
- Pariolius*, 188
- Pariolius armillatus*, 188
- parkeri*, *Aspistor*, 36
- parkeri*, *Silurus*, 36
- parkii*, *Arius*, 32
- parkoi*, *Ituglanis*, 407
- parkoi*, *Pygidium*, 407
- parma*, *Cetopsis*, 132
- parmocassis*, *Arius*, 50
- parnaguensis*, *Ageneiosus*, 70
- parnahybae*, *Brachyplatystoma*, 331
- parnahybae*, *Loricaria*, 269
- parnahybae*, *Loricariichthys*, 266
- parnahybae*, *Pimelodella*, 194
- parnaibae*, *Glyptoperichthys*, 292
- parnaibae*, *Pterygoplichthys*, 292
- Paroreoglanis*, 397
- Paroreoglanis delacouri*, 397
- Parotocinclus*, 281
- Parotocinclus amazonensis*, 281
- Parotocinclus aripuanensis*, 281
- Parotocinclus bahiensis*, 281
- Parotocinclus bidentatus*, 282
- Parotocinclus britskii*, 282
- Parotocinclus cearensis*, 282
- Parotocinclus cesarpintoi*, 282
- Parotocinclus collinsae*, 282
- Parotocinclus cristatus*, 282
- Parotocinclus doceanus*, 282
- Parotocinclus eppleyi*, 282
- Parotocinclus haroldoi*, 282
- Parotocinclus jimi*, 282
- Parotocinclus jumbo*, 282
- Parotocinclus longirostris*, 282
- Parotocinclus maculicauda*, 283
- Parotocinclus minutus*, 283
- Parotocinclus muriaensis*, 283
- Parotocinclus planicauda*, 283
- Parotocinclus polyochrus*, 283
- Parotocinclus prata*, 283
- Parotocinclus spilosoma*, 283
- Parotocinclus spilurus*, 283
- Parotocinclus steindachneri*, 283
- Parrae*, *Galeichthys*, 38
- parryi*, *Rhamdia*, 199
- parva*, *Hemiloricaria*, 246
- parva*, *Loricaria*, 246
- parva*, *Paracanthopoma*, 410
- parva*, *Pimelodella*, 194
- parva*, *Pseudotatia*, 77
- parvanalis*, *Kryptopterus*, 375
- parvanalis*, *Phalacronotus*, 375
- parvicanata*, *Farlowella*, 239
- parvimanus*, *Clarias*, 140
- parvipinnis*, *Arius*, 47
- parvus*, *Otolithus* (*Arius* ?), 435
- parvus*, *Paradoxoglanis*, 303
- parvus*, *Pimelodus* (*Rhamdia*), 180
- paseae*, *Parauchenipterus*, 79

- passany*, *Bagrurus*, 54
passany, *Sciades*, 54
passarelli, *Homodiaetus*, 406
passarelli, *Stegophilus*, 406
passensis, *Ituglanis*, 408
pastazensis, *Corydoras*, 122
pastazensis orcesi, *Corydoras*, 122
patagoniensis, *Hatcheria*, 405
pataxo, *Microglanis*, 444
pati, *Luciopimelodus*, 334
pati, *Pimelodus*, 334
pati, *Silurus*, 335
patiae, *Chaetostoma*, 230
pattersoni, *Troglomelanis*, 216
paucerna, *Corydoras*, 122
paucidens, *Metaloricaria*, 272
paucidens, *Pangasius*, 325
paucimaculatus, *Hypostomus*, 258
paucipunctatus, *Hypostomus*, 258
pauciradiatum, *Megalonema*, 335
pauciradiatus, *Aspidoras*, 109
pauciradiatus, *Corydoras*, 109
pauciradiatus, *Nemuroglanis*, 188
pauciradiatus, *Trichomycterus*, 440
paucispinis, *Chaetostoma*, 230
paucispinis, *Chaetostomus*, 230
paucisquamatus, *Megalodoras*, 173
paucus, *Arius*, 48
paucus, *Neoarius*, 48
paulensis, *Parastegophilus*, 410
paulensis, *Pseudostegophilus*, 410
Paulicea, 344
Paulicea jahu, 344
paulina, *Loricaria*, 294
paulinus, *Hisonotus*, 249
paulinus, *Hypostomus*, 258
paulinus, *Otocinclus*, 249
paulimus, *Plecostomus*, 258
paviei, *Pseudecheneis*, 400
pavimentatus, *Arius*, 105
paynei, *Mochokiella*, 310
paysanduanus, *Auchenipterus*, 72
pearsei, *Chaetostoma*, 230
pearsei, *Chaetostomus*, 230
pearsoni, *Cetopsis*, 132
pearsoni, *Tridentopsis*, 426
Peckoltia, 283
Peckoltia arenaria, 284
Peckoltia bachi, 284
Peckoltia braueri, 284
Peckoltia brevis, 284
Peckoltia cavatica, 284
Peckoltia filicaudata, 284
Peckoltia furcata, 284
Peckoltia kuhlmanni, 284
Peckoltia oligospila, 284
Peckoltia sabaji, 284
Peckoltia snethlageae, 285
Peckoltia ucayalensis, 285
Peckoltia vermiculata, 285
Peckoltia vittata, 285
Peckoltichthys, 283
Peckoltichthys filicaudatus, 283, 284
Peckoltichthys kuhlmanni, 284
pecten, *Copionodon*, 404
pectinata, *Conta*, 384
pectinatus, *Ameiurus*, 206
pectinatus, *Ictalurus*, 206
pectinatus, *Tetracamphilius*, 27
pectinidens, *Pimelodus*, 46
pectinifer, *Pimelodella*, 194
pectinifrons, *Agamyxis*, 168
pectinifrons, *Doras*, 168
pectinopterus, *Glyptosternon*, 392
pectinopterus, *Glyptothonax*, 392
pectorale, *Leptoplosternum*, 128
pectoralis, *Arius*, 48
pectoralis, *Callichthys*, 128
pectoralis, *Neoarius*, 48
pediculatus, *Dolichancistrus*, 235
pediculatus, *Pseudancistrus*, 235
pediculatus cobrensis, *Pseudancistrus*, 235
peguensis, *Hemibagrus*, 90
peguensis, *Macrones*, 90
pellegrini, *Andersonia*, 24
pellegrini, *Plecostomus*, 289
pellopterygius, *Microglanis*, 354
pellucida, *Parailia*, 360
pellucida, *Physailia*, 360
Pelodichthys, 215
peloichthys, *Trachelyopterus*, 80
peloichthys, *Trachycorystes insignis*, 80
peltatus, *Rhineastes*, 430
Pelteobagrus, 81, 99
Pelteobagrus argentivittatus, 100
Pelteobagrus brashnikowi, 100
Pelteobagrus eupogon, 100
Pelteobagrus fulvidraco, 100
Pelteobagrus intermedius, 100
Pelteobagrus kyphus, 82
Pelteobagrus mica, 100
Pelteobagrus microps, 100
Pelteobagrus nitidus, 82
Pelteobagrus nudiceps, 100
Pelteobagrus ramentosus, 108
Pelteobagrus ussuriensis, 100
Pelteobagrus vachellii, 101
Pelteobagrus virgatus, 101
Pelteobagrus virgatus vinhensis, 101
Peltura, 26
Peltura Bovei, 26
pelusi, *Mystus*, 96
pelusi, *Silurus*, 96
pemucus, *Bagrurus*, 53
pemon, *Cetopsidium*, 131
Penesilurus, 375
Penesilurus bokorensis, 376
Penesilurus palawanensis, 375, 377
pentamaculata, *Rineloricaria*, 295
pentapterus, *Clarias*, 146
pentlandi, *Trichomycterus*, 423
Pentlandii, *Pimelodus*, 200
peregrinus, *Ictalurus*, 207
perforated, *Microlepidogaster*, 272
perforatus, *Microlepidogaster*, 272
perijae, *Cordylancistrus*, 231
perporosus, *Hypophthalmus*, 334
Perrunichthys, 335
Perrunichthys perruno, 335, 336
perruno, *Perrunichthys*, 335, 336
persimilis, *Chrysichthys*, 158
personatus, *Callychthys*, 129
peruana, *Pimelodella*, 194
peruanus, *Arges*, 66
peruanus, *Astroblepus*, 66
peruanus, *Chasmocranus*, 183
peruanus, *Duopalatinus*, 332
peruense, *Pimelodella*, 194
peruensis, *Pimelodella*, 194
Perugia, 339
Perugia argentina, 335
Perugia xanthus, 335
perugiae, *Anyperistius*, 347
perugiae, *Centromochlus*, 73
peruvianus, *Galeichthys*, 43
pestai, *Corydoras*, 116
Petacara, 62
petenensis, *Pimelodus*, 198
petleyi, *Limatulichthys*, 266
petleyi, *Rhineloricaria*, 266
petricola, *Clariallabes*, 138
petricola, *Synodontis*, 319
petroleus, *Hoplomyzon atrizona*, 61
Peyeria, 428
peyeria, *Ariopsis*, 432
pfefferi, *Synodontis*, 311
phaeus, *Noturus*, 214
Phagorus, 139
Phagorus cataractus, 141
phaiosoma, *Silurichthys*, 378
phaiosoma, *Silurus*, 377, 378
phalacra, *Brachyglanis*, 181
Phalacronotini, 367
Phalacronotus, 367, 374
Phalacronotus apogon, 374
Phalacronotus bleekeri, 374
Phalacronotus micronemus, 375
Phalacronotus micruropterus, 375
Phalacronotus parvanalis, 375
phalacronotus, *Silurus*, 374, 375
phaneronema, *Branchioica*, 411
phaneronema, *Paravandellia*, 411
phantasia, *Cetopsorhamdia*, 182
phelpsi, *Astroblepus*, 66

phelpsi, *Spatularicaria*, 296, 297
Phenacorhamdia, 188
Phenacorhamdia anisura, 188
Phenacorhamdia boliviensis, 188
Phenacorhamdia hoehnei, 188
Phenacorhamdia macarenensis, 188, 189
Phenacorhamdia nigrolineata, 189
Phenacorhamdia somnians, 189
Phenacorhamdia tenebrosa, 189
Phenacorhamdia unifasciata, 189
philippinus, *Pseudarius*, 35
phillipsi, *Clarias*, 145
pholeter, *Astroblepus*, 66
phoxocephala, *Hemiloricaria*, 246
phoxocephala, *Loricaria*, 246
Phractocephalus, 336
Phractocephalus bicolor, 336
Phractocephalus gogra, 105
Phractocephalus hemiolopterus, 336
Phractocephalus itchkeea, 386
Phractocephalus kuturnee, 105
Phractocephalus nassi, 336
Phractura, 26
Phractura ansorgii, 26
Phractura bovei, 26
Phractura brevicauda, 26
Phractura clauseni, 26
Phractura fasciata, 26
Phractura gladyuae, 26
Phractura ineac, 28
Phractura intermedia, 26
Phractura lindica, 26
Phractura longicauda, 26
Phractura lukugae, 26
Phractura macrura, 27
Phractura scaphyrhynchura, 27
Phractura tenuicauda, 27
Phreatobinae, 429
Phreatobius, 429
Phreatobius cisternarum, 429, 430
phreatophila, *Prietella*, 215
phrixosoma, *Plecostomus*, 298
phrixosoma, *Squaliforma*, 298
phrygiatus, *Arius*, 32
Phytonemus, 164
Phytonemus brichardi, 164
Phytonemus filinemus, 164
Phytonemus typus, 164
physacanthus, *Arius*, 36
Physailia, 360
Physailia ansorgii, 360
Physailia pellucida, 360
Physailia somalensis tanensis, 360
Physailia villiersi, 360
Physopyxis, 176
Physopyxis ananas, 176
Physopyxis cristata, 176
Physopyxis lyra, 176

piauhiae, *Loricaria*, 269
picinguabae, *Listrura*, 444
picklei, *Cetopsorhamdia*, 182
picta, *Dekeyseria*, 234
picta, *Megalechis*, 129
pictus, *Akysis*, 14
pictus, *Amphilinus*, 21
pictus, *Ancistrus*, 234, 265
pictus, *Bagrus (Sciades)*, 334
pictus, *Callichthys*, 129
pictus, *Hypostomus*, 265
pictus, *Leiarius*, 334
pictus, *Pimelodus*, 338
pictus, *Trichomycterus*, 423
pidada, *Arius*, 55
pietschmanni, *Allabenchelys*, 138
pietschmanni, *Auchenoglanis*, 162
pietschmanni, *Clariallabes*, 138
pipperi, *Cetopsorhamdia*, 186
pipperi, *Imparfinis*, 186
pilosus, *Neblinichthys*, 273
Pimelenotus, 197
Pimelenotus Vilsoni, 197, 200
Pimeletropis, 332
Pimeletropis lateralis, 332
Pimelode livrée, 213
Pimelodela garbei, 189
Pimelodella, 189
Pimelodella altipinnis, 189
Pimelodella australis, 189
Pimelodella avanhandavae, 189
Pimelodella boliviiana, 189
Pimelodella boschmai, 190
Pimelodella brasiliensis, 190, 192
Pimelodella breviceps, 190
Pimelodella buckleyi, 190
Pimelodella chagresi, 190
Pimelodella chagresi odynea, 193
Pimelodella chaparae, 190
Pimelodella cinerascens, 201
Pimelodella cochabambae, 185
Pimelodella conguetaensis, 190
Pimelodella copei, 190
Pimelodella cristata, 190
Pimelodella cruxenti, 190
Pimelodella cyanostigma, 190
Pimelodella dorseyi, 190
Pimelodella eigenmanni, 191, 193, 340
Pimelodella eigenmanniorum, 191
Pimelodella elongata, 191
Pimelodella enochi, 191
Pimelodella eutaenia, 191
Pimelodella figueroai, 191
Pimelodella garbei, 189
Pimelodella geryi, 191
Pimelodella gracilis, 191
Pimelodella griffini, 191
Pimelodella grisea, 191
Pimelodella hartii, 191
Pimelodella hartwelli, 192
Pimelodella hasemani, 192
Pimelodella howesi, 192
Pimelodella insignis, 190
Pimelodella itapicuruensis, 192
Pimelodella kronei, 192
Pimelodella lateristriga, 192
Pimelodella laticeps, 192
Pimelodella laticeps australis, 189
Pimelodella laurenti, 192
Pimelodella linami, 192
Pimelodella macrocephala, 192
Pimelodella macturki, 192
Pimelodella martinezii, 193
Pimelodella meeki, 193
Pimelodella megalops, 193
Pimelodella megalura, 193
Pimelodella metae, 193
Pimelodella modesta, 193
Pimelodella montana, 193
Pimelodella mucosa, 193
Pimelodella nigrofasciata, 193
Pimelodella notomelas, 193
Pimelodella odynea, 193
Pimelodella ophthalmica, 193
Pimelodella pallida, 194
Pimelodella pappenheimeri, 194
Pimelodella parnahybae, 194
Pimelodella parva, 194
Pimelodella pectinifer, 194
Pimelodella peruviana, 194
Pimelodella peruvense, 194
Pimelodella peruensis, 194
Pimelodella procera, 194
Pimelodella rambarrani, 181
Pimelodella rendahli, 196
Pimelodella reyesi, 194
Pimelodella roccae, 194
Pimelodella rudolphi, 194
Pimelodella serrata, 194
Pimelodella spelaea, 195
Pimelodella steindachneri, 195
Pimelodella taeniophora, 195
Pimelodella taenioptera, 195
Pimelodella tapatapae, 195
Pimelodella transitoria, 195
Pimelodella vittata, 195
Pimelodella wesselii, 195
Pimelodella witmeri, 195
Pimelodella yuncensis, 195
Pimelodes filamentosus, 330
Pimelodes fossor, 431
Pimelodes lateristrigus, 192
Pimelodes macropterus, 332
Pimelodidae, 329
Pimelodina, 336
Pimelodina flavipinnis, 336
Pimelodina goeldii, 332

- Pimelodina nasus*, 336
Pimelodinae, 329
Pimelodini, 329
Pimelodon, 211
Pimelodon insignarius, 211, 213
Pimelodus, 336
Pimelodus (Bagrus) maculatus, 345
Pimelodus (Pimelodella) eigenmanni, 191, 193
Pimelodus (Pimelodella) griseus, 191
Pimelodus (Pimelodella) taeniophorus, 195
Pimelodus (Pimelodina) flavipinnis, 336
Pimelodus (Pimelodus) grosskopfii, 338
Pimelodus (Pseudopimelodus) cottooides, 354
Pimelodus (Pseudopimelodus) pulcher, 355
Pimelodus (Pseudorhamdia) brasiliensis, 190
Pimelodus (Pseudorhamdia) Chagresi, 190
Pimelodus (Pseudorhamdia) Hartii, 191
Pimelodus (Pseudorhamdia) nigribarbis, 335
Pimelodus (Pseudorhamdia) Wesselii, 195
Pimelodus (Rhamdia) Baronis Müller, 201
Pimelodus (Rhamdia) brachypterus, 198
Pimelodus (Rhamdia) Cuyabae, 201
Pimelodus (Rhamdia) Knerii, 199
Pimelodus (Rhamdia) longicauda, 185
Pimelodus (Rhamdia) Parahybae, 201
Pimelodus (Rhamdia) parvus, 180
Pimelodus (Rhamdia) Queleni cuprea, 201
Pimelodus absconditus, 336
Pimelodus aeneus, 215
Pimelodus affinis, 208
Pimelodus ailurus, 205
Pimelodus albicans, 337
Pimelodus albidus, 40, 204
Pimelodus albofasciatus, 337
Pimelodus altipinnis, 189
Pimelodus altissimus, 337
Pimelodus angius, 359
Pimelodus anisurus, 98
Pimelodus antiquus, 207
Pimelodus antoniensis, 205
Pimelodus aor, 106
Pimelodus arekaima, 334
Pimelodus argenteus, 53
Pimelodus argyrus, 209
Pimelodus arius, 34
Pimelodus asperus, 395
Pimelodus atrarius, 205
Pimelodus atrobrunneus, 337
Pimelodus auratus, 155
Pimelodus bagarius, 383
Pimelodus bahianus, 195
Pimelodus balayi, 162
Pimelodus barbancho, 339
Pimelodus barbus, 44
Pimelodus batasio, 84, 85
Pimelodus Batasius, 85, 98
Pimelodus bathyurus, 201
Pimelodus bilineatus, 107
Pimelodus biscutatus, 154
Pimelodus blochii, 336, 337
Pimelodus borneensis, 41
Pimelodus botius, 388
Pimelodus Boucardi, 201
Pimelodus brachycephalus, 198
Pimelodus breviceps, 190
Pimelodus brevis, 337
Pimelodus buckleyi, 190
Pimelodus Bufonius, 355
Pimelodus caerulescens, 81
Pimelodus cœrulescens, 81
Pimelodus Cantonensis, 431
Pimelodus carcio, 85, 98
Pimelodus Carnaticus, 383
Pimelodus catulus, 205
Pimelodus cauda-furcatus, 209
Pimelodus cavasius, 95
Pimelodus cavia, 388
Pimelodus cenia, 386
Pimelodus chandramara, 86
Pimelodus charus, 355
Pimelodus cinerascens, 200
Pimelodus clarias, 320
Pimelodus clarias coprophagus, 337
Pimelodus coerulescens, 81
Pimelodus Commersonii, 43, 44
Pimelodus confinis, 205
Pimelodus conirostris, 428
Pimelodus conta, 383, 384
Pimelodus coprophagus, 337
Pimelodus cranchii, 156
Pimelodus cristatus, 189, 190
Pimelodus ctenodus, 332
Pimelodus cupreoides, 205
Pimelodus cupreus, 215
Pimelodus cyanochloros, 392
Pimelodus cyclopum, 64
Pimelodus dekayi, 206
Pimelodus Deppei, 200
Pimelodus eigenmanni, 340
Pimelodus elongatus, 191
Pimelodus eques, 184
Pimelodus exsudans, 196
Pimelodus felinus, 205
Pimelodus felis, 206
Pimelodus foina, 197
Pimelodus fur, 337
Pimelodus furcatus, 208, 209
Pimelodus furcifer, 210
Pimelodus gagata, 385, 386
Pimelodus gagora, 34
Pimelodus gambensis, 54
Pimelodus garciabarrigai, 337
Pimelodus garcia-barrigai, 337
Pimelodus gaudryi, 432
Pimelodus genidens, 43, 44
Pimelodus godmanni, 200
Pimelodus gracilis, 191, 196, 210
Pimelodus graciosus, 210
Pimelodus grosskopfii, 338
Pimelodus grosskopfii navarroi, 338
Pimelodus grunnieni, 336
Pimelodus guatemalensis, 200
Pimelodus guirali, 162
Pimelodus gulio, 93, 95
Pimelodus guttatus, 88, 162, 163
Pimelodus Hammondii, 210
Pimelodus hara, 395, 396
Pimelodus heraldoi, 338
Pimelodus heteropleurus, 181
Pimelodus Hilarii, 200
Pimelodus holomelas, 199
Pimelodus houghi, 210
Pimelodus hoyi, 206
Pimelodus humilis, 198
Pimelodus hypselurus, 198
Pimelodus indicus, 19
Pimelodus insigne, 213
Pimelodus insignis, 339
Pimelodus jatius, 41
Pimelodus javus, 431
Pimelodus jenynsii, 196
Pimelodus jivaro, 338
Pimelodus labrosus, 334
Pimelodus lateristrigus, 192
Pimelodus laticaudus, 198
Pimelodus laticeps, 160
Pimelodus lemniscatus, 213
Pimelodus leptus, 332
Pimelodus limosus, 215
Pimelodus lividus, 205
Pimelodus longifilis, 338
Pimelodus lupus, 209
Pimelodus lutescens, 215
Pimelodus lynx, 204
Pimelodus maculatus, 209, 336, 338, 345
Pimelodus managuensis, 199
Pimelodus mangois, 18
Pimelodus mangurus, 355
Pimelodus Manillensis, 41
Pimelodus marginatus, 206
Pimelodus marmoratus, 206
Pimelodus megalops, 210

- Pimelodus melanogaster*, 12, 13
Pimelodus melas, 205
Pimelodus membranaceus, 309
Pimelodus menoda, 89
Pimelodus micropterus, 200
Pimelodus microstoma, 338
Pimelodus modestus, 193
Pimelodus mong, 55
Pimelodus motaguensis, 198
Pimelodus mülleri, 199
Pimelodus multiradiatus, 334
Pimelodus murius, 358
Pimelodus musculus, 200
Pimelodus mustelinus, 184
Pimelodus mysteriosus, 338
Pimelodus namdia, 200
Pimelodus nangra, 396, 397
Pimelodus natalis, 205
Pimelodus navarroi, 338
Pimelodus nebulosus, 205, 215
Pimelodus nella, 52
Pimelodus nenga, 47
Pimelodus nicaraguensis, 199
Pimelodus nigricans, 208, 209, 210
Pimelodus nigrodigitatus, 158
Pimelodus nigrofasciatus, 193
Pimelodus notatus, 210, 339
Pimelodus occidentalis, 154
Pimelodus octocirrhus, 431
Pimelodus olivaceus, 210
Pimelodus ophthalmicus, 194
Pimelodus ornatus, 338
Pimelodus ortmanni, 338
Pimelodus pallidus var. *Lateralis*, 210
Pimelodus pallidus var. *Leucoptera*, 210
Pimelodus pallidus var. *Marginata*, 210
Pimelodus pangasius, 325, 327
Pimelodus paranaensis, 338
Pimelodus pati, 334
Pimelodus pectinidens, 46
Pimelodus Pentlandii, 200
Pimelodus petenensis, 198
Pimelodus pictus, 338
Pimelodus pinirampus, 339
Pimelodus pirinampu, 339
Pimelodus platanus, 335
Pimelodus platespogon, 383
Pimelodus platicirris, 339
Pimelodus platycephalus, 206
Pimelodus platychir, 21, 22
Pimelodus platypogon, 392
Pimelodus platypogonides, 392
Pimelodus pleurostigma, 13
Pimelodus polycaulus, 198
Pimelodus pullus, 205
Pimelodus puma, 205
Pimelodus punctatus, 339
Pimelodus punctulatus, 216
Pimelodus pusillus, 431
Pimelodus quelen, 200
Pimelodus rama, 105
Pimelodus raninus, 352, 353
Pimelodus rigidus, 339
Pimelodus rita, 106
Pimelodus Rogersi, 198
Pimelodus rugosus, 13
Pimelodus Sadlieri, 431
Pimelodus sagor, 45
Pimelodus salvini, 198
Pimelodus sapo, 200
Pimelodus sebae, 197, 200
Pimelodus seengtee, 97
Pimelodus Sellonis, 200
Pimelodus silondia, 366
Pimelodus sona, 45
Pimelodus Spegazzinii, 335
Pimelodus spixii, 40
Pimelodus Stegelichii, 200
Pimelodus synodontis, 313
Pimelodus tachisurus, 431
Pimelodus telchitta, 394
Pimelodus telchitta, 394
Pimelodus tengana, 86
Pimelodus tengara, 97
Pimelodus thunberg, 35
Pimelodus urua, 359
Pimelodus vacha, 358
Pimelodus valenciennis, 335
Pimelodus variegatus, 13, 14, 15
Pimelodus velifer, 203
Pimelodus versicolor, 44
Pimelodus viridescens, 395
Pimelodus viscosus, 215
Pimelodus vulgaris, 206
Pimelodus vulpeculus, 206
Pimelodus vulpes, 211
Pimelodus wagneri, 201
Pimelodus Westermannii, 329, 330
Pimelodus wuchereri, 200
Pimelodus Xanthocephalus, 207
Pimelodus zonatus, 13
Pimelodus zungaro, 344
Pimelotus, 197
pinheiroi, Corydoras, 122
Pinirampidae, 329
Pinirampus, 329, 339
Pinirampus pirinampu, 339
Pinirampus typus, 339
pinirampus, *Pimelodus*, 339
pinnatus, Ompok, 373
pinnimaculatus, *Aelurichthys*, 38
pinnimaculatus, *Bagre*, 38
Pinnivallago, 375
Pinnivallago kanpurensis, 375
piperata, *Tympanopleura*, 68, 70
piperatus, Ageneiosus, 70
piperatus, Imparfinis, 185, 186
piperatus Kryptopterus, 370
pipri, Eretistoides, 384
pipri, Eretistoides montana, 384
pique, Hatcheria, 405
piracicabae, Loricaria, 269
Piramutana, 330
Piramutana macospila, 339
Pirarara, 336
Pirarara bicolor, 336
pirareta, Ancistrus, 224
piratatu, Hypostomus, 258
Piratinga, 330
Piratinga pirá-aíba, 330
Pirauáca, Sorubim, 343
pirá-aíba, Piratinga, 330
piresi, Glanidium, 78
piresi, Tocantinsia, 78
piriformis, Ancistrus, 224
pirinampu, Pimelodus, 339
pirinampu, Pinirampus, 339
Pirinampus, 339
Pirinampus agassizii, 339
pirrense, Cyclopium, 66
pirrensis, Astroblepus, 66
piscatrix, Pseudorhamdia, 337
pitmani, Chrysichthys, 160
piurae, Pygidium punctulatum, 421
piurae, Trichomycterus, 421
placidus, Noturus, 214
planicauda, Parotocinclus, 283
planiceps, Ancistrus, 264
planiceps, Arius, 50
planiceps, Bagrus, 90
planiceps, Clarias, 147
planiceps, Hemibagrus, 90
planiceps, Notarius, 50
planiceps, Platystoma, 343
planiceps, Sorubimichthys, 343
planifrons, Genidens, 44
planifrons, Netuma, 44
Planiloricaria, 217, 285
Planiloricaria cryptodon, 285
Planiloricariina, 217
planquettei, Lithoxus, 267
planquettei, Lithoxus (Paralithoxus), 267
planus, Otolithus (Arius), 435
planus, Tachysurus, 434, 435
platana, Bergiaria, 330
platana, Bergiella, 330
platanum, Megalonema, 335
platanus, Pimelodus, 335
platespogon, Pimelodus, 383
platicirris, Pimelodus, 339
platorynchus, Farlowella, 239
platus, Amarginops, 153
Platyallabes, 152
Platyallabes tihoni, 152

Platycephaloidea, 139
platycephalum, *Megalonema*, 335
platycephalum psammium,
 Megalonema, 335
platycephalus, *Acrochordonichthys*,
 12, 13
platycephalus, *Amblyceps*, 18
platycephalus, *Ameiurus*, 206
platycephalus, *Chaetostomus*, 231
platycephalus, *Chrysichthys*, 158
platycephalus, *Clarias*, 139, 147
platycephalus, *Cordylancistrus*, 23
platycephalus, *Hemiodon*, 288
platycephalus, *Heterobranchus*, 151
platycephalus, *Miuroglanis*, 409
platycephalus, *Noturus*, 212
platycephalus, *Pimelodus*, 206
platycephalus, *Pseudohemiodon*, 288
platychair, *Amphilius*, 22
platychair, *Pimelodus*, 21, 22
platychair cubangoensis, *Amphilius*, 23
Platyclarias, 152
Platyclarias machadoi, 152
Platydoras, 176
Platydoras armatus, 176
Platydoras costatus, 176
Platyglanis, 164
Platyglanis depierrei, 164
platymetopon, *Loricariichthys*, 271
platynema, *Brachyplatystoma*, 331
Platynematicichthys, 339
Platynematicichthys notatus, 339
platynemum, *Brachyplatystoma*, 331
Platypogon, 339
Platypogon caerulorostris, 339, 340
platypogon, *Arius*, 54
platypogon, *Glyptothorax*, 392
platypogon, *Kryptopterus*, 371
platypogon, *Micropogon*, 371
platypogon, *Pimelodus*, 392
platypogon, *Sciades*, 54
platypogonides, *Glyptothorax*, 392
platypogonides, *Pimelodus*, 392
platypogonoides, *Glyptothorax*, 392
platyprosopos, *Clariallabes*, 138
platyrhynchos, *Hemisorubim*, 333
platyrhynchos, *Platystoma*, 333
platyrhynchus, *Cordylancistrus*, 232
platyrhynchus, *Hemiancistrus*, 232
platyrhynchus, *Ompok*, 373
Platysilurus, 340
Platysilurus barbatus, 340
Platysilurus malarmo, 340
Platysilurus mucosus, 340
Platysilurus olallae, 340
Platysomatos, 61
Platystacinae, 57
Platystacus, 57, 61
Platystacus anguillaris, 350, 351

Platystacus chaca, 135, 136
Platystacus cotylephorus, 61
Platystacus laevis, 58
Platystacus nematophorus, 61
Platystacus verrucosus, 58, 60
Platystoma, 342
Platystoma affine, 330
Platystoma artedii, 341
Platystoma corruscans, 341
Platystoma coruscans, 341
Platystoma emarginatum, 332
Platystoma juruense, 331
Platystoma Luceri, 342
Platystoma lütkeni, 344
Platystoma mucosa, 340
Platystoma orbignianus, 341
Platystoma orbignyanum, 341
Platystoma panthale, 341, 342
Platystoma parahybae, 344
Platystoma pardale, 341
Platystoma pardalis, 341, 342
Platystoma planiceps, 343
Platystoma platyrhynchos, 333
Platystoma punctifer, 342
Platystoma seenghala, 107
Platystoma spatula, 343
Platystoma sturio, 340
Platystoma tigrinum, 341
Platystoma truncatum, 341
Platystoma Vaillantii, 331
platystoma, *Cteniloricaria*, 233
platystoma, *Loricaria*, 233
Platystomatichthys, 340
Platystomatichthys sturio, 340
platystomus, *Arius*, 52
platystomus, *Plicofollis*, 52
Platytropius, 361
Platytropius longianalis, 357
Platytropius siamensis, 361
Platytropius sinensis, 358
platyura, *Hemiloricaria*, 246
platyura, *Loricaria*, 246
plazaii, *Vandellia*, 427
plecostomoides, *Cochliodon*, 258
plecostomoides, *Hypostomus*, 258
Plecostomus, 250
Plecostomus (Carinotus) carinotus,
 234, 235
Plecostomus (Rhinelepis) microps, 281
Plecostomus aculeatus, 224
Plecostomus affinis, 251
Plecostomus āgnā, 251
Plecostomus albopunctatus, 251
Plecostomus ancistroides, 251
Plecostomus angipinnatus, 251
Plecostomus angulicauda, 235
Plecostomus annae, 297
Plecostomus argus, 251
Plecostomus aspilogaster, 251
Plecostomus bicirrus, 259
Plecostomus biseriatus, 297
Plecostomus boliviensis, 252
Plecostomus borellii, 252
Plecostomus boulengeri, 252
Plecostomus brasiliensis, 259
Plecostomus brevicauda, 252
Plecostomus brevis, 252
Plecostomus carinatus, 252
Plecostomus carvalhoi, 252
Plecostomus chaparae, 226
Plecostomus commersonii scabriceps,
 260
Plecostomus commersonoides, 255
Plecostomus cordovae, 258
Plecostomus derbyi, 253
Plecostomus festae, 263
Plecostomus flagellaris, 268
Plecostomus fluvialis, 253
Plecostomus francisci, 254
Plecostomus garmani, 254
Plecostomus gomesi, 298
Plecostomus goyazensis, 254
Plecostomus gymnorhynchus, 254
Plecostomus hemiurus, 254
Plecostomus Hermanni, 254
Plecostomus heylandi, 263
Plecostomus hondae, 250, 254
Plecostomus iheringi, 298
Plecostomus iheringii, 255, 298
Plecostomus interruptus, 255
Plecostomus jaguribensis, 255
Plecostomus johnii, 255
Plecostomus lacerta, 263
Plecostomus laplatae, 255
Plecostomus latirostris, 255
Plecostomus lexi, 255
Plecostomus lima, 256
Plecostomus lima atropinnis, 251
Plecostomus limosus, 253
Plecostomus longiradiatus, 256
Plecostomus luteomaculatus, 256
Plecostomus luteus, 256
Plecostomus Lütkeni, 251
Plecostomus macrops, 256
Plecostomus madeiræ, 225
Plecostomus margaritifer, 256
Plecostomus margaritifer butantanis,
 256
Plecostomus meleagris, 256
Plecostomus microps, 273, 281
Plecostomus micropunctatus, 225
Plecostomus myersi, 257
Plecostomus niger, 257
Plecostomus nigromaculatus, 257
Plecostomus niveatus, 234
Plecostomus nudiventris, 257
Plecostomus obtusirostris, 218
Plecostomus paulinus, 258

- Plecostomus pellegrini*, 289
Plecostomus phrixosoma, 298
Plecostomus plecostomus panamensis, 243
Plecostomus plecostomus papariae, 258
Plecostomus popoi, 225
Plecostomus pusarum, 259
Plecostomus rachovii, 255
Plecostomus regani, 259
Plecostomus Robini, 261
Plecostomus rondini, 259
Plecostomus rondoni, 259
Plecostomus scaphiceps, 260
Plecostomus scaphiceps, 260
Plecostomus scopularius, 298
Plecostomus seminudus, 260
Plecostomus spilosoma, 283
Plecostomus spilurus, 283
Plecostomus spiniger, 252
Plecostomus spinosissimus, 263
Plecostomus strigaticeps, 260
Plecostomus tenuicauda, 298
Plecostomus ternetzi, 261
Plecostomus tietensis, 261
Plecostomus topavae, 261
Plecostomus Una, 261
Plecostomus unicolor, 225
Plecostomus vaillanti, 261
Plecostomus variipictus, 261
Plecostomus varimaculosus, 261
Plecostomus variostictus, 261
Plecostomus vermicularis, 262
Plecostomus villarsi, 298
Plecostomus virescens, 298
Plecostomus Wertheimeri, 285
Plecostomus winzi, 262
Plecostomus wuchereri, 262
plecostomus, *Acipencer*, 258
plecostomus, *Hypostomus*, 258
plecostomus, *Plecostomus*, 243
plecostomus panamensis, *Plecostomus*, 243
plecostomus papariae, *Plecostomus*, 258
Plectrochilus, 411
Plectrochilus diabolicus, 411
Plectrochilus machadoi, 411
Plectrochilus wieneri, 411
Pleurophysus, 410
Pleurophysus hydrostaticus, 410
pleuros, *Arius*, 39
pleuros, *Synodontis*, 319
pleurostigma, *Pimelodus*, 13
pleurotaenia, *Farlowella*, 237
pleurotaenia, *Pangasius*, 328
pleurotaenia, *Pseudolais*, 328
Plicofollis, 32, 51
Plicofollis argyroleuron, 51
Plicofollis crossocheilos, 52
Plicofollis dussumieri, 52
Plicofollis nella, 52
Plicofollis platystomus, 52
Plicofollis polystrophodon, 52
Plicofollis tenuispinis, 52
Plicofollis tonggol, 52
pliocaenicus, *Silurus*, 432
Pliosilurus, 430
Pliosilurus primus, 430
Plotoseus, 350
Plotoseus ikapor, 350
Plotosichthyoidei, 345
Plotosidae, 345
Plotosis, 350
Plotosius, 350
Plotosus, 345, 350
Plotosus (Clarias) hamiltonii, 140
Plotosus (Tandanus) tandanus, 351, 352
Plotosus abbreviatus, 351
Plotosus albilabris, 349
Plotosus argenteus, 345
Plotosus brevibarbus, 351
Plotosus caesi, 350
Plotosus canius, 350
Plotosus castaneoides, 351
Plotosus castaneus, 351
Plotosus elongatus, 346
Plotosus fisadoha, 350
Plotosus flavolineatus, 345
Plotosus horridus, 350
Plotosus laticeps, 345
Plotosus limbatus, 108, 346, 350
Plotosus lineatus, 350
Plotosus lineatus, 351, 352
Plotosus macrocephalus, 346
Plotosus macropthalmus, 349
Plotosus malignus, 351
Plotosus marginatus, 350
Plotosus megastomus, 345, 346
Plotosus microceps, 346
Plotosus multiradiatus, 350
Plotosus nigricans, 352
Plotosus nkunga, 351
Plotosus papuensis, 351
Plotosus thunbergianus, 350
Plotosus unicolor, 350, 352
Plotosus vittatus, 350
Plotosus viviparus, 350
plumbea, *Cetopsis*, 133
plumbeus, *Cetopsis*, 132, 133
plumbeus, *Trichomycterus*, 421
plumbeus motatanensis,
Pseudocetopsis, 132
plumbeus orinoco, *Pseudocetopsis*, 132
pluriradiatus, *Hemibagrus*, 90
pluriradiatus, *Macrones*, 90
pluriradiatus, *Ompok*, 373
poecilopterus, *Bagrus*, 93
poecilopterus, *Leiocassis*, 93
poecilus, *Aspidoras*, 109
poecilus, *Microglanis*, 354
poensis, *Clarias*, 141
poeyanus, *Trichomycterus*, 423
poeyi, *Rhamdia*, 199
Pogonopoma, 285
Pogonopoma obscurum, 285
Pogonopoma parahybae, 285
Pogonopoma wertheimeri, 285
Pogonopomoides, 285
poilanei, *Pareuchiloglanis*, 398, 399
pojeri, *Chiloglanis*, 307
polli, *Chrysichthys*, 159
polli, *Malapterurus*, 302
polli, *Microsynodontis*, 310
polli, *Synodontis*, 319
polycaulus, *Pimelodus*, 198
polygramma, *Doras*, 168
polyochrus, *Parotocinclus*, 283
polyodon, *Chiloglanis*, 307
polyodon, *Synodontis*, 319
polypogon, *Chiloglanis*, 307
polystaphylodon, *Arius*, 52
polystaphylodon, *Plicofollis*, 52
polystictus, *Agenciosus*, 70
polystictus, *Ageneius*, 70
polystictus, *Ageneiosus*, 70
polystictus, *Corydoras*, 123
polystigma, *Synodontis*, 319
polyuranodon, *Pangasius*, 325, 327
ponderosus, *Pangasius*, 327
poonaensis, *Glyptothorax conirostre*, 395
popoi, *Plecostomus*, 185
Porcinae, 81
Porcus, 81, 83
Porcus auratus, 155
Porcus bayad, 83
Porcus bayad macropterus, 83
Porochilus, 351
Porochilus meraukensis, 351
Porochilus obbesi, 351
Porochilus rendahli, 345
porosus, *Silurus*, 95
porosus, *Trachycorystes*, 81
porphyreus, *Ageniosus*, 70
pospisili, *Cochliodon*, 254
Potamarius, 53
Potamarius grandoculis, 53
Potamarius izabalensis, 53
Potamarius nelsoni, 53
potaroensis, *Corydoras*, 123
potaroensis, *Myoglanis*, 187
potaroënsis, *Myoglanis*, 187
potschi, *Trichomycterus*, 421
pradensis, *Trichomycterus*, 422

praecox, *Pseudocetopsis*, 133
praecox, *Denticetopsis*, 133
praelitorum, *Astroblepus*, 66
praelongus, *Hassar*, 173
praelongus, *Leptodoras*, 173
Prajadhipokia, 94
Prajadhipokia rex, 94
prashadi, *Akysis*, 14
prashadi, *Glyptothorax*, 393
prata, *Parotocinclus*, 283
prateri, *Clariasoma*, 357
pratti, *Macrones*, 102
pratti, *Pseudobagrus*, 102
prenadilla, *Brontes*, 63, 66
prenadillus, *Astroblepus*, 66
prentissgrayi, *Dinopterooides*, 139, 146
pretoriae, *Chiloglanis*, 307
prianiomus, *Rhinodoras*, 176
pricei, *Ictalurus*, 209
pricei, *Villarius*, 208, 209
Prietella, 215
Prietella lundbergi, 215
Prietella phreatophila, 215
primaevus, *Ameiurus*, 207
primaevus, *Propygidium*, 433
primus, *Pliosilurus*, 430
prionotos, *Corydoras*, 130
prionotos, *Scleromystax*, 130
Pristiancistrus, 219
Pristiancistrus eustictus, 219, 221
pristos, *Imparfinis*, 186
procera, *Pimelodella*, 194
productus, *Chiloglanis*, 443
Proeutropiichthys, 361
Proeutropiichthys buchanani, 361
Proeutropiichthys goongwaree, 362
Proeutropiichthys macrophthalmos, 362
Proeutropiichthys taakree, 362
Proeutropiichthys taakree burmanicus, 362
Proeutropius, 362
prolatus, *Encheloclarias*, 150
prolixa, *Loricaria*, 286
prolixa, *Proloricaria*, 286
prolixa lentiginosa, *Loricaria*, 286
Proloricaria, 286
Proloricaria lentiginosa, 286
Proloricaria prolixa, 286
promagdalena, *Brachyplatystoma*, 331
proops, *Bagrus*, 54
proops, *Ituglanis*, 408
proops, *Sciades*, 54
proöps, *Trichomycterus*, 408
proops parahybae, *Pygidium*, 406, 407
Prophagorus, 139
Propimelodus, 340

Propimelodus caesi, 443
Propimelodus eigenmanni, 340
Propseudecheneis, 400
Propseudecheneis tchangi, 400, 401
Propygidium, 403, 433
Propygidium primaevus, 433
prosthistius, *Amiurus*, 205
proxima, *Netuma*, 49
proximus, *Arius*, 48, 49
psammatides, *Aspidoras*, 110
psammium, *Megalonema*, 335
psammium, *Megalonema platycephalum*, 335
Psammophiletria, 27
Psammophiletria delicata, 27
Psammophiletria nasuta, 27
Pseudacanthicini, 217
Pseudacanthicus, 217, 286
Pseudacanthicus (Lithoxus) fimbriatus, 237
Pseudacanthicus fimbriatus, 236
Pseudacanthicus fordii, 286
Pseudacanthicus histrix, 286
Pseudacanthicus leopardus, 286
Pseudacanthicus serratus, 286
Pseudacanthicus spinosus, 286
Pseudogeneios, 68
Pseudancistrus, 286
Pseudancistrus atratoensis, 235
Pseudancistrus barbatus, 287
Pseudancistrus brevispinis, 287
Pseudancistrus carnegiei, 235
Pseudancistrus coquenani, 287
Pseudancistrus depressus, 287
Pseudancistrus genisetiger, 287
Pseudancistrus guentheri, 287
Pseudancistrus longispinis, 287
Pseudancistrus luderwaldti, 280
Pseudancistrus niger, 287
Pseudancistrus nigrescens, 287
Pseudancistrus orinoco, 287
Pseudancistrus papariae, 287
Pseudancistrus pediculatus, 235
Pseudancistrus pediculatus cobrensis, 235
Pseudancistrus sidereus, 288
Pseudancistrus torbesensis, 231, 232
Pseudariodes, 336
Pseudariodes pantherinus, 339
Pseudarius, 34
Pseudarius microcephalus, 35
Pseudarius philippinus, 35
Pseudauchenipterini, 68
Pseudauchenipterus, 68, 75
Pseudauchenipterus affinis, 76
Pseudauchenipterus flavescens, 76
Pseudauchenipterus guppyi, 76
Pseudauchenipterus jequitinhonhae, 76

Pseudauchenipterus nigrolineatus, 76
Pseudauchenipterus nodosus, 76
Pseudecheneidina, 382
Pseudecheneis, 382, 400
Pseudecheneis crassicauda, 400
Pseudecheneis eddsi, 442
Pseudecheneis immaculata, 400
Pseudecheneis immaculatus, 400
Pseudecheneis intermedius, 400
Pseudecheneis paviei, 400
Pseudecheneis serracula, 400
Pseudecheneis stenura, 442
Pseudecheneis sulcata, 400, 442
Pseudecheneis sulcatoides, 400
Pseudecheneis suppaetula, 442
Pseudecheneis sympelvica, 401
Pseudecheneis sympelvicus, 401
Pseudecheneis tchangi, 401
Pseudepapterus, 76
Pseudepapterus cucuhyensis, 76
Pseudepapterus gracilis, 76
Pseudepapterus hasemani, 76
Pseudeutropichthys, 165
Pseudeutropichthys multiradiatus, 165
Pseudeutropius, 356, 357, 362
Pseudeutropius acutirostris, 359
Pseudeutropius atherinoides, 359
Pseudeutropius atherinoides walkeri, 359
Pseudeutropius brachypterus, 362
Pseudeutropius longimanus, 362
Pseudeutropius megalops, 362
Pseudeutropius mitchelli, 362
Pseudeutropius moolenburghae, 362
Pseudeutropius murius batarensis, 357
Pseudeutropius siamensis, 361
Pseudeutropius verbeekii, 326
Pseudexostoma, 401
Pseudexostoma brachysoma, 401
Pseudexostoma yunnanense, 401
Pseudexostoma yunnanensis, 401
Pseudocanthicus (Lithoxus) fimbriatus, 237
Pseudobagarius, 16
Pseudobagarius alfredi, 16
Pseudobagarius baramensis, 16
Pseudobagarius filifer, 16
Pseudobagarius fuscus, 16
Pseudobagarius hardmani, 16
Pseudobagarius inermis, 16
Pseudobagarius leucorhynchus, 16
Pseudobagarius macronemus, 16
Pseudobagarius meridionalis, 17
Pseudobagarius nitidus, 17
Pseudobagarius pseudobagarius, 17
Pseudobagarius similis, 17
Pseudobagarius sinensis, 17
pseudobagarius, *Akysis*, 16, 17

pseudobagarius, *Pseudobagarius*, 17
Pseudobagrichthys, 82
Pseudobagrichthys macracanthus, 82
Pseudobagrus, 86, 100, 101
Pseudobagrus adiposalis, 101
Pseudobagrus albomarginatus, 101
Pseudobagrus analis, 101
Pseudobagrus aurantiacus, 101, 108
Pseudobagrus brachysoma, 429
Pseudobagrus brevianalis, 101
Pseudobagrus brevicaudatus, 102
Pseudobagrus changi, 103
Pseudobagrus chinensis, 101
Pseudobagrus chryseus, 429
Pseudobagrus emarginatus, 100
Pseudobagrus eupogoides, 107
Pseudobagrus eupogon, 100
Pseudobagrus fangi, 101
Pseudobagrus fui, 107
Pseudobagrus gracilis, 102
Pseudobagrus henryi, 100
Pseudobagrus ichikawai, 86
Pseudobagrus ikiensis, 102
Pseudobagrus intermedius, 100
Pseudobagrus koreanus, 102
Pseudobagrus kyphus, 82
Pseudobagrus medianalis, 102
Pseudobagrus nitidus, 82
Pseudobagrus nudiceps, 100
Pseudobagrus omeihensis, 102
Pseudobagrus ondon, 102
Pseudobagrus ornatus, 92
Pseudobagrus pratti, 102
Pseudobagrus taeniatus, 102
Pseudobagrus taiwanensis, 102
Pseudobagrus tenuis, 102
Pseudobagrus tokiensis, 103
Pseudobagrus trilineatus, 103
Pseudobagrus truncatus, 103
Pseudobagrus ussuriensis, 101
Pseudobagrus virgatus, 101
Pseudobagrus wangi, 108
Pseudobagrus wittenburgii, 108
Pseudobagrus wui, 101
Pseudocallophysus, 332
Pseudocanthicus (*Lithoxus*) *fimbriatus*, 237
Pseudocetopsis, 131
Pseudocetopsis bauodensis, 131
Pseudocetopsis jurubidae, 132
Pseudocetopsis orientale, 130
Pseudocetopsis plumbeus
 motatanensis, 132
Pseudocetopsis plumbeus orinoco, 132
Pseudocetopsis praecox, 133
Pseudodoras, 175
pseudogladiolus, *Farlowella*, 237
Pseudohemiodon, 288

Pseudohemiodon (*Planiloricaria*)
 cryptodon, 285
Pseudohemiodon amazonus, 288
Pseudohemiodon apithanos, 288
Pseudohemiodon devincenzi, 288
Pseudohemiodon laminus, 288
Pseudohemiodon laticeps, 288
Pseudohemiodon platycephalus, 288
Pseudohemiodon thorectes, 288
pseudohemiurus, *Hypostomus*, 259
pseudohemiurus macrophthalmus,
 Hypostomus, 256
Pseudohypophthalmus, 333
Pseudolaguvia, 382, 401
Pseudolaguvia ferula, 442
Pseudolaguvia foveolata, 401
Pseudolaguvia inornata, 401
Pseudolaguvia kapuri, 401
Pseudolaguvia muricata, 401
Pseudolaguvia ribeiroi, 401
Pseudolaguvia shawi, 401, 444
Pseudolaguvia tenebricosa, 402
Pseudolaguvia tuberculata, 402
Pseudolais, 328
Pseudolais micronemus, 328
Pseudolais pleurotaenia, 328
Pseudolais tetranema, 328
pseudoleiacanthus, *Clarias*, 147
Pseudolithoxus, 288
Pseudolithoxus anthrax, 288
Pseudolithoxus dumus, 289
Pseudolithoxus nicoi, 289
Pseudolithoxus tigris, 289
Pseudoloricaria, 289
Pseudoloricaria laeviuscula, 289
Pseudomystus, 103
Pseudomystus bombooides, 103
Pseudomystus breviceps, 103
Pseudomystus carnosus, 103
Pseudomystus flavipinnis, 103
Pseudomystus fumosus, 103
Pseudomystus fuscus, 103
Pseudomystus inornatus, 103
Pseudomystus leiacanthus, 104
Pseudomystus mahakamensis, 104
Pseudomystus moeschii, 104
Pseudomystus myersi, 104
Pseudomystus ornatus, 104
Pseudomystus robustus, 104
Pseudomystus rugosus, 104
Pseudomystus siamensis, 104
Pseudomystus sobrinus, 104
Pseudomystus stenogrammus, 104
Pseudomystus stenomus, 104
Pseudomystus vaillanti, 105
pseudonemacheir, *Imparfinis*, 186
pseudonieuhofii, *Clarias*, 147
Pseudopangasius, 325
Pseudopangasius nasutus, 327
Pseudopimelodidae, 352
Pseudopimelodinae, 352
Pseudopimelodus, 352, 355
Pseudopimelodus acanthochirus, 353
Pseudopimelodus Agassizi, 353
Pseudopimelodus albomarginatus, 353
Pseudopimelodus apurensis, 353
Pseudopimelodus bufonius, 355
Pseudopimelodus charus, 355
Pseudopimelodus mangurus, 355
Pseudopimelodus nigricauda, 353
Pseudopimelodus parahybae, 354
Pseudopimelodus pulcher, 355
Pseudopimelodus roosevelti, 355
Pseudopimelodus schultz, 355
Pseudopimelodus transmontanus, 353
Pseudopimelodus variolosus, 355
Pseudopimelodus villosus, 353
Pseudopimelodus villosus butcheri, 352
Pseudoplatystoma, 340
Pseudoplatystoma corruscans, 341
Pseudoplatystoma fasciatum, 341
Pseudoplatystoma fasciatum
 intermedium, 341
Pseudoplatystoma fasciatum nigricans, 341
Pseudoplatystoma fasciatum
 reticulatum, 341
Pseudoplatystoma tigrinum, 341
Pseudorhamdia, 336
Pseudorhamdia fur, 337
Pseudorhamdia macronema, 337
Pseudorhamdia piscatrix, 337
Pseudorhamdia vittatus, 195
Pseudorinelepis, 289
Pseudorinelepis genibarbis, 289
Pseudosilurus, 371
Pseudosilurus macrophthalmos, 374
pseudosilvinichthys, *Trichomycterus*, 422
pseudospinosus, *Neosilurus*, 349
Pseudostegophilus, 411
Pseudostegophilus haemomyzon, 412
Pseudostegophilus nemurus, 412
Pseudostegophilus paulensis, 410
Pseudostegophilus scarificator, 410
Pseudosynodontis, 311
Pseudotatia, 77
Pseudotatia parva, 77
Pseudotocinclus, 289
Pseudotocinclus intermedius, 290
Pseudotocinclus juquiae, 290
Pseudotocinclus parahybae, 290
Pseudotocinclus ribeiroi, 295
Pseudotocinclus tietensis, 290
Pseudotothyris, 290
Pseudotothyris janeirensis, 290
Pseudotothyris obtusa, 290

- Psilichthys*, 247
Psilichthys cameroni, 247, 279
psilogaster, *Hypoptopoma*, 250
Ptedoras angelis, 178
Pterobunocephalus, 62
Pterobunocephalus depressus, 62
Pterobunocephalus dolichurus, 62
Pterocryptis, 367, 375
Pterocryptis anomala, 375
Pterocryptis barakensis, 441
Pterocryptis berdmorei, 376, 377
Pterocryptis bokorensis, 376
Pterocryptis buccata, 376
Pterocryptis burmanensis, 376
Pterocryptis cochininchinensis, 376
Pterocryptis crenula, 376
Pterocryptis cucphuongensis, 376
Pterocryptis furnessi, 376
Pterocryptis gangelica, 375, 376
Pterocryptis inusitata, 377
Pterocryptis taytayensis, 377
Pterocryptis torrentis, 377
Pterocryptis verecunda, 377
Pterocryptis wynnaadensis, 377
Pterodoras, 177
Pterodoras angeli, 177, 178
Pterodoras granulosus, 177
Pterodoras rivasi, 177
Pteroglanis, 343, 387
Pteroglanis horai, 387, 390
Pteroglanis manni, 343
Pteronotidae, 180
Pteronotus, 180, 197
Pteropangasius, 328
Pteropsoglanis, 387
Pterosturisoma, 290
Pterosturisoma microps, 290
Pterygoplichthys, 290
Pterygoplichthys, 217, 290
Pterygoplichthys (*Ancistrus*)
parananus, 272
Pterygoplichthys aculeatus, 272
Pterygoplichthys ambrosetti, 291
Pterygoplichthys disjunctivus, 291
Pterygoplichthys etentaculatus, 291
Pterygoplichthys gibbiceps, 291
Pterygoplichthys joselimaianus, 291
Pterygoplichthys juvens, 291
Pterygoplichthys litoratus, 291
Pterygoplichthys multiradiatus, 291
Pterygoplichthys pardalis, 292
Pterygoplichthys parnaibae, 292
Pterygoplichthys punctatus, 292
Pterygoplichthys scrophus, 292
Pterygoplichthys undecimalis, 292
Pterygoplichthys xinguensis, 292
Pterygoplichthys zuliaensis, 292
pubescens, *Rhamdia*, 201
pucaí, *Hoffstetterichthys*, 30
puganensis, *Loricaria*, 297
puganensis, *Spatuloricaria*, 297
pulcher, *Amphilius*, 23
pulcher, *Aneistrus* (*Hemiancistrus*),
234
pulcher, *Auchenoglanis*, 162
pulcher, *Corydoras*, 123
pulcher, *Clarias*, 147
pulchra, *Dekeyseria*, 234
pulcher, *Macrones*, 97
pulcher, *Mystus*, 97
pulcher, *Pimelodus*
(Pseudopimelodus), 355
pulcher, *Pseudopimelodus*, 355
pulcher, *Synodontis*, 319
pulcher ephippiata, *Amphilius*, 23
pulex, *Ammoglanis*, 403
pulicaris, *Clarias*, 142
pullus, *Pimelodus*, 205
puma, *Pimelodus*, 205
pumilus, *Arius*, 386
pumilus, *Chiloglanis*, 30
punctata, *Ailia*, 356
punctata, *Chaetostomus* (*Ancistrus*)
cirrhosus, 222
punctata, *Harttia*, 241
punctata, *Loricaria*, 266, 292
punctata, *Steindachneria scripta*, 344
punctata, *Nangra*, 395
punctata, *Tatia*, 73
punctatissimus, *Chaetostomus*, 274
punctatissimus, *Oligancistrus*, 274
punctatissimus, *Trichomycterus*, 422
punctatum, *Glyptosternon*, 389
punctatum, *Hoplosternum*, 127, 128
punctatum, *Hypostoma*, 220
punctatum, *Megalonema*, 339
punctatum, *Steindachneridion*, 344
punctatus, *Doras* (*Corydoras*), 167
punctatus, *Auchenipterichthys*, 71
punctatus, *Auchenipterus*, 71
punctatus, *Auchenoglanis*, 163
punctatus, *Bagrus*, 91
punctatus, *Cataphractus*, 111, 123
punctatus, *Centromochlus*, 73
punctatus, *Chrysichthys*, 159
punctatus, *Clarias*, 140
punctatus, *Corydoras*, 123
punctatus, *Doras*, 167
punctatus, *Hemibagrus*, 91
punctatus, *Henonemus*, 406
punctatus, *Hypostomus*, 220, 259
punctatus, *Ictalurus*, 208, 209, 211
punctatus, *Malapterurus*, 302
punctatus, *Pachypterus*, 358
punctatus, *Parauchenoglanis*, 163
punctatus, *Pimelodus*, 339
punctatus, *Pterygoplichthys*, 266, 292
punctatus, *Silurus*, 209, 377, 379
punctatus, *Stegophilus*, 406
punctatus argentina, *Corydoras*, 122
punctatus sipaliwini, *Corydoras*, 125
puncticulatus, *Arius*, 41
punctifer, *Mystus*, 97
punctifer, *Platystoma*, 342
punctifer, *Synodontis*, 319
punctulata, *Synodontis*, 320
punctulatum piurae, *Pygidium*, 421
punctulatus, *Bagrus*, 331, 339, 340
punctulatus, *Hemiancistrus*, 244
punctulatus, *Pimelodus*, 216
punctulatus, *Synodontis*, 320
punctulatus, *Trichomycterus*, 422
punjabensis, *Glyptothonax*, 393
punjabensis, *Glyptothonax conirostris*,
393
purusiensis, *Panaqolus*, 277
purusiensis, *Panaque*, 277
pusarum, *Hypostomus*, 259
pusarum, *Plecostomus*, 259
Pusichthys, 362
pusillus, *Erethistes*, 384
pusillus, *Pimelodus*, 431
pusillus, *Trichomycterus*, 411
pussilus, *Erethistes*, 384
Pygidianops, 412
Pygidianops cuao, 412
Pygidianops eigenmanni, 412
Pygidianops magoi, 412
Pygidiidae, 427
Pygidium, 427
Pygidium alternatum, 414
Pygidium alterum, 415
Pygidium angustirostris, 413
Pygidium arleoi, 415
Pygidium atocha, 423
Pygidium bananeui, 415, 420
Pygidium bananeui maracaiboensis,
420
Pygidium barbouri, 415
Pygidium bogotense, 415
Pygidium bomboizanum, 416
Pygidium boylei, 416
Pygidium Burmeisteri, 405
Pygidium caliense, 416
Pygidium chapmani, 417
Pygidium chiltoni, 417
Pygidium conradi, 417
Pygidium davisii, 417
Pygidium dispar, 417
Pygidium dorsostriatum, 417
Pygidium dorsotriatum, 417
Pygidium emanueli emanueli, 418
Pygidium emanueli motatanensis, 421
Pygidium fassli, 418
Pygidium fasslii, 418
Pygidium florense, 414

Pygidium fuscum, 427
Pygidium gabrieli, 418
Pygidium gracilior, 407
Pygidium guianense, 418
Pygidium guianensis, 418
Pygidium hasemani, 418
Pygidium heterodontum, 418
Pygidium iheringi, 419
Pygidium immaculatum, 419
Pygidium johnsoni, 419
Pygidium latidens, 420
Pygidium latistriatum, 420
Pygidium metae, 407
Pygidium metae guayaberensis, 407
Pygidium migrans, 420
Pygidium mondolfi, 421
Pygidium oroyae, 423
Pygidium paolence, 421
Pygidium paquequerense, 421
Pygidium parkoi, 407
Pygidium proops parahybae, 406, 407
Pygidium punctulatum piurae, 421
Pygidium quechuorum, 423
Pygidium regani, 422
Pygidium reinhardti, 422
Pygidium riojanum, 422
Pygidium romeroi, 423
Pygidium santae-ritae, 423
Pygidium Schmidti, 416
Pygidium septentrionale, 424
Pygidium Spegazzinii, 423
Pygidium spilosoma, 423
Pygidium stawiarski, 423
Pygidium stellatum, 424
Pygidium stramineum, 424
Pygidium straminium, 424
Pygidium striatum, 424
Pygidium taenia transandianum, 424
Pygidium tenue, 424
Pygidium tiraquae, 423
Pygidium totae, 412
Pygidium travassosi, 414
Pygidium triguttatum, 425
Pygidium unicolor, 425
Pygidium venulosum, 425
Pygidium vermiculatum, 425
Pygidium weyrauchi, 425
Pygidium zonatum, 425
pygmaeus, *Corydoras*, 123
pygmaeus, *Silurus*, 132
Pylodictis, 215
Pylodictis limosus, 215
Pylodictis olivaris, 215
pyrineusi, *Cochliodon*, 259
pyrineusi, *Hypostomus*, 259
Pyxiloricaria, 292
Pyxiloricaria menezesi, 292
Q
quadrensis, *Rineloricaria*, 295

quadricostatus, *Silurus*, 122
quadrifilis, *Ageneiosus*, 78
quadrifilis, *Tetranemachthys*, 78
quadrimaculatus, *Silurus*, 200
quadriocellatum, *Glyptosternon*, 393
quadriocellatus, *Glyptothorax*, 393
quadriradiatus, *Bunocephalus*, 60
quadriradiatus, *Dysichthys*, 60
quadriscutis, *Arius*, 37
quadriscutis, *Aspistor*, 37
quadrizonatus, *Chasmocranus*, 183
quechuorum, *Pygidium*, 423
quelen, *Pimelodus*, 200
quelen, *Rhamdia*, 199
Queleni cuprea, *Pimelodus (Rhamdia)*, 201
Quirixys, 240
R
rabauti, *Corydoras*, 123
rabdophorus, *Sisor*, 402
Rabida, 211
rachovii, *Plecostomus*, 255
radiata, *Netuma*, 435
radiatus, *Glyptocephalus*, 432, 433
radiosus, *Entomocorus*, 443
radulus, *Rhineastes*, 430
raimundi, *Aspidoras*, 110
raimundi, *Corydoras*, 110
Raja similis, 434
Rama, 105
Rama buchanani, 105
Rama chandramara, 86
Rama rama, 105
rama, *Pimelodus*, 105
rama, *Rama*, 105
rambarrani, *Brachyrhamdia*, 181
rambarrani, *Pimelodella*, 181
ramentosus, *Bagrus (Bagrus)*, 108
ramentosus, *Pelteobagrus*, 108
ramirezi, *Hoplodoras*, 174
ramiroi, *Ituglanis*, 408
ramosus, *Trichomycterus*, 422
raninus, *Batrochoglanis*, 353
raninus, *Pimelodus*, 352, 353
ranunculus, *Ancistrus*, 224
rebeli, *Synodontis*, 320, 323
recavus, *Akysis*, 14
reddelli, *Rhamdia*, 202
Reganella, 217, 292
Reganella depressa, 293
Reganellina, 217
regani, *Anadoras*, 169
regani, *Arges*, 66
regani, *Astroblepus*, 66
regani, *Doras*, 169
regani, *Hypostomus*, 259
regani, *Hemipsilichthys*, 280
regani, *Leiocassis*, 93
regani, *Pareiorraphis*, 280

regani, *Plecostomus*, 259
regani, *Pygidium*, 422
regani, *Rhamdia*, 198
regani, *Trichomycterus*, 422
reinhardtii, *Bagropsis*, 329
reinhardtii, *Pygidium*, 422
reinhardtii, *Trichomycterus*, 422
reinhardtii, *Ochmacanthus*, 410
Reinhardtii, *Stegophilus*, 410
reini, *Liobagrus*, 19, 20
reisi, *Ancistrus*, 224
rendahli, *Aoria*, 108
rendahli, *Copidoglanis*, 345
rendahli, *Neosilurus*, 345
rendahli, *Pimelodella*, 196
rendahli, *Porochilus*, 345
rengifoii, *Astroblepus*, 66
resimus, *Astrophus*, 207
resupinata, *Synodontis*, 320
resupinatus, *Synodontis*, 320
reticulata, *Farlowella*, 239
reticulata, *Tatia*, 73
reticulatum, *Glyptosternon*, 387
reticulatum, *Pseudoplatystoma fasciatum*, 341
reticulatus, *Ameiurus*, 206
reticulatus, *Bagrus*, 330, 331
reticulatus, *Centromochlus*, 73
reticulatus, *Chiloglanis*, 307
reticulatus, *Corydoras*, 123
reticulatus, *Glyptosternon*, 387
retropinna, *Arges*, 66
retropinnatus, *Horiomyzon*, 185
retropinnis, *Bunocephalus*, 59
retropinnis, *Trichomycterus*, 422
retropinnus, *Astroblepus*, 66
reticulatus, *Glyptosternon*, 387
revelatus, *Corydoras*, 123
rex, *Prajadhipokia*, 94
reyesi, *Pimelodella*, 194
reynoldsi, *Corydoras*, 123
rhabdophorus, *Sisor*, 402
rhabdostigma, *Megalonema*, 338
rhabdurus, *Pareuchiloglanis*, 399
Rhadinoloricaria, 293
Rhadinoloricaria macromystax, 293
rhadinurus, *Ompok*, 373
rhaeas, *Ictalurus*, 210
rhaeas, *Rhineastes*, 210
Rhamdella, 196
Rhamdella aymarae, 196
Rhamdella eriarcha, 196
Rhamdella exsudans, 196
Rhamdella gilli, 196
Rhamdella ignobilis, 196
Rhamdella jenynsii, 196
Rhamdella lemai, 196
Rhamdella leptosoma, 196
Rhamdella longipinnis, 196

Rhamdella longiuscula, 196
Rhamdella montana, 196
Rhamdella papariae, 197
Rhamdella robinsoni, 197
Rhamdella rusbyi, 197
Rhamdella schultzi, 187
Rhamdella straminea, 196
Rhamdella wolfi, 197
Rhamdia, 197
Rhamdia alfaroi, 199
Rhamdia amatitlanensis, 198
Rhamdia barbata, 201
Rhamdia branneri, 201
Rhamdia branneri voulezi, 201
Rhamdia bransfordii, 201
Rhamdia cabrerae, 198
Rhamdia cyanostigma, 190
Rhamdia depressa, 201
Rhamdia dorsalis, 201
Rhamdia duquei, 202
Rhamdia eigenmanniorum, 191
Rhamdia enfurnada, 197
Rhamdia eriarcha, 196
Rhamdia foina, 197
Rhamdia gilli, 196
Rhamdia guairensis, 198
Rhamdia guasarensis, 197
Rhamdia guatemalensis decolor, 202
Rhamdia guatemalensis muriei, 202
Rhamdia guatemalensis stygaea, 202
Rhamdia heteracantha, 201
Rhamdia holomelas rupununi, 197
Rhamdia humilis, 198
Rhamdia itacaiunas, 198
Rhamdia javanica, 431
Rhamdia jequitinhonha, 198
Rhamdia laluchensis, 198
Rhamdia laticauda, 198, 199, 202
Rhamdia laticauda typhla, 198
Rhamdia laukidi, 199
Rhamdia lehmanni, 202
Rhamdia luigiana, 199
Rhamdia macuspanensis, 199
Rhamdia marthae, 181
Rhamdia micayi, 197
Rhamdia microcephala, 202
Rhamdia microps, 201
Rhamdia minuta, 186
Rhamdia mounseyi, 201
Rhamdia muelleri, 199
Rhamdia nasuta, 201
Rhamdia nicaraguensis, 199
Rhamdia oaxacae, 201
Rhamdia obesa, 199
Rhamdia ortoni, 201
Rhamdia parryi, 199
Rhamdia poeyi, 199
Rhamdia pubescens, 201
Rhamdia quelen, 199

Rhamdia reddelli, 202
Rhamdia regani, 198
Rhamdia riojae, 201
Rhamdia sacrificii, 198
Rhamdia saijaensis, 202
Rhamdia sebae Martyi, 202
Rhamdia tenella, 199
Rhamdia underwoodi, 198
Rhamdia xetequepeque, 202
Rhamdia zongolicensis, 199
Rhamdiae, 180
Rhamdioglanis, 202
Rhamdioglanis frenatus, 202
Rhamdioglanis transfasciatus, 202
Rhamdiopsis, 202
Rhamdiopsis microcephala, 202
Rhamdiopsis moreirai, 202, 203
rhami, Crossoloricaria, 232
rhegma, Mystus, 97
Rheoglanis, 164
Rheoglanis dendrophorus, 164
rheophilus, Amphilius, 23
rheophilus, Pangasius, 328
rheophilus, Sisor, 402
Rhineastes, 207, 430
Rhineastes arcuatus, 207
Rhineastes calvus, 207
Rhineastes grangeri, 432
Rhineastes peltatus, 430
Rhineastes radulus, 430
Rhineastes rhaeas, 210
Rhineastes smithii, 430
Rhinelepis, 217, 293
Rhinelepis Agassizii, 289
Rhinelepis aspera, 293
Rhinelepis levii, 255
Rhinelepis lophophanes, 276
Rhinelepis parahybae, 285
Rhinelepis rudolphi, 281
Rhinelepis strigosa, 293
Rhineloricaria morrowi, 246
Rhineloricaria petleyi, 266
Rhineloricaria wolfei, 247
Rhinobagrus, 92
Rhinobagrus dumerili, 92, 93
Rhinodoras, 178
Rhinodoras amazonum, 170
Rhinodoras boehlkei, 178
Rhinodoras dorbignyi, 178
Rhinodoras prianomus, 176
Rhinodoras teffeanus, 176
Rhinodoras thomersoni, 178
Rhinoglanina, 303
Rhinoglanis, 303, 310
Rhinoglanis typus, 310, 311
Rhinoglanis Vannutellii, 311
Rhinolepis paraguensis, 275
Rhizosomichthys, 412
Rhizosomichthys totae, 412

rhodonotus, Bagrus, 49
rhodopterygius, Bagrus, 95
rhombocephala, Harttia, 242
Rhynchodoras, 178
Rhynchodoras woodsi, 178
Rhynchodoras xingui, 178
ribeirae, Microcambeva, 409
ribeirensis, Neoplectostomus, 274
ribeiroi, Glanidium, 75
ribeiroi, Laguvia, 401
ribeiroi, Pseudolaguvia, 401
ribeiroi, Pseudotocinclus, 295
ribeiroi kapuri, Laguvia, 401
riberae, Astroblepus, 66
ribularis, Silurus, 201
ricardoae, Synodontis, 320
Ricola, 217, 293
Ricola macrops, 293
Ricolina, 217
rigidus, Pimelodus, 339
Rinelepis acanthicus, 218
Rinelepis genibarbis, 289
Rinelepis histrix, 218, 286
Rineloricaria, 217, 244, 293
Rineloricaria aequalicuspis, 293
Rineloricaria cadeae, 293
Rineloricaria castroi, 245
Rineloricaria catamarcensis, 294
Rineloricaria cubataonis, 294
Rineloricaria felipponei, 294
Rineloricaria formosa, 245
Rineloricaria hasemani, 245
Rineloricaria henselii, 294
Rineloricaria heteroptera, 293, 294
Rineloricaria jaraguensis, 294
Rineloricaria kronei, 294
Rineloricaria lanceolata, 246
Rineloricaria latirostris, 294
Rineloricaria lima, 294
Rineloricaria longicauda, 294
Rineloricaria maquinensis, 294
Rineloricaria microlepidogaster, 295
Rineloricaria microlepidota, 295
Rineloricaria misionera, 295
Rineloricaria pareiacantha, 295
Rineloricaria pentamaculata, 295
Rineloricaria quadrensis, 295
Rineloricaria steindachneri, 295
Rineloricaria strigilata, 295
Rineloricaria thrissiceps, 295
Rineloricaria uracantha, 240
Rineloricariina, 217
ringueleti, Hisonotus, 249
riojae, Rhamdia, 201
riojanum, Pygidium, 422
riojanus, Trichomycterus, 422
rios, Pangasius, 328
Rita, 81, 105
Rita Buchanani, 105, 106

- Rita chrysea*, 105
Rita gogra, 105
Rita grandis cutata, 105
Rita kuturnee, 105
Rita macracanthus, 106
Rita rita, 106
Rita sacerdotum, 106
rita, *Pimelodus*, 106
rita, Rita, 106
Ritae, 81
ritoides, *Arius*, 106
rivasi, *Apureadoras*, 177
rivasi, *Pterodoras*, 177
rivularis, *Silurus*, 201
rivulatus, *Trichomycterus*, 422
roae, *Cetopsidium*, 131
robbianus, *Synodontis*, 320
robecchii, *Clarias*, 143
robertsi, *Ariopsis*, 48
robertsi, *Arius*, 48
robertsi, *Synodontis*, 320
robinea, *Corydoras*, 124
robinii, *Hypostomus*, 259
Robinii, *Plecostomus*, 261
robinsoni, *Rhamdella*, 197
robusta, *Nangra*, 397
robusta, *Oxyloricaria*, 299
robusta, *Pareuchiloglanis*, 399
robusta, *Sundagagata*, 387, 392
robustum, *Megalonema*, 339
robustum, *Sturisoma*, 299
robustus, *Auchenipterus*, 80
robustus, *Corydoras*, 124
robustus, *Ichthaelurus*, 210
robustus, *Leiocassis*, 104
robustus, *Neosilurus*, 348
robustus, *Pareuchiloglanis*, 399
robustus, *Pseudomystus*, 104
roccae, *Pimelodella*, 194
rochai, *Aspidoras*, 108, 110
rodriguesi, *Glaphyropoma*, 405
rogersae, *Leptodoras*, 173
Rogersi, *Pimelodus*, 198
roigi, *Trichomycterus*, 423
romani, *Centromochlus*, 73
romani, *Tatia*, 73
romeroi, *Pygidium*, 423
romeroi, *Trichomycterus*, 423
roncallii, *Farlowella*, 240
rondoni, *Ageneiosus*, 69
rondini, *Hypostomus*, 259
rondoni, *Plecostomus*, 259
roosae, *Clupisoma*, 357
roosevelti, *Pseudopimelodus*, 355
roraima, *Neblinichthys*, 273
rosae, *Chasmocranus*, 183
rosei, *Astroblepus*, 66
roseopunctatus, *Hypostomus*, 259
rostrata, *Loricaria*, 245, 298, 300
rostrata, *Otothyris*, 276
rostratum, *Sturisoma*, 300
rostratus, *Arius*, 55
rostratus, *Loricariichthys*, 271
rothschildi, *Xenocara*, 221
rotundatus, *Otolithus (Arius)*, 435
rotundiceps, *Gephyroglanis*, 28
rotundiceps, *Zaireichthys*, 28
rotundifrons, *Bathyclarias*, 136
rotundifrons, *Dinopterus*, 136
rousseauxii, *Bagrus*, 331
rousseauxii, *Brachyplatystoma*, 331
royauxi, *Euchilichthys*, 309
royeroi, *Denticetopsis*, 133
ruandae, *Synodontis*, 320
rubermentus, *Glyptothonax*, 394
rudis, *Diplomystes*, 435
rudolphi, *Pareiorhina*, 281
rudolphi, *Pimelodella*, 194
rudolphi, *Rhinelepis*, 281
rueppelli, *Chrysichthys*, 159
rufescens, *Macrones*, 97
rufescens, *Mystus*, 94, 97
rufigiensis, *Synodontis*, 320
rugispinis, *Arius*, 32
rugosa, *Farlowella*, 239
rugosus, *Acrochordonichthys*, 13
rugosus, *Bunocephalus*, 60
rugosus, *Liocassis*, 104
rugosus, *Pimelodus*, 13
rugosus, *Pseudomystus*, 104
rukwaensis, *Chiloglanis*, 308
rukwaensis, *Synodontis*, 320
rukwaensis, *Synodontis zambezensis*, 320
rupestris, *Loricaria uracantha*, 240
rupestris, *Fonchiichthys*, 240
rupununi, *Rhamdia holomelas*, 197
rusbyi, *Rhamdella*, 197
Russellii, *Wallago*, 381
russi, *Arius*, 57
rutilus, *Bagarius*, 383
rutschi, *Arius*, 435
ruziziensis, *Chiloglanis*, 308
S
sabahensis, *Pangasius*, 328
sabaji, *Peckoltia*, 284
sabalo, *Arges*, 63, 67
sabalo, *Astroblepus*, 67
sabanus, *Hemibagrus*, 91
sabanus, *Kryptopterus*, 370
sabanus, *Mystus*, 91
sabanus, *Ompok*, 371
Sacchii, *Oxyglanis*, 154
Saccobranchus, 151
Saccobranchus microcephalus, 151
Saccobranchus microps, 151
sacerdotum, *Rita*, 106
Sachsedoras, 177
- Sachsedoras apurensis*, 177, 178
sacrificii, *Rhamdia*, 198
Sadlieri, *Pimelodus*, 431
saetiger, *Lasiancistrus*, 265
sagor, *Hexanematicthys*, 45
sagor, *Pimelodus*, 45
sagoroides, *Arius*, 45
saharsai, *Hara*, 396
saijaensis, *Rhamdia*, 202
saisii, *Glypto sternum*, 393
saisii, *Glyptothonax*, 393
salae, *Clarias*, 147
salathei, *Bunocephalus*, 59
salgadae, *Ancistrus*, 300
salmacis, *Scleromystax*, 130
salvini, *Pimelodus*, 198
sanagaensis, *Chiloglanis*, 308
sanches, *Corydoras*, 124
sandrae, *Cetopsis*, 133
sandrae, *Euristhmus*, 442
sanghensis, *Trachyglanis*, 28
sanguinea, *Vandellia*, 427
sanguineus, *Silurichthys*, 378
sanitwongsei, *Pangasius*, 328
santaerita, *Trichomycterus*, 423
santae-ritae, *Pygidium*, 423
santanderensis, *Astroblepus*, 67
santanderensis, *Astroblepus cyclopus*, 67
sapito, *Dupouyichthys*, 60
sapo, *Pimelodus*, 200
saramaccensis, *Corydoras*, 124
saramaccensis, *Hypostomus*, 259
sarareensis, *Corydoras*, 124
saravacensis, *Leiocassis*, 93
saravacensis, *Liocassis*, 93
sarcodes, *Cetopsis*, 133
Sarcogenys, 48
Sarcoglanidinae, 403
Sarcoglanis, 403, 412
Sarcoglanis simplex, 412
sardinhai, *Chiloglanis*, 308
sarmiento, *Stenolicmus*, 414
saropterix, *Brachyspondylus*, 82, 432
Satan, 216
Satan eurystomus, 216
satparanus, *Arius*, 56
sauli, *Denticetopsis*, 133, 134
sauteri, *Clarias*, 148
savorgnani, *Atopochilus*, 303, 304
sawrockensis, *Ameiurus*, 206
sawrockensis, *Ictalurus*, 206
saxicola, *Isbrueckerichthys*, 441
scabriceps, *Bunocephalus*, 58, 60
scabriceps, *Hypostomus*, 260
scabriceps, *Plecostomus commersonii*, 260
scaphirhynch, *Dekeyseria*, 234
scaphirhynchus, *Ancistrus*, 234

- scaphyceps**, *Hypostomus*, 260
scaphyceps, *Plecostomus*, 260
scaphyrhynchura, *Doumea*, 27
scaphyrhynchura, *Phractura*, 27
scaplyceps, *Plecostomus*, 260
scarificator, *Pseudostegophilus*, 410
schall, *Silurus*, 320
schall, *Synodontis*, 315, 320
Schilbe, 356, 362
Schilbe angolensis, 363
Schilbe auratus, 364
Schilbe banguensis, 363
Schilbe bocagii, 363
Schilbe bouvieri, 364
Schilbe brevianalis, 363
Schilbe congensis, 363
Schilbe congolensis, 364
Schilbe djemeri, 363
Schilbe durinii, 363
Schilbe emini, 364
Schilbe grenfelli, 363
Schilbe Hasselquistii, 365
Schilbe intermedius, 363
Schilbe Isidori, 365
Schilbe laticeps, 364
Schilbe mandibularis, 364
Schilbe marmoratus, 364
Schilbe micropogon, 364
Schilbe moebiusii, 364
Schilbe multitaeniatus, 365
Schilbe mystus, 365
Schilbe nyongensis, 365
Schilbe palmeri, 364
Schilbe senegalensis, 364
Schilbe senegalensis fasciata, 364
Schilbe Senegallus, 364
Schilbe steindachneri, 364
Schilbe tumbanus, 365
Schilbe uranoscopus, 362, 365
Schilbe yangambianus, 365
Schilbe zairensis, 366
Schilbeichthys, 357
Schilbeidae, 356
schilbeides, *Bagrurus*, 362, 365
schilbeides, *Hemisilurus*, 371
schilbeides, *Kryptopterus*, 371
Schilbeini, 356
Schilbeodes, 211
Schilbeodes gallowayi, 212
Schilbeodes hildebrandi, 213
Schilbeodes marginatus atrorus, 213
Schilbidae, 324, 356, 429
schilboides, *Bagrurus*, 362
schilby, *Silurus*, 431
Schillee Sykesii, 362
Schizolecis, 295
Schizolecis guntheri, 295
schlegeli, *Arius*, 56
Schlegelii, *Bagrurus*, 36, 95
Schmidelia graciliformis, 432
Schmidtii, *Callomyxstax*, 392
Schmidtii, *Pygidium*, 416
scheidei, *Silurichthys*, 378
Schomburgkii, *Davalla*, 68, 69
schomburgkii, *Chaetostomus*, 265
schomburgkii, *Lasiancistrus*, 265
schoutedeni, *Synodontis*, 321
schreineri, *Hoplosternum*, 128
schreitmüllerii, *Farlowella*, 239
schubarti, *Imparfinis*, 186
schubarti, *Nannorhamdia*, 186
schultzei, *Corydoras*, 112
schultzii, *Centromochlus*, 73
schultzii, *Leptorhamdia*, 187
schultzii, *Pseudopimelodus*, 355
schultzii, *Rhamdella*, 187
schultzii, *Zungaro zungaro*, 355
Schultzichthys, 413
Schultzichthys bondi, 413
Schultzichthys gracilis, 413
schwartzi, *Corydoras*, 124
schwartzi surinamensis, *Corydoras*, 126
schweinfurthi, *Fajumia*, 428, 429
Sciadeichthys, 53
Sciadeichthys (Selenaspis) walrechti, 53
Sciadeoides, 334
Sciadeops, 53
Sciades, 53
Sciades couma, 53
Sciades dowii, 53
Sciades herzbergii, 53
Sciades marmoratus, 334
Sciades passany, 54
Sciades platypogon, 54
Sciades proops, 54
Sciades troschelii, 53, 54
sciurus, *Arius*, 35
sclateri, *Austroglanis*, 81
sclateri, *Gephyroglanis*, 81
Scleromystax, 112, 129
Scleromystax barbatus, 129
Scleromystax macropterus, 130
Scleromystax prionotos, 130
Scleromystax salmacis, 130
Scleronema, 413
Scleronema angustirostre, 413
Scleronema minutum, 413
Scleronema operculatum, 413
scleronema, *Hemisilurus*, 368
scleronemus, *Ceratoglanis*, 368
Scobinancistrus, 296
Scobinancistrus aureatus, 296
Scobinancistrus parolispos, 296
scolopacina, *Loricaria*, 237
Scoloplacidae, 366
Scoloplacinae, 366
Scoloplax, 366, 367
Scoloplax dicra, 367
Scoloplax distolothrix, 367
Scoloplax dolicholophia, 367
Scoloplax empousa, 367
scolymus, *Lasiancistrus*, 265
scopularia, *Squaliforma*, 298
scopularius, *Plecostomus*, 298
Scorpiodoras, 178
Scorpiodoras heckelii, 178
scripta, *Steindachneria*, 344
scripta punctata, *Steindachneria*, 344
scriptum, *Steindachneridion*, 344
scrophus, *Liposarcus*, 292
scrophus, *Pterygoplichthys*, 292
sculpodon, *Hypostomus*, 260
scutatus, *Aelurichthys*, 38
sebae, *Ageneiosus*, 69
sebae, *Pimelodus*, 197, 200
sebae Martyi, *Rhamdia*, 202
secundus, *Microglanis*, 355
seducta, *Denticetopsis*, 134
seemanni, *Ariopsis*, 33
seemanni, *Arius*, 33
seenghala, *Platystoma*, 107
seenghala, *Sperata*, 107
seengee, *Mystus*, 97
Seengtee, *Pimelodus*, 97
Selenaspis, 53
Sellonis, *Pimelodus*, 200
semiaquilus, *Corydoras*, 124
semicultratus, *Sinopangasius*, 325, 326
seminuda, *Dasyloricaria*, 234
seminuda, *Loricaria filamentosa*, 234
seminudus, *Hypostomus*, 260
seminudus, *Plecostomus*, 260
semiscutatus, *Corydoras*, 125
Senegalensis, *Clarias*, 140
Senegalensis, *Heterobranchus*, 150
senegalensis, *Schilbe*, 364
senegalensis fasciata, *Schilbe*, 364
Senegallus, *Schilbe*, 364
sepikensis, *Lambertichthys ater*, 347
septentrionale, *Pygidium*, 424
septentrionalis, *Acrochordonichthys*, 13
septentrionalis, *Corydoras*, 124
septentrionalis, *Stegophilus*, 414
Septobranchus, 42
Septobranchus johannae, 42
seraoi, *Eutropius*, 363
Serdicensis, *Silurus*, 432
sericeum, *Chaetostoma*, 230
sericeus, *Chaetostomus*, 230
serpentis, *Synodontis*, 321
serracanthus, *Ameiurus*, 206
serracanthus, *Ictalurus*, 206
serracula, *Pseudecheneis*, 400

- serralabium, Callichthys*, 111
serrata, Hara, 396
serrata, Pimelodella, 194
serrata, Synodontis, 321
serratum, Amblyceps, 18
serratus, Arius, 49
serratus, Corydoras, 124
serratus, Hara, 396
serratus, Hypostomus, 286
serratus, Pseudacanthicus, 286
serratus, Synodontis, 311, 321
serratus tanganicae, Synodontis, 322
setifera, Loricaria, 268
setiger, Oreoglanis, 398
setosus, Chaetostomus, 235
setosus, Dolichancistrus, 235
seussi, Corydoras, 124
sexcirrhis, Aspredo, 61
sexpapilostoma, Hoplomyzon, 61
sextentaculatus, Heterobranchus, 197, 200
sexualis, Gagata, 386
sharavatiensis, Batasio, 86
sharpii, Chrysichthys, 159
shawi, Laguvia, 395, 402
shawi, Pseudolaguvia, 401, 444
shermani, Cetopsorhamdia, 182
shirensis, Malapterurus, 302
shirui, Hoplosternum, 128
shuwaiensis, Bagrus, 84
sialis, Noturus, 213
siamensis, Glyptothorax, 393
siamensis, Hemipimelodus, 41
siamensis, Liocassis, 104
siamensis, Oreoglanis, 397, 398
siamensis, Pangasius, 327
siamensis, Platytropius, 361
siamensis, Pseudeutropius, 361
siamensis, Pseudomystus, 104
sianenna, Chrysichthys, 159
sichuanensis, Pareuchiloglanis, 399
sicuephorus, Aspredo, 58
sicula, Eretistoides, 384
sicyephorus, Aspredo, 58
Sieboldii, Bagrus, 90
sifontesi, Oxydoras, 176
Silondia, 366
silondia, Silonia, 366
Silonia, 366
Silonia childreni, 366
Silonia silondia, 366
Silonopangasius, 366
Silundia, 366
Silundia gangetica, 366
Silundia Sykesii, 366
Siluranodon, 356, 366
Siluranodon auritus, 366
Siluranodontinae, 356
Silurichthys, 377
Silurichthys australis, 348
Silurichthys basilewskii, 381
Silurichthys berdmorei, 376
Silurichthys citatus, 377
Silurichthys gibbiceps, 377
Silurichthys hasseltii, 377
Silurichthys indragiriensis, 378
Silurichthys leucopodus, 378
Silurichthys marmoratus, 378
Silurichthys phaiosoma, 378
Silurichthys sanguineus, 378
Silurichthys schneideri, 378
Siluridae, 367
Silurodes, 371
Silurodes macronema, 372
Silurodon, 380
Silurodon hexanema, 380, 381
siluroides, Ompok, 371
Siluroides, 367
siluroides, Ompok, 373
Silurus, 375, 378
Silurus (?)gaudryi, 432
Silurus (Acanthonotus) cuvieri, 356
Silurus (Callichrus) affinis, 374
Silurus (Callichrus) erythrogaster, 373
Silurus (Callichrus) immaculatus, 374
Silurus (Callichrus) macrostomus, 381
Silurus (Callichrus) nebulosus, 374
Silurus (Callichrus) vittatus, 372
Silurus (Clarias) argentata, 357
Silurus (Heterobranchus) gariepinus, 143
Silurus (Parasilurus) aristotelis, 378
Silurus (Pimelodus) nigrescens, 210
Silurus 11-radiatus, 335
Silurus 12-radiatus, 177
Silurus 16-radiatus, 44
Silurus 38-radiatus, 70
Silurus 7-radiatus, 122
Silurus 9-radiatus, 201
Silurus afghana, 376
Silurus altus, 381
Silurus anguillaris, 138, 140, 143
Silurus anostomus, 374
Silurus apogon, 374
Silurus arab, 351
Silurus argentinus, 209
Silurus aristotelis, 378
Silurus armatus, 177
Silurus ascita, 54
Silurus asotus, 378, 381
Silurus aspredo, 58
Silurus Atherinoides, 359
Silurus athu, 380
Silurus attu, 380
Silurus auritus, 366
Silurus bagre, 37
Silurus bajad, 83
Silurus batrachus, 140
Silurus bedfordi, 379
Silurus bicirrhos, 369
Silurus bimaculatus, 372
Silurus biserratus, 151
Silurus biwaensis, 379
Silurus boalis, 380
Silurus burmanensis, 376
Silurus callarias, 313
Silurus callichthys, 110
Silurus calvarius, 99, 100
Silurus canio, 373, 374
Silurus carinatus, 170, 171
Silurus cataphractus, 167
Silurus catus, 204
Silurus cæcutiens, 131
Silurus Cerulescens, 207, 208, 211
Silurus cerulescens melanurus, 209
Silurus cerulescens var. melanurus, 209
Silurus chantrei, 379
Silurus chechra, 374
Silurus chilensis, 165
Silurus cinereus, 379
Silurus clarias, 311, 313, 321, 336, 337
Silurus Cochinchinensis, 376
Silurus coecutiens, 131, 132
Silurus congensis, 362, 363
Silurus costatus, 176
Silurus coues, 31, 387, 389
Silurus cryptopterus, 369
Silurus cucphuongensis, 376
Silurus cupreus, 215
Silurus dauuricus, 379
Silurus docmak, 84
Silurus duanensis, 379
Silurus duda, 374
Silurus dukai, 377
Silurus Egertoni, 56
Silurus electricus, 301
Silurus fasciatus, 340, 341
Silurus felis, 33
Silurus ferox, 431
Silurus fossilis, 151
Silurus galeatus, 79
Silurus garua, 357
Silurus gilberti, 375, 376
Silurus glanis, 378, 379
Silurus glanis aralensis, 379
Silurus glanis atavus, 382
Silurus goae, 374
Silurus grahami, 379
Silurus gurgu, 431
Silurus gyrinus, 211, 213, 214
Silurus hemiolopterus, 336
Silurus Herzbergii, 53
Silurus hexadactylus, 61
Silurus hexapterus, 371
Silurus hypophthalmus, 372
Silurus ichneumon, 431

- Silurus imberbis*, 70, 379
Silurus indicus, 374
Silurus inermis, 69, 379
Silurus japonicus, 378, 379
Silurus lais, 370
Silurus lamghur, 373
Silurus lanzhouensis, 380
Silurus laticeps, 151
Silurus leptonema, 374
Silurus lima, 342
Silurus limosus, 215
Silurus limpok, 370
Silurus lineatus, 350, 351
Silurus lithophilus, 380
Silurus lividus, 204, 205
Silurus lividus Fuscatus, 205
Silurus lividus var. *Fuscatus*, 205
Silurus luvur, 431
Silurus macrocephalus, 341
Silurus macronema, 371, 372
Silurus maculatus, 35, 207, 209
Silurus maculatus Erythroptera, 209
Silurus maculatus var. *Erythroptera*, 209
Silurus Malabaricus, 372
Silurus mangrullo, 341
Silurus marinus, 37, 38, 44
Silurus melas, 205
Silurus mento, 380
Silurus meridionalis, 380
Silurus microcephalus, 374
Silurus microdorsalis, 380
Silurus micronemus, 375
Silurus micropogon, 374
Silurus militaris, 51, 68, 69
Silurus minutus, 431
Silurus mollis, 214
Silurus mononema, 370
Silurus morehensis, 376
Silurus Müller, 380, 381
Silurus muticus, 337
Silurus Mysoricus, 374
Silurus mystus, 362, 365
Silurus nebulosus, 215
Silurus nodosus, 75, 76
Silurus ocellatus, 35
Silurus olivaris, 215
Silurus pabda, 371, 372
Silurus pabo, 373
Silurus palembangensis, 369, 370
Silurus pallidus, 210
Silurus pallidus Lateralis, 210
Silurus pallidus Leucoptera, 210
Silurus pallidus Marginatus, 210
Silurus pallidus var. *Lateralis*, 210
Silurus pallidus var. *Leucoptera*, 210
Silurus pallidus var. *Marginatus*, 210
Silurus parkeri, 36
Silurus pati, 335
Silurus pelusius, 96
Silurus phaiosoma, 377, 378
Silurus phalacronotus, 374, 375
Silurus pliocaenicus, 432
Silurus porosus, 95
Silurus punctatus, 209, 377, 379
Silurus pygmaeus, 132
Silurus quadricostatus, 122
Silurus quadrimaculatus, 200
Silurus ribularis, 201
Silurus rivularis, 201
Silurus schall, 320
Silurus schilbe niloticus, 431
Silurus schilby, 431
Silurus Serdicensis, 432
Silurus silurus, 379
Silurus sinensis, 375, 376, 381
Silurus singio, 151
Silurus soldatovi, 380
Silurus soldatovi meridionalis, 380
Silurus stenocephalus, 432
Silurus torrentis, 377
Silurus triostegus, 380
Silurus undecimalis, 431
Silurus unitius, 350
Silurus Vaillantii, 331
Silurus verrucosus, 60
Silurus viscosus, 215
Silurus vittatus, 98
Silurus wallagoo, 381
Silurus wynaadensis, 375, 377
Silurus Xanthocephalus, 207
Silurus xanthosteus, 379
silurus, *Silurus*, 379
Silvaichthys, 76
Silvaichthys aguilerae, 76
silvasantosi, Steindachneridion, 344
silviae, *Glyptothorax*, 393
Silvinichthys, 413
Silvinichthys bortayro, 413
Silvinichthys mendozensis, 413
simeonsi, Clariablabes, 138
similis, *Akysis*, 17
similis, Corydoras, 124
similis, *Leiocassis*, 108
similis, Pseudobagarius, 17
similis, *Raja*, 434
simillima, Loricaria, 269
simios, Hypostomus, 260
simonsi, *Galeichthys*, 33
simonsii, *Arges*, 67
simonsii, Astroblepus, 67
simplex, Sarcoglanis, 412
simplex, Tatia, 78
simpsonii, *Ictalurus*, 210
simulatus, Corydoras, 124
Simulidentinae, 303
Sindensis, *Callichrous*, 373
sindensis, Ompok, 373
sinesis, *Akysis*, 17
sinense, Clupisoma, 358
sinense, *Glyptosternum*, 393
sinensis, *Akysis*, 17
sinensis, *Arius*, 55
sinensis, *Bagrus*, 108
sinensis, *Cranoglanis*, 164, 165
sinensis, *Euchiloglanis*, 399
sinensis, Glyptothorax, 393
sinensis, *Macrones*, 165
sinensis, Pareuchiloglanis, 399
sinensis, *Platytropius*, 358
sinensis, Pseudobagarius, 17
sinensis, *Silurus*, 375, 376, 381
sinensis, *Tachysurus*, 431
singaringan, *Bagrus*, 97
singaringan, Mystus, 97
Sinopangasius, 325
Sinopangasius semicultratus, 325, 326
sinyanensis, *Leiocassis*, 108
sipaliwini, *Bunocephalus amaurus*, 58
sipaliwini, Corydoras, 125
sipaliwini, *Corydoras punctatus*, 125
sipaliwini, Hypostomus, 260
Sisor, 382, 402
Sisor barakensis, 402
Sisor chennuah, 402
Sisor rhabdophorus, 402
Sisor rheophilus, 402
Sisor tortosus, 402
Sisorichthyoidei, 382
Sisoridae, 382
Slatinia, 24
Slatinia mongallensis, 24
smithi, Farlowella, 239
smithii, *Clarias*, 143
smithii, Rhineastes, 430
smithii, *Synodontis*, 321
smiti, Synodontis, 321
sneiderni, Hemiloricaria, 246
sneiderni, *Loricaria*, 246
snethlageae, *Ancistrus*, 283, 285
snethlageae, *Peckoltia*, 285
sobrinus, Pseudomystus, 104
Socnopaea, 430
Socnopaea grandis, 430
Socnopaea horai, 431
sodalis, Corydoras, 125
soldatovi, Silurus, 380
soldatovi meridionalis, *Silurus*, 380
solidus, *Arius*, 39
solidus, Brustiarius, 39
soloni, Synodontis, 321
solox, Corydoras, 125
somalensis, *Ailia*, 360
somalensis, Parailia, 360
somalensis tanensis, *Physailia*, 360
somereni, Chiloglanis, 308

- somnians*, *Heptapterus*, 189
somnians, *Phenacorhamdia*, 189
sona, *Hemiarius*, 45
sona, *Pimelodus*, 45
sondaicus, *Bagrus*, 45
songdaensis, *Pareuchiloglanis*, 400
songmaensis, *Pareuchiloglanis*, 400
soniae, *Hypostomus*, 260
Sophiancistrus, 283
sorex, *Synodontis*, 321
Sorubim, 281, 342
Sorubim caparary, 341
Sorubim cuspicaudus, 342
Sorubim elongatus, 342
Sorubim infraoculare, 342
Sorubim jandia, 342, 343
Sorubim latirostris, 342
Sorubim lima, 342
Sorubim maniradii, 342
Sorubim Pirauáca, 343
Sorubim trigonocephalus, 342
Sorubimichthys, 342
Sorubimichthys ortoni, 343
Sorubimichthys planiceps, 343
Sorubinae, 329
Sorubium, 329, 342
Sosia, 12
Sosia chamaeleon, 12, 13
Sosia chamaeleon pallida, 13
Sosia chamaeleon var. *pallida*, 13
Sovichthys, 332
Sovichthys abuelo, 332
sovichthys, *Chaetostoma*, 230
sovichthys, *Chaetostoma anomala*, 230
spatula, *Arius*, 42, 43
spatula, *Cochlefelis*, 43
spatula, *Platystoma*, 343
Spatularicaria, 296
Spatularicaria atratoensis, 296
Spatularicaria caquetae, 296
Spatularicaria curvispina, 296
Spatularicaria euacanthagenys, 296
Spatularicaria evansii, 296
Spatularicaria fimbriata, 297
Spatularicaria gymnogaster, 297
Spatularicaria lagoichthys, 297
Spatularicaria nudiventris, 297
Spatularicaria phelpsi, 296, 297
Spatularicaria puganensis, 297
spectabile, *Nannoptopoma*, 272
spectabilis, *Corydoras*, 125
spectabilis, *Otocinclus*, 272, 273
Spectracanthicina, 217
Spectracanthicus, 217, 297
Spectracanthicus murinus, 297
spectrum, *Aspredo*, 61
spectrum, *Glyptothorax*, 393
Spegazzinii, *Pimelodus*, 335
Spegazzinii, *Pygidium*, 423
- spiegazzinii*, *Trichomycterus*, 423
spelaea, *Pimelodella*, 195
spelaeus, *Trichomycterus*, 423
Sperata, 106
Sperata acicularis, 106
Sperata aor, 106
Sperata aorella, 107
Sperata sarwari, 107
Sperata seenghala, 107
spilomma, *Hemiancistrus*, 244
spilopterus, *Hemibagrus*, 91
spilosoma, *Parotocinclus*, 283
spilosoma, *Plecostomus*, 283
spilosoma, *Pygidium*, 423
spilosoma, *Trichomycterus*, 423
spilotus, *Aspidoras*, 110
spilurus, *Batusio*, 442
spilurus, *Corydoras*, 125
spilurus, *Parotocinclus*, 283
spilurus, *Plecostomus*, 283
spiniger, *Plecostomus*, 252
spinipectoralis, *Mystus*, 97
spiniserrata, *Parailia*, 360
spinosae, *Loricaria*, 286
spinosisimus, *Acanthodoras*, 168
spinosisimus, *Doras*, 168
spinosisimus, *Hemiancistrus*, 244
spinosisima, *Isorineloricaria*, 263
spinosisimus, *Plecostomus*, 263
spinosus, *Ancistrus*, 224
spinosus, *Hypostomus*, 286
spinosus, *Pseudacanthicus*, 286
spixii, *Cathorops*, 40
Spixii, *Cetopsis*, 131
Spixii, *Hypophthalmus*, 333
Spixii, *Loricaria*, 270
spixii, *Pimelodus*, 40
splendens, *Brochis*, 125
splendens, *Callichthys*, 125
splendens, *Corydoras*, 125
splendens, *Hemipsilichthys*, 280
splendens, *Pareiorhaphis*, 280
spodium, *Ictalurus*, 211
spurrellii, *Imparfinis*, 186
spurrellii, *Nannorhamdia*, 185, 186
Squaliforma, 297
Squaliforma annae, 297
Squaliforma biseriata, 297
Squaliforma emarginata, 297
Squaliforma gomesi, 298
Squaliforma horrida, 298
Squaliforma phrixosoma, 298
Squaliforma scopularia, 298
Squaliforma squalina, 298
Squaliforma tenuicauda, 298
Squaliforma tenuis, 298
Squaliforma villarsi, 298
Squaliforma virescens, 298
squalina, *Squaliforma*, 298
- squalinum*, *Hypostoma*, 298
squalus, *Arius*, 166
stanauli, *Noturus*, 214, 442
stanneus, *Galeichthys*, 49
stanni, *Chaetostoma*, 230
Stannii, *Chaetostomus*, 230
stappersii, *Chrysichthys*, 159
stappersii, *Clarias*, 147
starnesi, *Cetopsis*, 133
stauroforus, *Arius*, 32
stauroforus, *Felichthys*, 32
Stauroglanis, 413
Stauroglanis gouldingi, 413
stawiarski, *Pygidium*, 423
stawiarski, *Trichomycterus*, 423
Stearopterus, 37
Stearopterus bagre, 37
steerei, *Taenionema*, 330, 331
Stegelichii, *Pimelodus*, 200
Stegophilina, 403
Stegophilus, 403, 414
Stegophilus insidiosus, 414
Stegophilus intermedius, 405
Stegophilus macrops, 406
Stegophilus maculatus, 410
Stegophilus nemurus, 412
Stegophilus panzeri, 414
Stegophilus passarelli, 406
Stegophilus punctatus, 406
Stegophilus Reinhardtii, 410
Stegophilus septentrionalis, 414
steinbachi, *Ixinandria*, 263
Steinbachi, *Loricaria*, 263
steindachneri, *Cathorops*, 40, 48
Steindachneri, *Centromochlus*, 73
steindachneri, *Corydoras*, 125
steindachneri, *Glyptosternum*, 393
steindachneri, *Glyptothorax*, 393
steindachneri, *Hemipsilichthys*, 280
steindachneri, *Hypoptopoma*, 250
steindachneri, *Loricaria*, 295
Steindachneri, *Oxydoras*, 179
steindachneri, *Opsodoras*, 169
steindachneri, *Pareiorhaphis*, 280
steindachneri, *Parotocinclus*, 283
steindachneri, *Pimelodella*, 195
steindachneri, *Rineloricaria*, 295
steindachneri, *Schilbe*, 364
steindachneri, *Synodontis*, 321
steindachneri, *Tachysurus*, 40
steindachneri, *Trachydoras*, 169
Steindachneria, 343
Steindachneria amblyurus, 343
Steindachneria doceana, 343
Steindachneria scripta, 344
Steindachneria scripta punctata, 344
Steindachneridion, 343
Steindachneridion amblyurum, 343
Steindachneridion doceana, 343

- Steindachneridion iheringi*, 343
Steindachneridion melanodermatum, 343
Steindachneridion parahybae, 344
Steindachneridion punctatum, 344
Steindachneridion scriptum, 344
Steindachneridion silvasantosi, 344
stellatum, *Leptoplosternum*, 128
stellatum, Pygidium, 424
stellatus, *Nanobagrus*, 98
stellatus, *Trichomycterus*, 424
stenocephalus, *Corydoras*, 125
stenocephalus, *Osteogeniosus*, 51
stenocephalus, *Silurus*, 432
stenogrammus, *Pseudomystus*, 104
Stenolicmus, 414
Stenolicmus sarmientoi, 414
stenomus, *Bagrus*, 103, 104
stenomus, *Pseudomystus*, 104
stenopeltis, *Doras* (*Oxydoras*), 171, 172
stenopeltis, *Hemidoras*, 172
stenura, *Pseudecheneis*, 442
stephanus, *Pareiorhaphis*, 280
sterbai, *Corydoras*, 125
sternoptychum, *Nannoptopoma*, 273
stewardii, *Hoplosternum*, 128
stewarti, *Hemiloricaria*, 246
stewarti, *Heptapterus*, 184
stewarti, *Loricaria*, 247
sthenocephalus, *Osteogeniosus*, 51
stiassnyae, *Malapterurus*, 301
stibaros, *Lamontichthys*, 264
stictonotus, *Imparfinis*, 186
stictonotus, *Nannorhamdia*, 186
stigmaticus, *Ancistrus*, 224
stigmaturus, *Mystus*, 85
stigmosus, *Noturus*, 214
stirlingi, *Arius*, 46
stocki, *Glyptothorax*, 393
stocki, *Lithoxus*, 268
Stolickae, *Glyptosternum*, 393
stolickae, *Glyptothorax*, 393
stoliczkae, *Exostoma*, 387
stomias, *Hemipsilichthys*, 281
stomias, *Pareiorhaphis*, 280
Stoneiella, 286
Stoneiella leopardus, 286
Stormi, *Hemiarius*, 45
Stormii, *Cephalocassis*, 44, 45
stormii, *Hemiarius*, 45
straminea, *Rhamdella*, 196
stramineum, Pygidium, 424
straminum, Pygidium, 424
straminius, *Trichomycterus*, 424
Strephon, 63
striatulus, *Auchenipterus* (*Pseudauchenipterus*), 80
striatulus, *Trachelyopterus*, 80
striatum, Pygidium, 424
Striatus, *Glyptosternon*, 387, 394
striatus, *Glyptothorax*, 394
striatus, *Trichomycterus*, 424
stricticassis, *Arius*, 50
strigata, *Tatia*, 78
strigaticeps, *Hypostomus*, 260
strigaticeps, *Plecostomus*, 260
strigilata, *Loricaria*, 295
strigilata, *Rineloricaria*, 295
strigosa, *Rhinelepis*, 293
strigosus, *Acrochordonichthys*, 13
stromeri, *Fajumia*, 429
stroudi, *Gelanoglanis*, 74
stroumpoulos, *Chaetostoma*, 444
stuarti, *Glyptosternum*, 385
stübeli, *Arges*, 67
Stübelii, *Loricaria*, 271
Stübelii, *Oxydoras*, 175
stuebeli, *Astroblepus*, 67
stuebelii, *Loricariichthys*, 271
stuebelii, *Opsodoras*, 175
sturio, *Platystoma*, 340
sturio, *Platystomatichthys*, 340
Sturiosoma nigrirostrum, 299
Sturisoma, 298
Sturisoma aureum, 298
Sturisoma barbatum, 299
Sturisoma brevirostre, 299
Sturisoma dariense, 299
Sturisoma festivum, 299
Sturisoma frenatum, 299
Sturisoma guentheri, 299
Sturisoma kneri, 299
Sturisoma lyra, 299
Sturisoma monopeltis, 299
Sturisoma nigrirostrum, 299
Sturisoma panamense, 299
Sturisoma robustum, 299
Sturisoma rostratum, 300
Sturisoma tenuirostre, 300
Sturisomatichthys, 300
Sturisomatichthys caquetae, 300
Sturisomatichthys citurensis, 300
Sturisomatichthys leightoni, 300
Sturisomatichthys tamanae, 300
styani, *Liobagrus*, 20
stygaea, *Rhamdia guatemalensis*, 202
Stygogenes, 63
Stygogenes guentheri, 64
Stygogenes humboldtii, 64
Suarezi, *Incaichthys*, 30
subcarinatus, *Hypostomus*, 260
submarginatus, *Clarias*, 147
submarginatus, *Loricaria*, 246
submarginatus, *Thysvillensis*, *Clarias* (*Allabenchelys*), 141
subrostratus, *Arius*, 35
subteres, *Kronichthys*, 263
subviridis, *Hemiancistrus*, 244
suchus, *Bagarius*, 383
sufii, *Glyptothorax*, 394
sufii, *Glyptothorax telchitta*, 394
sugubii, *Liobagrus*, 20
sulcata, *Pseudecheneis*, 400
sulcatooides, *Pseudecheneis*, 400
sulcatus, *Callichthys*, 129
sulcatus, *Clarias*, 147
sulcatus, *Glyptosternon*, 400
sumatrarus, *Arius*, 36
sumatrarus, *Bagrus*, 36
Sundagagata, 387
Sundagagata robusta, 387, 392
sundaicus, *Hexanematicthys*, 45
sundanensis, *Hemipimelodus*, 56
Superglyptothorax, 388
suppaetula, *Pseudecheneis*, 442
supramollis, *Astroblepus*, 67
surinamensis, *Chasmocranus*, 183
surinamensis, *Corydoras*, 126
surinamensis, *Corydoras schwartzii*, 126
surinamensis, *Harttia*, 242
surinamensis, *Heptapterus*, 183
surinamensis, *Hexanematicthys*, 33
surinamensis, *Hoplosternum thoracatum*, 129
surinamensis, *Hypostomus*, 260
surinamensis, *Lithoxus*, 268
surinamensis, *Lithoxus (Paralithoxus)*, 268
sutchi, *Pangasius*, 325
suttoni, *Panaque*, 278
suttonorum, *Panaque*, 278
swierstrai, *Chiloglanis*, 308
sychri, *Corydoras*, 126
sykesi, *Glyptosternum*, 394
sykesii, *Glyptothorax*, 394
sykesii, *Gogrius*, 105
Sykesii, *Schillee*, 362
Sykesii, *Silundia*, 366
sympelvica, *Pseudecheneis*, 401
sympelvicus, *Pseudecheneis*, 401
sympterygium, *Heptapterus*, 184
Synechoglanis, 207
Synechoglanis Beadlei, 207, 210
synodon, *Arius*, 166
Synodontes, 311
Synodontini, 303
Synodontis, 303, 311
Synodontis acanthomias, 311
Synodontis acanthoperca, 441
Synodontis afrofischeri, 311
Synodontis Afro-fischeri, 311
Synodontis alberti, 311
Synodontis albolineata, 311
Synodontis albolineatus, 311
Synodontis angelica, 311

- Synodontis angelicus*, 311
Synodontis angelicus var. *zonatus*, 311
Synodontis angelicus zonatus, 311
Synodontis annectens, 312
Synodontis ansorgii, 312
Synodontis arabi, 321
Synodontis arnoulti, 312
Synodontis aterrima, 312
Synodontis aterrimus, 312
Synodontis augierasi, 314
Synodontis bastiani, 312
Synodontis batensoda, 304
Synodontis batesi longispinis, 316
Synodontis batesi var. *longispinis*, 316
Synodontis batesii, 312
Synodontis brichardi, 312
Synodontis budgetti, 312
Synodontis camelopardalis, 312
Synodontis caudalis, 312
Synodontis caudovittata, 312
Synodontis caudovittatus, 313
Synodontis centralis, 313
Synodontis citernii, 314
Synodontis clarias, 313, 320
Synodontis colyeri, 318
Synodontis comoensis, 313
Synodontis congica, 313
Synodontis congicus, 313
Synodontis contracta, 313
Synodontis contractus, 313
Synodontis courteti, 313
Synodontis cuangoana, 313
Synodontis cuangoanus, 313
Synodontis dageti, 312
Synodontis davidi, 313
Synodontis decora, 314
Synodontis decorus, 314
Synodontis dekimepi, 314
Synodontis depauwi, 314
Synodontis dhonti, 314
Synodontis dorsomaculata, 314
Synodontis dorsomaculatus, 314
Synodontis eburneensis, 312
Synodontis euptera, 314
Synodontis eupterus, 314
Synodontis eurystomus, 305, 319
Synodontis fascipinna, 314
Synodontis filamentosa, 314
Synodontis filamentosus, 314
Synodontis flavaeniata, 314
Synodontis flavitaeniatus, 314
Synodontis frontosa, 314
Synodontis frontosus, 314
Synodontis fueleborni, 315
Synodontis galinae, 323
Synodontis gambiensis, 315
Synodontis gambiensis latifrons, 315
Synodontis geledensis, 315
Synodontis gobroni, 315
Synodontis grandiops, 445
Synodontis granulosa, 315
Synodontis granulosus, 315
Synodontis greshoffi, 315
Synodontis guentheri, 309
Synodontis guttata, 315
Synodontis guttatus, 315
Synodontis haugi, 315
Synodontis Hollyi, 323
Synodontis Hollyi ntemensis, 322
Synodontis holopercnus, 315
Synodontis humeratus, 321
Synodontis ilebrevis, 445
Synodontis irsacae, 314
Synodontis ituriensis, 315
Synodontis iturii, 315
Synodontis jallae, 316
Synodontis katangae, 315
Synodontis khartoumensis, 316
Synodontis koensis, 316
Synodontis labeo, 323
Synodontis lacustricolus, 322
Synodontis laesoei, 316
Synodontis leopardina, 316
Synodontis leopardinus, 316
Synodontis leoparda, 316
Synodontis leopardus, 316
Synodontis levequei, 316
Synodontis longirostris, 316
Synodontis longispinis, 316
Synodontis loppei, 318
Synodontis lucipinnis, 445
Synodontis lufirae, 316
Synodontis macrepipterus, 314
Synodontis macrodon, 313
Synodontis macrophthalma, 316
Synodontis macrophtalmus, 316
Synodontis macrops, 316
Synodontis macrostigma, 317
Synodontis macrostoma, 317
Synodontis maculatus, 318
Synodontis maculipinna, 320
Synodontis maculosus, 311, 318, 321
Synodontis manni, 317
Synodontis marmorata, 317
Synodontis marmoratus, 317
Synodontis marmoratus truncatus, 317
Synodontis matthesi, 317
Synodontis melanoptera, 317
Synodontis melanopterus, 317
Synodontis melanostictus, 318
Synodontis melanostictus iturii, 315
Synodontis melanostictus var. *iturii*, 315
Synodontis multimaculata, 317
Synodontis multamaculatus, 317
Synodontis multipunctata, 317
Synodontis multipunctatus, 317
Synodontis nebulosa, 317
Synodontis nebulosus, 317
Synodontis nigrita, 317
Synodontis nigriventralis, 318
Synodontis nigromaculata, 318
Synodontis nigromaculatus, 318
Synodontis njassae, 318
Synodontis notata, 318
Synodontis notatus, 318
Synodontis notatus binotata, 318
Synodontis notatus ocellatus, 318
Synodontis notatus var. *binotata*, 318
Synodontis notatus var. *ocellatus*, 318
Synodontis nummifer, 318
Synodontis obesus, 318, 323
Synodontis ocellifer, 318
Synodontis omias, 319
Synodontis ornatipinnis, 319
Synodontis ornatissima, 319
Synodontis ornatissimus, 319
Synodontis ornatus, 317, 319
Synodontis ovidius, 316
Synodontis pantherinus, 315
Synodontis pardalis, 319
Synodontis petricola, 319
Synodontis pfefferi, 311
Synodontis pleurops, 319
Synodontis polli, 319
Synodontis polyodon, 319
Synodontis polystigma, 319
Synodontis pulcher, 319
Synodontis punctifer, 319
Synodontis punctulata, 320
Synodontis punctulatus, 320
Synodontis rebeli, 320, 323
Synodontis resupinata, 320
Synodontis resupinatus, 320
Synodontis ricardoae, 320
Synodontis robbianus, 320
Synodontis robertsi, 320
Synodontis ruandae, 320
Synodontis rufigensis, 320
Synodontis rukwaensis, 320
Synodontis schall, 315, 320
Synodontis schoutedeni, 321
Synodontis serpentis, 321
Synodontis serrata, 321
Synodontis serratus, 311, 321
Synodontis serratus tanganaiae, 322
Synodontis smithii, 321
Synodontis smiti, 321
Synodontis soloni, 321
Synodontis sorex, 321
Synodontis steindachneri, 321
Synodontis tanganaiae, 321
Synodontis tanganyicae, 322
Synodontis tenuis, 321
Synodontis tessmanni, 322
Synodontis thamalakanensis, 322
Synodontis tholloni, 311

- Synodontis thomasi*, 313
Synodontis thysi, 322
Synodontis tourei, 322
Synodontis unicolor, 322
Synodontis vaillanti, 323
Synodontis vanderwaali, 322
Synodontis velifer, 322
Synodontis vermiculata, 322
Synodontis vermiculatus, 322
Synodontis victoriae, 322
Synodontis violacea, 322
Synodontis violaceus, 322
Synodontis vittatus, 314
Synodontis voltae, 323
Synodontis wamiensis, 320
Synodontis waterloti, 323
Synodontis wernerii, 311
Synodontis woosnami, 323
Synodontis xiphias, 323
Synodontis zambesensis, 323
Synodontis zambezensis, 323
Synodontis zambezensis rukwaensis, 320
Synodontis zanzibarica, 323
Synodontis zanzibaricus, 316, 320, 323
synodontis, *Pimelodus*, 313
Syriacus, *Clarias*, 143
- T**
- taakree*, *Hypophthalmus*, 362
taakree, *Proeutropiichthys*, 362
taakree burmanicus,
Proeutropiichthys, 362
tachiraense, *Chaetostoma*, 230
tachiraensis, *Chaetostoma*, 230
Tachisurus agassizii, 39
Tachisurus gulosus, 40
Tachisurus jordani, 33
Tachisurus lentiginosus, 33
Tachisurus longicephalus, 39
Tachisurus longicephalus, 39
Tachisurus upsulonophorus, 44
tachisurus, *Pimelodus*, 431
Tachysurus, 48, 431
Tachysurus (Pararius) berneyi, 47
Tachysurus (Pararius) godfreyi, 45
Tachysurus atroplumbeus, 32
Tachysurus broadbenti, 51
Tachysurus crassus, 434, 435
Tachysurus emmelane, 40
Tachysurus equatorialis, 40
Tachysurus evermanni, 40
Tachysurus landanensis, 57
Tachysurus liropus, 41
Tachysurus machadoi, 44
Tachysurus oblongus, 435
Tachysurus planus, 434, 435
Tachysurus sinensis, 431
Tachysurus steindachneri, 40
taczanowskii, *Arges*, 67
taczanowskii, *Astroblepus*, 67
taczanowskii, *Chaetostoma*, 231
Taczanowskii, *Chaetostomus*, 231
taczanowskii, *Trichomycterus*, 424
taenia, *Trichomycterus*, 407, 424
taenia transandianum, *Pygidium*, 424
taeniatus, *Macrones (Liocassis)*, 102
taeniatus, *Plecostomus*, 255
taeniatus, *Pseudobagrus*, 102
taeniatus, *Trachelyopterichthys*, 78
taeniatus, *Trachelyopterus*, 78
Taenionema, 330
Taenionema steerei, 330, 331
taeniophora, *Pimelodella*, 195
taeniophorus, *Pimelodus*
(Pimelodella), 195
taeniopterus, *Trichomycterus*, 424
taenioptera, *Pimelodella*, 195
taeniura, *Pangasius*, 326
taimensis, *Hisonotus*, 249
taimensis, *Microlepidogaster*, 249
taiosh, *Callichthys*, 125
taiwanensis, *Pseudobagrus*, 102
tamanae, *Oxylicaria*, 300
tamanae, *Sturisomatichthys*, 300
tamboensis, *Ancistrus*, 224
tamoata, *Callichthys*, 111
Tandanus, 351
Tandanus bostocki, 352
Tandanus coatesi, 348
Tandanus tandanus, 352
tandanus, *Tandanus*, 352
tanensis, *Physailia somalensis*, 360
tanganicae, *Synodontis*, 321
tanganicae, *Synodontis serratus*, 322
tanganicanus, *Auchenoglanis*
occidentalis, 154
Tanganikallabes, 152
Tanganikallabes mortiauxi, 152
tanganyicae, *Synodontis*, 322
tanganyikaensis, *Malapterurus*, 302
tanoensis, *Malapterurus*, 302
tapanahoniensis, *Heptapterus*, 184
tapanahoniensis, *Hypostomus*, 260
tapanahoniensis, *Hypostomus*
gymnorhynchus, 261
tapatapae, *Pimelodella*, 195
tapeinopterus, *Encheloclarias*, 150
tapeinopterus, *Heterobranchus*, 149,
150
taphorni, *Cochliodon*, 261
taphorni, *Farlowella*, 239
taphorni, *Hypostomus*, 261
taphophilus, *Hemibagrus*, 108
tapijara, *Hypostomus*, 261
tapirope, *Otocinclus*, 275
tarabinii, *Pardiglanis*, 163
taroba, *Trichomycterus*, 424
Tatia, 77
- Tatia aulopygia*, 77
Tatia boemia, 77
Tatia brunnea, 77
Tatia concolor, 73
Tatia creutzbergi, 77
Tatia dunni, 77
Tatia galaxias, 77
Tatia gyrina, 77
Tatia intermedia, 77
Tatia musaica, 77
Tatia neivai, 77
Tatia perugiae, 73
Tatia punctata, 73
Tatia reticulata, 73
Tatia romani, 73
Tatia simplex, 78
Tatia strigata, 78
taunayi, *Ancistrus*, 224
Taunayia, 203
Taunayia bifasciata, 203
Taunayia marginata, 203
taurus, *Aspidoras*, 110
taxistigma, *Henonemus*, 406
taxistigma, *Ochmacanthus*, 405, 406
taylori, *Arius*, 40, 48
taylori, *Hemipimelodus*, 48
taylori, *Neoarius*, 48
taylori, *Noturus*, 214
taytayensis, *Hito*, 375, 377
taytayensis, *Hitoichthys*, 375, 377
taytayensis, *Pterocryptis*, 377
tchadiensis, *Auchenoglanis*
occidentalis, 154
tchangi, *Pseudecheneis*, 401
teaguei, *Parabranchioica*, 410, 411
teaguei, *Trachelyopterus*, 80
teaguei, *Trachycorystes*, 80
tectirostris, *Chaetostomus*, 221
teffiana, *Hemiloricaria*, 247
teffiana, *Loricaria*, 247
teffeanus, *Rhinodoras*, 176
teijsmanni, *Clarias*, 147
telchita, *Pimelodus*, 394
telchitta, *Glyptothorax*, 394
telchitta, *Pimelodus*, 394
telchitta, *sufii*, *Glyptothorax*, 394
temminckianus, *Bagrus*, 55
temminckii, *Ancistrus*, 224
Temminckii, *Bagrus*, 55
Temminckii, *Hypostomus*, 224
tenebricosa, *Pseudolaguvia*, 402
tenebricus, *Leiocassis*, 443
tenebrosa, *Phenacorhamdia*, 189
tenebrosus, *Imparfinis*, 189
tenella, *Rhamdia*, 199
tengana, *Batasio*, 86, 442
tengana, *Pimelodus*, 86
tengara, *Mystus*, 97, 98
tengara, *Pimelodus*, 97

tenius, *Otolithus (Arius)*, 435
tentaculatus, *Lasiancistrus*, 265
tenue, *Pygidium*, 424
tenuicauda, *Paraphractura*, 26, 27
tenuicauda, *Phractura*, 27
tenuicauda, *Plecostomus*, 298
tenuicauda, *Squaliforma*, 298
tenuifurcatus, *Leiocassis*, 93
tenuirostre, *Sturisoma*, 300
tenuirostris, *Oxyloricaria*, 300
tenuis, *Belonoglanis*, 24
tenuis, *Clarias*, 148
tenuis, *Heptapterus*, 184
tenuis, *Hypostomus*, 298
tenuis, *Macrones (Pseudobagrus)*, 103
tenuis, *Otolithus (Arius)*, 435
tenuis, *Pseudobagrus*, 102
tenuis, *Squaliforma*, 298
tenuis, *Synodontis*, 321
tenuis, *Trichomycterus*, 424
tenuispinis, *Amblyceps*, 18
tenuispinis, *Arius*, 52, 56
tenuispinis, *Plicofollis*, 52
ternetzi, *Hypostomus*, 261
ternetzi, *Opsodoras*, 175
ternetzi, *Plecostomus*, 261
ternetzi, *Typhlobelus*, 426, 427
tessmanni, *Synodontis*, 322
Tetracamphilius, 27
Tetracamphilius angustifrons, 27
Tetracamphilius clandestinus, 27
Tetracamphilius notatus, 27
Tetracamphilius pectinatus, 27
tetranema, *Bathybagrus*, 154, 155
tetranema, *Pseudolais*, 328
Tetranemichthys, 78
Tetranemichthys quadrifilis, 78
Tetranemichthys wallacei, 444
Tetranesodon, 42
Tetranesodon conorrhynchus, 42
tetraradiata, *Listrura*, 408
teugelsi, *Chrysichthys*, 159
teugelsi, *Clariallabes*, 138
teugelsi, *Malapterurus*, 302
teugelsi, *Paramphilius*, 25
thalassina, *Netuma*, 49
thalassina jacksonensis, *Netuma*, 49
thalassinus, *Bagrus*, 49
thamalakanensis, *Synodontis*, 322
theobaldi, *Chrysichthys*, 105
theodorae, *Clarias*, 139, 147
theresiae, *Arges*, 67
theresiae, *Astroblepus*, 67
therezinae, *Ageneiosus (Pseudageneiosus)*, 69
thienemanni, *Clarias*, 148
tholloni, *Synodontis*, 311
thomasi, *Chrysichthys delhezi*, 156
thomasi, *Notoglanidium*, 162

thomasi, *Synodontis*, 313
thomersoni, *Rhinodoras*, 178
thomsoni, *Chaetostoma*, 231
thomsoni, *Chaetostomus*, 231
thonneri, *Chrysichthys*, 159
thoracata, *Megalechis*, 129
thoracatum, *Hypoptopoma*, 249, 250
thoracatum cayennae, *Hoplosternum*, 128
thoracatum surinamensis,
 Hoplosternum, 129
thoracatus, *Auchenipterichthys*, 71
thoracatus, *Auchenipterus*, 71
thoracatus, *Callichthys*, 129
thoracicus, *Auchenipterus*, 71
thorectes, *Pseudohemiodon*, 288
Thrichomycterus, 404
Thrichomycterus macraei, 405
thrissocope, *Loricaria*, 295
thrissocope, *Rineloricaria*, 295
thunberg, *Pimelodus*, 35
thunbergianus, *Plotosus*, 350
Thysanocara, 219
thysi, *Chrysichthys*, 159
thysi, *Doumea*, 25
thysi, *Malapterurus*, 301
thysi, *Synodontis*, 322
thysvillensis, *Clarias (Allabenchelys)*
 submarginatus, 141
tianquanensis, *Pareuchiloglanis*, 400
tibicen, *Aspredinichthys*, 58
tibicen, *Aspredo*, 57, 58
tietensis, *Hypostomus*, 261
tietensis, *Otocinclus*
 (*Microlepidogaster*), 290
tietensis, *Plecostomus*, 261
tietensis, *Pseudotocinclus*, 290
tigrinum, *Brachyplatystoma*, 331
tigrinum, *Pseudoplatystoma*, 341
tigrinus, *Batasio*, 86
tigrinus, *Merodontotus*, 330, 331
tigrinus, *Trichomycterus*, 415
tigris, *Lasiancistrus*, 288, 289
tigris, *Pseudolithoxus*, 289
tihoni, *Gymnallabes*, 152
tihoni, *Platyallabes*, 152
tilhoi, *Gephyroglanis*, 155
tiong, *Glyptosternum*, 395
tiraquae, *Pygidium*, 423
titcombi, *Hatcheria*, 405
tocantinensis, *Corumbataia*, 232
tocantinsensis, *Aguarunichthys*, 329
tocantinsi, *Tridentopsis*, 426
Tocantinsia, 78
Tocantinsia depressa, 78
Tocantinsia piresi, 78
tokiensis, *Pseudobagrus*, 103
tombador, *Ancistrus*, 225
tonggol, *Ariodes*, 52

tonggol, *Arius*, 52
tonggol, *Plicofollis*, 52
topavae, *Hypostomus*, 261
topavae, *Plecostomus*, 261
torbesensis, *Cordylancistrus*, 232
torbesensis, *Pseudancistrus*, 231, 232
tordilho, *Leptoplosternum*, 129
torobo, *Oloplotosus*, 349
torosilabris, *Liocassis*, 108
torosus, *Aguarunichthys*, 329
Torpedo, 301
torrenticola, *Harttia*, 242
torrentis, *Pterocryptis*, 377
torrentis, *Silurus*, 377
tortosus, *Sisor*, 402
totae, *Pygidium*, 412
totae, *Rhizosomichthys*, 412
tourei, *Synodontis*, 322
trachacanthus, *Bagrus*, 89
Trachelyichthys, 78
Trachelyichthys decaradiatus, 78
Trachelyichthys exilis, 78
Trachelyopterichthys, 78
Trachelyopterichthys anduzei, 78
Trachelyopterichthys taeniatus, 78
Trachelyopterini, 68
Trachelyopterus, 68, 79
Trachelyopterus albicrux, 79
Trachelyopterus amblops, 79
Trachelyopterus ceratophysis, 79
Trachelyopterus coriaceus, 79
Trachelyopterus coriaceus maculosus, 79
Trachelyopterus fisheri, 79
Trachelyopterus galeatus, 79
Trachelyopterus insignis, 79
Trachelyopterus lacustris, 79
Trachelyopterus leopardinus, 80
Trachelyopterus lucenai, 80
Trachelyopterus peloichthys, 80
Trachelyopterus striatulus, 80
Trachelyopterus taeniatus, 78
Trachelyopterus teaguei, 80
trachipomus, *Bagrus*, 55
Trachycorystes, 80
Trachycorystes albicrux, 79
Trachycorystes analis, 80
Trachycorystes coracoideus, 71
Trachycorystes cratensis, 80
Trachycorystes fisheri, 79
Trachycorystes insignis badeli, 80
Trachycorystes insignis peloichthys, 80
Trachycorystes jokeannae, 80
Trachycorystes leopardinus, 80
Trachycorystes magdalena, 79
Trachycorystes porosus, 81
Trachycorystes teaguei, 80
Trachycorystes trachycorystes, 80
Trachycorystes typus, 81

- trachycorystes*, *Auchenipterus*, 80
trachycorystes, *Trachycorystes*, 80
Trachydoras, 179
Trachydoras atripes, 179
Trachydoras brevis, 179
Trachydoras microstomus, 179
Trachydoras nattereri, 179
Trachydoras paraguayensis, 179
Trachydoras steindachneri, 179
Trachyglanis, 28
Trachyglanis ineac, 28
Trachyglanis intermedius, 28
Trachyglanis minutus, 28
Trachyglanis sangensis, 28
Trachymochlus, 431
Trachymochlus cupidio, 431, 432
trachyparia, *Oxydoras*, 179
Trachypoma, 404
Trachypoma marmoratum, 404
transandianum, *Pygidium taenia*, 424
transandianus, *Trichomycterus*, 424
transfasciatus, *Rhamdioglanis*, 202
transitoria, *Pimelodella*, 195
transmontanus, *Batrachoglanis*, 353
transmontanus, *Pseudopimelodus*, 353
transvaaliensis, *Amphilius*, 23
trautmani, *Noturus*, 214
travancoria, *Batasio*, 86
travassosi, *Otothyris*, 276
travassosi, *Pygidium*, 414
trefauti, *Trichomycterus*, 425
treitlii, *Corydoras*, 126
trewavasae, *Glyptothorax*, 394
Trewavasia, 433
Trewavasia carinata, 433
triacanthopomus, *Henonemus*, 440
triactis, *Leporacanthicus*, 266
Trichogenes, 403, 414
Trichogenes longipinnis, 414
Trichogeninae, 403
Trichomycteridae, 403
Trichomycterini, 403
trichomycteroidea, *Paramphilius*, 25
Trichomycterus, 403, 413
Trichomycterus aguarague, 441
Trichomycterus albinotatus, 414
Trichomycterus alternatus, 414
Trichomycterus alterus, 415
Trichomycterus amazonicus, 406
Trichomycterus areolatus, 415
Trichomycterus arleoi, 415
Trichomycterus auroguttatus, 415
Trichomycterus bahianus, 415
Trichomycterus banneai, 415
Trichomycterus barbatula, 422
Trichomycterus barbouri, 415
Trichomycterus belensis, 415
Trichomycterus bogotensis, 415
Trichomycterus bomboizanus, 416
Trichomycterus borellii, 416
Trichomycterus boylei, 416
Trichomycterus brasiliensis, 416
Trichomycterus brasiliensis itatiayae, 419
Trichomycterus brasiliensis tristis, 416
Trichomycterus caliensis, 416
Trichomycterus candidus, 416
Trichomycterus castroi, 416
Trichomycterus catamarcensis, 416
Trichomycterus caudofasciatus, 416
Trichomycterus celsae, 416
Trichomycterus chaberti, 417
Trichomycterus chapmani, 417
Trichomycterus chiltoni, 417
Trichomycterus chungaraensis, 417
Trichomycterus concolor, 417
Trichomycterus conradi, 417
Trichomycterus corduvensis, 417
Trichomycterus cubataonis, 425
Trichomycterus dawsoni, 417
Trichomycterus diabolus, 417
Trichomycterus dispar, 417
Trichomycterus dorsotriatus, 417
Trichomycterus duellmani, 418
Trichomycterus eichorniarum, 407
Trichomycterus eigenmanni, 423
Trichomycterus emanueli, 418
Trichomycterus fassli, 418
Trichomycterus gabrieli, 418
Trichomycterus giganteus, 418
Trichomycterus goeldii, 418
Trichomycterus gorgona, 418
Trichomycterus gracilis, 422
Trichomycterus guaraquessaba, 418
Trichomycterus guianensis, 418
Trichomycterus hasemani, 418
Trichomycterus herberti, 407
Trichomycterus heterodontus, 418
Trichomycterus iheringi, 419
Trichomycterus immaculatus, 419
Trichomycterus incae, 423
Trichomycterus inermis, 324
Trichomycterus itacambirussu, 419
Trichomycterus itacarambiensis, 419
Trichomycterus itatiayae, 419
Trichomycterus jacupiranga, 419
Trichomycterus jequitinhonhae, 419
Trichomycterus johnsoni, 419
Trichomycterus knerii, 419
Trichomycterus landinga, 419
Trichomycterus laticeps, 407
Trichomycterus latidens, 419
Trichomycterus latistriatus, 420
Trichomycterus laucaensis, 420
Trichomycterus lewi, 420
Trichomycterus longibarbatus, 420
Trichomycterus maculatus, 415
Trichomycterus maracaiboensis, 420
Trichomycterus marmoratus, 415
Trichomycterus mboyey, 420
Trichomycterus mendozensis, 413
Trichomycterus meridae, 420
Trichomycterus migrans, 420
Trichomycterus mimonha, 420
Trichomycterus minutus, 413
Trichomycterus mirissumba, 420
Trichomycterus mondolfi, 421
Trichomycterus motatanensis, 421
Trichomycterus naipi, 421
Trichomycterus nigricans, 414, 421
Trichomycterus nigromaculatus, 421
Trichomycterus palleus, 415
Trichomycterus pantherinus, 421
Trichomycterus paolencis, 421
Trichomycterus papilliferus, 421
Trichomycterus paquequerensis, 421
Trichomycterus pardus, 423
Trichomycterus pauciradiatus, 440
Trichomycterus pentlandi, 423
Trichomycterus pictus, 423
Trichomycterus piurae, 421
Trichomycterus plumbeus, 421
Trichomycterus poeyanus, 423
Trichomycterus potschi, 421
Trichomycterus pradensis, 422
Trichomycterus proöps, 408
Trichomycterus pseudosilvinichthys, 422
Trichomycterus punctatissimus, 422
Trichomycterus punctulatus, 422
Trichomycterus pusillus, 411
Trichomycterus ramosus, 422
Trichomycterus regani, 422
Trichomycterus reinhardti, 422
Trichomycterus retropinnis, 422
Trichomycterus riojanus, 422
Trichomycterus rivulatus, 422
Trichomycterus roigi, 423
Trichomycterus romeroi, 423
Trichomycterus santaeritae, 423
Trichomycterus spegazzinii, 423
Trichomycterus spelaeus, 423
Trichomycterus spilosoma, 423
Trichomycterus stawiarski, 423
Trichomycterus stellatus, 424
Trichomycterus straminus, 424
Trichomycterus striatus, 424
Trichomycterus taczanowskii, 424
Trichomycterus taenia, 407, 424
Trichomycterus taeniops, 424
Trichomycterus taroba, 424
Trichomycterus tenuis, 424
Trichomycterus tigrinus, 415
Trichomycterus transandianus, 424
Trichomycterus trefauti, 425
Trichomycterus triguttatus, 425
Trichomycterus tupinamba, 425

- Trichomycterus unicolor*, 425
Trichomycterus variegatus, 425
Trichomycterus venulosus, 425
Trichomycterus vermiculatus, 425
Trichomycterus vittatus, 425
Trichomycterus weyrauchi, 425
Trichomycterus yuska, 425
Trichomycterus zonatus, 425
tricornis, *Hopliancistrus*, 249
Tridens, 403, 426
Tridens brevis, 426
Tridens melanops, 426
Tridensimilis, 426
Tridensimilis brevis, 426
Tridensimilis venezuelae, 426
Tridentinae, 403
Tridentopsis, 426
Tridentopsis cahuali, 426
Tridentopsis pearsoni, 426
Tridentopsis tocantinsi, 426
trifasciatum, *Cyclopium*, 67
trifasciatus, *Astroblepus*, 67
trifasciatus, *Pachypterus*, 359
trigonocephalus, *Sorubim*, 342
triguttatum, *Pygidium*, 425
triguttatus, *Trichomycterus*, 425
trilineatoides, *Glyptothonax*, 394
trilineatus, *Corydoras*, 126
trilineatus, *Glyptothonax*, 394
trilineatus, *Leiocassis*, 103
trilineatus, *Pseudobagrus*, 103
trilobatus, *Chiloglanis*, 308
trimaculatus, *Leptodoras*, 174
trimaculatus, *Nemadoras*, 174
trimaculatus, *Opsodoras*, 174
trimaculatus, *Oxydoras*, 174
trinitatis, *Ancistrus*, 225
trinitatis, *Chaetostomus*, 225
triostegus, *Silurus*, 380
triradiatus, *Ancistrus*, 225
triradiatus martini, *Ancistrus*, 223
triseriatus, *Corydoras nattereri*, 121
tristis, *Trichomycterus brasiliensis*, 416
troelli, *Claibornichthys*, 434, 435
Trogloglanis, 216
Trogloglanis pattersoni, 216
trombetensis, *Harttia*, 242
troschelii, *Sciades*, 53, 54
truncatorstris, *Chasmocranus*, 183
truncatum, *Platystoma*, 341
truncatus, *Amblydoras*, 168
truncatus, *Arius*, 43
truncatus, *Belodontichthys*, 368
truncatus, *Cryptarius*, 43
truncatus, *Liocassis*, 103
truncatus, *Pseudobagrus*, 103
truncatus, *Synodontis marmoratus*, 317
tsanensis, *Clarias*, 143
tubbi, *Pangasius*, 328
tucumanensis, *Loricaria*, 269
tukano, *Corydoras*, 126
tumbanus, *Eutropius*, 365
tumbanus, *Schilbe*, 365
tupinamba, *Trichomycterus*, 425
tuyra, *Arius*, 40
tuyrensis, *Dasylophicaria*, 234
tuyrensis, *Loricaria*, 234
tweediei, *Wallagonia*, 381
Tympanopleura, 68
Tympanopleura alta, 69
Tympanopleura nigricollis, 68
Tympanopleura piperata, 68, 70
typhla, *Rhamdia laticauda*, 198
Typhlobagrus, 189
Typhlobagrus kronei, 189, 192
Typhlobelus, 426
Typhlobelus guacamaya, 426
Typhlobelus lundbergi, 426
Typhlobelus macromycter, 426
Typhlobelus ternetzi, 426
typica, *Doumea*, 24, 25
typus, *Gagata*, 386
typus, *Gymnallabes*, 159
typus, *Helicophagus*, 324, 325
typus, *Ketengus*, 45
typus, *Micronema*, 375
typus, *Parahemiodon*, 269, 270
typus, *Phyllonemus*, 164
typus, *Pinirampus*, 339
typus, *Rhinoglanis*, 310, 311
typus, *Trachycorystes*, 81
typus heterocercalis, *Gymnallabes*, 150
- U**
- uatumensis*, *Harttia*, 242
ubangensis, *Auchenoglanis*, 162
ubangensis, *Bagrus*, 84
ubidai, *Astroblepus*, 67
ubidai, *Cyclopium*, 67
ucamara, *Leptoplosternum*, 129
ucayalensis, *Ageneiosus*, 70
ucayalensis, *Hemiancistrus*, 283, 285
ucayalensis, *Hassar*, 171
ucayalensis, *Loricariichthys*, 271
ucayalensis, *Peckoltia*, 285
Uegitlanis, 152
Uegitlanis zammaranoi, 152, 153
uelensis, *Clariallabes*, 138
uelensis, *Clarias (Allabenchelys)*, 138
umbrosa, *Cetopsis*, 133
unae, *Hypostomus*, 261
Unae, *Plecostomus*, 261
uncinatus, *Arius*, 36
undecimalis, *Chaetostomus*, 292
undecimalis, *Pterygoplichthys*, 292
undecimalis, *Silurus*, 431
- underwoodi*, *Rhamdia*, 198
undulatus, *Corydoras*, 126
unicolor, *Aphanotorulus*, 225
unicolor, *Plecostomus*, 225
unicolor, *Plotosus*, 350, 352
unicolor, *Pygidium*, 425
unicolor, *Synodontis*, 322
unicolor, *Trichomycterus*, 425
unidorsalis, *Helogenes*, 134
unifasciata, *Phenacorhamdia*, 189
unifasciatum, *Cyclopium*, 67
unifasciatus, *Astroblepus*, 67
uniformis, *Amphilus baudoni*, 21
uniformis, *Chrysichthys*, 159
unitius, *Silurus*, 350
upiensis, *Pangasius pangasius*, 327
Upsilodus, 247
Upsilodus victori, 247
upsulonophorus, *Tachisurus*, 44
uracantha, *Loricaria*, 240
uracantha, *Rineloricaria*, 240
uracantha rupestris, *Loricaria*, 240
uracanthus, *Fonchiichthys*, 240
uranoscopus, *Amphilus*, 23
uranoscopus, *Anoplopterus*, 21, 23
uranoscopus, *Doras*, 173, 174
uranoscopus, *Megalodoras*, 174
uranoscopus, *Schilbe*, 362, 365
urbaini, *Cryptopterus*, 373
urbaini, *Ompok*, 373
urichi, *Caecorhamdia*, 197, 202
Urinophilus, 427
Urinophilus diabolicus, 411
Urinophilus erythrurus, 427
urostigma, *Bagrus*, 84
urostriatum, *Decapogon*, 127
urostriatum, *Dianema*, 127
urotriatum, *Decapogon*, 127
urua, *Pimelodus*, 359
uruguayensis, *Ageneiosus*, 70
uruguayensis, *Hypostomus*, 261
uryensis, *Helogenes*, 134
uryensis, *Helogenes marmoratus*, 134
ussuriensis, *Bagrus*, 92, 100
ussuriensis, *Leiocassis*, 101
ussuriensis, *Pelteobagrus*, 100
ussuriensis, *Pseudobagrus*, 101
utarus, *Arius*, 48
utarus, *Neoarius*, 48
utik, *Arius*, 35
- V**
- vacha*, *Eutropiichthys*, 358
vacha, *Pimelodus*, 358
vachellii, *Bagrus*, 101
vachellii, *Pelteobagrus*, 101
vagum, *Chaetostoma*, 231
vagus, *Chaetostoma*, 231
vaillanti, *Arges*, 67
vaillanti, *Astroblepus*, 67

- vaillanti**, *Hypostomus*, 261
vaillanti, *Liocassis*, 105
Vaillanti, *Plecostomus*, 261
vaillanti, *Pseudomyctus*, 105
vaillanti, *Synodontis*, 323
vaillantii, *Bagrichthys*, 83
Vaillantii, *Bagroides*, 83
vaillanti, *Brachyplatystoma*, 331
Vaillanti, *Platystoma*, 330, 331
vaillantii, *Silurus*, 331
valencia, *Lithogenes*, 267
valenciennesi, *Ageneiosus*, 69
valenciennesi, *Osteogeneiosus*, 51
valenciennesii, *Genidens*, 44
Valenciennesii, *Loricaria*, 247
valenciennis, *Parapimelodus*, 335
valenciennis, *Pimelodus*, 335
valeya, *Wallago attu*, 381
vanceae, *Astroblepus*, 67
vanceae, *Cyclopium*, 67
vandeli, *Arius*, 50
Vandellia, 403, 427
Vandellia Balzani, 427
Vandellia beccarii, 427
Vandellia cirrhosa, 427
Vandellia gigantea, 427
Vandellia hasemani, 411
Vandellia hematophaga, 410
Vandellia plazaii, 427
Vandellia sanguinea, 427
Vandellia wieneri, 411
Vandelliini, 403
vandenhoulei, *Clarias*, 144
vanderwaali, *Synodontis*, 322
vandeweyeri, *Eutropiellus*, 361
vanigonis, *Otolithus (Arius)*, 434
Vannutellii, *Rhinoglanis*, 311
variabilis, *Clariablabes*, 138
variegata, *Crossoloricaria*, 233
variegata, *Loricaria*, 232, 233
variegata venezuelae, *Loricaria*, 233
variegatum, *Amblyceps*, 19
variegatus, *Pimelodus*, 13, 14, 15
variegatus, *Akysis*, 15
variegatus, *Akysis variegatus*, 14
variegatus, *Hemibagrus*, 91
variegatus, *Microglanis*, 355
variegatus, *Trichomycterus*, 425
variipictus, *Hypostomus*, 261
variipictus, *Neoplecostomus*, 274
variipictus, *Plecostomus*, 261
varimaculatus, *Hypostomus*, 261
varimaculatus, *Plecostomus*, 261
variolosus, *Arius*, 41
variolosus, *Pseudopimelodus*, 355
variolus, *Ancistrus*, 225
variolus, *Chaetostomus*, 225
variostictus, *Hypostomus*, 261
variostictus, *Plecostomus*, 261
varispinis, *Clarias*, 147
varius, *Acrochordonichthys*, 13
varius, *Akysis*, 15
varius, *Liposarcus*, 292
vasquezii, *Chaetostoma*, 231
vazferreiraiai, *Homodiaetus*, 406
velatus, *Encheloclarias*, 150
velifer, *Chrysichthys*, 157, 158
velifer, *Gephyroglanis*, 157
velifer, *Pimelodus*, 203
velifer, *Synodontis*, 322
velites, *Aspidoras*, 110
velox, *Hemibagrus*, 91
velutinus, *Hemipimelodus*, 48
velutinus, *Neoarius*, 48
venaticus, *Bagrus*, 55
venezuelae, *Chaetostoma*, 231
venezuelae, *Corymbophanes*, 231
venezuelae, *Crossoloricaria*, 233
venezuelae, *Haemomaster*, 405
venezuelae, *Loricaria variegata*, 233
venezuelae, *Tridensimilis*, 426
venezuelanus, *Corydoras*, 112
venezuelensis, *Farlowella*, 239
venezuelensis, *Farlowella acus*, 239
venosus, *Arius*, 36
ventrale, *Cyclopium*, 68
ventralis, *Astroblepus*, 68
ventralis, *Cetopsis*, 135
ventromaculatus, *Hypostomus*, 262
venulosum, *Pygidium*, 425
venulosus, *Trichomycterus*, 425
verbeekii, *Pseudeutropius*, 326
vereunda, *Pterocryptis*, 377
verecundus, *Ancistrus*, 225
verissimi, *Decapogon*, 127
vermicularis, *Hypostomus*, 262
vermicularis, *Plecostomus*, 262
vermiculata, *Ancistrus vittatus*, 285
vermiculata, *Peckoltia*, 285
vermiculata, *Synodontis*, 322
vermiculatum, *Pygidium*, 425
vermiculatus, *Synodontis*, 322
vermiculatus, *Trichomycterus*, 425
verres, *Hypostomus*, 262
verrucosa, *Aspredo*, 60
verrucosa, *Parakysis*, 15, 16
verrucosus, *Arius*, 45
verrucosus, *Bunocephalus*, 60
verrucosus, *Hemiarius*, 45
verrucosus, *Parakysis*, 16
verrucosus, *Platystacus*, 58, 60
verrucosus, *Silurus*, 60
versicolor, *Pimelodus*, 44
vertagus, *Bagrus*, 55
vespa, *Akysis*, 15
vespertinus, *Ameiurus*, 206
vespertinus, *Ictalurus*, 206
vestigipinnis, *Hemipsilichthys*, 281
vestigipinnis, *Pareiorraphis*, 281
vestitus, *Otocinclus*, 274, 275
vetula, *Loricaria*, 278
vetula, *Paraloricaria*, 278
vicinus, *Hypostomus*, 278
victori, *Upsilodus*, 247
victoriae, *Synodontis*, 322
viedmensis, *Diplomystes*, 166
viedmensis, *Olivaichthys*, 166
viedmensis cuyanus, *Diplomystes*, 166
viedmensis mesembrinus, *Diplomystes*, 166
vietnamicus, *Hemibagrus*, 91
vietnamicus, *Hemibagrus*, 91
vigilis, *Microsynodontis*, 310
Villarius, 208
Villarius pricei, 208, 209
villarsi, *Plecostomus*, 298
villarsi, *Squaliforma*, 298
villiersi, *Physailia*, 360
villosus, *Arius*, 166
villosus, *Batrachoglanis*, 353
villosus, *Lithogenes*, 267
villosus, *Pseudopimelodus*, 353
villosus butcheri, *Pseudopimelodus*, 352
Vilsoni, *Pimelenotus*, 197, 200
vinciguerrae, *Clarias*, 143
Vinciguerrae, *Exostoma*, 385
vinhensis, *Pelteobagrus virgatus*, 101
violacea, *Synodontis*, 322
violaceus, *Synodontis*, 322
virescens, *Corydoras*, 123
virescens, *Plecostomus*, 298
virescens, *Squaliforma*, 298
virgatus, *Aoria*, 101
virgatus, *Leiocassis*, 101
virgatus, *Pelteobagrus*, 101
virgatus, *Pseudobagrus*, 101
virgatus vinhensis, *Pelteobagrus*, 101
virginiae, *Corydoras*, 126
virgo, *Ageneiosus*, 70
virgulatus, *Aspidoras*, 110
viridescens, *Gogangra*, 395
viridescens, *Pimelodus*, 395
viscosus, *Pimelodus*, 215
viscosus, *Silurus*, 215
vitata, *Ansorgia*, 361
vittata, *Ansorgia*, 360, 361
vittata, *Farlowella*, 239
vittata, *Macrones*, 106
vittata, *Peckoltia*, 285
vittata, *Pimelodella*, 195
vittata bistriata, *Ansorgia*, 361
vittatus, *Ageneiosus*, 70
vittatus, *Chaetostomus*, 283, 284, 285
vittatus, *Corydoras*, 126
vittatus, *Corydoras blochi*, 126
vittatus, *Mystus*, 98

vittatus, *Otocinclus*, 275
vittatus, *Plotosus*, 350
vittatus, *Pseudorhamdia*, 195
vittatus, *Silurus*, 98
vittatus, *Silurus* (*Callichrus*), 372
vittatus, *Synodontis*, 314
vittatus, *Trichomycterus*, 425
vittatus horai, *Mystus* (*Mystus*), 95
vittatus vermiculata, *Ancistrus*, 285
viviparus, *Arius*, 55
viviparus, *Plotosus*, 350
vogti, *Atopochilus*, 304
volcanensis, *Lasiancistrus*, 264
voltae, *Chiloglanis*, 308
voltae, *Irvineia*, 358
voltae, *Synodontis*, 323
Vorhisia, 434
Vorhisia vulpes, 434, 435
Vorhisidae, 434
votouro, *Hemiancistrus*, 244
voulezi, *Rhamdia branneri*, 201
vulgaris, *Pimelodus*, 206
vulpeculus, *Pimelodus*, 206
vulpes, *Pimelodus*, 211
vulpes, *Vorhisia*, 434, 435
W
waandersii, *Helicophagus*, 325
wagenaari, *Chrysichthys*, 159
wagneri, *Pimelodus*, 201
waiampi, *Hypostomus*, 262
walkeri, *Chrysichthys*, 159, 160
walkeri, *Clarias*, 141
walkeri, *Notoglanidium*, 162
walkeri, *Pseudeutropius atherinoides*, 359
wallacei, *Tetranematicthys*, 444
Wallago, 380
Wallago attu, 380
Wallago attu valeya, 381
Wallago dinema, 368
Wallago heterorhynchus, 368
Wallago krattensis, 374
Wallago leerii, 380, 381
Wallago leiacanthus, 371, 372
Wallago maculatus, 381
Wallago micropogon, 381
Wallago miostoma, 372
Wallago nebulosus, 381
Wallago Russellii, 381
Wallagonia, 380
Wallagonia tweediei, 381
wallagoo, *Silurus*, 381
walrechti, *Sciadeichthys* (*Selenaspis*), 53
wamiensis, *Leptoglanis*, 29
wamiensis, *Synodontis*, 320
wamiensis, *Zaireichthys*, 29
wangi, *Pseudobagrus*, 108
Watawata, 250

waterhousii, *Neoplotosus*, 346
waterloti, *Chiloglanis niloticus*, 307
waterloti, *Synodontis*, 323
watwata, *Hypostomus*, 262
weberi, *Callichrous*, 373
weberi, *Ompok*, 373
weddellii, *Anadoras*, 169
weddellii, *Doras*, 169
weitzmani, *Corydoras*, 126
wernerii, *Clarias*, 148
wernerii, *Synodontis*, 311
Wertheimeri, *Plecostomus*, 285
wertheimeri, *Pogonopoma*, 285
Wertheimeria, 179
Wertheimeria maculata, 179
wesselii, *Pimelodella*, 195
Wesselii, *Pimelodus* (*Pseudorhamdia*), 195
westermannii, *Bergiaria*, 330
Westermannii, *Pimelodus*, 329, 330
weyrauchi, *Pygidium*, 425
weyrauchi, *Trichomycterus*, 425
whymperi, *Arges*, 68
whymperi, *Astroblepus*, 68
wieneri, *Plectrochilus*, 411
wieneri, *Vandellia*, 411
wilderi, *Hassar*, 171
wilsoni, *Eomacrones*, 87
wilsoni, *Hemiancistrus*, 244
wilsoni, *Macronoides*, 87
winzi, *Hypostomus*, 262
winzi, *Plecostomus*, 262
witmeri, *Pimelodella*, 195
Wittei, *Auchenoglanis*, 154
wittenburgii, *Pseudobagrus*, 108
wolfei, *Hemiloricaria*, 247
wolfei, *Rhineloricaria*, 247
Wolfii, *Bagrus*, 98
wolfii, *Mystus*, 98
wolfi, *Rhamdella*, 197
woodi, *Hassar*, 171
woodsi, *Rhynchodoras*, 178
woosnami, *Synodontis*, 323
worthingtoni, *Bathyclarias*, 136
worthingtoni, *Dinotopterus*, 136
wotroi, *Corydoras*, 120
wrightiana, *Oxyropsis*, 276
wrightii, *Oxyropsis*, 276
wuchereri, *Hypostomus*, 262
wuchereri, *Pimelodus*, 200
wuchereri, *Plecostomus*, 262
wui, *Pseudobagrus*, 101
wurnoense, *Nigerium*, 98, 99
Wyckii, *Bagrus*, 91
wyckii, *Hemibagrus*, 91
wyckioides, *Hemibagrus*, 91
wyckioides, *Macrones* (*Hemibagrus*), 91
wynaadensis, *Pterocryptis*, 377
wynaadensis, *Silurus*, 375, 377
X
xakriaba, *Otocinclus*, 275
Xanthocephalus, *Pimelodus*, 207
Xanthocephalus, *Silurus*, 207
xanthosteus, *Silurus*, 379
xanthum, *Megalonema*, 335
xanthus, *Perugia*, 335
xenauchen, *Galeichthys*, 33
Xenocara, 219
Xenocara boliviiana, 219
Xenocara brevipinnis, 219
Xenocara buforia, 219
Xenocara damasceni, 220
Xenocara fulva, 221
Xenocara heterorhynchus, 221
Xenocara macrophthalmus, 222
Xenocara montana, 223
Xenocara multispinis, 223
Xenocara occidentalis, 223
Xenocara rothschildi, 221
Xenoclarias, 153
Xenoclarias eupogon, 153
Xenoclarias holobranchus, 153
xenodon, *Clarias*, 143
xenognathus, *Leptoglanis*, 25
Xenomystus, 247
Xenomystus gobio, 247
Xenopholis, 433
Xenopholis carinatus, 433
Xenopholoides, 433
xetequepeque, *Rhamdia*, 202
Xiliphius, 62
Xiliphius kryptos, 62
xinguensis, *Corydoras*, 126
xinguensis, *Glyptoperichthys*, 292
xinguensis, *Pterygoplichthys*, 292
xingui, *Rhynchodoras*, 178
xiphias, *Synodontis*, 323
Xiurenbagrus, 20
Xiurenbagrus gigas, 20
Xiurenbagrus xiurenensis, 20
xiurenensis, *Xiurenbagrus*, 20
Xyliphius, 62
Xyliphius barbatus, 62
Xyliphius kryptos, 62
Xyliphius labrosus, 62
Xyliphius lepturus, 62
Xyliphius lombarderoi, 62
Xyliphius magdaleneae, 62
Xyliphius melanopterus, 62
Y
yangambianus, *Eutropius*, 365
yangambianus, *Schilbe*, 365
yaravi, *Ancistrus* (*Hemiancistrus*), 273
yaravi, *Neblinichthys*, 273
yarrelli, *Bagarius*, 383
Yarrelli, *Bagrus*, 383
yasi, *Epactionotus*, 236

- youngicus*, *Clarias*, 145
youssoufii, *Gagata*, 386
yuncensis, *Pimelodella*, 195
yunnanense, *Pseudexostoma*, 401
yunnanensis, *Glyptosternum*, 401
yunnanensis brachysoma,
 Pseudexostoma, 401
yurubiene, *Chaetostoma*, 231
yuska, *Trichomycterus*, 425
- Z**
- zainaensis*, *Glyptothorax*, 394
Zaireichthys, 25, 28
Zaireichthys brevis, 28
Zaireichthys camerunensis, 28
Zaireichthys dorae, 28
Zaireichthys flavomaculatus, 28
Zaireichthys heterurus, 28
Zaireichthys mandevillei, 28
Zaireichthys rotundiceps, 28
Zaireichthys wamiensis, 29
Zaireichthys zonatus, 28, 29
zairensis, *Amphilophus*, 23
zairensis, *Schilbe*, 366
zambesensis, *Synodontis*, 323
zambezensis, *Synodontis*, 323
zambezensis rukwaensis, *Synodontis*,
 320
zambeziensis, *Malapterurus*, 302
zammaranoi, *Uegitglanis*, 152, 153
zanaensis, *Glyptothorax*, 394
zanzibarica, *Synodontis*, 323
zanzibaricus, *Synodontis*, 316, 320,
 323
Zathorax, 168
Zathorax gonzalezi, 168
Zathorax monitor, 168, 169
Zathorax nauticus, 169
zebra, *Hypancistrus*, 249
zhuijangensis, *Glyptothorax*, 394
Zonancistrus, 234
zonatum, *Pygidium*, 425
zonatus, *Acrochordonichthys*, 13
zonatus, *Caelatoglanis*, 383
zonatus, *Microglanis*, 355
zonatus, *Pimelodus*, 13
zonatus, *Synodontis angelicus*, 311
zonatus, *Trichomycterus*, 425
zonatus, *Zaireichthys*, 28, 29
zongolicensis, *Rhamdia*, 199
zuliaensis, *Pterygoplichthys*, 292
zuloagai, *Doraops*, 170
Zungaro, 344
Zungaro humboldtii, 344
Zungaro jahu, 344
Zungaro mathisoni, 356
Zungaro zungaro, 344
Zungaro zungaro schultzi, 355
zungaro, *Zungaro*, 344
zungaro schultzi, *Zungaro* 355